

## Review Article

# The Impact of Immune System in Regulating Bone Metastasis Formation by Osteotropic Tumors

Lucia D'Amico<sup>1</sup> and Ilaria Roato<sup>2</sup>

<sup>1</sup>Department of Orthopedics, Washington University School of Medicine, St. Louis, MO 63130, USA

<sup>2</sup>CeRMS, San Giovanni Battista General Hospital, University of Turin, Via Santena 5, 10126 Turin, Italy

Correspondence should be addressed to Ilaria Roato; roato78@libero.it

Received 13 October 2014; Accepted 2 December 2014

Academic Editor: Roberta Faccio

Copyright © 2015 L. D'Amico and I. Roato. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Bone metastases are frequent and debilitating consequence for many tumors, such as breast, lung, prostate, and kidney cancer. Many studies report the importance of the immune system in the pathogenesis of bone metastasis. Indeed, bone and immune system are strictly linked to each other because bone regulates the hematopoietic stem cells from which all cells of the immune system derive, and many immunoregulatory cytokines influence the fate of bone cells. Furthermore, both cytokines and factors produced by immune and bone cells promote the growth of tumor cells in bone, contributing to supporting the vicious cycle of bone metastasis. This review summarizes the current knowledge on the interactions among bone, immune, and tumor cells aiming to provide an overview of the osteoimmunology field in bone metastasis from solid tumors.

## 1. Introduction

The skeleton is the most common site of metastasis and bone in turn is the main responsible of death since the presence of bone metastases makes the primary disease no longer curable [1]. Symptoms like bone pain, hypercalcemia, fracture, and spinal cord compression appear in this type of metastasis, causing a decline in the quality of life [2]. Some types of tumors are characterized by a selective bone tropism, out of which are prostate, breast, lung, and kidney cancers. Bone metastases can give osteolytic, osteosclerotic, or mixed lesions. Osteolytic metastases are due to an enhanced activity of bone-resorbing cells, the osteoclasts (OCs), which cause bone destruction [3, 4]. Typically, breast, lung, and kidney cancers metastasize to bone with osteolytic lesions, whereas prostate cancer metastasizes with osteosclerotic ones. Lung and kidney tumors metastasize in an early phase of the disease, while breast cancer metastasizes with a slower and less aggressive growth. At least 65–75% of breast and prostate cancer patients develop bone metastases during the course of their disease, and breast cancer patients show a relatively long median survival time after diagnosis of bone metastases [5–7]. Bone metastases from prostate cancer are typically osteosclerotic and are caused by an increased

activity of bone-forming cells, the osteoblasts (OBs), leading to enhanced bone formation [3, 4].

Approximately 30–40% of NSCLC patients develop bone metastasis during their disease, with a median survival time measured in months [1]. About 20–35% of kidney cancer patients develop bone metastases, which are particularly destructive, with a rate of skeletal complications higher than other tumors [8, 9]. Accumulating evidences suggest the importance of the immune cell response to factors in the tumor microenvironment as main regulator of cancer progression and metastases. The bone marrow is a reservoir for immune cells such as macrophages, dendritic cells (DCs), myeloid derived suppressor cells (MDSCs), and different T cell subsets that can directly impair the so called “tumor/bone vicious cycle” [10]. This review focuses on the current knowledge of the role of the immune cells in controlling tumor spreading to bone.

## 2. Bone Marrow Is an Attractive Soil for Cancer Cells

Bone marrow (BM) microenvironment is a fertile soil for homing, survival, and proliferation of circulating cancer cells. It provides both endosteal and vascular niches, which support



This phenomenon is called osteomimicry and it has been observed in bone metastatic prostate and breast cancer cell lines [32–34].

Progenitors and mature cells in the BM frequently expressed the receptor Notch [35], whereas the Notch ligand Jagged is overexpressed by bone metastatic tumor cells [36]. Thus, cancer cells directly activate osteolysis through the Notch-Jagged interactions in the BM. In particular, Jagged1, which is a downstream mediator of the prometastatic TGF- $\beta$ , directly activates OC differentiation and promotes tumor growth stimulating IL-6 production by OBs [37].

### 3. Bone and Immune System Cross Talk

Bone and immune and hematopoietic systems are tightly linked since bone cells and hematopoietic cells are in deep physical contact, are reciprocally regulated, are interconnected in their function, and share several common pathways [38]. Indeed, bone cells express surface molecules regulating the expansion of HSCs from which all cells of the mammalian immune system derive, whereas many immunoregulatory cytokines directly act on bone cells [39, 40]. OBs and OCs both affect the maintenance and the mobilization of HSCs [13, 14]. OBs control the proliferation of hematopoietic progenitors [41] and support commitment and differentiation of all stages of B cell development. Indeed *in vitro* production of B cell precursors from progenitors required contact with OBs and expression of CXCL12 and interleukin-7 (IL-7), which was induced by PTH [13, 42]. Moreover, OC precursors, T, B, and NK cells originate from the same stem cell; thus, some of the receptors and ligands that mediate the immune process also regulate the maturation of OC precursors and the ability of OCs to degrade bone. Circulating OC precursors are a reservoir of the pre-OC pool in the BM, but they are also an abundant source of pre-OCs that can be recruited into bone or joint tissue in response to reparative or pathological signals.

RANKL, its receptor RANK, and the natural decoy receptor osteoprotegerin (OPG) [43] form a crucial molecular link between the immune system and bone [44]. The membrane RANKL is expressed by OBs/stromal cells; the soluble RANKL was originally cloned from T cells [45, 46], but it is also expressed by B cells [47], whereas the receptor RANK is expressed by DCs, monocytes, macrophages, and tumor cells [44, 48]. Activated T cells produce RANKL, which directly regulates osteoclastogenesis and bone remodelling, explaining why different pathological conditions, such as cancer, result in systemic and local bone loss. The RANKL to OPG ratio in serum is a determinant factor for OC activation at bone level: a higher serum RANKL to OPG ratio is an index for upregulation of osteoclastogenesis [49].

Many immune factors, including costimulatory receptors and cytokines such as interferon- $\gamma$  (IFN- $\gamma$ ) and TNF regulate bone cell development, bone turnover, and pathogenesis of bone diseases [50]. The role of IFN- $\gamma$  in osteoclastogenesis is controversial; indeed, in studies conducted *in vitro* [51] and *in vivo*, in animal model, it shows an antiosteoclastogenic effect [52], whereas, in humans, IFN- $\gamma$  increases in presence of bone loss due to oestrogen deficiency and rheumatoid arthritis

[53, 54]. IFN- $\gamma$  influences osteoclastogenesis directly by blocking OC formation through inhibition of OC maturation [55] and indirectly by stimulating T cell activation with a consequent increase of proosteoclastogenic factors [56].

*3.1. The Interactions between T Cells and Osteoclast Precursors Regulate Bone Resorption in Bone Metastasis.* A direct regulation of bone resorption by T cell has been widely described for bone metastasis by both solid tumors and multiple myeloma (MM) [57, 58]. Indeed, studies on peripheral blood mononuclear cells (PBMCs), isolated from patients affected by breast, prostate, and lung cancer with bone metastases, demonstrated an increase of circulating OC precursors in these patients compared to patients without bone metastases and healthy controls [58, 59]. In bone metastatic patients, OC precursors differentiate into mature OCs *in vitro* in presence of T cells without adding M-CSF and RANKL, but T cell depletion results in the absence of OC formation without exogenous stimulation [58].

Another important mediator of the interactions between T and bone cells is IL-7, a cytokine produced by stromal cells and by cells at inflammatory site, with different effects on hematopoietic and immunologic systems [60]. The main function of IL-7 is the control of B and T lymphopoiesis [61], but it is also important for the tumor process [62] and the correct bone homeostasis [63, 64]. According to the model considered, IL-7 displays either inhibitory or activator effects on OCs [63, 65]. Some studies demonstrated that IL-7, produced by T cells, promotes osteoclastogenesis by upregulating T cell-derived cytokines, such as RANKL and TNF $\alpha$  [66–68], and that its production is increased by oestrogen deficiency [69]. Furthermore, in bone metastatic patients, IL-7 serum levels were significantly higher than those in nonbone metastatic patients and in healthy controls [59, 68, 70]. This increase of serum IL-7 is at least in part dependent on IL-7 production by tumor cells as demonstrated in a human-in-mice model of bone metastasis from lung cancer [71]. All these data confirm the T cell modulatory activity on OCs. Nevertheless, also OCs affect T cell activity, because they present antigenic peptides to T cells and induce FoxP3 expression in CD8 T cells, which regulate an inappropriate activation of the immune response [72]. The cellular responses in cell-to-cell interactions between T cells and OCs are regulated through reciprocal CD137/CD137L and RANK/RANKL interactions [73]. CD137 is a costimulatory member of the TNF receptor induced by T cell receptor activation. Its ligand CD137L is expressed on OC precursors: *in vitro* CD137L ligation suppresses osteoclastogenesis through the inhibition of OCs precursor fusion. On the other hand, RANKL expressed on T cells binds to RANK on OCs, producing a reverse signal in T cells able to enhance apoptosis [73].

*3.2. T Cells Regulate Tumor Growth in Bone.* Many data suggest that T cells can regulate tumor growth in bone also independently from their interaction with bone cells. Indeed, memory T cells have been found in the BM of breast cancer patients suggesting their role in cancer immune surveillance

[74]. Moreover, the RANKL-RANK interaction between CD4 T cells and breast cancer cells promotes invasion, dissemination, and metastasis formation in an animal model [75]. Some antitumor metastatic therapies show immunomodulatory effects; for instance, the blockade of TGF- $\beta$  at metastatic sites may locally activate an antitumor T cell response, because, normally, TGF- $\beta$ , released in BM by OC activity, inhibits T cell proliferation [76].

Zoledronic acid, an antiresorptive agent, can activate cytotoxic  $\gamma/\delta$ -T cells and inhibit populations of myeloid derived cells with T cell suppressor capabilities [77]. Modulation of antitumor T cell responses alters tumor growth in bone. Indeed, by using mice models *Lyn*<sup>-/-</sup>, which have more OCs and a hyperactive myeloid population with an increased T cell responses, Zhang et al. reported a reduced tumor growth in bone despite enhanced osteolysis [78]. *Lyn* is a member of the Src family tyrosine kinases, which inhibits OC differentiation by downregulating PLC $\gamma$ 2 activation, which regulates the OC formation and function [79]. PLC $\gamma$ 2<sup>-/-</sup> mice have an increased bone tumor burden despite protection from bone loss, because they have dysfunctional OCs and impaired T cell activation mediated by DCs. Importantly, injection of antigen-specific wild-type cytotoxic CD8 T cells in both these mice models reduces the growth of tumor cells in the bone, regardless of OC functionality. According to these data, a condition of immune deficiency can interfere with the antitumor effects of OC blockade [78]. In particular, cytotoxic CD8 T cells seem to be critical regulators of tumor growth in bone, since their activation diminishes and their depletion enhances bone metastases, even with zoledronic acid.

**3.3. Myeloid Derived Suppressor Cells Regulate Cancer Progression.** Myeloid derived suppressor cells are a heterogeneous population of immature myeloid cells identified by the coexpression of Gr-1 and CD11b in mice and CD11b and CD33 in humans [80–83]. MDSCs are significantly overproduced in tumor-bearing mice and cancer patients and they represent a prognostic indicator in various osteotropic tumors including breast, lung and MM [84, 85]. Emerging evidences suggest the importance of the MDSCs in driving the progression of cancer disease by suppressing both the innate and the adaptive immune response. Thus, MDSCs exert their proneoplastic effects through the impairment of T cell/antigen recognition, the release of small soluble oxidizers, and depletion of essential amino acids from the local extracellular environment [86–88]. Besides suppressing CD4 and CD8 T cell populations, MDSCs promote the activation and expansion of regulatory T cells (Treg) and thus mediate immunosuppression.

Finally, all these mechanisms contribute to tumor progression and metastasis spreading to many organs, especially to bone (Figure 1).

Bone metastases are associated with an increase in OC activation and since MDSCs are progenitors of the OC precursors, it is not surprising that they are found to be largely increased in bone metastatic patients. Strikingly, Sawant et al.

confirmed that MDSCs isolated from tumor-bone microenvironment can differentiate into mature and functional OCs *in vitro* and *in vivo* in a mouse model of breast cancer bone metastases [89]. MDSCs from mice bearing bone metastases also induce osteolysis in syngenic animals, indicating that these cells are primed as OC progenitors and the bone microenvironment triggers their activation in functional OCs. It has been also suggested that cancer cells release different soluble factors in the bone, which promotes MDSCs to differentiate into OCs. Thus, breast cancer cells can secrete CCL2, CCL5, or osteopontin which promotes the expression of cathepsin K and matrix metalloproteinase 9 (MMP9), thus enhancing OC functions [90]. On the other hand, MDSC expresses several proosteoclastogenic factors as CCR2, the receptor of CCL2, showing the responsiveness of these cells to the chemokine. Similarly, in the MM model, Zhuang et al. discovered that tumor induced MDSCs were responsible to induce osteolytic lesions by acting as OC precursors [91]. Additionally, only MDSCs isolated from bone are capable of becoming active OCs, suggesting the importance of the bone microenvironment in driving OC maturation.

Despite the critical role for OCs in the establishment of bone metastatic vicious cycle, the PLC $\gamma$ 2<sup>-/-</sup> mouse model, bearing severe OC defects, suggests that MDSCs can enhance tumor growth in bone independently of their ability to differentiate into OCs [78]. Interestingly, the increased PLC $\gamma$ 2<sup>-/-</sup> tumor growth was the result of a higher MDSCs accumulation in secondary lymphoid organs, leading to a strong inhibition of the antitumor T cell response (Figure 1). Despite the importance of MDSCs expansion as a crucial event in the pathogenesis of tumor progression, little is known about the mechanisms leading to this process. Capietto et al. have recently shown  $\beta$ -catenin as a crucial modulator of MDSC accumulation in response to tumor [92]. The downregulation of  $\beta$ -catenin signaling in MDSC promotes their expansion and consequently increases tumor growth in both mice and humans. On the contrary, expression of a constitutively activate form of  $\beta$ -catenin in mice decreased the number of MDSCs and tumor growth. Importantly, the downregulation of  $\beta$ -catenin can also occur in MDSCs from WT mice during tumor dissemination to bone, indicating that  $\beta$ -catenin pathway modulates MDSC expansion in both primary and metastatic solid tumors.

## 4. Conclusions

The rapidly developing field of osteoimmunology shows the importance of the deep interconnection between skeletal and immune system. This relationship results in the generation of several cellular pathways, which provides the discovery of new potential targets for the prevention and treatment of bone metastasis.

The bone marrow represents an active and hospitable microenvironment, allowing multiple cell interactions which are critical in the pathogenesis of tumor progression. Thus, additional studies to elucidate new mechanisms promoting the accumulation of bone marrow derived cells such as MDSCs are mandatory to address the critical steps of tumor

progression in bone. The design of new drugs must consider the potential effects on both immune system and bone; thus further investigations to understand the osteoimmune system are even more important.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

## Acknowledgments

The authors thank the organizers of the Benish Trophy and the Italian Ministry of Health: Ricerca Sanitaria Finalizzata e Giovani Ricercatori 2009 (GR 2009-1584485), for supporting this work.

## References

- [1] R. E. Coleman, "Clinical features of metastatic bone disease and risk of skeletal morbidity," *Clinical Cancer Research*, vol. 12, part 2, pp. 6243s–6249s, 2006.
- [2] N. Sethi and Y. Kang, "Dysregulation of developmental pathways in bone metastasis," *Bone*, vol. 48, no. 1, pp. 16–22, 2011.
- [3] G. R. Mundy, "Metastasis to bone: causes, consequences and therapeutic opportunities," *Nature Reviews Cancer*, vol. 2, no. 8, pp. 584–593, 2002.
- [4] G. D. Roodman, "Mechanisms of bone metastasis," *The New England Journal of Medicine*, vol. 350, no. 16, pp. 1655–1664, 2004.
- [5] R. E. Coleman, "Metastatic bone disease: clinical features, pathophysiology and treatment strategies," *Cancer Treatment Reviews*, vol. 27, no. 3, pp. 165–176, 2001.
- [6] A. Lipton, "Bisphosphonates and breast carcinoma: present and future," *Cancer*, vol. 88, no. 12, pp. 3033–3037, 2000.
- [7] R. E. Coleman, "Bisphosphonates: clinical experience," *Oncologist*, vol. 9, supplement 4, pp. 14–27, 2004.
- [8] S. L. Wood and J. E. Brown, "Skeletal metastasis in renal cell carcinoma: current and future management options," *Cancer Treatment Reviews*, vol. 38, no. 4, pp. 284–291, 2012.
- [9] M. R. Smith, "Zoledronic acid to prevent skeletal complications in cancer: corroborating the evidence," *Cancer Treatment Reviews*, vol. 31, supplement 3, pp. 19–25, 2005.
- [10] L. M. Cook, G. Shay, A. Aruajo, and C. C. Lynch, "Integrating new discoveries into the "vicious cycle" paradigm of prostate to bone metastases," *Cancer and Metastasis Reviews*, vol. 33, no. 2–3, pp. 511–525, 2014.
- [11] S. Azizdoost, S. Babashah, F. Rahim, M. Shahjahani, and N. Saki, "Bone marrow neoplastic niche in leukemia," *Hematology*, vol. 19, no. 4, pp. 232–238, 2014.
- [12] O. Kollet, A. Dar, S. Shviti et al., "Osteoclasts degrade endosteal components and promote mobilization of hematopoietic progenitor cells," *Nature Medicine*, vol. 12, no. 6, pp. 657–664, 2006.
- [13] L. M. Calvi, G. B. Adams, K. W. Weibrecht et al., "Osteoblastic cells regulate the haematopoietic stem cell niche," *Nature*, vol. 425, no. 6960, pp. 841–846, 2003.
- [14] J. Zhang, C. Niu, L. Ye et al., "Identification of the haematopoietic stem cell niche and control of the niche size," *Nature*, vol. 425, no. 6960, pp. 836–841, 2003.
- [15] S. I. Park, C. Lee, W. David Sadler et al., "Parathyroid hormone-related protein drives a CD11b<sup>+</sup>Gri1<sup>+</sup> cell-mediated positive feedback loop to support prostate cancer growth," *Cancer Research*, vol. 73, no. 22, pp. 6574–6583, 2013.
- [16] Y. Shiozawa, E. A. Pedersen, A. M. Havens et al., "Human prostate cancer metastases target the hematopoietic stem cell niche to establish footholds in mouse bone marrow," *The Journal of Clinical Investigation*, vol. 121, no. 4, pp. 1298–1312, 2011.
- [17] J. E. Compston, "Bone marrow and bone: a functional unit," *Journal of Endocrinology*, vol. 173, no. 3, pp. 387–394, 2002.
- [18] M. K. Herroon, E. Rajagurubandara, A. L. Hardaway et al., "Bone marrow adipocytes promote tumor growth in bone via FABP4-dependent mechanisms," *Oncotarget*, vol. 4, no. 11, pp. 2108–2123, 2013.
- [19] B. F. Boyce, E. Rosenberg, A. E. de Papp, and L. T. Duong, "The osteoclast, bone remodelling and treatment of metabolic bone disease," *European Journal of Clinical Investigation*, vol. 42, no. 12, pp. 1332–1341, 2012.
- [20] L. J. Suva, C. Washam, R. W. Nicholas, and R. J. Griffin, "Bone metastasis: mechanisms and therapeutic opportunities," *Nature Reviews Endocrinology*, vol. 7, no. 4, pp. 208–218, 2011.
- [21] A. C. Chiang and J. Massagué, "Molecular basis of metastasis," *The New England Journal of Medicine*, vol. 359, no. 26, pp. 2814–2823, 2008.
- [22] X. Lu, E. Mu, Y. Wei et al., "VCAM-1 promotes osteolytic expansion of indolent bone micrometastasis of breast cancer by engaging alpha4beta1-positive osteoclast progenitors," *Cancer Cell*, vol. 20, no. 6, pp. 701–714, 2011.
- [23] I. Pécheur, O. Peyruchaud, C.-M. Serre et al., "Integrin alpha(v)beta3 expression confers on tumor cells a greater propensity to metastasize to bone," *The FASEB Journal*, vol. 16, no. 10, pp. 1266–1268, 2002.
- [24] J. M. Reuben, B.-N. Lee, H. Gao et al., "Primary breast cancer patients with high risk clinicopathologic features have high percentages of bone marrow epithelial cells with ALDH activity and CD44<sup>+</sup>CD24<sup>lo</sup> cancer stem cell phenotype," *European Journal of Cancer*, vol. 47, no. 10, pp. 1527–1536, 2011.
- [25] S. A. Patel, S. H. Ramkissoon, M. Bryan et al., "Delineation of breast cancer cell hierarchy identifies the subset responsible for dormancy," *Scientific Reports*, vol. 2, article 906, 2012.
- [26] L. D'Amico, S. Patanè, C. Grange et al., "Primary breast cancer stem-like cells metastasize to bone, switch phenotype and acquire a bone tropism signature," *British Journal of Cancer*, vol. 108, no. 12, pp. 2525–2536, 2013.
- [27] M. K. Conley-LaComb, A. Saliganan, P. Kandagatla, Y. Q. Chen, M. L. Cher, and S. R. Chinni, "PTEN loss mediated Akt activation promotes prostate tumor growth and metastasis via CXCL12/CXCR4 signaling," *Molecular Cancer*, vol. 12, no. 1, article 85, 2013.
- [28] R. S. Taichman, C. Cooper, E. T. Keller, K. J. Pienta, N. S. Taichman, and L. K. McCauley, "Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone," *Cancer Research*, vol. 62, no. 6, pp. 1832–1837, 2002.
- [29] P. V. Hauschka, A. E. Mavrikos, M. D. Iafrazi, S. E. Doleman, and M. Klagsbrun, "Growth factors in bone matrix. Isolation of multiple types by affinity chromatography on heparin-sepharose," *The Journal of Biological Chemistry*, vol. 261, no. 27, pp. 12665–12674, 1986.
- [30] A. Bellahcène, M.-P. Merville, and V. Castronovo, "Expression of bone sialoprotein, a bone matrix protein, in human breast cancer," *Cancer Research*, vol. 54, no. 11, pp. 2823–2826, 1994.

- [31] A. Bellahcene and V. Castronovo, "Increased expression of osteonectin and osteopontin, two bone matrix proteins, in human breast cancer," *The American Journal of Pathology*, vol. 146, no. 1, pp. 95–100, 1995.
- [32] K. S. Koeneman, F. Yeung, and L. W. Chung, "Osteomimetic properties of prostate cancer cells: a hypothesis supporting the predilection of prostate cancer metastasis and growth in the bone environment," *Prostate*, vol. 39, no. 4, pp. 246–261, 1999.
- [33] D.-L. Lin, C. P. Tarnowski, J. Zhang et al., "Bone metastatic LNCaP-derivative C4-2B prostate cancer cell line mineralizes in vitro," *The Prostate*, vol. 47, no. 3, pp. 212–221, 2001.
- [34] R. F. Cox, A. Jenkinson, K. Pohl, F. J. O'Brien, and M. P. Morgan, "Osteomimicry of mammary adenocarcinoma cells *in vitro*; increased expression of bone matrix proteins and proliferation within a 3D collagen environment," *PLoS ONE*, vol. 7, no. 7, Article ID e41679, 2012.
- [35] S. Chiba, "Notch signaling in stem cell systems," *Stem Cells*, vol. 24, no. 11, pp. 2437–2447, 2006.
- [36] S. Santagata, F. Demichelis, A. Riva et al., "JAGGED1 expression is associated with prostate cancer metastasis and recurrence," *Cancer Research*, vol. 64, no. 19, pp. 6854–6857, 2004.
- [37] N. Sethi, X. Dai, C. G. Winter, and Y. Kang, "Tumor-derived JAGGED1 promotes osteolytic bone metastasis of breast cancer by engaging notch signaling in bone cells," *Cancer Cell*, vol. 19, no. 2, pp. 192–205, 2011.
- [38] H. L. Aguila and D. W. Rowe, "Skeletal development, bone remodeling, and hematopoiesis," *Immunological Reviews*, vol. 208, pp. 7–18, 2005.
- [39] H. Takayanagi, "New developments in osteoimmunology," *Nature Reviews Rheumatology*, vol. 8, no. 11, pp. 684–689, 2012.
- [40] S. Kasagi and W. Chen, "TGF-beta1 on osteoimmunology and the bone component cells," *Cell and Bioscience*, vol. 3, article 4, 2013.
- [41] F. Arai, A. Hirao, M. Ohmura et al., "Tie2/angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche," *Cell*, vol. 118, no. 2, pp. 149–161, 2004.
- [42] J. Zhu, R. Garrett, Y. Jung et al., "Osteoblasts support B-lymphocyte commitment and differentiation from hematopoietic stem cells," *Blood*, vol. 109, no. 9, pp. 3706–3712, 2007.
- [43] P. J. Kostenuik and V. Shalhoub, "Osteoprotegerin: a physiological and pharmacological inhibitor of bone resorption," *Current Pharmaceutical Design*, vol. 7, no. 8, pp. 613–635, 2001.
- [44] L. C. Hofbauer, C. A. Kühne, and V. Viereck, "The OPG/RANKL/RANK system in metabolic bone diseases," *Journal of Musculoskeletal Neuronal Interactions*, vol. 4, no. 3, pp. 268–275, 2004.
- [45] B. R. Wong, R. Josien, S. Y. Lee et al., "TRANCE (Tumor necrosis factor [TNF]-related Activation-induced Cytokine), a new TNF family member predominantly expressed in T cells, is a dendritic cell-specific survival factor," *The Journal of Experimental Medicine*, vol. 186, no. 12, pp. 2075–2080, 1997.
- [46] D. M. Anderson, E. Maraskovsky, W. L. Billingsley et al., "A homologue of the TNF receptor and its ligand enhance T-cell growth and dendritic-cell function," *Nature*, vol. 390, no. 6656, pp. 175–179, 1997.
- [47] Y.-Y. Kong, H. Yoshida, I. Sarosi et al., "OPGL is a key regulator of osteoclastogenesis, lymphocyte development and lymph-node organogenesis," *Nature*, vol. 397, no. 6717, pp. 315–323, 1999.
- [48] D. Santini, G. Perrone, I. Roato et al., "Expression pattern of receptor activator of NF $\kappa$ B (RANK) in a series of primary solid tumors and related bone metastases," *Journal of Cellular Physiology*, vol. 226, no. 3, pp. 780–784, 2011.
- [49] L. C. Hofbauer, S. Khosla, C. R. Dunstan, D. L. Lacey, W. J. Boyle, and B. L. Riggs, "The roles of osteoprotegerin and osteoprotegerin ligand in the paracrine regulation of bone resorption," *Journal of Bone and Mineral Research*, vol. 15, no. 1, pp. 2–12, 2000.
- [50] R. Pacifici, "The immune system and bone," *Archives of Biochemistry and Biophysics*, vol. 503, no. 1, pp. 41–53, 2010.
- [51] H. Takayanagi, K. Ogasawara, S. Hida et al., "T-cell-mediated regulation of osteoclastogenesis by signalling cross-talk between RANKL and IFN- $\gamma$ ," *Nature*, vol. 408, no. 6812, pp. 600–605, 2000.
- [52] K. Sato, T. Satoh, K. Shizume et al., "Prolonged decrease of serum calcium concentration by murine  $\gamma$ -interferon in hypercalcemic, human tumor (EC-GI)-bearing nude mice," *Cancer Research*, vol. 52, no. 2, pp. 444–449, 1992.
- [53] Y. Gao, F. Grassi, M. R. Ryan et al., "IFN- $\gamma$  stimulates osteoclast formation and bone loss in vivo via antigen-driven T cell activation," *The Journal of Clinical Investigation*, vol. 117, no. 1, pp. 122–132, 2007.
- [54] S. Cenci, G. Toraldo, M. N. Weitzmann et al., "Estrogen deficiency induces bone loss by increasing T cell proliferation and lifespan through IFN- $\gamma$ -induced class II transactivator," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 18, pp. 10405–10410, 2003.
- [55] S. Abbas and Y. Abu-Amer, "Dominant-negative I $\kappa$ B facilitates apoptosis of osteoclasts by tumor necrosis factor- $\alpha$ ," *Journal of Biological Chemistry*, vol. 278, no. 22, pp. 20077–20082, 2003.
- [56] R. Pacifici, "Estrogen deficiency, T cells and bone loss," *Cellular Immunology*, vol. 252, no. 1-2, pp. 68–80, 2008.
- [57] S. Colucci, G. Brunetti, R. Rizzi et al., "T cells support osteoclastogenesis in an in vitro model derived from human multiple myeloma bone disease: the role of the OPG/TRAIL interaction," *Blood*, vol. 104, no. 12, pp. 3722–3730, 2004.
- [58] I. Roato, M. Grano, G. Brunetti et al., "Mechanisms of spontaneous osteoclastogenesis in cancer with bone involvement," *The FASEB Journal*, vol. 19, no. 2, pp. 228–230, 2005.
- [59] I. Roato, E. Gorassini, L. Buffoni et al., "Spontaneous osteoclastogenesis is a predictive factor for bone metastases from non-small cell lung cancer," *Lung Cancer*, vol. 61, no. 1, pp. 109–116, 2008.
- [60] P. M. Appasamy, "Biological and clinical implications of interleukin-7 and lymphopolesis," *Cytokines, Cellular and Molecular Therapy*, vol. 5, no. 1, pp. 25–39, 1999.
- [61] A. E. Namen, S. Lupton, K. Hjerrild et al., "Stimulation of B-cell progenitors by cloned murine interleukin-7," *Nature*, vol. 333, no. 6173, pp. 571–573, 1988.
- [62] M. A. A. Al-Rawi, K. Rmali, G. Watkins, R. E. Mansel, and W. G. Jiang, "Aberrant expression of interleukin-7 (IL-7) and its signalling complex in human breast cancer," *European Journal of Cancer*, vol. 40, no. 4, pp. 494–502, 2004.
- [63] C. Miyaura, Y. Onoe, M. Inada et al., "Increased B-lymphopoiesis by interleukin 7 induces bone loss in mice with intact ovarian function: similarity to estrogen deficiency," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 17, pp. 9360–9365, 1997.
- [64] M. N. Weitzmann, C. Roggia, G. Toraldo, L. Weitzmann, and R. Pacifici, "Increased production of IL-7 uncouples bone formation from bone resorption during estrogen deficiency," *The Journal of Clinical Investigation*, vol. 110, no. 11, pp. 1643–1650, 2002.

- [65] S.-K. Lee, J. F. Kalinowski, S. L. Jastrzebski, L. Puddington, and J. A. Lorenzo, "Interleukin-7 is a direct inhibitor of in vitro osteoclastogenesis," *Endocrinology*, vol. 144, no. 8, pp. 3524–3531, 2003.
- [66] G. Toraldo, C. Roggia, W.-P. Qian, R. Pacific, and M. N. Weitzmann, "IL-7 induces bone loss in vivo by induction of receptor activator of nuclear factor  $\kappa$ B ligand and tumor necrosis factor  $\alpha$  from T cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 1, pp. 125–130, 2003.
- [67] N. Giuliani, S. Colla, R. Sala et al., "Human myeloma cells stimulate the receptor activator of nuclear factor- $\alpha$ B ligand (RANKL) in T lymphocytes: a potential role in multiple myeloma bone disease," *Blood*, vol. 100, no. 13, pp. 4615–4621, 2002.
- [68] I. Roato, G. Brunetti, E. Gorassini et al., "IL-7 up-regulates TNF- $\alpha$ -dependent osteoclastogenesis in patients affected by solid tumor," *PLoS ONE*, vol. 1, article e124, 2006.
- [69] P. D'Amelio, A. Grimaldi, S. Di Bella et al., "Estrogen deficiency increases osteoclastogenesis up-regulating T cells activity: a key mechanism in osteoporosis," *Bone*, vol. 43, no. 1, pp. 92–100, 2008.
- [70] I. Roato, E. Gorassini, G. Brunetti et al., "IL-7 modulates osteoclastogenesis in patients affected by solid tumors," *Annals of the New York Academy of Sciences*, vol. 1117, pp. 377–384, 2007.
- [71] I. Roato, D. Caldo, L. Godio et al., "Bone invading NSCLC cells produce IL-7: mice model and human histologic data," *BMC Cancer*, vol. 10, article 12, 2010.
- [72] J. R. Kiesel, Z. S. Buchwald, and R. Aurora, "Cross-presentation by osteoclasts induces FoxP3 in CD8<sup>+</sup> T cells," *The Journal of Immunology*, vol. 182, no. 9, pp. 5477–5487, 2009.
- [73] R. Senthikumar and H.-W. Lee, "CD137L- and RANKL-mediated reverse signals inhibit osteoclastogenesis and T lymphocyte proliferation," *Immunobiology*, vol. 214, no. 2, pp. 153–161, 2009.
- [74] M. Feuerer, M. Rocha, L. Bai et al., "Enrichment of memory T cells and other profound immunological changes in the bone marrow from untreated breast cancer patients," *International Journal of Cancer*, vol. 92, no. 1, pp. 96–105, 2001.
- [75] W. Tan, W. Zhang, A. Strasner et al., "Tumour-infiltrating regulatory T cells stimulate mammary cancer metastasis through RANKL-RANK signalling," *Nature*, vol. 470, no. 7335, pp. 548–553, 2011.
- [76] S. H. Wrzesinski, Y. Y. Wan, and R. A. Flavell, "Transforming growth factor- $\beta$  and the immune response: implications for anticancer therapy," *Clinical Cancer Research*, vol. 13, no. 18, pp. 5262–5270, 2007.
- [77] K. Schilbach, A. Geiselhart, and R. Handgretinger, "Induction of proliferation and augmented cytotoxicity of  $\gamma\delta$  T lymphocytes by bisphosphonate clodronate," *Blood*, vol. 97, no. 9, pp. 2917–2918, 2001.
- [78] K. Zhang, S. Kim, V. Cremasco et al., "CD8<sup>+</sup> T cells regulate bone tumor burden independent of osteoclast resorption," *Cancer Research*, vol. 71, no. 14, pp. 4799–4808, 2011.
- [79] S.-H. Yoon, Y. Lee, H.-J. Kim et al., "Lyn inhibits osteoclast differentiation by interfering with PLC $\gamma$ 1-mediated Ca<sup>2+</sup> signaling," *FEBS Letters*, vol. 583, no. 7, pp. 1164–1170, 2009.
- [80] V. Bronte, E. Apolloni, A. Cabrelle et al., "Identification of a CD11b<sup>+</sup>/Gr-1<sup>+</sup>/CD31<sup>+</sup> myeloid progenitor capable of activating or suppressing CD8<sup>+</sup> T cells," *Blood*, vol. 96, no. 12, pp. 3838–3846, 2000.
- [81] D. I. Gabrilovich, M. P. Velders, E. M. Sotomayor, and W. M. Kast, "Mechanism of immune dysfunction in cancer mediated by immature Gr-1<sup>+</sup> myeloid cells," *Journal of Immunology*, vol. 166, no. 9, pp. 5398–5406, 2001.
- [82] G. Gallina, L. Dolcetti, P. Serafini et al., "Tumors induce a subset of inflammatory monocytes with immunosuppressive activity on CD8<sup>+</sup> T cells," *The Journal of Clinical Investigation*, vol. 116, no. 10, pp. 2777–2790, 2006.
- [83] S. Kusmartsev, S. Nagaraj, and D. I. Gabrilovich, "Tumor-associated CD8<sup>+</sup> T cell tolerance induced by bone marrow-derived immature myeloid cells," *The Journal of Immunology*, vol. 175, no. 7, pp. 4583–4592, 2005.
- [84] B. Almand, J. I. Clark, E. Nikitina et al., "Increased production of immature myeloid cells in cancer patients: a mechanism of immunosuppression in cancer," *The Journal of Immunology*, vol. 166, no. 1, pp. 678–689, 2001.
- [85] W. Li, K. Wu, E. Zhao et al., "HMGB1 recruits myeloid derived suppressor cells to promote peritoneal dissemination of colon cancer after resection," *Biochemical and Biophysical Research Communications*, vol. 436, no. 2, pp. 156–161, 2013.
- [86] S. Kusmartsev, Y. Nefedova, D. Yoder, and D. I. Gabrilovich, "Antigen-specific inhibition of CD8<sup>+</sup> T cell response by immature myeloid cells in cancer is mediated by reactive oxygen species," *The Journal of Immunology*, vol. 172, no. 2, pp. 989–999, 2004.
- [87] Y. Liu, J. A. Van Ginderachter, L. Brys, P. De Baetselier, G. Raes, and A. B. Geldhof, "Nitric oxide-independent CTL suppression during tumor progression: association with arginase-producing (M2) myeloid cells," *The Journal of Immunology*, vol. 170, no. 10, pp. 5064–5074, 2003.
- [88] A. Mazzoni, V. Bronte, A. Visintin et al., "Myeloid suppressor lines inhibit T cell responses by an NO-dependent mechanism," *The Journal of Immunology*, vol. 168, no. 2, pp. 689–695, 2002.
- [89] A. Sawant, J. Deshane, J. Jules et al., "Myeloid-derived suppressor cells function as novel osteoclast progenitors enhancing bone loss in breast cancer," *Cancer Research*, vol. 73, no. 2, pp. 672–682, 2013.
- [90] S. Das, R. S. Samant, and L. A. Shevde, "Hedgehog signaling induced by breast cancer cells promotes osteoclastogenesis and osteolysis," *The Journal of Biological Chemistry*, vol. 286, no. 11, pp. 9612–9622, 2011.
- [91] J. Zhuang, J. Zhang, S. T. Lwin et al., "Osteoclasts in multiple myeloma are derived from Gr-1+CD11b+myeloid-derived suppressor cells," *PLoS ONE*, vol. 7, no. 11, Article ID e48871, 2012.
- [92] A.-H. Capietto, S. Kim, D. E. Sanford et al., "Down-regulation of PLC $\gamma$ 2- $\beta$ -catenin pathway promotes activation and expansion of myeloid-derived suppressor cells in cancer," *Journal of Experimental Medicine*, vol. 210, no. 11, pp. 2257–2271, 2013.



**Hindawi**  
Submit your manuscripts at  
<http://www.hindawi.com>

