Research Article

Evaluation of Four Rapid Tests for Detection of Hepatitis B Surface Antigen in Ivory Coast

Bamory Dembele^{1,2}, **Roseline Affi-Aboli**^{2,3}, **Mathieu Kabran**^{2,3}, **Daouda Sevede**⁴,
Vanessa Goha², **Aimé Cézaire Adiko**^{1,2}, **Rodrigue Kouamé**², **Emile Allah-Kouadio**⁵,
and **Andre Inwoley**^{2,3}

¹NBTS (National Blood Transfusion Center), Côte d'Ivoire

²Félix Houphouët-Boigny University, Abidjan, Côte d'Ivoire

³Research and Diagnosis Center for AIDS and other infectious diseases (CeDReS), CHU (University Hospital) of Treichville, Abidjan, Côte d'Ivoire

⁴Institut Pasteur, Côte d'Ivoire

⁵National Hepatitis Program, Côte d'Ivoire

Correspondence should be addressed to Bamory Dembele; bamory.dembele225@gmail.com

Received 19 February 2020; Revised 4 May 2020; Accepted 11 June 2020; Published 26 June 2020

Academic Editor: Ravirajsinh Jadeja

Copyright © 2020 Bamory Dembele et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Hepatitis B virus (HBV) infection is a leading cause of liver disease worldwide. Hepatitis B surface antigen (HBsAg) rapid diagnostic tests (RDTs) could be an ideal tool for a large-scale HBV screening in settings with high endemicity but limited infrastructure. The aim of this study was to evaluate the diagnosis performance of such RDTs for screening HBV infection in Ivory Coast. **Methods.** From September 2018 to January 2019, a cross-sectional phase I evaluation study of RDTs was conducted in three laboratories of Abidjan (CeDReS, CNTS and IPCI), on a panel of 405 whole blood samples and 699 plasmas. Four HBsAg RDTs (Determine™ HBsAg, SD Bioline HBsAg WB®, Standard Q HBsAg® and Vikia HBsAg®) were evaluated. The diagnostic performance (sensitivity and specificity) was calculated in comparison to the reference sequential algorithms of two EIA tests (Dia.Pro HBsAg® one version ULTRA and Monolisa™ HBsAg ULTRA). **Results.** The Determine™ HBsAg and Vikia HBsAg® tests performed well, with 100% of sensitivity, specificity both on plasma and on whole blood. For SD Bioline HBsAg WB® and Standard Q HBsAg®, the specificities were 99.8% and the sensitivities 99.3% and 97.1% respectively. Finally, there were a total of 19 false negative results: 3 with SD Bioline HBsAg WB® and 16 with Standard Q HBsAg®. **Conclusion.** Determine HBsAg® from Alere and Vikia HBsAg® from Biomérieux are the most suitable RDTs for screening for HBV in Ivory Coast. A phase II evaluation must be initiated.

1. Introduction

Hepatitis B virus (HBV) is a serious public health problem worldwide and a major cause of chronic hepatitis, cirrhosis, and hepatocellular carcinoma (HCC). The World Health Organization (WHO) estimated that in 2015, 257 million persons, were living with chronic HBV infection in the world with 890 000 deaths from cirrhosis, and HCC [1]. Most countries in Africa are of higher-intermediate endemicity or highly endemic for HBV with a mean prevalence of 6.1%

[1, 2]. In Ivory Coast, a sub-Saharan country, hepatitis B virus surface antigen (HBsAg) prevalence was estimated at 4.3 among schoolchildren [3], 8.48% in the general population [4] and 11% in the blood donors [5].

Early diagnosis is critical in reducing hepatitis-related morbidity and mortality, as well as identifying candidates for HBV vaccination. HBsAg is used as the marker of infection for both HBV screening and to detect suspected acute cases or chronic hepatitis B in any community [6]. HBsAg is typically detected using sensitive immunoassays with an

immunoassay analyzer in a hospital laboratory setting. Although such enzyme immunoassays (EIAs) can effectively detect the viral antigens, they have high costs, require dedicated facilities, sophisticated equipment, trained technicians, and a continuous supply of electricity, and have long turnaround times [7]. These drawbacks of laboratory-based immunoassays limit their usefulness in resource-limited settings [8]. For these reasons, WHO recommends the use of inexpensive simple and rapid tests with performance comparable to the immunoassays that can be performed by the laboratories of peripheral health centers in resource-limited countries. Rapid diagnostic tests (RDTs) are quantitative immunochromatographic tests for the purpose of massive screening in non-laboratory environment. It could be designed to detect HBsAg with a very little turnaround time for the test result to be made available for the patients. Several RDTs are circulating and used for hepatitis B screening in Ivory Coast during clinical research and for routine diagnosis without the approval of the Direction of Pharmacy, Drug and Laboratory (DPML) which is the regulatory body of the health ministry of Ivory Coast.

Although some evaluations have been carried out in others countries [9, 10], no evaluation has been made in Ivory Coast as recommended by WHO before the marketing to certify their reliability. This study was carried out to evaluate the performance of four different HBsAg RDTs for marketing in Ivory Coast.

2. Materials and Methods

2.1. Study Design. From September 2018 to January 2019, a cross-sectional phase I evaluation study of RDTs was conducted at the Institut Pasteur of Ivory Coast (IPCI), the Center for Diagnosis and Research on AIDS and other infectious diseases (CeDRes) and the National Blood Transfusion Center (NBTC). It was initiated by the Direction of Pharmacy, Drug and Laboratory (DPML) in association with the National Program of fight against the viral hepatitis according to the procedures used at national level. The selection criteria of tests were: colorimetric reading between one and two steps; performing tests on serum / plasma and whole blood obtained by venous or capillary sampling; sensitivity and specificity known in other regions of the world on plasma / serum / whole blood; test life greater than 12 months; storage and stability at room temperature (20-30°C).

2.2. Assessment Panel. This study was carried out on serum / plasma and whole blood samples. According to WHO RDTs evaluation guide [11], six hundred ninety-nine (699) serum and plasma samples came from the biobank of IPCI and CeDRes, while four hundred and five (405) whole blood samples were collected from blood donors at the National Blood Transfusion Center. Approximately, 4 ml of whole blood was collected into a tube containing an anticoagulant (heparin or ethylene diaminetetracetic) in addition to the usual tubes provided for the biological qualification of donations. Part of the blood collected was centrifuged to carry out the reference tests. The status (positive or negative) of all specimens was established according to the algorithms used in each lab-

oratory. Briefly, laboratories performed Enzyme Immunoassay (EIA) and or Chemiluminescence immunoassay (CLIA).

2.3. Rapid Tests Evaluated. We evaluated four HBsAg RDTs: Determine™ HBsAg (Alere International Limited, Ballybrit Galway, Ireland), SD Bioline HBsAg WB® (Standard Diagnostics Inc, Korea), Standard Q HBsAg® (SD Biosensor, India) and Vikia HBsAg® (Biomérieux, Marcy l'étoile, France). These qualitative tests are based on immunochromatographic techniques for lateral association of monoclonal and polyclonal antibodies specific for HBsAg. We performed the test according to manufacturer's procedure. All these tests give visual readout.

2.4. References Tests. HBsAg status of the panel serum/plasma samples has been determined using a sequential algorithm consisted of two commercially available enzyme-linked immuno-assay (ELISA) tests: Dia.Pro HBsAg® one version ULTRA (Diagnostic BioProbes Srl, Italy) and Monolisa™ HBsAg ULTRA (BIO-RAD, Marnes-la-coquette, France). The result unit used is a ratio of the sample optical density (OD) to the threshold value (TV). Samples with OD/TV values higher than or equal to 1.00 are considered reactive. Positive results obtained with Dia.Pro Ag were confirmed with the Monolisa AgHBs ULTRA®.

2.5. Statistical Analysis. The sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV) and accuracy of these four kits were calculated with 95% confidence interval according to the existing formula [12]. Kappa concordance value was also calculated and interpreted according to the criteria proposed by Landis and Koch [13]. The diagnostic performances were compared between the plasma/serum and whole blood samples and statistical significance of differences in diagnostic performances were determined using Fisher's exact test. $P \leq 0.05$ were considered statistically significant.

3. Results

3.1. Diagnostic Performance and Accuracy. The performance of each of the four RDTs evaluated in this study is shown in Table 1. The Determine™ HBsAg and Vikia HBsAg® tests have shown better performance, with 100% of a sensitivity, specificity, PPV, NPV and an accuracy rate. However, all the four tests showed a good agreement ($k > 0.97$). In both plasma/serum and whole blood samples, the performances of each of the four commercially available rapid kits were comparable (Table 2).

3.2. Discordance Results. Globally, there were a total of 19 false negative results: 3 with SD Bioline HBsAg WB® and 16 with Standard Q HBsAg® for a discordance rate of 0.36% and 1.54% respectively. Details are resumed in Table 3.

4. Discussion

Many HBsAg rapid tests using immunochromatographic assays have been commercialized worldwide. The major challenge for these tests is to detect the low levels of the target

TABLE 1: Performances of Determine™, SD Bioline WB®, Standard Q® and Vikia® for HBsAg detection compared with EIA tests.

	HBsAg serology EIA		Sensitivity, % (95% CI)	Specificity, % (95% CI)	PPV, % (95% CI)	NPV, % (95% CI)	Accuracy, % (95% CI)	Kappa (95% CI)
	Positive	Negative						
Determine™								
Positive	551	0	100	100	100	100	100	1
Negative	0	553	(99.17-100)	(99.17-100)	(99.17-100)	(99.17-100)	(99.17-100)	
SD Bioline WB®								
Positive	548	1	99.46	99.82	99.82	99.46	99.64	0.99
Negative	3	552	(98.33-99.89)	(98.88-100)	(98.87-100)	(98.34-99.89)	(99.04-99.89)	(0.98-1)
Standard Q®								
Positive	535	1	97.1	99.82	99.81	97.18	98.46	0.97
Negative	16	552	95.3-98.24	98.88-100	98.84-100	95.44-98.29	97.53-99.06	0.95-0.98
Vikia®								
Positive	551	0	100	100	100	100	100	1
Negative	0	553	99.17-100	99.17-100	99.17-100	99.17-100	99.17-100	

Abbreviations: PPV, positive predictive value, NPV, negative predictive value, CI, confidence interval.

TABLE 2: Comparison of Diagnostic accuracy between the plasma/serum and whole of Determine™, SD Bioline WB®, Standard Q® and Vikia® for HBsAg detection.

	Sensitivity, % (95% CI)	Specificity, % (95% CI)	PPV, % (95% CI)	NPV,% (95% CI)	Accuracy, % (95% CI)	P*
Determine™						
plasma/serum	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	0,91
Wholeblood	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	
SD Bioline WB®						
plasma/serum	98.84 (95.61-99.95)	99.82 (98.88-100)	99.82 (98.87-100)	98.93 (95.93-99.96)	99.64 (99.04-99.89)	0,38
Wholeblood	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	
Standard Q®						
plasma/serum	96.53 (92.47-98.57)	99.82 (98.88-100)	99.81 (98.84-100)	97.18 (95.44-98.29)	98.46 (97.53-99.06)	0,42
Wholeblood	98.05 (94.91-99.42)	100 (99.17-100)	100 (99.17-100)	98.04 (94.89-99.41)	99.01 (97.33-99.71)	
Vikia®						
plasma/serum	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	0,91
Wholeblood	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	100 (99.17-100)	

Abbreviations: PPV, positive predictive value, NPV, negative predictive value, CI, confidence interval. * Calculated by Fisher's test.

TABLE 3: Results observed with the Monolisa test for samples giving false results with any of the rapid diagnosis tests.

Identity	OD/TV ^a with Monolisa AgHBs Ultra®	Status HBsAg ^b	Determine™ HBsAg	Results of rapid diagnosis tests		
				SD Bioline HBsAg WB®	Standard Q HBsAg®	Vikia HBsAg®
7402926	33,7	Positive	Positive	Positive	Negative	Positive
9449856	43,8	Positive	Positive	Positive	Negative	Positive
1457809	32,0	Positive	Positive	Positive	Negative	Positive
9441806	41,2	Positive	Positive	Positive	Negative	Positive
ETR 036B/18	31,9	Positive	Positive	Positive	Negative	Positive
ETR 037B/18	33,1	Positive	Positive	Negative	Negative	Positive
ETR 045B/18	34,0	Positive	Positive	Positive	Negative	Positive
ETR 048B/18	34,6	Positive	Positive	Positive	Negative	Positive
ETR 052B/18	27,9	Positive	Positive	Positive	Negative	Positive
ETR 164B/18	21,1	Positive	Positive	Positive	Negative	Positive
C-019	32,3	Positive	Positive	Positive	Negative	Positive
C-345	32,3	Positive	Positive	Negative	Negative	Positive
S-262	29,5	Positive	Positive	Positive	Positive	Positive
S-169	23,6	Positive	Positive	Positive	Negative	Positive
YOP HGE 022	32,9	Positive	Positive	Negative	Negative	Positive
H-3569	32,3	Positive	Positive	Positive	Negative	Positive
HGT HGE 011	12,5	Positive	Positive	Positive	Negative	Positive
ETR 144B/18	1,3	Negative	Negative	Positive	Positive	Negative

^aRatio of the sample optical density (OD) to the threshold value (TV), as calculated by Monolisa AgHBs Ultra®. Samples with OD/TV values greater than or equal to 1.00 are considered reactive. ^b Determined with two EIA tests: Dia.Pro HBsAg® one version ULTRA for detection of HBsAg; positive samples were confirmed using Monolisa™ HBsAg ULTRA Confirmation test.

antigen that are present in a relatively high proportion of asymptomatic carriers [14, 15]. Therefore, in a poor resource setting where EIA is unavailable, rapid diagnosis test is a reasonable alternative for epidemiologic surveys due to the lower cost of testing and simpler logistics [8]. As recommended by WHO, in the present study, we evaluated the performance of four HBsAg RDTs.

We found that Determine™ HBsAg and Vikia HBsAg® tests had best technical performance. Global specificities

obtained with Determine™ HBsAg and Vikia HBsAg® were similar to those reported by previous studies (100%) [9, 10, 16]. By contrast, sensitivities were slightly higher than those reported in previous evaluations between 82.5% and 98% [9, 16, 17].

Many studies revealed that the sensitivity and specificity of the SD Bioline WB kits were 94.1 to 100% and 99.3 to 100% for the HBsAg kit [18, 19], values which were similar to our findings. However, Farooq et al. have found low

sensitivity (17,4%) with SD Bioline HBsAg WB in healthy blood donors of Pakistan using PCR as gold standard [20].

The disparity of results on the sensitivity and specificity of rapid tests could be explained by the existence of mutant viruses which have modified surface antigens (HBsAg), thus making their detection by routine immunological techniques impossible [20, 21]. There are many HBsAg-immune-escape mutants those can be found both within and without the “a” determinant. The first mutant described, and which remains the most prevalent is G145R. Indeed, Studies reported HBsAg mutant prevalence ranging from 0,7% to 14,8% depending on the populations studied [22–25].

Also lower HBsAg concentration and viral load could lead to a false negative reaction [9, 20, 21]. Antibodies included in reagents do not take into account all mutated antigens.

Regarding the Standard Q HBsAg, no previous study, apart from the one reported by the manufacturer, has been documented, not allowing us to make comparison.

According to current WHO procurement eligibility for HBsAg assays that require that RDTs assays might have a diagnostic sensitivity and specificity of > 99% and > 98% respectively [26], three of the four evaluated tests (Determine™ HBsAg, Vikia HBsAg® and SD Bioline HBsAg WB®) can be accepted as HBsAg in vitro Diagnostic Tests.

We did not observe any significant difference in the performance of the tests evaluated with whole blood and serum/plasma. Njai et al. reported higher sensitivity in serum (88.5% vs 95.3%) but higher specificity in whole blood (100% vs 93.3%) [10]. This difference could be explained by the fact that the dried blood spots (DBS) has been used as a medium for the reference standard instead of serum or plasma.

An important consideration of definitive laboratory diagnosis also relates to controlling for false-positive and false negative results. We noticed a low rate of false negative results only with SD Bioline WB® (0.5%) and Standard Q® (2.8%) in this study. On the contrary, other studies have reported false-negative results of Determine and Vikia HBsAg tests. They associated this with a low HBsAg concentration, HBsAg mutants, low viral load, and certain viral genotypes [9, 10, 21]. Moreover false-negative results have a threat of silent transmission and spreading of diseases among people and also create more interest for sensitive assays like EIA.

In our study, the inter-reader variability and variability of results over time were not investigated. These parameters would have allowed us to appreciate the stability of the test results over time.

5. Conclusion

Among the four RDTs evaluated in this study, Determine™ HBsAg and Vikia HBsAg® tests should be appropriate for HBV screening and marketed in Ivory Coast. These two tests are rapid, simple and very suitable for peripheral laboratories and provided excellent performances both on serum / plasma and on whole blood. As recommended by the World Health Organization, Ivory Coast must now start the phase II of

evaluation of these two RDTs to validate these performances under field conditions in an environment characterized by limited resources and multiple public health priorities.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Authors' Contributions

Bamory Dembélé, André Inwoley, Daouda Sévédé and Emile Allah-Kouadio contributed to the design and implementation of the research

Roseline Affi-Aboli, Mathieu Kabran, Aimé Cézaire Adiko, Rodrigue Kouamé and Vanessa Goha managed the collection and manipulation of specimens.

André Inwoley, Bamory Dembélé, Daouda Sévédé contributed to the analysis of the results

Bamory Dembélé, Roseline Affi-Aboli and André Inwoley wrote the paper with input from all authors.

Funding

The authors state that they did not receive any specific funding for this research. Study was performed as part of the employment of the authors at Research and Diagnosis Center for AIDS and other infectious diseases (CeDReS), CHU (University Hospital) of Treichville, NBTS (National Blood Transfusion Center), Ivory Coast and Institut Pasteur, Ivory Coast (IPCI)

Acknowledgments

The authors would like to thank the Direction of Pharmacy, Drug and Laboratory in association with the National Program of fight against the viral hepatitis for initiating the study. We would like to thank all laboratory participants which supported this evaluation. Most importantly, we would like to thank all the participants of the study, without whom, this study would not have been possible.

References

- [1] World Health Organization, World Health Organization, and Global Hepatitis Programme, *Global hepatitis report, 2017*, 2017, December 2019, <http://apps.who.int/iris/bitstream/10665/255016/1/9789241565455-eng.pdf?ua=1>.
- [2] A. Schweitzer, J. Horn, R. T. Mikolajczyk, G. Krause, and J. J. Ott, “Estimations of worldwide prevalence of chronic hepatitis B virus infection: a systematic review of data published between 1965 and 2013,” *The Lancet*, vol. 386, no. 10003, pp. 1546–1555, 2015.

- [3] M.-J. Lohouès-Kouacou, C. Assi, A. Simen-Kapeu et al., "Prevalence of HBV Sero-markers in two Different Socioeconomic Groups of Schoolchildren from Abidjan, Côte d'Ivoire," *Journal of Gastroenterology and Hepatology Research*, vol. 2, no. 9, pp. 798–802, 2013.
- [4] C. Assi, A. Ouattara, S. Kone et al., "Screening for Hepatitis B and C in Occupational Settings: Cross-Sectional Study about 4268 Corporates Agents in Côte d'Ivoire," *Open Journal of Gastroenterology*, vol. 7, no. 3, pp. 89–95, 2017.
- [5] B. Seri, A. Minga, D. Gabillard et al., "Twenty-Year Evolution of Hepatitis B Virus and Human Immunodeficiency Virus Prevalence and Incidence in Voluntary Blood Donors in Côte d'Ivoire," *Open Forum Infectious Diseases*, vol. 5, no. 4, article ofy060, 2018.
- [6] W. Levinson, *Review of Medical Microbiology and Immunology*, McGraw-Hill Medical, Estados Unidos, 10 edition, 2008.
- [7] M. S. Khuroo, N. S. Khuroo, and M. S. Khuroo, "Accuracy of rapid point-of-care diagnostic tests for hepatitis B surface antigen—a systematic review and meta-analysis," *Journal of Clinical and Experimental Hepatology*, vol. 4, no. 3, pp. 226–240, 2014.
- [8] R. G. Gish, J. A. Gutierrez, N. Navarro-Cazarez et al., "A simple and inexpensive point-of-care test for hepatitis B surface antigen detection: serological and molecular evaluation," *Journal of Viral Hepatitis*, vol. 21, no. 12, pp. 905–908, 2014.
- [9] J. Bottero, A. Boyd, J. Gozlan et al., "Performance of rapid tests for detection of HBsAg and anti-HBsAb in a large cohort, France," *Journal of Hepatology*, vol. 58, no. 3, pp. 473–478, 2013.
- [10] H. F. Njai, Y. Shimakawa, B. Sanneh et al., "Validation of Rapid Point-of-Care (POC) Tests for Detection of Hepatitis B Surface Antigen in Field and Laboratory Settings in the Gambia, Western Africa," *Journal of Clinical Microbiology*, vol. 53, no. 4, pp. 1156–1163, 2015.
- [11] WHO, *Directives pour l'évaluation des techniques de dépistage du VIH en Afrique* May 2020, https://www.who.int/diagnostics_laboratory/publications/FR_HIVEval_Guide.pdf.
- [12] A.-M. Šimundić, "Measures of diagnostic accuracy: basic definitions," *EJIFCC*, vol. 19, no. 4, pp. 203–211, 2009.
- [13] J. R. Landis and G. G. Koch, "An application of hierarchical kappa-type statistics in the assessment of majority agreement among multiple observers," *Biometrics*, vol. 33, no. 2, pp. 363–374, 1977.
- [14] A. S. Patil and A. Shankarkumar, "Hepatitis B diagnosis in blood bank: evaluation and challenges," *MGM Journal of Medical Sciences*, vol. 2, no. 2, pp. 83–89, 2015.
- [15] M. Subic and F. Zoulim, "How to improve access to therapy in hepatitis B patients," *Liver International*, vol. 38, Supplement 1, pp. 115–121, 2018.
- [16] H. M. Cruz, L. P. Scalioni, V. S. de Paula et al., "Evaluating HBsAg rapid test performance for different biological samples from low and high infection rate settings & populations," *BMC Infectious Diseases*, vol. 15, no. 1, 2015.
- [17] A. Amini, O. Varsaneux, H. Kelly et al., "Diagnostic accuracy of tests to detect hepatitis B surface antigen: a systematic review of the literature and meta-analysis," *BMC Infectious Diseases*, vol. 17, Supplement 1, p. 698, 2017.
- [18] O. J. Kweon, Y. K. Lim, H. R. Kim, T. H. Kim, and M. K. Lee, "Analytical performance of newly developed rapid point-of-care test for the simultaneous detection of hepatitis A, B, and C viruses in serum samples," *Journal of Medical Virology*, vol. 91, no. 6, pp. 1056–1062, 2019.
- [19] S. R. Upreti, S. Gurung, M. Patel et al., "Prevalence of chronic hepatitis B virus infection before and after implementation of a hepatitis B vaccination program among children in Nepal," *Vaccine*, vol. 32, no. 34, pp. 4304–4309, 2014.
- [20] A. Farooq, U. Waheed, H. A. Zaheer, F. Aldakheel, S. Alduraywish, and M. Arshad, "Detection of HBsAg mutants in the blood donor population of Pakistan," *PLoS One*, vol. 12, no. 11, article e0188066, 2017.
- [21] H. Scheiblaue, M. el-Nageh, S. Diaz et al., "Performance evaluation of 70 hepatitis B virus (HBV) surface antigen (HBsAg) assays from around the world by a geographically diverse panel with an array of HBV genotypes and HBsAg subtypes," *Vox Sanguinis*, vol. 98, 3, part 2, pp. 403–414, 2010.
- [22] T. Bian, H. Yan, L. Shen et al., "Change in Hepatitis B Virus Large Surface Antigen Variant Prevalence 13 Years after Implementation of a Universal Vaccination Program in China," *Journal of Virology*, vol. 87, no. 22, pp. 12196–12206, 2013.
- [23] L. Sticchi, P. Caligiuri, R. Cacciani, C. Alicino, and B. Bruzzone, "Epidemiology of HBV S-gene mutants in the Liguria Region, Italy," *Human Vaccines & Immunotherapeutics*, vol. 9, no. 3, pp. 568–571, 2014.
- [24] S. P. Kamat, P. R. Mehta, S. M. Paranjpe, and N. A. Ingole, "Hepatitis B Virus (HBV) Infection in Liver Disease Patients in Mumbai, India with Special Reference to Hepatitis B Surface Antigen (HBsAg) Mutant Detection," *Journal of Clinical and Diagnostic Research*, vol. 8, no. 3, pp. 19–21, 2014.
- [25] P. Perazzo, N. Eguibar, R. H. Gonzalez, A. D. Nusblat, and M. L. Cuestas, *Hepatitis B Virus (HBV) and S-Escape Mutants: From the Beginning until Now*, 2015, May 2020, <http://ri.conicet.gov.ar/handle/11336/47548>.
- [26] World Health Organization, *WHO Performance Evaluation Acceptance Criteria for HBsAg In vitro diagnostics in the context of WHO Prequalification*, 2016, January 2019, https://www.who.int/diagnostics_laboratory/evaluations/hepb/161125.