

# Review Article **The Immunomodulation Potential of Exosomes in Tumor Microenvironment**

# Meng Wang in and Bo Zhang in the second seco

Reproductive Medicine Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

Correspondence should be addressed to Bo Zhang; bo.zhang@tjh.tjmu.edu.cn

Received 9 August 2021; Accepted 15 September 2021; Published 27 September 2021

Academic Editor: Xiao-Jie Lu

Copyright © 2021 Meng Wang and Bo Zhang. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Exosomes are lipid bilayer particles that originated from almost all types of cells and play an important role in intercellular communication. Tumor-derived exosomes contain large amounts of noncoding RNA, DNA, and proteins, which can be transferred into recipient cells as functional components in exosomes. These exosomal functional constituents depend on the originating cells, and it has been proved that types and numbers of exosomel components differ in cancer patients and healthy individuals. This review summarizes the role of tumor-derived exosomes in immunomodulation and discusses the application of exosomes in immunotherapy in cancers. Overall, exosomes isolated from cancer cells are turned out to promote immune evasion and interfere with immune responses in tumors through inducing apoptosis of CD8+ T cells, facilitating generation of Tregs, suppressing natural killer (NK) cell cytotoxicity, inhibiting maturation and differentiation of monocyte, and enhancing suppressive function of myeloid-derived suppressor cells (MDSCs). Mechanistically, exosomal functional components play a significant role in the immunomodulation in cancers. Moreover, based on the existing studies, exosomes could potentially serve as therapeutic delivery vehicles, noninvasive biomarkers, and immunotherapeutic vaccines for various types of cancers.

## 1. Introduction

Cancer is a global public health problem, and the incidence and mortality of cancer are dramatically increasing. It is reported that there are more than 18 million new cases and 9 million deaths each year worldwide [1–3]. There are several strategies for tumor development and survival, and tumor immune evasion, as one of the important mechanisms, enables tumors to escape from immune surveillance, inhibit antitumor immune responses, and finally, grow progressively [4]. Immune cells in tumor microenvironment play important roles in tumorigenesis [5], and exosomes released into tumor microenvironment have also been proved to be able to regulate immune responses in tumors [6].

Extracellular vesicle is membrane-bound vesicle in all body fluids for crosstalk between cells. Exosome, a subtype of extracellular vesicle, refers to lipid bilayer particles with a diameter of 50-150 mm [7]. They are released by almost

all types of cells into microenvironment and have emerged as a novel method for intercellular communication through either functional component transfer or membrane receptor-mediated signaling transduction [8]. Cancerderived exosomes are widely distributed in body fluids from tumor-xenograft animal models and cancer patients, such as plasma, ascites, pleural effusion, and other fluids [9]. It has been demonstrated that types and numbers of exosomal constituents differ in cancer patients and healthy individuals [10]. Exosomes isolated from cancer cells can interfere with immune responses and correlate with the development and progression of tumors, including tumor growth, invasion, and metastasis [11, 12]. However, the intercellular communication of cancer cells and noncancer cells via exosomes has not been fully illustrated. Therefore, this review is aimed at summarizing the functional components in exosomes, reviewing the role of tumor-derived exosomes in immunomodulation, and discussing the application of exosomes in immunotherapy in cancers.

## 2. Formation and Contents of Cancer Exosomes tas

The biogenesis of exosomes is attributed to endocytosis and exocytosis of all cell types [13]. The endocytosis of cell membrane can form early endosomes, which are located near the cell membrane with a tubular appearance. After acidification, protein contents change, and fusion with other membrane components and early endosomes develop into late endosomes, which are located near the nucleus and exhibit a spherical shape. The multivesicular bodies (MVBs) then develop from late endosomes through reversed budding. The process of budding of late endosomes into their lumen is limited, resulting in vesicle enrichment in internal lumen [14]. Subsequently, the vesicles, also named exosomes, are released into extracellular space mediated by the exocytic fusion of the external membrane of MVBs and the cell membrane (Figure 1).

Since exosomes are originated from endocytosis and exocytosis of membrane vesicles, the molecular compositions of exosomes are dependent on the originating cells (Figure 1). Structurally, exosomes are with a lipid bilayer that encloses cytoplasm; thus, proteins and nucleic acids, main components in cytoplasm, are also the contents of exosomes (Table 1). Cancer exosomes regulate tumor biological processes, including tumor progression, metastasis, and angiogenesis.

2.1. RNAs in Cancer Exosomes. MicroRNA (miRNA) in cancer exosomes has been demonstrated to be involved in tumor progression. miRNAs, which inhibit targeted mRNA translation process, can be absorbed by cells through exosomes and act as tumor promoters or suppressors, finally interfering with other stromal cells in tumor microenvironment [15]. In breast cancer, tumor-derived exosomes have been proved to be specifically enriched in miRNA, which is able to stimulate a transformation of nontumor-originated breast cells into tumors [16]. Similarly, miR-105 secreted by exosomes isolated from the same type of cancer cells is reported to promote metastasis by destroying vascular endothelial barriers [17]. High levels of miR-122 in exosomes can also reprogram glucose metabolism in nontumor cells to facilitate metastasis [18]. Except for the significant role in tumor development and metastasis, miRNA may also serve as biomarkers in cancer, especially colorectal cancer [19]. A study has demonstrated that exosomal miR-320d in serum may act as a noninvasive diagnostic marker for metastatic colorectal cancer [20]. Also, the expression level of circulating exosomal miR-25-3p is also associated with the metastasis of colorectal cancer [21]. In addition, exosomal miRNAs are also significant in other types of tumors, such as esophageal squamous cell carcinoma [22] and breast cancer [15], and serve as molecular biomarkers [23].

Long noncoding RNA (lncRNA), another type of RNA content in cancer exosomes [24], also participates in biological processes of genetic transcription, mRNA translation, and protein modification [25]. It has been proved that lncRNA from cancer cell-originated exosomes is able to promote tumorigenesis by facilitating angiogenesis, suppressing immune functions, and inducing metas-

tasis [26]. lncRNA HOTAIR can be conveyed into endothelial cells via exosomes secreted by glioma cells, and HOTAIR is known to promote angiogenesis [27]. Similarly, lncRNA H19, which facilitates hepatocarcinogenesis [28], is enriched in exosomes from liver cancer cells and can be internalized by endothelial cells, resulting in cell adhesion and angiogenesis [29]. Moreover, exosomal lncRNA SNHG16 in breast cancer is reported to suppress immune functions by inducing  $CD73^{+}\gamma\delta1$  Treg cells [30], revealing a close relationship between exosomal lncRNA and immunosuppression in cancers. Furthermore, many studies have also confirmed the involvement of exosomal lncRNA in tumor metastasis in different cancer types. lncRNA MALAT-1 is abundant in exosomes isolated from the serum of non-small-cell lung cancer patients, and it facilitates tumor migration and lymphatic node metastasis [31]. Meanwhile, serous exosomal lncRNA 91H, which is proved to promote tumor migration, also dramatically decreases after operation in colorectal cancer patients, indicating the significance of exosomal lncRNA in cancers.

Although circRNA is a novel type of noncoding RNA, the role of exosomal circRNA in cancer tissues has also been emphasized. The presence of circular RNA (circRNA) has been demonstrated to play a critical role in cancer growth and metastasis. Exosomal circPACRGL is upregulated in colorectal cancer, and promotes tumor cell proliferation, migration, and invasion *in vitro* [32]. Exosomal circRNAs are also found to promote cancer progression in gastric cancer [33] and tumor metastasis in hepatocellular carcinoma [34].

2.2. DNAs in Cancer Exosomes. Recently, it has been reported that DNA also exists in cancer-derived exosomes [35, 36], and exosomal DNA may be crucial in cell communication as RNA components do. Although the exact mechanism of DNA presence in exosomes is still under investigation, mitochondrial DNA (mtDNA) was observed in exosomes isolated from astrocytes and glioblastoma cells for the first time [37]. Interestingly, a large abundance of mtDNA can be detected on the surface of exosomes [38], and it is speculated that it may be associated with exosomal aggregation and recipient cell dysfunction [39]. Moreover, other types of DNA, including single-stranded DNA (ssDNA) and double-stranded DNA (dsDNA), are also overserved in exosomes derived from cancer cells in recent years [40, 41]. What should be noted is that exosomal dsDNA exhibits a tumor-specific manner, whose amount is much higher originated from cancer cells than normal cells [42]. It revealed that exosomal DNA may act as a noninvasion biomarker for early cancer detection. Moreover, exosomal DNA in cancers also allows for treatment monitoring. The horizontal transfer of mtDNA from exosomes can result in therapy resistance in oxidative phosphorylationdependent breast cancer and lead to metastasis [43]. However, in another study, the so-called exosomal DNA is just a nucleic acid-histone complex resulting from autophagy [44]. The presence of exosomal DNA needs to be further investigated and explored.



FIGURE 1: Formation and contents of exosomes. The biogenesis of exosomes is attributed to endocytosis and exocytosis of cells. The endocytosis of cell membrane can form early endosomes, which then develop into late endosomes. The process of budding of late endosomes into their lumen is limited, resulting in vesicle enrichment in internal lumen in multivesicular bodies (MVBs). Subsequently, exosomes, with a diameter of 50-150 nm, are released into extracellular space mediated by the exocytic fusion of the external membrane of MVBs and the cell membrane. The molecular compositions of exosomes are dependent on the originating cells, and structurally, exosomes are lipid bilayer vehicles with proteins and nucleic acids. MVBs: multivesicular bodies; HSP: heat shock proteins.

Content	Molecular types	Function	
RNA	MicroRNA (miRNA)	Inducing metastasis, invasion, and transformation and formation of tumors in noncancerous cells; serving as biomarkers in cancers	
	Long noncoding RNA (IncRNA)	Promoting tumorigenesis by facilitating angiogenesis, suppressing immune functions, and inducing metastasis	
	Circular RNA (circRNA)	Promoting carcinoma growth and metastasis	
DNA	DNA in exosomes (exoDNA)	Being associated with abnormal DNA replication in cancer cells, and reverse transcription of cellular RNA; mediating immune response activation	
Protein	Targeting/adhesion proteins		
	Membrane transport and fusion proteins	Regulating tumor proliferation, growth, metastasis,	
	Heat shock proteins	migration, angiogenesis, adhesion, immune suppression, and	
	Enzymes	many other biological processes of tumor;	
	Receptor proteins	having potential immunotherapeutic effects	
	Cell type-specific markers of the originating cells		

2.3. Proteins in Cancer Exosomes. The protein components in cancer exosomes are complicated and have an endocytic origin. A recent study identified 232 unique proteins via exosomal proteome using ion trap mass spectrometry [45]. Generally, targeting/adhesion proteins, membrane transport and fusion proteins, heat shock proteins, enzymes, receptor proteins, cell type-specific markers of the originating cells are the main constitutive protein components of exosomes. In hepatocarcinoma, exosomal proteins are associated with cell migration, invasion, and angiogenesis [46]. Threspanin-8 in gastric cancer cell-derived exosomes can act as a biomarker for cancer growth and metastasis [47]. Another study also reported that exosomal CD317 and EGFR might also be used as early biomarkers for non-small-cell lung cancer [48]. Some tumor-specific markers, such as prostate-specific antigen (PSA) and survivin, have



FIGURE 2: The functions of cancer exosomes in immunomodulation. Exosomes isolated from cancer cells are turned out to promote immune evasion and interfere with immune responses in tumors through inducing apoptosis of CD8+ T cells, facilitating generation of Tregs, suppressing NK cells cytotoxicity, inhibiting maturation and differentiation of monocyte, and enhancing suppressive function of MDSCs. NK cells: natural killer cells; DCs: dendritic cells; MDSCs: myeloid-derived suppressor cells.

also been reported to highly express in exosomes, revealing a clinical significance of exosomal markers in diagnosis and differential diagnosis in prostate cancer [49]. In addition, the function of exosomal proteins from metastatic and nonmetastatic murine breast cancer cells is totally different. The former one is mainly involved in cell proliferation, migration, and angiogenesis, while the latter mainly participated in cell adhesion [50]. Moreover, immunosuppressive proteins are highly expressed in melanoma cell-derived exosomes, resulting in cell apoptosis, T cells proliferation suppression, and NK cells dysfunction [51]. These results emphasize that exosomal proteins in cancer play a critical role in tumor progression via immunomodulation.

## 3. Cancer Exosomes in Immunomodulation

3.1. Cancer Exosomes Induce Apoptosis of  $CD8^+$  T Cells. Cytotoxic  $CD8^+$  T cells and  $CD4^+$  Th1 cells are the main antitumor immune response effector cells [52, 53]. Cancerderived exosomes regulate functions of T cells mainly through impairing proliferation and facilitating apoptosis of  $CD8^+$  T cells [54], while immune cell-derived exosomes intend to promote T cell proliferation [55] (Figure 2). Fas ligand (FasL) has been reported to be highly expressed in various types of tumor cell-derived exosomes, including ovarian cancer [56], melanoma [54], prostate cancer [57], and oral squamous cell carcinoma [58]. The activation of Fas/FasL signaling pathway can promote apoptosis and downregulate immune response [59]. Several studies have also demonstrated that FasL-expressed cancer exosomes

can induce CD8<sup>+</sup> T cell apoptosis and be correlated with poor prognosis [54, 60]. The interaction between programmed death-ligand 1 (PD-L1) in cancer cells and programmed cell death protein 1 (PD-1) receptor in activated T cells is another mechanism of immune evasion in cancer [61]. On the one hand, exosomal PD-L1 can inhibit T cell activation and proliferation [62]; on the other hand, it can be transferred into cancer cells [63], finally resulting in immunosuppression amplification. In non-small-cell lung cancer, the exosomal PD-L1 level in serum is much higher in patients suffering from cancers with a higher TMN stage, indicating that the level of exosomal PD-L1 might be a potential cancer progression monitoring [64]. Similarly, exosomal PD-L1 expression in serum is also regarded as a reliable marker for treatment response prediction in melanoma [61]. Apart from FasL and PD-L1, it is also reported that exosomes can induce immunosuppression through many mechanisms. For instance, galectin-9 is detected in exosomes isolated from nasopharyngeal carcinoma patients. The binding of exosomal galectin-9 and mucin-domain containing-3 (Tim-3) in T cells promotes apoptosis of CD4<sup>+</sup> T cells, and this process can be reserved by neutralizing these two components [65].

3.2. Cancer Exosomes Facilitate Generation of Tregs. Regulation T cells (Tregs) help tumor cells escape from attacks of immune system by releasing immunosuppressive cytokines, such as IL-10 and TGF- $\beta$ 1 [66], and the expression of tumor-infiltrating Tregs has a close relationship with the prognosis of cancer. It has been demonstrated that exosomes

isolated from tumor cells facilitate generation and expansion of Tregs [67] (Figure 2). CD4<sup>+</sup> CD25<sup>-</sup> T cells can be converted into CD4<sup>+</sup> CD25<sup>high</sup> FoxP3<sup>+</sup> Tregs after coculture with tumor-derived exosomes. In terms of mechanism, exosomal miR-214 inhibits the expression of phosphatase and tensin homolog (PTEN) in T cells and induce Tregs to secret IL-10, finally promoting tumor growth [68]. Similar phenomena can also be investigated in another study. Exosomes isolated from mutant KRAS lung cancer cells are found to facilitate the switch of naïve CD4<sup>+</sup> T cells into Tregs, which is also observed after the transfection of mutant KRAS cDNA. It indicates that exosomal mutant KRAS DNA leads to this conversion, and it is further confirmed by the enrichment of FoxP3<sup>+</sup> Tregs in tumor tissues from mutant KRAS patients, compared to the WT KRAS controls [69]. Moreover, lncRNA SNHG16 transmitted by exosomes is also proved to induce CD73<sup>+</sup> $y\delta$ 1 Treg in breast cancer by sponging miR-16-4p, which enables the downregulation of SMAD5, the subsequent enhancement of TGF-b1/SMAD5 pathway, and finally, the promotion of CD73 expression [30]. Furthermore, Th17 cells can either induce angiogenesis and immunosuppression to facilitate tumor progression or recruit immune cells to promote antitumor immune response [70]. An upregulated Treg/Th17 ratio can be observed in ovarian cancer patients, and the imbalance of Treg/Th17 is induced by the transfer of miR-29a-3p and miR-21-5p mediated by cancer-derived exosomes. These exosomal miRNAs can directly induce signal transducer and activator of transcription 3 (STAT3) inhibition, promoting cancer progression and metastasis [71].

3.3. Cancer Exosomes Suppress Cytotoxicity of NK Cells. Natural killer (NK) cells are innate lymphocytes involved in antitumor immune response and immune surveillance [72]. Natural killer group 2 member D (NKG2D) is an activating receptor for NK cells, loss of which is crucial in immune evasion [73]. It is found that the activity of NK cells is frequently decreased in cancer patients compared with healthy controls [74], so the expression of NKG2D in NK cells treated with exosomes isolated from cancer cells [75] (Figure 2). Studies have demonstrated that cancer-derived exosomes express NKG2D ligands to depress NKG2D expression and inhibit NK cell cytotoxicity [76]. The presence of TGF- $\beta$ 1, as a cytokine that can suppress NK cells cytotoxicity, in tumor-derived exosomes may contribute to the suppression of NK cells activity in cancers [75], which can be verified by the rescue of exosome-mediated NKG2D reduction in NK cells upon TGF- $\beta$ 1 neutralization [74]. Some other studies have also suggested that exosomes from cancers can promote tumor growth by impairing NK functions [77]. Noncoding RNAs in exosomes, such as circUHRF1, are also proved to be associated with NK cell exhaustion in cancers [78].

3.4. Cancer Exosomes Inhibit Maturation and Differentiation of Monocyte. Monocytes are innate myeloid cells that can differentiate into macrophages and dendritic cells (DCs). Cancer exosomes have been proved to induce immunosuppression by impairing maturation and differentiation of

monocytes [79] (Figure 2). DCs, as specialized antigenpresenting cells, have significant functions in innate and adaptive immune responses. However, DC functions are suppressed in tumor environment, and thereby immunesuppressive DCs are recruited and infiltrated in tumor tissues [80]. Cancer exosomes are first reported to inhibit monocyte differentiation by Valenti et al. [81]. Exosomes from lung and breast cancers block the process of myeloid precursor cells into DCs [82]. It has been reported that exosomes isolated from colorectal cancer and melanoma cells inhibit the differentiation of monocytes to DCs [81]. The possible mechanism of the suppression in immune system may attribute to the protein components in exosomes, such as TGF- $\beta$ , IL-6, and PGE2 [83]. It has been demonstrated that tumor-derived exosomes lead to inhibition of the differentiation of bone marrow myeloid precursors into DCs via secreting IL-6 and activating Stata3 signaling [84]. The intake of exosomal TGF- $\beta$ 1 released by tumor cells by immature DCs can also block DC maturation [85]. Meanwhile, the differentiation of monocytes to macrophages in colorectal cancer can also be regulated by exosomes [86]. Exosomes from glioblastoma-derived stem cells can foster the differentiation of monocytes to M2 macrophages, resulting in suppression in immune response [87]. Detailed mechanisms may focus on noncoding RNA in exosomes. For instance, the levels of miR-222-3p released by exosomes are higher in ovarian cancer patients, and it can be transferred into macrophages to induce tumor-promoting M2 population, resulting in tumor growth promotion [88]. Exosomal miR-200b is also upregulated in serums of ovarian cancer patients and promotes proliferation and invasion of cancer cells via inducing M2 macrophage polarization [89]. Immunosuppressive monocytes are gained by the fusion of tumor-derived exosomes and monocytes, exhibiting as high CD14 expression without HLA-DA expression [90], and CD14<sup>+</sup>HLA-DR<sup>lo/neg</sup> monocytes, as tumor-induced immunosuppressive mediator, have been proved to increase in serum of many cancers, such as pancreatic cancer [91]. Moreover, horizon transfer of tumor antigens into antigenpresenting cells, such as monocytes, macrophages, and DCs mediated by tumor-derived exosomes may activate tumor immune response, interfering with processes of intercellular communication [92]. In summary, tumor-derived exosomes mediate immunosuppression by interfering with the differentiation of monocytes into DCs and macrophages.

3.5. Cancer Exosomes Enhance Suppressive Function of MDSCs. Myeloid-derived suppressor cells (MDSCs), equipped with strong immune suppressive activity, are immature myeloid cells with multiple phenotypes in tumor microenvironment [93]. The accumulation of MDSCs negatively impacts the process of antigen processing and presentation and produces abundant immunosuppressive factors to interfere with immune responses [94]. Several studies have indicated that the suppressive function of MDSCs on T cells can be potentiated by cancer exosomes (Figure 2). In renal cancer, exosomal HSP70 facilitates proliferation and enhances activation of MDSCs via activating TLR2 signaling [95]. Meanwhile, miR-9 and miR-181a from breast

Exosome types	Exosome source	Functional cargo	Immune response	Reference
SMART-Exo	Expi293F	Anti-CD3, anti-HER2, anti-EGFR	Activating and redirecting T cells toward HER2- or EGFR-expressing breast cancer cells	[105]
B7-1 and B7-2 Exo	Leukemia cells	B7 costimulatory proteins and leukemia-associated antigens	Facilitating T cell-mediated antitumor responses	[106]
Cell-free vaccine	DCs differentiated from autologous monocytes	MHC and antigenic peptide	Initiating T cell responses	[124]
CD40L-Exo	3LL Lewis lung cells	CD40L, TAA	Activating DCs-mediated antitumor immunity in 3LL tumors	[125]
TEX-N1ND	HCC, breast and pancreatic cancer cells	N1ND and TAA	Activating DCs-mediated antitumor immune response	[126]
IFNγ-Exo vaccine	RM-1 cancer cells	IFN- $\gamma$ and TAA	Activating M1-mediated antitumor immune response in RM-1 tumors	[127]
Decoy for TNF-α	HEK293	Extracellular domain of TNFR1	Antagonizing TNF- $\alpha$ in vitro	[128]

TABLE 2: Engineered exosomes for cancer immunotherapy.

SMART-Exo: synthetic multivalent antibodies retargeted exosome; HER2: human epidermal growth factor receptor 2; EGFR: epidermal growth factor receptor; DCs: dendritic cells; MHC: major histocompatibility complex; CD40L: CD40 ligand; TAA: tumor-associated antigens; TEX: tumor-derived exosome; N1ND: N-terminus domain of HMGN1; HCC: hepatocellular carcinoma; IFN- $\gamma$ : interferon gamma; M1: macrophage 1; TNFR1: tumor necrosis factor receptor 1; TNF- $\alpha$ : tumor necrosis factor alpha.

cancer-derived exosomes facilitate tumor growth and immune evasion via enhancing MDSC function through activating JAK/STAT signaling pathway by separately inhibiting SOCS3 and PIAS3 [96]. TGF- $\beta$ 1 and PGE2 from exosomes isolated from breast cancers help with the accumulation of MDSCs and the enhancement of tumor growth [97]. Similarly, under hypoxia conditions, miR-10a and miR-21 in glioma-derived exosomes mediate MDSC expansion and activation via RORA and PTEN, respectively [98]. Exosomal miR-21 also plays a significant role in the modulation of MDSC function in oral squamous cell carcinoma microenvironment. Exosomes derived from hypoxic oral squamous cell carcinoma enhance immunosuppressive function of MDSCs to interfere with moderation functions of  $\gamma\delta$  T cell via miR-21/PTEN/PD-L1 signaling pathway [99].

#### 4. Cancer Exosomes in Immunotherapy

Immunotherapy has become a popular therapeutic option for cancer patients. Exosomes, membranous vesicles secreted by almost all types of cells, can be absorbed and internalized by recipient cells via membrane fusion, receptor transportation, and many other pathways [100]. Given the excellent modifiability, biocompatibility, and cyclic half-life, exosomes are regarded as potential therapeutic delivery vehicles for biological components such as antibodies and chemotherapeutic drugs [101, 102]. Abundant tumor peptide antigens, such as MHC I and MHC II, are capsulized in exosomes and can be used to stimulate antitumor responses as cell-free vaccines. In animal models, administration of exosomes-loaded DCs can improve the therapeutic effect of cytotoxic drugs and prolong the survival time [103]. To improve the targeting efficiency of naturally occurring exosomes, exosome reprogramming is becoming popu-

lar. A novel exosome platform named synthetic multivalent antibodies retargeted exosome (SMART-Exo) is designed and developed to enable exosome genetic modification [104, 105]. Exosomes are reprogrammed to express CD3specific antibodies for T cells and EGFR antibodies for EGFR-expressing breast cancer cells [104]. In another study, exosomes are also engineered to display both anti-CD3 and anti-HER2 antibodies to target cytotoxic T cells and HER2-expressing breast cancer cells [105]. In addition to breast cancer, CD80 and CD86 are also packed in engineered exosomes to facilitate immune responses and secret immune-associated cytokines in leukemia [106]. These studies demonstrate that exosome reprogramming might be potential targeted immunotherapy for cancers. Another exosome-based drug delivery system named exosomebased superparamagnetic nanoparticle cluster (SMNC-EXO) is also developed to help exosomes with targeted drug delivery in the presence of external magnetic fields [107]. In a word, engineered exosome-based drug delivery systems might be a promising candidate for antitumor immunotherapy in the future (Table 2).

It has been shown that activated immune cells can secret exosomes containing miRNAs that can be used as biomarkers for immunotherapy. A study investigates exosomal miRNA profiles in non-small-cell lung cancer patients receiving PD-1/PD-L1 immunotherapy and healthy controls. More than 150 unique exosomal miRNAs are identified in cancer patients, and hsa-miR-320d, hsa-miR-320c, and hsa-miR-320b may be potential biomarkers to predict treatment efficacy of PD-1/PD-L1 immunotherapy in lung cancers [108]. The levels of plasma exosomal caveolin-1 are found to be downregulated in ovarian cancer, and they are positively associated with prognosis and overall survival as a biomarker [109]. Similarly, plasma-derived exosomal miR-4732-5p is also highly expressed in epithelial ovarian cancer patients and could be used to monitor cancer progression [110]. The above studies suggest the significance of exosomes as biomarkers in immunotherapy and prognosis monitor.

However, most of the current studies that investigated the predictor value of exosomal components in cancer progression and immunotherapy are laboratory studies; the clinical values should be confirmed in clinical trials. The first exosome phase I trial reported the administration of autologous exosomes derived from DCs in 15 cancer patients, and the results confirm the safety of exosome administration, despite the fact that no specific T cell responses are observed in circulating [111]. Meanwhile, some other clinical trials also prove that plasma DC-derived exosomes in cancer patients are also proved to be effective. It is worthy noted that although exosomes are expected to be potential immunotherapeutic vaccines in cancer treatment, it still will be a long time for exosome-based immunotherapy in clinical practices [112].

# 5. Challenge in Clinical Application of Exosomes for Cancer Treatment

It seems that exosomes isolated from tumors have an excellent prospect and are emerged as a potential tool for cancer therapy, yet the future of exosomes in clinical application still present many challenges. Currently, there is no gold standard for exosome isolation and purification, but new methodologies seemingly emerge in an endless stream. It has been reported that the purity and concentration of exosomes isolated vary with isolation methods [113]. A comparison of miRNA profiles of exosomes isolated by different methods suggested that the discrepancies of the content and amount of miRNA result from methodological differences [114]. Similar results are also reported in other studies, in which the circulating exosomal miRNA varies due to the isolated methods used [115]. Moreover, the proteins in exosomes may also be differential in results of the difference of isolation techniques [116]. Ultracentrifugation is the most widely used approach for exosome isolation worldwide [117]; however, several shortcomings, including the existence of nonexosomal impurity, potential damage, low yield of exosomes, and RNA components, are still investigated [118]. In addition to isolation methods, concentration and components of exosomes can also be influenced by microenvironments. It has been found that hypoxic conditions can increase the production of exosomes [119]. Meanwhile, exosomal miRNAs are also reported to be influenced by the oxygen concentration in microenvironment in terms of expression level [120]. The inconsistency of productions and the drawbacks in different isolation methods limit the clinical utilization of exosomes in diagnostic, prognostic, and therapeutic applications. Furthermore, although some components in exosomes derived from cancer cells, such miRNAs and proteins, are thought to be potential biomarkers for diagnosis and prognosis of cancers [121-123], these biomarkers are lack of specificity on screening, which may narrow their application. Focusing on the limitations of characterizations of exosomes as well as isolation methods is conductive to the development of exosomes in human diseases.

#### 6. Conclusion

Tumor-derived exosomes contain large amounts of noncoding RNA, DNA, and proteins, which can be transferred into recipient cells as functional components in exosomes. Exosomes isolated from cancer cells are turned out to promote immune evasion and interfere with immune responses in tumors through inducing apoptosis of CD8+ T cells, facilitating generation of Tregs, suppressing NK cells cytotoxicity, inhibiting maturation and differentiation of monocyte, and enhancing suppressive function of MDSCs. Mechanistically, exosomal functional components play a significant role in the immunomodulation in cancers. Moreover, based on the existing studies, exosomes could potentially serve as therapeutic delivery vehicles, noninvasive biomarkers, and immunotherapeutic vaccines for various types of cancers.

## **Conflicts of Interest**

All authors have no conflicts of interest to declare.

# Acknowledgments

We would like to express heartfelt gratitude to our colleagues in Reproductive Medicine Center, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology. This study was supported by the National Natural Science Foundation of China (81601348) and the Fundamental Research Funds for the Central Universities (2021yjsCXCY095).

#### References

- F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," *CA: A Cancer Journal for Clinicians*, vol. 68, no. 6, pp. 394–424, 2018.
- [2] P. Subedi, S. Nembrini, Q. An et al., "Telomere length and cancer mortality in American Indians: the Strong Heart Study," *Geroscience*, vol. 41, no. 3, pp. 351–361, 2019.
- [3] A. Csiszar, P. Balasubramanian, S. Tarantini et al., "Chemically induced carcinogenesis in rodent models of aging: assessing organismal resilience to genotoxic stressors in geroscience research," *Geroscience*, vol. 41, no. 2, pp. 209– 227, 2019.
- [4] X. Lei, Y. Lei, J. K. Li et al., "Immune cells within the tumor microenvironment: biological functions and roles in cancer immunotherapy," *Cancer Letters*, vol. 470, pp. 126–133, 2020.
- [5] T. F. Gajewski, H. Schreiber, and Y. X. Fu, "Innate and adaptive immune cells in the tumor microenvironment," *Nature Immunology*, vol. 14, no. 10, pp. 1014–1022, 2013.
- [6] T. L. Whiteside, "Exosomes and tumor-mediated immune suppression," *The Journal of Clinical Investigation*, vol. 126, no. 4, pp. 1216–1223, 2016.
- [7] R. Kalluri and V. S. LeBleu, "The biology, function, and biomedical applications of exosomes," *Science*, vol. 367, no. 6478, p. eaau6977, 2020.
- [8] L. Gao, L. Wang, T. Dai et al., "Tumor-derived exosomes antagonize innate antiviral immunity," *Nature Immunology*, vol. 19, no. 3, pp. 233–245, 2018.

- [10] F. G. Kugeratski and R. Kalluri, "Exosomes as mediators of immune regulation and immunotherapy in cancer," *The FEBS Journal*, vol. 288, no. 1, pp. 10–35, 2021.
- [11] C. Kahlert and R. Kalluri, "Exosomes in tumor microenvironment influence cancer progression and metastasis," *Journal of Molecular Medicine (Berlin, Germany)*, vol. 91, no. 4, pp. 431–437, 2013.
- [12] Y. Liu, Y. Gu, and X. Cao, "The exosomes in tumor immunity," Oncoimmunology, vol. 4, no. 9, article e1027472, 2015.
- [13] R. C. Piper and D. J. Katzmann, "Biogenesis and function of multivesicular bodies," *Annual Review of Cell and Developmental Biology*, vol. 23, no. 1, pp. 519–547, 2007.
- [14] T. Döring, K. Gotthardt, J. Stieler, and R. Prange, "y2-Adaptin is functioning in the late endosomal sorting pathway and interacts with ESCRT-I and -III subunits," *Biochimica et Biophysica Acta*, vol. 1803, no. 11, pp. 1252–1264, 2010.
- [15] Q. Liu, F. Peng, and J. Chen, "The role of exosomal micro-RNAs in the tumor microenvironment of breast cancer," *International Journal of Molecular Sciences*, vol. 20, no. 16, p. 3884, 2019.
- [16] S. A. Melo, H. Sugimoto, J. T. O'Connell et al., "Cancer exosomes perform cell-independent microRNA biogenesis and promote tumorigenesis," *Cancer Cell*, vol. 26, no. 5, pp. 707–721, 2014.
- [17] W. Zhou, M. Y. Fong, Y. Min et al., "Cancer-secreted miR-105 destroys vascular endothelial barriers to promote metastasis," *Cancer Cell*, vol. 25, no. 4, pp. 501–515, 2014.
- [18] M. Y. Fong, W. Zhou, L. Liu et al., "Breast-cancer-secreted miR-122 reprograms glucose metabolism in premetastatic niche to promote metastasis," *Nature Cell Biology*, vol. 17, no. 2, pp. 183–194, 2015.
- [19] Ó. Rapado-González, A. Álvarez-Castro, R. López-López, J. Iglesias-Canle, M. M. Suárez-Cunqueiro, and L. Muinelo-Romay, "Circulating microRNAs as promising biomarkers in colorectal cancer," *Cancers*, vol. 11, no. 7, p. 898, 2019.
- [20] Y. Tang, Y. Zhao, X. Song, X. Song, L. Niu, and L. Xie, "Tumor-derived exosomal miRNA-320d as a biomarker for metastatic colorectal cancer," *Journal of Clinical Laboratory Analysis*, vol. 33, no. 9, article e23004, 2019.
- [21] Z. Zeng, Y. Li, Y. Pan et al., "Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis," *Nature Communications*, vol. 9, no. 1, p. 5395, 2018.
- [22] D. Luo, Z. Huang, H. Lv, Y. Wang, W. Sun, and X. Sun, "Upregulation of microRNA-21 indicates poor prognosis and promotes cell proliferation in esophageal squamous cell carcinoma via upregulation of lncRNA SNHG1," *Cancer Management and Research*, vol. 12, pp. 1–14, 2020.
- [23] V. C. Kok and C. C. Yu, "Cancer-derived exosomes: their role in cancer biology and biomarker development," *International Journal of Nanomedicine*, vol. 15, pp. 8019–8036, 2020.
- [24] M. Wang, L. Zhou, F. Yu, Y. Zhang, P. Li, and K. Wang, "The functional roles of exosomal long non-coding RNAs in cancer," *Cellular and Molecular Life Sciences*, vol. 76, no. 11, pp. 2059–2076, 2019.

- [25] F. Chen, N. Wang, H. Y. Tan, W. Guo, C. Zhang, and Y. Feng, "The functional roles of exosomes-derived long non-coding RNA in human cancer," *Cancer Biology & Therapy*, vol. 20, no. 5, pp. 583–592, 2019.
- [26] Z. Sun, S. Yang, Q. Zhou et al., "Emerging role of exosomederived long non-coding RNAs in tumor microenvironment," *Molecular Cancer*, vol. 17, no. 1, p. 82, 2018.
- [27] S. K. Tan, C. Pastori, C. Penas et al., "Serum long noncoding RNA HOTAIR as a novel diagnostic and prognostic biomarker in glioblastoma multiforme," *Molecular Cancer*, vol. 17, no. 1, p. 74, 2018.
- [28] I. J. Matouk, N. DeGroot, S. Mezan et al., "The H19 noncoding RNA is essential for human tumor growth," *PLoS One*, vol. 2, no. 9, article e845, 2007.
- [29] A. Conigliaro, V. Costa, A. Lo Dico et al., "CD90<sup>+</sup> liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 lncRNA," *Molecular Cancer*, vol. 14, no. 1, p. 155, 2015.
- [30] C. Ni, Q. Q. Fang, W. Z. Chen et al., "Breast cancer-derived exosomes transmit lncRNA SNHG16 to induce CD73+γδ1 Treg cells," *Signal Transduction and Targeted Therapy*, vol. 5, no. 1, p. 41, 2020.
- [31] R. Zhang, Y. Xia, Z. Wang et al., "Serum long non coding RNA MALAT-1 protected by exosomes is up-regulated and promotes cell proliferation and migration in non-small cell lung cancer," *Biochemical and Biophysical Research Communications*, vol. 490, no. 2, pp. 406–414, 2017.
- [32] A. Shang, C. Gu, W. Wang et al., "Exosomal circPACRGL promotes progression of colorectal cancer via the miR-142-3p/miR-506-3p- TGF-β1 axis," *Molecular Cancer*, vol. 19, no. 1, p. 117, 2020.
- [33] M. Xie, T. Yu, X. Jing et al., "Exosomal circSHKBP1 promotes gastric cancer progression via regulating the miR-582-3p/HUR/VEGF axis and suppressing HSP90 degradation," *Molecular Cancer*, vol. 19, no. 1, p. 112, 2020.
- [34] X. Y. Huang, Z. L. Huang, J. Huang et al., "Exosomal circRNA-100338 promotes hepatocellular carcinoma metastasis via enhancing invasiveness and angiogenesis," *Journal of Experimental & Clinical Cancer Research*, vol. 39, no. 1, p. 20, 2020.
- [35] X. Qu, Q. Li, J. Yang et al., "Double-stranded DNA in exosomes of malignant pleural effusions as a novel DNA source for EGFR mutation detection in lung adenocarcinoma," *Frontiers in Oncology*, vol. 9, p. 931, 2019.
- [36] Z. Song, Z. Cai, J. Yan, Y. W. Shao, and Y. Zhang, "Liquid biopsies using pleural effusion-derived exosomal DNA in advanced lung adenocarcinoma," *Translational Lung Cancer Research*, vol. 8, no. 4, pp. 392–400, 2019.
- [37] M. Guescini, S. Genedani, V. Stocchi, and L. F. Agnati, "Astrocytes and glioblastoma cells release exosomes carrying mtDNA," *Journal of Neural Transmission*, vol. 117, no. 1, pp. 1–4, 2010.
- [38] Y. Kawamura, Y. Yamamoto, T. A. Sato, and T. Ochiya, "Extracellular vesicles as trans-genomic agents: emerging roles in disease and evolution," *Cancer Science*, vol. 108, no. 5, pp. 824–830, 2017.
- [39] B. Konečná, Ľ. Tóthová, and G. Repiská, "Exosomes-associated DNA-new marker in pregnancy complications?," *International Journal of Molecular Sciences*, vol. 20, no. 12, p. 2890, 2019.
- [40] L. L. Wang, W. Q. Chen, Y. R. Wang et al., "Numerous long single-stranded DNAs produced by dual amplification

reactions for electrochemical detection of exosomal micro-RNAs," *Biosensors & Bioelectronics*, vol. 169, article 112555, 2020.

- [41] C. Kahlert, S. A. Melo, A. Protopopov et al., "Identification of double-stranded genomic DNA spanning all chromosomes with mutated *KRAS* and *p53* DNA in the serum exosomes of patients with pancreatic cancer," *The Journal of Biological Chemistry*, vol. 289, no. 7, pp. 3869–3875, 2014.
- [42] B. K. Thakur, H. Zhang, A. Becker et al., "Double-stranded DNA in exosomes: a novel biomarker in cancer detection," *Cell Research*, vol. 24, no. 6, pp. 766–769, 2014.
- [43] P. Sansone, C. Savini, I. Kurelac et al., "Packaging and transfer of mitochondrial DNA via exosomes regulate escape from dormancy in hormonal therapy-resistant breast cancer," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 114, no. 43, pp. E9066–E9075, 2017.
- [44] S. Pluchino and J. A. Smith, "Explicating exosomes: reclassifying the rising stars of intercellular communication," *Cell*, vol. 177, no. 2, pp. 225–227, 2019.
- [45] D. D. Taylor and C. Gercel-Taylor, "Exosomes/microvesicles: mediators of cancer-associated immunosuppressive microenvironments," *Seminars in Immunopathology*, vol. 33, no. 5, pp. 441–454, 2011.
- [46] R. Sasaki, T. Kanda, O. Yokosuka, N. Kato, S. Matsuoka, and M. Moriyama, "Exosomes and hepatocellular carcinoma: from bench to bedside," *International Journal of Molecular Sciences*, vol. 20, no. 6, p. 1406, 2019.
- [47] W. Li, C. Li, T. Zhou et al., "Role of exosomal proteins in cancer diagnosis," *Molecular Cancer*, vol. 16, no. 1, p. 145, 2017.
- [48] K. R. Jakobsen, B. S. Paulsen, R. Bæk, K. Varming, B. S. Sorensen, and M. M. Jørgensen, "Exosomal proteins as potential diagnostic markers in advanced non-small cell lung carcinoma," *Journal of Extracellular Vesicles*, vol. 4, no. 1, article 26659, 2015.
- [49] B. Pang, Y. Zhu, J. Ni et al., "Extracellular vesicles: the next generation of biomarkers for liquid biopsy-based prostate cancer diagnosis," *Theranostics*, vol. 10, no. 5, pp. 2309– 2326, 2020.
- [50] L. Gangoda, M. Liem, C. S. Ang et al., "Proteomic profiling of exosomes secreted by breast cancer cells with varying metastatic potential," *Proteomics*, vol. 17, no. 23-24, article 1600370, 2017.
- [51] P. Sharma, B. Diergaarde, S. Ferrone, J. M. Kirkwood, and T. L. Whiteside, "Melanoma cell-derived exosomes in plasma of melanoma patients suppress functions of immune effector cells," *Scientific Reports*, vol. 10, no. 1, p. 92, 2020.
- [52] B. Farhood, M. Najafi, and K. Mortezaee, "CD8<sup>+</sup> cytotoxic T lymphocytes in cancer immunotherapy: a review," *Journal of Cellular Physiology*, vol. 234, no. 6, pp. 8509–8521, 2019.
- [53] J. Borst, T. Ahrends, N. Bąbała, C. J. M. Melief, and W. Kastenmüller, "CD4<sup>+</sup> T cell help in cancer immunology and immunotherapy," *Nature Reviews Immunology*, vol. 18, no. 10, pp. 635–647, 2018.
- [54] E. U. Wieckowski, C. Visus, M. Szajnik, M. J. Szczepanski, W. J. Storkus, and T. L. Whiteside, "Tumor-derived microvesicles promote regulatory T cell expansion and induce apoptosis in tumor-reactive activated CD8<sup>+</sup> T lymphocytes," *Journal of Immunology*, vol. 183, no. 6, pp. 3720–3730, 2009.
- [55] E. Wieckowski and T. L. Whiteside, "Human tumor-derived vs dendritic cell-derived exosomes have distinct biologic roles

9

and molecular profiles," *Immunologic Research*, vol. 36, no. 1-3, pp. 247–254, 2006.

- [56] D. D. Taylor, C. Gerçel-Taylor, K. S. Lyons, J. Stanson, and T. L. Whiteside, "T-cell apoptosis and suppression of T-cell receptor/CD3-zeta by Fas ligand-containing membrane vesicles shed from ovarian tumors," *Clinical Cancer Research*, vol. 9, pp. 5113–5119, 2003.
- [57] A. J. Abusamra, Z. Zhong, X. Zheng et al., "Tumor exosomes expressing Fas ligand mediate CD8<sup>+</sup> T-cell apoptosis," *Blood Cells, Molecules & Diseases*, vol. 35, no. 2, pp. 169–173, 2005.
- [58] J. W. Kim, E. Wieckowski, D. D. Taylor, T. E. Reichert, S. Watkins, and T. L. Whiteside, "Fas ligand-positive membranous vesicles isolated from sera of patients with oral cancer induce apoptosis of activated T lymphocytes," *Clinical Cancer Research*, vol. 11, pp. 1010–1020, 2005.
- [59] R. M. Siegel, F. Ka-Ming Chan, H. J. Chun, and M. J. Lenardo, "The multifaceted role of Fas signaling in immune cell homeostasis and autoimmunity," *Nature Immunology*, vol. 1, no. 6, pp. 469–474, 2000.
- [60] G. Andreola, L. Rivoltini, C. Castelli et al., "Induction of lymphocyte apoptosis by tumor cell secretion of FasL-bearing microvesicles," *The Journal of Experimental Medicine*, vol. 195, no. 10, pp. 1303–1316, 2002.
- [61] T. Okazaki and T. Honjo, "PD-1 and PD-1 ligands: from discovery to clinical application," *International Immunology*, vol. 19, no. 7, pp. 813–824, 2007.
- [62] G. Chen, A. C. Huang, W. Zhang et al., "Exosomal PD-L1 contributes to immunosuppression and is associated with anti-PD-1 response," *Nature*, vol. 560, no. 7718, pp. 382– 386, 2018.
- [63] M. Poggio, T. Hu, C. C. Pai et al., "Suppression of exosomal PD-L1 induces systemic anti-tumor immunity and memory," *Cell*, vol. 177, no. 2, pp. 414–27.e13, 2019.
- [64] C. Li, C. Li, C. Zhi et al., "Clinical significance of PD-L1 expression in serum-derived exosomes in NSCLC patients," *Journal of Translational Medicine*, vol. 17, no. 1, p. 355, 2019.
- [65] J. Klibi, T. Niki, A. Riedel et al., "Blood diffusion and Th1suppressive effects of galectin-9-containing exosomes released by Epstein-Barr virus-infected nasopharyngeal carcinoma cells," *Blood*, vol. 113, no. 9, pp. 1957–1966, 2009.
- [66] J. Wada, H. Onishi, H. Suzuki et al., "Surface-bound TGFbeta1 on effusion-derived exosomes participates in maintenance of number and suppressive function of regulatory Tcells in malignant effusions," *Anticancer Research*, vol. 30, pp. 3747–3757, 2010.
- [67] M. Szajnik, M. Czystowska, M. J. Szczepanski, M. Mandapathil, and T. L. Whiteside, "Tumor-derived microvesicles induce, expand and up-regulate biological activities of human regulatory T cells (Treg)," *PLoS One*, vol. 5, no. 7, article e11469, 2010.
- [68] Y. Yin, X. Cai, X. Chen et al., "Tumor-secreted miR-214 induces regulatory T cells: a major link between immune evasion and tumor growth," *Cell Research*, vol. 24, no. 10, pp. 1164–1180, 2014.
- [69] A. Kalvala, P. Wallet, L. Yang et al., "Phenotypic switching of naïve T cells to immune-suppressive Treg-like cells by mutant KRAS," *Journal of Clinical Medicine*, vol. 8, no. 10, p. 1726, 2019.
- [70] L. Guéry and S. Hugues, "Th17 cell plasticity and functions in cancer immunity," *BioMed Research International*, vol. 2015, Article ID 314620, 11 pages, 2015.

- [71] J. Zhou, X. Li, X. Wu et al., "Exosomes released from tumorassociated macrophages transfer miRNAs that induce a Treg/Th17 cell imbalance in epithelial ovarian cancer," *Cancer Immunology Research*, vol. 6, no. 12, pp. 1578–1592, 2018.
- [72] E. Vivier, S. Ugolini, D. Blaise, C. Chabannon, and L. Brossay, "Targeting natural killer cells and natural killer T cells in cancer," *Nature Reviews Immunology*, vol. 12, no. 4, pp. 239–252, 2012.
- [73] P. Y. Lam, M. D. Nissen, and S. R. Mattarollo, "Invariant natural killer T cells in immune regulation of blood cancers: harnessing their potential in immunotherapies," *Frontiers in Immunology*, vol. 8, p. 1355, 2017.
- [74] A. Clayton, J. P. Mitchell, J. Court, S. Linnane, M. D. Mason, and Z. Tabi, "Human tumor-derived exosomes downmodulate NKG2D expression," *Journal of Immunology*, vol. 180, no. 11, pp. 7249–7258, 2008.
- [75] M. J. Szczepanski, M. Szajnik, A. Welsh, T. L. Whiteside, and M. Boyiadzis, "Blast-derived microvesicles in sera from patients with acute myeloid leukemia suppress natural killer cell function via membrane-associated transforming growth factor-1," *Haematologica*, vol. 96, no. 9, pp. 1302–1309, 2011.
- [76] L. Mincheva-Nilsson and V. Baranov, "Cancer exosomes and NKG2D receptor-ligand interactions: impairing NKG2Dmediated cytotoxicity and anti-tumour immune surveillance," Seminars in Cancer Biology, vol. 28, pp. 24–30, 2014.
- [77] C. Liu, S. Yu, K. Zinn et al., "Murine mammary carcinoma exosomes promote tumor growth by suppression of NK cell function," *Journal of Immunology*, vol. 176, no. 3, pp. 1375– 1385, 2006.
- [78] P. F. Zhang, C. Gao, X. Y. Huang et al., "Cancer cell-derived exosomal circUHRF1 induces natural killer cell exhaustion and may cause resistance to anti-PD1 therapy in hepatocellular carcinoma," *Molecular Cancer*, vol. 19, no. 1, p. 110, 2020.
- [79] K. E. Kunigelis and M. W. Graner, "The dichotomy of tumor exosomes (TEX) in cancer immunity: is it all in the Con-TEXt?," *Vaccine*, vol. 3, no. 4, pp. 1019–1051, 2015.
- [80] J. M. Tran Janco, P. Lamichhane, L. Karyampudi, and K. L. Knutson, "Tumor-infiltrating dendritic cells in cancer pathogenesis," *Journal of Immunology*, vol. 194, no. 7, pp. 2985– 2991, 2015.
- [81] R. Valenti, V. Huber, M. Iero, P. Filipazzi, G. Parmiani, and L. Rivoltini, "Tumor-released microvesicles as vehicles of immunosuppression: figure 1," *Cancer Research*, vol. 67, no. 7, pp. 2912–2915, 2007.
- [82] Y. Ning, K. Shen, Q. Wu et al., "Tumor exosomes block dendritic cells maturation to decrease the T cell immune response," *Immunology Letters*, vol. 199, pp. 36–43, 2018.
- [83] S. Yu, C. Liu, K. Su et al., "Tumor exosomes inhibit differentiation of bone marrow dendritic cells," *Journal of Immunol*ogy, vol. 178, no. 11, pp. 6867–6875, 2007.
- [84] Y. Liu, X. Xiang, X. Zhuang et al., "Contribution of MyD88 to the tumor exosome-mediated induction of myeloid derived suppressor cells," *The American Journal of Pathology*, vol. 176, no. 5, pp. 2490–2499, 2010.
- [85] C. Yang, S. H. Kim, N. R. Bianco, and P. D. Robbins, "Tumorderived exosomes confer antigen-specific immunosuppression in a murine delayed-type hypersensitivity model," *PLoS One*, vol. 6, no. 8, article e22517, 2011.
- [86] M. Baj-Krzyworzeka, B. Mytar, R. Szatanek et al., "Colorectal cancer-derived microvesicles modulate differentiation of

human monocytes to macrophages," *Journal of Translational Medicine*, vol. 14, no. 1, p. 36, 2016.

- [87] K. Gabrusiewicz, X. Li, J. Wei et al., "Glioblastoma stem cellderived exosomes induce M2 macrophages and PD-L1 expression on human monocytes," *Oncoimmunology*, vol. 7, no. 4, article e1412909, 2018.
- [88] X. Ying, Q. Wu, X. Wu et al., "Epithelial ovarian cancersecreted exosomal miR-222-3p induces polarization of tumor-associated macrophages," *Oncotarget*, vol. 7, no. 28, pp. 43076–43087, 2016.
- [89] J. Xiong, X. He, Y. Xu, W. Zhang, and F. Fu, "MiR-200b is upregulated in plasma-derived exosomes and functions as an oncogene by promoting macrophage M2 polarization in ovarian cancer," *Journal of Ovarian Research*, vol. 14, no. 1, p. 74, 2021.
- [90] A. E. Mengos, D. A. Gastineau, and M. P. Gustafson, "The CD14<sup>+</sup>HLA-DR<sup>lo/neg</sup> monocyte: an immunosuppressive phenotype that restrains responses to cancer immunotherapy," *Frontiers in Immunology*, vol. 10, p. 1147, 2019.
- [91] N. Javeed, M. P. Gustafson, S. K. Dutta et al., "Immunosuppressive CD14<sup>+</sup>HLA-DR<sup>lo/neg</sup> monocytes are elevated in pancreatic cancer and "primed" by tumor-derived exosomes," *Oncoimmunology*, vol. 6, no. 1, article e1252013, 2017.
- [92] L. Czernek, A. Chworos, and M. Duechler, "The uptake of extracellular vesicles is affected by the differentiation status of myeloid cells," *Scandinavian Journal of Immunology*, vol. 82, no. 6, pp. 506–514, 2015.
- [93] V. Kumar, S. Patel, E. Tcyganov, and D. I. Gabrilovich, "The nature of myeloid-derived suppressor cells in the tumor microenvironment," *Trends in Immunology*, vol. 37, no. 3, pp. 208–220, 2016.
- [94] P. Filipazzi, M. Bürdek, A. Villa, L. Rivoltini, and V. Huber, "Recent advances on the role of tumor exosomes in immunosuppression and disease progression," *Seminars in Cancer Biology*, vol. 22, no. 4, pp. 342–349, 2012.
- [95] Y. Gao, H. Xu, N. Li et al., "Renal cancer-derived exosomes induce tumor immune tolerance by MDSCs-mediated antigen-specific immunosuppression," *Cell Communication* and Signaling, vol. 18, no. 1, p. 106, 2020.
- [96] M. Jiang, W. Zhang, R. Zhang et al., "Cancer exosomederived miR-9 and miR-181a promote the development of early- stage MDSCs via interfering with SOCS3 and PIAS3 respectively in breast cancer," *Oncogene*, vol. 39, no. 24, pp. 4681–4694, 2020.
- [97] X. Xiang, A. Poliakov, C. Liu et al., "Induction of myeloidderived suppressor cells by tumor exosomes," *International Journal of Cancer*, vol. 124, no. 11, pp. 2621–2633, 2009.
- [98] X. Guo, W. Qiu, Q. Liu et al., "Immunosuppressive effects of hypoxia-induced glioma exosomes through myeloid-derived suppressor cells via the miR-10a/*Rora* and miR-21/*Pten* pathways," *Oncogene*, vol. 37, no. 31, pp. 4239–4259, 2018.
- [99] L. Li, B. Cao, X. Liang et al., "Microenvironmental oxygen pressure orchestrates an anti- and pro-tumoral  $\gamma\delta$  T cell equilibrium via tumor-derived exosomes," *Oncogene*, vol. 38, no. 15, pp. 2830–2843, 2019.
- [100] L. A. Mulcahy, R. C. Pink, and D. R. Carter, "Routes and mechanisms of extracellular vesicle uptake," *Journal of Extracellular Vesicles*, vol. 3, no. 1, 2014.
- [101] L. Zhao, C. Gu, Y. Gan, L. Shao, H. Chen, and H. Zhu, "Exosome-mediated siRNA delivery to suppress postoperative

breast cancer metastasis," Journal of Controlled Release, vol. 318, pp. 1–15, 2020.

- [102] Z. Xunian and R. Kalluri, "Biology and therapeutic potential of mesenchymal stem cell-derived exosomes," *Cancer Science*, vol. 111, no. 9, pp. 3100–3110, 2020.
- [103] L. Xiao, U. Erb, K. Zhao, T. Hackert, and M. Zöller, "Efficacy of vaccination with tumor-exosome loaded dendritic cells combined with cytotoxic drug treatment in pancreatic cancer," *Oncoimmunology*, vol. 6, no. 6, article e1319044, 2017.
- [104] Q. Cheng, X. Shi, M. Han, G. Smbatyan, H. J. Lenz, and Y. Zhang, "Reprogramming exosomes as nanoscale controllers of cellular immunity," *Journal of the American Chemical Society*, vol. 140, no. 48, pp. 16413–16417, 2018.
- [105] X. Shi, Q. Cheng, T. Hou et al., "Genetically engineered cellderived nanoparticles for targeted breast cancer immunotherapy," *Molecular Therapy*, vol. 28, no. 2, pp. 536–547, 2020.
- [106] W. Hu, F. Huang, L. Ning, J. Hao, J. Wan, and S. Hao, "Enhanced immunogenicity of leukemia-derived exosomes via transfection with lentiviral vectors encoding costimulatory molecules," *Cellular Oncology*, vol. 43, no. 5, pp. 889– 900, 2020.
- [107] H. Qi, C. Liu, L. Long et al., "Blood exosomes endowed with magnetic and targeting properties for cancer therapy," ACS Nano, vol. 10, no. 3, pp. 3323–3333, 2016.
- [108] X. X. Peng, R. Yu, X. Wu et al., "Correlation of plasma exosomal microRNAs with the efficacy of immunotherapy inEG-FR/ALKwild-type advanced non-small cell lung cancer," *Journal for Immunotherapy of Cancer*, vol. 8, no. 1, 2020.
- [109] L. Yang, H. Wu, Y. Zhu, X. Chen, and Y. Chen, "Plasma exosomal caveolin-1 predicts poor prognosis in ovarian cancer," *Journal of Cancer*, vol. 12, no. 16, pp. 5005–5012, 2021.
- [110] J. Liu, J. Yoo, J. Y. Ho et al., "Plasma-derived exosomal miR-4732-5p is a promising noninvasive diagnostic biomarker for epithelial ovarian cancer," *Journal of Ovarian Research*, vol. 14, no. 1, p. 59, 2021.
- [111] B. Escudier, T. Dorval, N. Chaput et al., "Vaccination of metastatic melanoma patients with autologous dendritic cell (DC) derived-exosomes: results of the first phase I clinical trial," *Journal of Translational Medicine*, vol. 3, no. 1, p. 10, 2005.
- [112] J. Zhou, X. L. Li, Z. R. Chen, and W. J. Chng, "Tumor-derived exosomes in colorectal cancer progression and their clinical applications," *Oncotarget*, vol. 8, no. 59, pp. 100781– 100790, 2017.
- [113] R. J. Lobb, M. Becker, S. Wen Wen et al., "Optimized exosome isolation protocol for cell culture supernatant and human plasma," *Journal of Extracellular Vesicles*, vol. 4, no. 1, p. 27031, 2015.
- [114] Y. T. Tang, Y. Y. Huang, L. Zheng et al., "Comparison of isolation methods of exosomes and exosomal RNA from cell culture medium and serum," *International Journal of Molecular Medicine*, vol. 40, no. 3, pp. 834–844, 2017.
- [115] M. Ding, C. Wang, X. Lu et al., "Comparison of commercial exosome isolation kits for circulating exosomal microRNA profiling," *Analytical and Bioanalytical Chemistry*, vol. 410, no. 16, pp. 3805–3814, 2018.
- [116] E. Serrano-Pertierra, M. Oliveira-Rodríguez, M. Rivas et al., "Characterization of plasma-derived extracellular vesicles isolated by different methods: a comparison study," *Bioengineering*, vol. 6, no. 1, p. 8, 2019.

- [117] C. Gardiner, D. Di Vizio, S. Sahoo et al., "Techniques used for the isolation and characterization of extracellular vesicles: results of a worldwide survey," *Journal of Extracellular Vesicles*, vol. 5, no. 1, article 32945, 2016.
- [118] M. Y. Konoshenko, E. A. Lekchnov, A. V. Vlassov, and P. P. Laktionov, "Isolation of extracellular vesicles: general methodologies and latest trends," *BioMed Research International*, vol. 2018, Article ID 8545347, 27 pages, 2018.
- [119] N. Ludwig, B. M. Razzo, S. S. Yerneni, and T. L. Whiteside, "Optimization of cell culture conditions for exosome isolation using mini-size exclusion chromatography (mini-SEC)," *Experimental Cell Research*, vol. 378, no. 2, pp. 149– 157, 2019.
- [120] H. Namazi, E. Mohit, I. Namazi et al., "Exosomes secreted by hypoxic cardiosphere-derived cells enhance tube formation and increase pro-angiogenic miRNA," *Journal of Cellular Biochemistry*, vol. 119, no. 5, pp. 4150–4160, 2018.
- [121] J. Nilsson, J. Skog, A. Nordstrand et al., "Prostate cancerderived urine exosomes: a novel approach to biomarkers for prostate cancer," *British Journal of Cancer*, vol. 100, no. 10, pp. 1603–1607, 2009.
- [122] J. Skog, T. Würdinger, S. van Rijn et al., "Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers," *Nature Cell Biol*ogy, vol. 10, no. 12, pp. 1470–1476, 2008.
- [123] D. M. Smalley, N. E. Sheman, K. Nelson, and D. Theodorescu, "Isolation and identification of potential urinary microparticle biomarkers of bladder cancer," *Journal of Proteome Research*, vol. 7, no. 5, pp. 2088–2096, 2008.
- [124] L. Zitvogel, A. Regnault, A. Lozier et al., "Eradication of established murine tumors using a novel cell-free vaccine: dendritic cell derived exosomes," *Nature Medicine*, vol. 4, no. 5, pp. 594–600, 1998.
- [125] J. Wang, L. Wang, Z. Lin, L. Tao, and M. Chen, "More efficient induction of antitumor T cell immunity by exosomes from CD40L gene-modified lung tumor cells," *Molecular Medicine Reports*, vol. 9, no. 1, pp. 125–131, 2014.
- [126] B. Zuo, H. Qi, Z. Lu et al., "Alarmin-painted exosomes elicit persistent antitumor immunity in large established tumors in mice," *Nature Communications*, vol. 11, no. 1, p. 1790, 2020.
- [127] X. Shi, J. Sun, H. Li et al., "Antitumor efficacy of interferon-γmodified exosomal vaccine in prostate cancer," *The Prostate*, vol. 80, no. 11, pp. 811–823, 2020.
- [128] N. Duong, K. Curley, A. Brown et al., "Decoy exosomes as a novel biologic reagent to antagonize inflammation," *International Journal of Nanomedicine*, vol. 14, pp. 3413–3425, 2019.