

Research Article

Semiconductor Quantum Dots (CdX, X=S, Te, Se) Modify Titanium Dioxide Nanoparticles for Photodynamic Inactivation of Leukemia HL60 Cancer Cells

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Received 12 August 2021; Revised 24 October 2021; Accepted 3 November 2021; Published 15 December 2021

Academic Editor: Mazeyar Parvinzadeh Gashti

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Titanium dioxide nanoparticles (TiO₂-NPs) are highly efficient photosensitizers in traditional photodynamic therapy (PDT). The particle size of TiO₂-NPs is small, only about 20 nm. However, the demands of ultraviolet light (UV) excitation feature shallow tissue penetration depth and may lead to severe tissue photon damage. Thus, in this research, TiO₂-NPs are modified with semiconductor quantum dots (QDs) CdX (X = S, Te, Se) in various methods, such as ultrasonic, hydrothermal, sol-gel, aqueous phase, and hydrolysis precipitation. The transmission electron microscopy (TEM) images show that the size of CdSe-TiO₂ is ranging from 6 to 14 nm. The ultraviolet-visible (UV-Vis) spectrum demonstrates that the CdX (X = S, Te, Se) modification can successfully extend the absorption range of TiO₂-NPs into a different visible light region. CdSe QDs have the narrowest band gap compared with CdX (X = S, Te, Se) QDs. Visible light-activated CdSe-TiO₂ nanocomposite shows the highest PDT inactivation efficiency toward HL60 cells compared with CdX-TiO₂. The photogenerated carrier separation efficiency of CdSe-TiO₂ nanocomposite is the highest shown in a fluorescence spectrum (FS). Furthermore, when conjugated with folic acid (FA), the prepared FA-CdX-TiO₂ (X = S, Se) exhibits excellent cancer-targeting ability during PDT treatment. Optimum PDT efficiency of FA-CdSe-TiO₂ indicates that photocatalytic and targeting ability is much higher than pure TiO₂ and CdSe-TiO₂. Our results provided a detailed investigation on the PDT performance of CdX (X = S, Te, Se) modified TiO₂ and may act as a guide for further design of highly targeted performance visible-light response TiO₂-NPs.

1. Introduction

Photodynamic therapy (PDT) is a nontoxic, noninvasive treatment [1]. Photodynamic therapy is based on the local or systemic administration of photosensitizer, which accumulates intensively in pathological tissues [2–4]. TiO₂ is the most common photocatalyst and photosensitizer with the advantages of low cost, good chemical stability, low toxicity, etc. [5–8]. The holes and electrons generated by TiO₂ have excellent oxidation and reduction abilities, respectively, under the ultraviolet light irradiation [9, 10]. However, due to the high band gap energy (3.2 eV) of TiO₂, the visible light response is ineffective and fast recombination of the electron-hole generated on the surface, which

limits the PDT efficiency of titanium dioxide in practical application [11, 12].

As reported in our previous work, coupling TiO₂ with low band gap energy metal or nonmetal to form a heterostructure is a feasible strategy to extend the visible light response range of TiO₂-based nanocomposites and thus improve the photocatalytic performance [13–16]. CdS, CdSe, and CdTe belong to the II-VI group cadmium-based quantum dot material [17, 18]. Previous reports have shown that the physical size of QDs with a high specific surface area is smaller than the Bohr radius of excitons. With low band gap energy, semiconductor quantum dots CdX (X = S, Te, Se) are also more resistant to photobleaching than organic dyes and fluorescent proteins [19, 20]. A lot of luminescent

materials which were resistant to photobleaching such as lanthanide-doped upconversion nanoparticles were used in near-infrared (NIR) excitation as biolabelling in human cervical cancer (HeLa) cells [21, 22]. However, the main problem of these materials was the low quantum efficiency at visible emission. Ruan et al. studied that CdS QDs had an illumination wavelength for photoelectrochemical generation of about 410 nm and were applied in a biosensor [23]. Guo et al. investigated that Daunorubicin-Loaded CdTe QDs enhanced the therapeutic effects of myelodysplastic syndromes to prevent acute leukemia from occurring [24]. Corredor et al. found that CdSe showed the best performance for hydrogen production compared with CdTe QDs [25]. Hua et al. developed strongly coupled CdX (X=S, Se and Te) quantum dots/TiO₂ nanocomposites, which had good performance in photocatalytic degradation of benzene [26]. Nideep et al. investigated the good photovoltaic performance of CdX (X=S, Te, Se) QD solar cells [17]. Bansal et al. compared the visible light absorption spectra of CdX (X=S, Te, Se) nanospheres; CdTe was the most efficient at absorbing solar energy [27]. Due to the lack of research on photodynamic therapy about CdX-TiO₂ (X=S, Te, Se) NPs, therefore, it is necessary to research the CdX-TiO₂ (X=S, Te, Se) NP-mediated PDT.

At present, the relapse rate of leukemia is high, and the disease cure rate should be improved [28]. Acute myeloid leukemia (AML) stem cells have abilities of highly self-renewal [29]. HL60 cells are common human AML cells. Earlier studies reveal that folate receptor- β (FR- β) was overexpressed in promyelocytic leukemia [30–32]. Most investigators find that FA can specifically combine with folate receptors (FR) that are overexpressed on the cancer cell cytomembrane but are rarely expressed in normal blood cells [33–35]. Even though CdX (X=S, Te, Se) modified TiO₂-NPs have potential in various drug delivery applications, a major challenge is its toxicity and nonspecific targeting. Most photosensitizers accumulate in normal and tumor tissue indiscriminately in nonspecific PDT, leading to the decrease in therapeutic effectiveness. So HL60 cells were used to evaluate the PDT efficiency of FA-CdX-TiO₂ nanocomposite. The surface of nanocomposites modified with FA can achieve targeting performance. In order to demonstrate FA-CdX-TiO₂ nanocomposite with higher PDT efficiency, we should evaluate the size, morphology, spectral response range, lower dark toxicity, and other physicochemical properties by TEM, X-Ray Diffraction (XRD), UV-Vis, Fourier transform infrared spectroscopy (FT-IR), and so on.

Our studies further explore and reveal the mechanism of high photocatalytic activity, cancer-targeting functionality, and potential therapeutic effect of CdX-TiO₂ nanocomposite modified with folic acid on PDT inactivating HL60 cells in vitro (Figure 1). CdX (X=S, Te, Se) modification can improve the PDT inactivation efficiency of TiO₂-NPs in visible light irradiation. When CdX-TiO₂ nanocomposite is conjugated with FA, it can exhibit excellent cancer-targeting ability in PDT inactivation. Our results provided mass of data investigation on PDT performance of CdX (X=S, Te, Se) modified TiO₂ and may act as a guide for further design of high targeting ability TiO₂-NPs in visible light response.

2. Materials and Methods

2.1. Chemicals and Apparatus. HL60 cells were kindly provided by the Department of Medicine of Sun Yat-sen University. RPMI medium 1640 and fetal calf serum (FCS) were obtained from Gibco BRL (USA). Fluo-3 AM and phosphate-buffered saline (PBS) were obtained from Sigma (USA). Cell Counting Kit-8 (CCK-8) assays were purchased from Dojindo (Japan). TiO₂-NPs were purchased from Degussa (GER). Trypan blue was obtained from Invitrogen (USA).

The apparatuses used in this research are as follows: ZEISS Ultra-55 scanning electron microscope (Carl Zeiss, Germany), JEM-2100HR transmission electron microscope (JEOL, Japan), U-3010 UV-visible spectrophotometer (Hitachi, Japan), F-4500 Fluorescence Spectrophotometer (Hitachi, Japan), the Countess automated cell counter (Invitrogen, USA), Model 680 Microplate Reader (Bio-Rad, USA), HH.CP-TW80 CO₂ incubator (Bluepard, China), lab-assembled PDT light reaction chamber, lab-assembled 410 nm LED light source, BRUKER D8 ADVANCE X-ray powder diffractometer (XRD) (Bruker, Germany), BS124S Electronic Scales (Sartorius, GER), SK2510LHC Ultrasonic Cleaner (KUDOS, China), SW-CJ Standard Clean Bench (Suzhou Antai Airtech Co., Ltd., China), LPE-1A Laser Power Meter (Physcience Opto-Electronics, Beijing), Eppendorf (Finland), 96-well culture plates (Costar, USA), and so on.

2.2. Light Source. The emission spectrum of light-emitting diode (LED) light source in the photodynamic irradiation chamber used in this experiment shows that emitting light from 400 to 420 nm in the visible-light region and the emission peak is located at 409.97 nm (Figure 2). The luminous power at the position of the sample was 18 J/cm² as measured with a photodiode.

2.3. Preparation of CdS-Based Nanocomposite A. Isopropyl titanate (C₁₂H₂₈O₄Ti, CAS No. 546-68-9, purity 95%) was purchased from Shanghai Macklin Biochemical Technology Co., Ltd., China. Cadmium chloride hydrate (CdCl₂·xH₂O, CAS No. 654054-66-7, purity 95%) was purchased from Aladdin Industrial Co., China. Ethylenediamine (C₂H₈N₂, CAS No. 6780-13-8, purity 98%), carbon disulfide (CS₂, CAS No. 30860-31-2, purity 99%), and anhydrous ethanol (C₂H₆O, CAS No. 64-17-5) were purchased from Tianjin Zhiyuan Chemical Industry Ltd., China. All reagents were of analytical purity and used without further purification.

The water temperature in the ultrasonic cleaner was set to 35°C. Then, 40 mL of deionized water at 14.8°C was added to the beaker and the beaker was placed in the cleaner. Dropping 0.6 mL of ethylenediamine and 0.2 mL of carbon disulfide into the beaker. Ultrasonic dispersion was carried out for 2 minutes (40 kHz, power 100 W); then, 10 mL of 0.015 g/mL CdCl₂·xH₂O aqueous solution was dropped, and ultrasound continued for 10 minutes. The ultrasonic cleaner increased the water temperature from 35°C to 71°C, and the ultrasound continued for 30 minutes (40 kHz, power 100 W). Isopropyl titanate (0, 0.2, 0.4, 0.6, and 1.0 mL) was

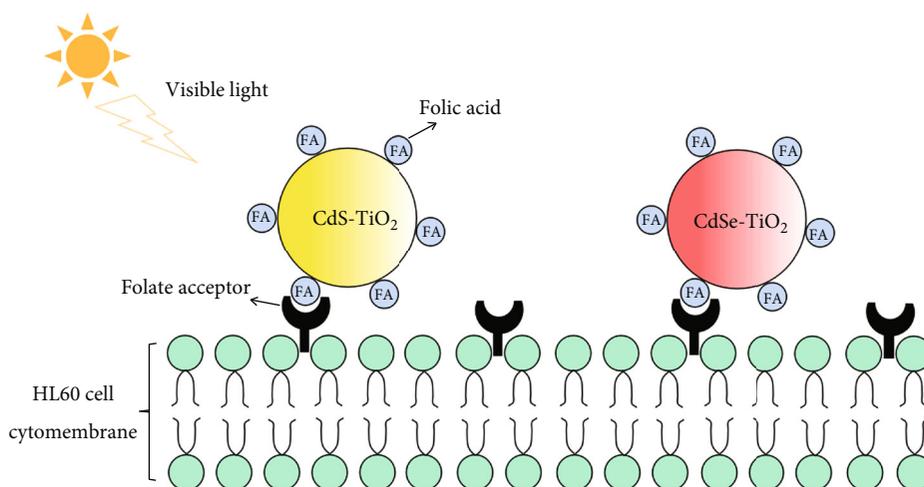


FIGURE 1: The possible mechanism of CdX (X=S, Te, Se)-based nanocomposites modified with folic acid with enhanced targeting functionality.

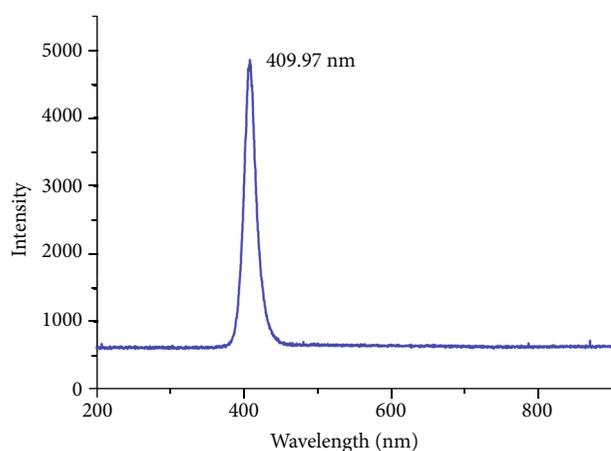


FIGURE 2: The emission spectra of blue LED array source.

added drop by drop, the temperature was controlled unchanged, and the ultrasound continued for 1.5 hours (40 kHz, power 100 W). The precipitate was centrifugally separated (10000 r/min, 10 minutes), washed once with deionized water, twice with anhydrous ethanol, and air-dried at room temperature. The obtained precipitate was calcined in the muffle furnace at 500°C for 4 hours, and grinding for 30 minutes, filtered through a 0.22 μm membrane filter, and sterilized to prepare the sample. The obtained samples were labeled as sample TiO_2 , 0.2, 0.4, 0.6, and 1.0, depending on the amount of TiO_2 precursor isopropyl titanate added.

2.4. Different Preparation Methods of CdTe-Based Titanium Dioxide Nanocomposites. CdTe-based nanocomposite A: the water-soluble CdTe quantum dots were prepared by the hydrothermal method. The TiO_2 -CdTe nanocomposite was prepared by ultrasonic mixing. CdTe-based nanocomposite B: the core-shell structure of CdTe/ TiO_2 was prepared by the sol-gel method. CdTe was coated inside the TiO_2 . CdTe-based nanocomposite C: CdTe was coated with carboxylic acid to form CdTe/HS- CH_2 -COOH nanoparticle.

2.4.1. Preparation of CdTe-Based Nanocomposite A. Tellurium (Te, CAS No. 13494-80-9, purity > 99.99%) and sodium hydroxide (NaOH, CAS No. 1310-73-2, A.R.) were purchased from Tianjin Zhiyuan Chemical Industry Ltd., China. Sodium borohydride (NaBH_4 , CAS No. 16940-66-2, purity 98%) and cadmium chloride (CdCl_2 , CAS No. 10108-64-2, A.R.) were purchased from Aladdin Industrial Co., China. Thioglycolic acid ($\text{C}_2\text{H}_4\text{O}_2\text{S}$, CAS No. 68-11-1, A.R.) was purchased from Sigma-Aldrich, USA. All reagents were of analytical purity and used without further purification.

Referring to the method of Zhang et al. and making appropriate adjustments [36], poured tellurium powder and NaBH_4 into the reaction flask, respectively, injected proper amount of water, reacting for half an hour under magnetic stirring. The reaction system is protected by nitrogen deoxygenation during the reaction process to obtain NaHTe solution. Thioglycolic acid was added into the aqueous solution of CdCl_2 saturated with nitrogen, and the pH value of the solution was adjusted to about 9.5 by NaOH. After stirring the solution with strong magnetic force for half an hour, NaHTe solution was added and magnetically stirred for reflux at 95°C for two hours. The experimental process was protected by nitrogen gas to obtain CdTe water-soluble QDs. After that, CdTe powder was obtained by centrifugation, drying, and grinding.

2.4.2. Preparation of CdTe-Based Nanocomposite B. Tellurium (Te, CAS No. 13494-80-9, purity > 99.99%), sodium hydroxide (NaOH, CAS No. 1310-73-2, A.R.), ethylene glycol ($\text{C}_2\text{H}_6\text{O}_2$, CAS No. 107-21-1, A.R.), and anhydrous ethanol ($\text{C}_2\text{H}_6\text{O}$, CAS No. 64-17-5, A.R.) were purchased from Tianjin Zhiyuan Chemical Industry Ltd., China. Sodium borohydride (NaBH_4 , CAS No. 16940-66-2, purity 98%), cadmium chloride (CdCl_2 , CAS No. 10108-64-2, A.R.), and acetylacetone ($\text{C}_5\text{H}_8\text{O}_2$, CAS No. 123-54-6, A.R.) were purchased from Aladdin Industrial Co., China. Isopropyl titanate ($\text{C}_{12}\text{H}_{28}\text{O}_4\text{Ti}$, CAS No. 546-68-9, purity 95%) was purchased from Shanghai Macklin Biochemical Technology

Co., Ltd., China. Thioglycolic acid ($C_2H_4O_2S$, CAS No. 68-11-1, A.R.) was purchased from Sigma-Aldrich, USA. All reagents were of analytical purity and used without further purification.

NaHTe solution was prepared by reduction of tellurium powder with sodium borohydride ($NaBH_4$) under nitrogen. Thioglycolic acid was added into the nitrogen-saturated $CdCl_2$ solution, and the pH value of the solution was adjusted to about 11 with 4 mol/L NaOH solution. Then, the NaHTe solution was quickly added, and the solution was stirred evenly and refluxed in a water bath at $100^\circ C$ for 1 hour to obtain water-soluble CdTe QDs. The whole process was in a nitrogen deoxygenation state.

CdTe/ TiO_2 nanocomposite was prepared by the sol-gel method. To synthesize titanium-containing solution with good dispersibility, the isopropyl titanate was added into ethylene glycol under magnetic stirring for 5 hours. After 2 mL of CdTe solution was added dropwise to 280 mL of acetylacetone under magnetic stirring and stirred evenly, then, 15 mL of titanium-containing solution was added dropwise slowly, and magnetic stirring was continued for 15 minutes and left to set for at least 30 minutes. Then, the samples were centrifuged in a 10000 r/min high-speed centrifuge for 10 minutes. After the precipitate was dissolved in anhydrous ethanol for ultrasonic dispersion, it was centrifuged again and repeated for three times to obtain CdTe/ TiO_2 nanocomposite.

2.5. Different Preparation Methods of CdSe-Based Nanocomposites. CdSe-based nanocomposite A: CdSe was coated with carboxylic acid to form CdSe/HS- CH_2 -COOH nanoparticle. CdSe-based nanocomposite B: the CdSe-doped anatase TiO_2 (CdSe/ TiO_2) was synthesized by the ultrasonic-driven method.

2.5.1. Preparation of CdSe-Based Nanocomposite B. The hydrazine monohydrate ($H_4N_2 \cdot H_2O$, CAS No. 7803-57-8, purity 85%) and the ammonium hydroxide solution ($NH_3 \cdot H_2O$, CAS No. 1336-21-6, purity 25%) were purchased from Tianjin Zhiyuan Chemical Industry Ltd., China. The sodium sulfite anhydrous (Na_2SO_3 , CAS No. 7757-83-7, A.R.), cadmium nitrate ($Cd(NO_3)_2 \cdot 4H_2O$, CAS No. 10022-68-1, A.R.), acetic acid (CH_3COOH , CAS No. 64-19-7, A.R.), and selenium (Se, CAS No. 7782-49-2, purity 99.99%) were purchased from Shanghai Macklin Biochemical Technology Co., Ltd., China. All reagents were of analytical purity and used without further purification.

In a 250 mL conical flask, anhydrous sodium sulfite (Na_2SO_3) (0.315 g) was dissolved in 50 mL distilled water, and selenium (Se) (0.1974 g) was added to the mixed solution to obtain selenosulfate solution (Na_2SeSO_3) (about 0.0025 mol). Then, 5 mL hydrazine hydrate ($H_4N_2 \cdot H_2O$) solution was added to a 250 mL conical flask of the prepared selenosulfate solution (Na_2SeSO_3) until the pH value of the solution reached 9.0 approximately. After cadmium nitrate $Cd(NO_3)_2 \cdot 4H_2O$ (1.5424 g) and $NH_3 \cdot H_2O$ (3 mL) were mixed, the mixed solution was added to the prepared Na_2SeSO_3 , and then, water-bath ultrasound was performed for 2 hours in an ultrasonic cleaning tank (operating fre-

quency: 59 kHz). It could be observed that the turbid solution gradually changed from black to bright yellow and then to bright orange which was the CdSe nanoparticle solution. Then, 0.9568 g, 0.4784 g, and 0.2392 g of TiO_2 were added into the prepared CdSe nanoparticle solution according to the doping mass ratios of CdSe doped TiO_2 of one to two, one to one, and two to one, respectively, and the mixed solution of CdSe nanoparticles with TiO_2 was subjected to ultrasonic radiation. Finally, the samples with different doping ratios of CdSe to TiO_2 equaled 0.5, CdSe to TiO_2 equaled one, and CdSe to TiO_2 equaled two were labeled as CdSe/ TiO_2 -0.5, CdSe/ TiO_2 -1, and CdSe/ TiO_2 -2, respectively, through centrifugation and vacuum drying.

2.6. Preparation of CdX (X = S, Se)-Based Nanocomposite Modified with Folic Acid

2.6.1. Preparation of CdS-Based Nanocomposite B. Folic acid ($C_{19}H_{19}N_7O_6$, CAS No. 59-30-3, purity $\geq 97\%$) was purchased from Sigma-Aldrich, USA. Sodium bicarbonate ($NaHCO_3$, CAS No. 144-55-8, A.R.) was purchased from Aladdin Industrial Co., China. Hydrochloric acid (HCl, CAS No. 7647-01-0, purity 37%) and sodium hydroxide (NaOH, CAS No. 1310-73-2, A.R.) were purchased from Tianjin Zhiyuan Chemical Industry Ltd., China. All reagents were of analytical purity and used without further purification.

Firstly, 420 mg of the sodium bicarbonate solution was weighed, then added into 50 mL of deionized water, and adjusted the pH of the solution to 5.5 with sodium hydroxide and hydrochloric acid. Then, a proper amount of folic acid was added into the prepared sodium bicarbonate solution, and the mixture was uniformly mixed with a magnetic stirrer, labbed as solution A. The same amount of the prepared CdS- TiO_2 nanocomposite (CdS-based nanocomposite A) was added into 4.5 mL of deionized water and dispersed by ultrasound for 15 minutes, labbed as solution B. The solution B was added dropwise to solution A while stirring in the ultrasound, and the mixed solution was treated in dark and continuously stirred at room temperature for 24 hours. The obtained turbidity mixture was centrifuged, washed twice with saturated sodium bicarbonate solution and deionized water, and then air-dried at room temperature. Finally, the sample was filtered through a $0.22 \mu m$ membrane filter and sterilized to obtain FA-CdS- TiO_2 nanocomposite (CdS-based nanocomposite B).

2.6.2. Preparation of CdSe-Based Nanocomposite C. Folic acid ($C_{19}H_{19}N_7O_6$, CAS No. 59-30-3, purity $\geq 97\%$) and thioglycolic acid ($C_2H_4O_2S$, CAS No. 68-11-1, A.R.) were purchased from Sigma-Aldrich, USA. Selenium (Se, CAS No. 7782-49-2, purity $> 99.99\%$) and sodium sulfite anhydrous (Na_2SO_3 , CAS No. 7757-83-7, A.R.) were purchased from Shanghai Macklin Biochemical Technology Co., Ltd., China. Tetrabutyl titanate ($C_{16}H_{36}O_4Ti$, CAS No. 5593-70-4, purity $\geq 99\%$), anhydrous ethanol (C_2H_6O , CAS No. 64-17-5, A.R.), hydrochloric acid (HCl, CAS No. 7647-01-0, 37%), and sodium hydroxide (NaOH, CAS No. 1310-73-2, A.R.) were purchased from Tianjin Zhiyuan Chemical Industry Ltd., China. Cadmium chloride ($CdCl_2$, CAS No.

10108-64-2, A.R.) and sodium bicarbonate (NaHCO_3 , CAS No. 144-55-8, A.R.) were purchased from Aladdin Industrial Co., China. All reagents were of analytical purity and used without further purification.

CdSe-TiO_2 nanocomposite was prepared by hydrolytic precipitation method. After 0.13196 g CdSe powder was weighed and added with 30 mL of anhydrous ethanol, the dispersion system A was obtained by ultrasonic vibration for 1 hour. Then, 1.761 mL tetrabutyl titanate was dropwise added to solution A with stirring for 15 minutes to get dispersion system B. Next, 10 mL of water and anhydrous ethanol mixture in a volume ratio of one to five was added dropwise to solution B with continuously stirring for 1 hour. The precipitate was separated by centrifugation and further washed twice with distilled water and anhydrous ethanol, respectively, and air-dried at room temperature. The CdSe-TiO_2 nanocomposite was prepared by calcination in the muffle furnace at 500°C for 4 hours, grinding for 30 minutes, filtration, and sterilization.

Firstly, 840 mg of sodium bicarbonate was dissolved in 100 mL of deionized water, 40 mL of the sodium bicarbonate solution was measured and adjusted the pH value to 5.5 using sodium hydroxide and hydrochloric acid. A proper amount of folic acid was added into the prepared carbonic acid solution and stirred on a magnetic stirrer. A proper amount of CdSe-TiO_2 nanocomposite was added into 5 mL deionized water for ultrasonic dispersion. The prepared CdSe-TiO_2 nanocomposite suspension was added step by step to the folic acid solution under the ultrasonic environment. And the mixed solution was treated in the dark and continuously stirred at room temperature for 24 hours. Finally, the prepared FA- CdSe-TiO_2 nanocomposite (CdSe-based nanocomposite C) was washed twice with saturated sodium bicarbonate solution and deionized water, respectively.

2.7. Selection of Nanoparticle Concentration. The concentration range selected in our experiment is from 0.2 to $320\ \mu\text{g}/\text{mL}$. Previous studies have shown that TiO_2 NPs did not affect HL60 cell viability at the concentration from 0 to $150\ \mu\text{g}/\text{mL}$ [37, 38]. Higher than $150\ \mu\text{g}/\text{mL}$ belongs to high concentration [39]. Working with high concentrations of nanoparticles possibly affects the normal cell viability and reduces the toxicological value of the results [40].

2.8. Cell Culture. Human leukemia HL60 cells were cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) in a humidified incubator with 5% CO_2 at 37°C . All experiments were performed using cells during the logarithmic growth phase. The cell concentration was measured using a Countess automated cell counter, and the cell density was adjusted to the required final concentration. Cell viability before treatment was always over 95%.

2.9. Cell Viability Assay. The cell activity detection is by using the Model 680 Microplate Reader, the method of CCK-8 (Cell Counting Kit-8) with simple operation, high sensitivity, and good repeatability compared with traditional methods. To make the results more precise, a method of

dual-wavelength testing is conducted, with 450 nm as the measuring wavelength and 630 nm as the reference wavelength to detect the cell activity of general steps: (1) the preparation of cell suspension, (2) cell suspension will join the 96-well culture plates and culture at a 37°C incubator for a period of time, (3) to join the appropriate amount of CCK-8 agent, at 37°C incubator shading cultivation in four hours, and (4) determination of absorbance (OD value).

2.10. Statistical Analysis. Data were presented as means \pm SD (standard deviation) from three independent groups. Each experiment was repeated three times. SPSS22.0 statistical software was used to conduct the analysis of variance and the *t*-test for the comparison of multiple local mean. The value of *P* smaller than 0.05 was considered statistically significant.

3. Results and Discussion

3.1. Characterization of CdX ($X = \text{S}, \text{Te}, \text{Se}$)-Based Nanocomposites

3.1.1. TEM and Particle Size Studies. The morphology and particle sizes were observed by a JEM-2100HR transmission electron microscope, dipping a small amount of nanocomposites into deionized water and conducting ultrasonic treatment for 2 minutes. To take a drop of copper wire with a rubber head dropper, experiment will be carried out after drying. TEM is used to observe particle size and dispersion [41, 42]. Figures 3(a) and 3(b) show CdS QDs and core-shell structure of CdS-TiO_2 (CdS-based nanocomposite A) with relatively uniform dispersibility in water solution. The particle size of CdS QDs is from 13 to 21 nm shown in Figure 4(a), and CdS-TiO_2 nanocomposite is from 16 to 23 nm shown in Figure 4(b).

Figure 3(c) indicates the TEM image of CdTe water-soluble quantum dots with good dispersibility and stability in aqueous solution. CdTe QDs feature a size of from 3 to 5 nm shown in Figure 4(c). Figure 3(d) is the TEM image of CdTe-TiO_2 (CdTe-based nanocomposite B) with a certain degree of dispersion. There are black particles inside the nanocomposites, and we suppose that CdTe is coated inside the TiO_2 . The size of CdTe-based nanocomposite B is about 80 nm as shown in Figure 4(d).

TEM images of CdSe QDs and CdSe-TiO_2 nanocomposite (CdSe-based nanocomposite B) are shown in Figures 3(e) and 3(f). It could be observed that the two kinds of nanoparticles are uniformly distributed with good monodispersity and spherical or quasi-spherical particle morphology. The size of CdSe nanoparticles mainly concentrated around 18 nm is shown in Figure 4(e), while the size of doped CdSe-TiO_2 nanocomposite mainly concentrated around 10 nm is shown in Figure 4(f). The ultrasonic driving method could effectively reduce the size of nanoparticles and promote the crystal transformation of nanoparticles, so as to obtain CdSe-TiO_2 nanocomposite with smaller particle size and improve the effective absorption of the material by cells.

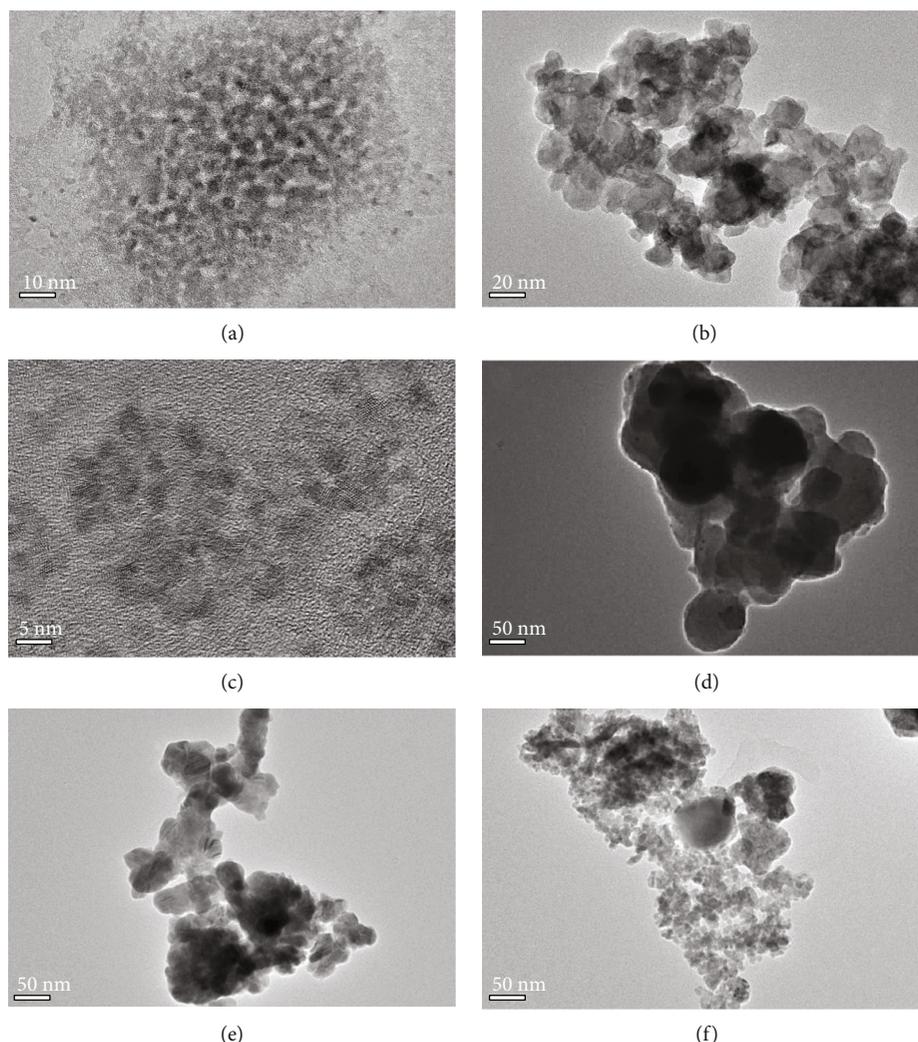


FIGURE 3: (a) The TEM image of CdS QDs; (b) the TEM image of CdS-TiO₂ nanocomposite sample (CdS-based nanocomposite A); (c) the TEM image of CdTe QDs; (d) the TEM image of CdTe/TiO₂ nanocomposite (CdTe-based nanocomposite B); (e) the TEM image of CdS QDs; (f) the TEM image of CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B).

Studies have shown that nanocomposites with particle size less than 100 nm can meet the condition of entering cells, so the above nanocomposites prepared in our laboratory meet the size requirements of a photosensitizer.

3.1.2. X-Ray Diffraction. The crystallite size is calculated by the Scherrer formula [43]:

$$D = \frac{K\lambda}{\beta \cos \theta}, \quad (1)$$

where D is the crystalline size, λ is the X-ray wavelength (0.1541 nm), K is the constant usually taken as 0.89, θ is Bragg's angle $2\theta = 25.3^\circ$ for anatase phase titanium dioxide, and β is the pure full width of the diffraction line at half of the maximum intensity.

XRD patterns of CdS-TiO₂ nanocomposites (CdS-based nanocomposite A) are shown in Figure 5(a). According to the above formula (1), it was calculated that the average crystallite sizes were 15.1 nm for CdS-TiO₂ sample 0.2 nano-

composites and 21.9 nm for CdS-TiO₂ sample 1.0 nanocomposites. The data obtained by TEM analysis of CdS-TiO₂ nanocomposites (Figure 3(b)) were consistent with XRD results. The average crystallite sizes were 17.0, 17.2, and 19.1 nm for CdS-TiO₂ sample pure TiO₂, 0.4, and 0.6, respectively. The XRD diffraction peaks of the synthesized CdS-TiO₂ nanocomposite (CdS-based nanocomposite A) around 2θ of 25.1° , 37.8° , 48° , 53.8° , and 55.1° could be indexed to the characteristic peaks (101), (004), (200), (105), and (211) of anatase TiO₂. Thus, the CdS-TiO₂ nanoparticles had the anatase phase. The crystal diffraction peak of CdS was mainly located between 25° and 35° . The 2θ peaks of 27.2° , 28.2° , 36.8° , 43.6° , and 51.8° indicated that the CdS in the sample was mainly hexagonal.

XRD pattern of CdTe QDs (CdTe-based nanocomposite A) is shown in Figure 5(b); it had two diffraction peaks which are obvious but not spike and conformed to the characteristics of the small size of quantum dots. The XRD diffraction peaks around 2θ of 24.38° , 40.31° , and 47.02° , which could be indexed to the characteristic peaks (111),

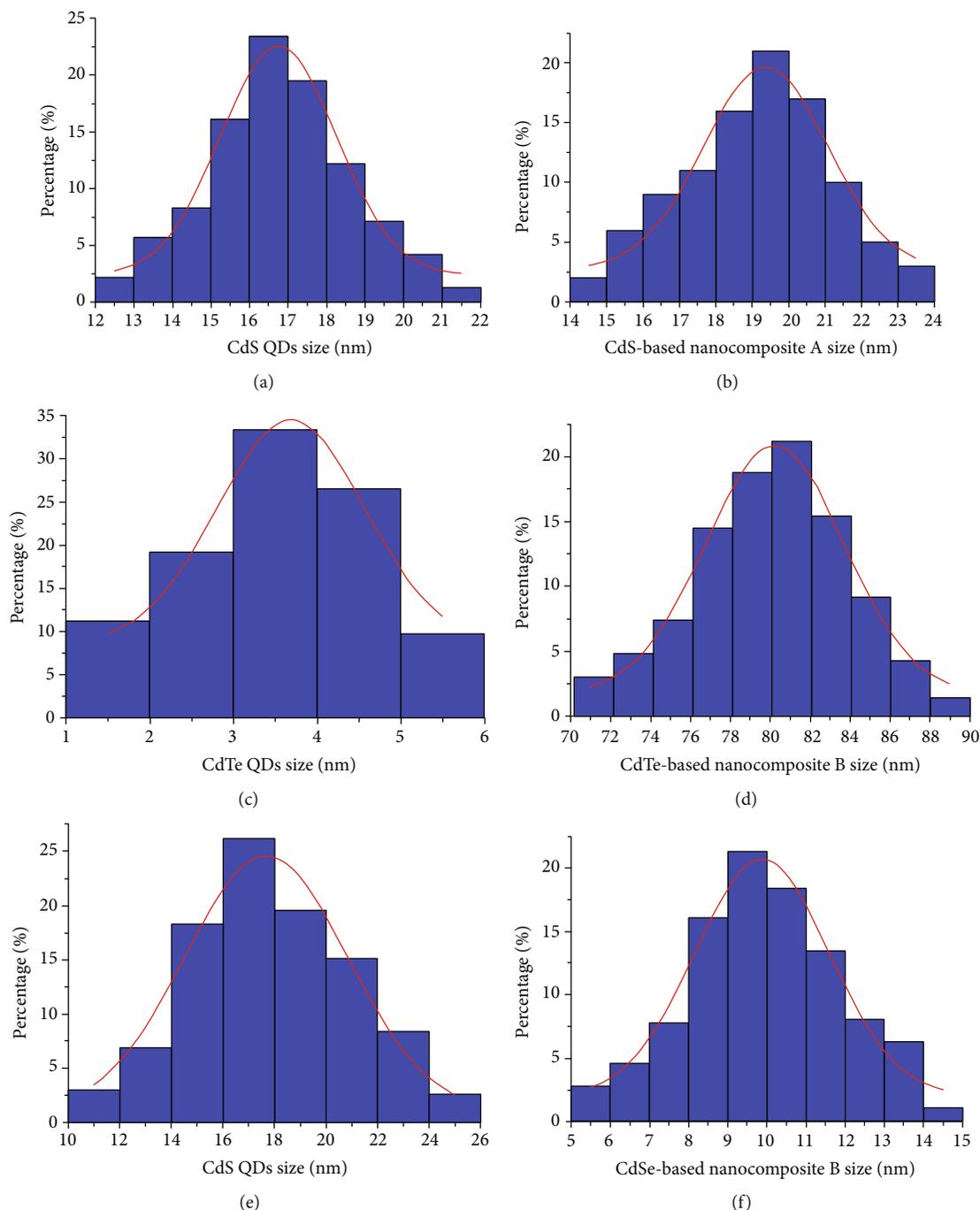


FIGURE 4: (a) Size distribution of CdS QDs; (b) size distribution of CdS-TiO₂ nanocomposite sample (CdS-based nanocomposite A); (c) size distribution of CdTe QDs; (d) size distribution of CdTe/TiO₂ nanocomposite (CdTe-based nanocomposite B); (e) size distribution of CdSe QDs; (f) size distribution of CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B).

(220), and (311) of cubic crystal structure of CdTe, belonged to the sphalerite cubic phase.

As shown in Figure 5(c), the XRD diffraction peaks of the synthesized CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B) around 2θ of 25.27°, 37.8°, 56°, and 62° could be indexed to the characteristic peaks (101), (004), (211), and (204) of anatase TiO₂. It was calculated that the average crystallite sizes are 21.5 nm for pure TiO₂, 17.5 nm for CdSe, and 10.6 nm for CdSe/TiO₂ nanocomposite. The

data obtained by TEM analysis of CdSe and CdSe/TiO₂ nanocomposite (Figure 3(f)) was consistent with it.

3.1.3. UV-Vis Spectroscopy. The nanoparticles were investigated by their UV-Vis absorption spectra.

As shown in Figure 6(a), the absorption thresholds were 387.5, 480, 491, 495, and 507 nm for the TiO₂, CdS-TiO₂ samples 1.0, 0.6, 0.4, and 0.2 (CdS-based nanocomposite A), respectively. It indicated that the absorption spectra of

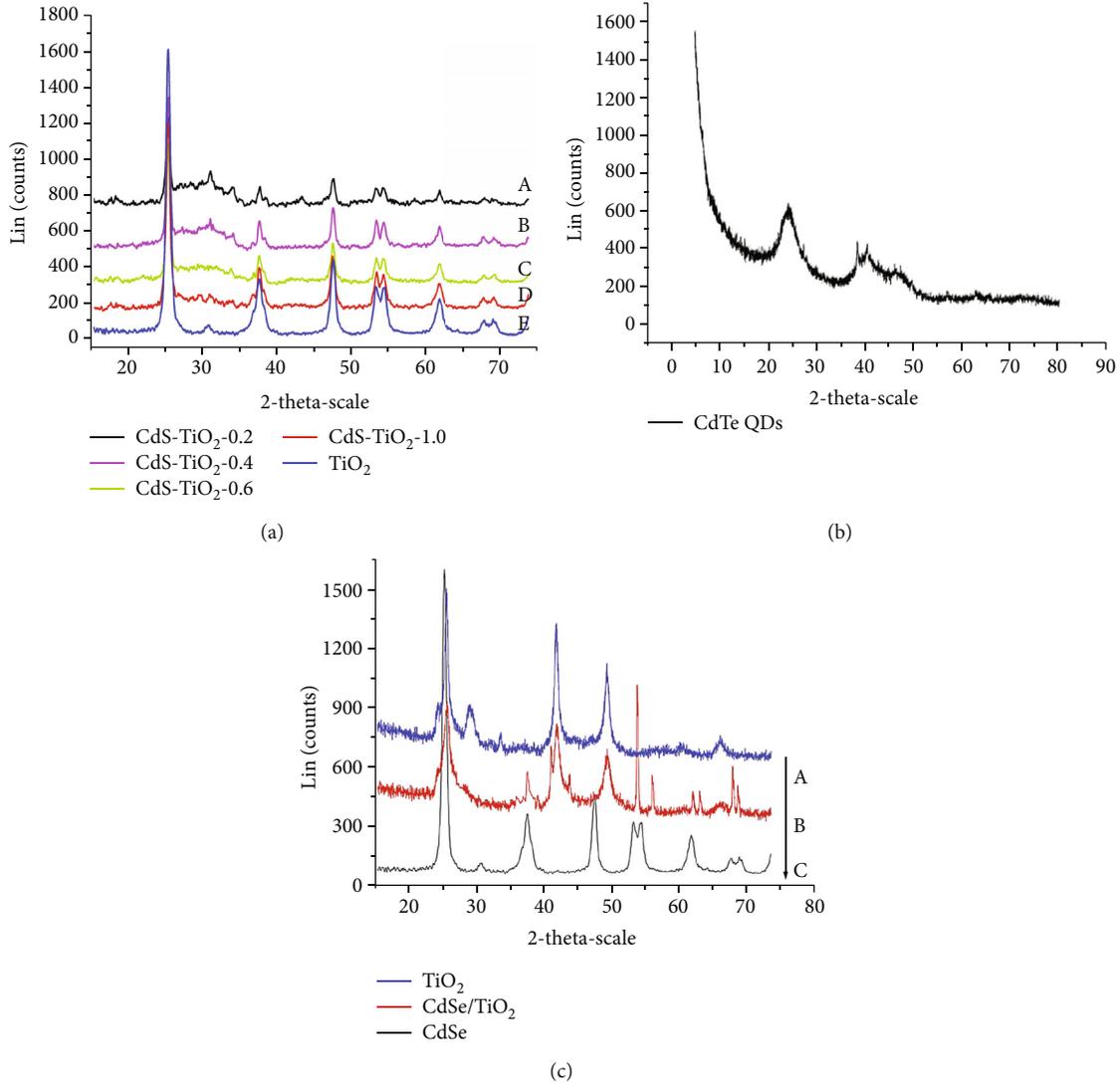


FIGURE 5: (a) The XRD patterns of TiO₂ and CdS-TiO₂ nanocomposites (CdS-based nanocomposite A); (b) the XRD pattern of CdTe quantum dots (CdTe-based nanocomposite A); (c) the XRD patterns of CdSe, TiO₂, and CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B).

CdS-TiO₂ samples were red-shifted to the visible region in varying degrees compared with pure TiO₂. According to the formula [44]:

$$\lambda = \frac{1240}{E_g}, \quad (2)$$

the band gap energy was 3.2, 2.58, 2.52, 2.50, and 2.45 eV for the TiO₂, CdS-TiO₂ samples 1.0, 0.6, 0.4, and 0.2, respectively. The results showed that the presence of CdS would narrow the band gap of nanoparticles, enhance the absorption of visible light, and result in the absorption spectrum of nanoparticles to red shift.

As shown in Figure 6(b), the absorption peak of CdTe was at 460 nm. The absorption spectrum range corresponded to CdTe/TiO₂ nanocomposite (CdTe-based nanocomposite B) was expanded to the visible region. The

results showed that the presence of CdTe QDs in CdTe/TiO₂ enhanced the absorption of visible light and caused the red shift of the absorption spectrum of TiO₂.

The absorption spectra of hydroxyacetic acid-coated CdTe (CdTe-based nanocomposite C) and CdSe (CdSe-based nanocomposite A) QDs are shown in Figure 6(c). The absorption wavelength of the two kinds of QDs was concentrated in the range of from 400 to 500 nm. In this band, the absorption of CdSe QDs was obviously greater than CdTe.

As shown in Figure 6(d), the absorption thresholds were 387.5, 415, 444, 487, and 637 nm for the pure TiO₂, CdSe/TiO₂ samples 0.5, 1, and 2 (CdSe-based nanocomposite B) and CdSe, respectively. It indicated that the absorption spectra of CdSe/TiO₂ samples were red-shifted to the visible region in varying degrees compared with pure TiO₂. According to the above formula (2), the band gap energy was 3.2, 2.99, 2.79, 2.55, and 1.95 eV for the pure TiO₂, CdSe/TiO₂ samples 0.5,

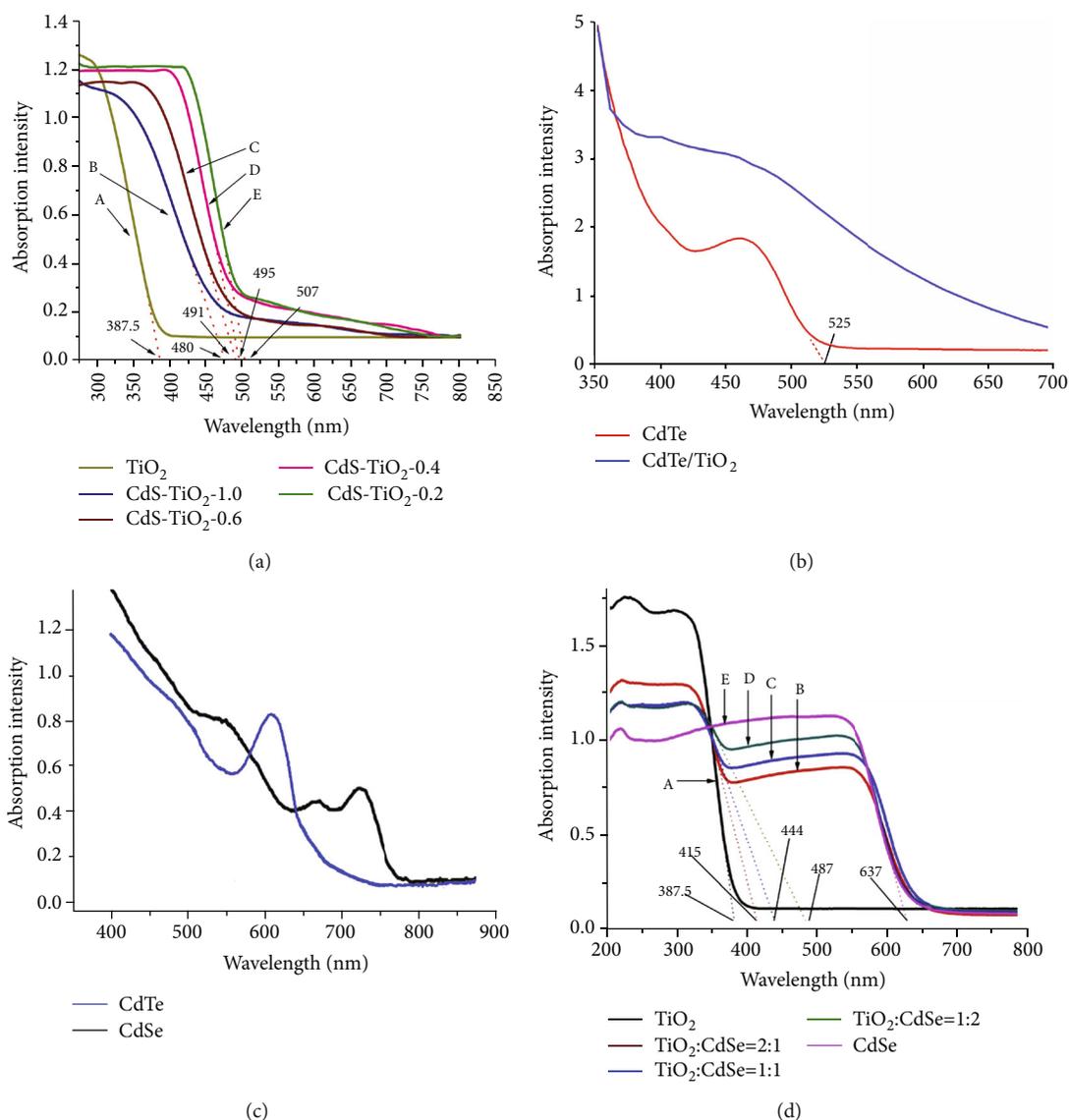


FIGURE 6: (a) The UV-Vis absorption spectra of pure TiO_2 and CdS-based nanocomposite A. (b) The UV-Vis absorption spectra of CdTe and CdTe-based nanocomposite B. (c) The UV-Vis absorption spectra of hydroxyacetic acid-coated CdTe QDs (CdTe-based nanocomposite C) and CdSe QDs (CdSe-based nanocomposite A). (d) The UV-Vis absorption spectra of pure TiO_2 , CdSe, and CdSe-based nanocomposite B.

1, and 2 and CdSe, respectively. The result showed that the presence of CdSe would narrow the band gap of nanoparticles, enhance the absorption of visible light, and result in the absorption spectrum of nanoparticles to red shift.

3.1.4. FS Analysis. Combination of holes and photogenerated electrons on the surface of semiconductor nanoparticles and the efficiency of photocatalytic activity can be demonstrated by FS. The lower the photoluminescence intensity, the lower the recombination rate of photoinduced electron-hole pairs, and the higher the photocatalytic activity of semiconductor photocatalyst. The reactive oxygen species will be produced through the reaction of the hole and the substance in the solution, and it will inactivate the cells. Therefore, the number of oxygen holes on the surface of TiO_2 determines the intensity of its photocatalytic activity [45].

As shown in Figure 7(a), it showed the fluorescence emission spectra of TiO_2 and CdS- TiO_2 nanocomposite, and the hole electron recombination rate on the TiO_2 surface was analyzed. The fluorescence intensity of TiO_2 changed significantly after the doping of CdS QDs. Compared with pure TiO_2 , the fluorescence emission intensity of CdS- TiO_2 samples 0.6, 0.4, and 1.0 decreased significantly, while the fluorescence intensity of sample 0.2 increased. Due to that, the emission fluorescence of semiconductor nanoparticles mainly depended on the combination of photogenerated holes and electrons. Therefore, it could be analyzed that effective electron transfer occurs between CdS QDs and TiO_2 shell of samples 0.6, 0.4, and 1.0, and the recombination rate of holes and electrons was significantly reduced under 410 nm irradiation. The fluorescence intensity of sample 0.6 was the most strongly weakened,

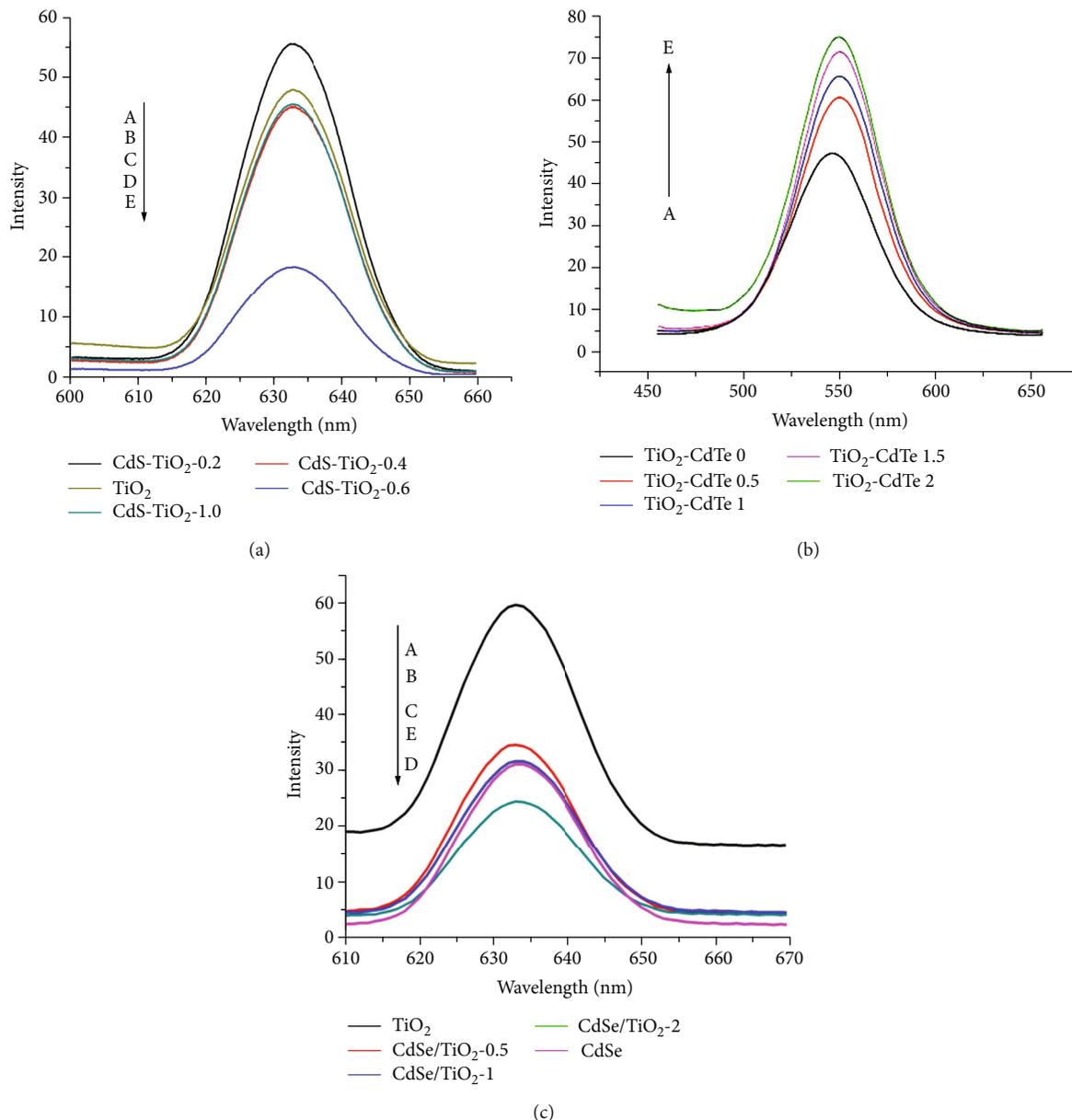


FIGURE 7: (a) The fluorescence emission spectrum of TiO₂ and CdS-based nanocomposite A with the excitation wavelength of 410 nm. (b) The fluorescence spectra of CdTe with the excitation wavelength of 345 nm. CdTe-based nanocomposite A (CdTe: 25 $\mu\text{g}/\text{mL}$; TiO₂ (A–E): 0 $\mu\text{g}/\text{mL}$, 0.5 $\mu\text{g}/\text{mL}$, 1 $\mu\text{g}/\text{mL}$, 1.5 $\mu\text{g}/\text{mL}$, and 2 $\mu\text{g}/\text{mL}$). (c) The fluorescence spectra of pure TiO₂, CdSe, and CdSe-based nanocomposite B with the excitation wavelength of 410 nm.

indicating that the inhibition of hole electron recombination was the most prominent. However, the fluorescence emission of sample 0.2 was enhanced which might be due to the high content of CdS which led to a lot of CdS QD aggregation on the surface of TiO₂ and reduced the number of oxygen holes.

As shown in Figure 7(b), the emission wavelength of CdTe was about from 500 to 600 nm when the excitation wavelength was 345 nm. While the concentration of CdTe was constant, the concentration of TiO₂ increased, and the fluorescence emission peak intensity of CdTe increased gradually. This indicated that CdTe could reduce the recombination of hole and electron on the surface of TiO₂.

As shown in Figure 7(c), the fluorescence intensity of CdSe/TiO₂ nanocomposite was significantly lower than that of pure TiO₂, and the fluorescence intensity of sample CdSe/TiO₂-2 was the weakest. The doping of CdSe QDs changed the efficiency of capture, migration, and transformation of the nanocomposite, reduced the recombination rate of electrons and holes, and improved the separation efficiency of photogenerated carriers, and photocatalytic activity.

3.2. Cytotoxicity of CdX (X=S, Te, Se)-Based Nanocomposites on HL60 Cells. The low dark toxicity is one of the important characteristics of a photosensitizer. The cell activity was detected by the CCK-8 method. The

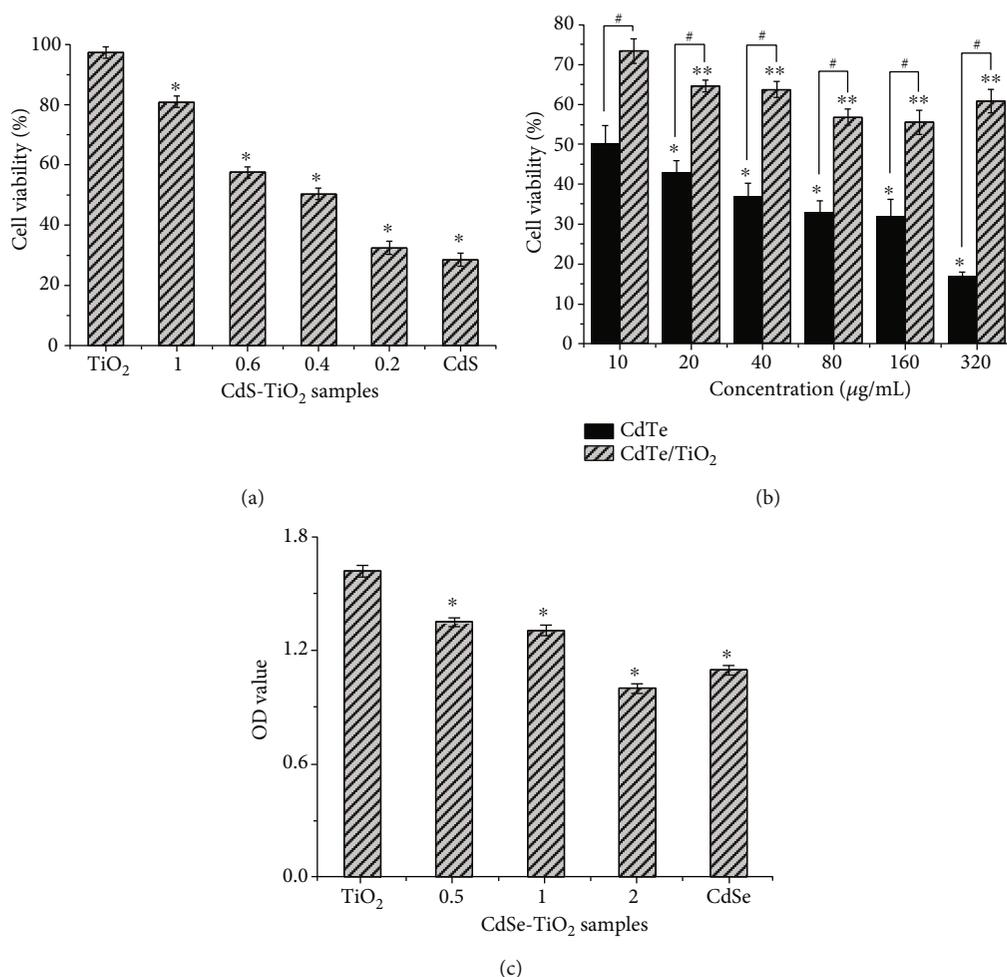


FIGURE 8: (a) Viability of cells treated with CdS-based nanocomposite A in different shell thickness at the concentration of 20 $\mu\text{g/mL}$. The without radiation group was kept in dark condition for 24 hours. Each data point represents mean \pm SD ($n = 3$). t -test was performed on the data of the TiO₂ group and the different sample group. * $P < 0.05$ indicated that there was a difference compared with groups of TiO₂ without irradiation. (b) The effects of relative survival of HL60 cells coincubated with CdTe and CdTe-based nanocomposite B for 24 hours in the dark. Each data point represents mean \pm SD ($n = 3$). t -test was performed on the data of the CdTe group and the CdTe/TiO₂ group. * $P < 0.05$ indicated that there was a difference compared with groups of CdTe in different concentrations, ** $P < 0.05$ indicated that there was a difference compared with groups of CdTe/TiO₂ in different concentrations, and # $P < 0.001$ indicated that there was a difference that CdTe compared with CdTe/TiO₂ in the same concentration. (c) Viability of HL60 cells treated with CdSe, TiO₂, and CdSe-based nanocomposite B with different doping ratios at the concentration of 4 $\mu\text{g/mL}$. The without irradiation group was kept in dark condition for 24 hours. Each data point represents mean \pm SD ($n = 3$). t -test was performed on the data of the TiO₂ group and the different sample group. * $P < 0.05$ indicated that there was a difference compared with groups of TiO₂ without irradiation. All the data shown in the figure are representative.

OD values of the treatment group (treatment with nanocomposites for one day) and the control group (without treatment with nanoparticles) were measured by the Model 680 Microplate Reader. The cell viability was calculated as follows:

$$R(\%) = \left(\frac{\text{OD}_{\text{treat}}}{\text{OD}_{\text{control}}} \right) \cdot 100\%, \quad (3)$$

where the OD_{treat} and $\text{OD}_{\text{control}}$ are the mean absorption values at 490 nm for the treatment and control group, respectively.

Studies have shown that the injection concentration of nanoparticles for mouse experiments is from 0 to 350 $\mu\text{g/mL}$ [46–48]. Therefore, we choose the experiment concentration from 0.2 to 320 $\mu\text{g/mL}$ as a reference and that conforms to toxicological value.

The effect of CdS-TiO₂ nanocomposites (CdS-based nanocomposite A) with different thickness of TiO₂ shell on HL60 cell activity in dark is shown in Figure 8(a). As can be seen from the figure, cell survival rates of treatment with CdS and CdS-TiO₂ groups 0.2, 0.4, 0.6, and 1.0 and TiO₂ were 28.5%, 32.4%, 50.4%, 57%, 80%, and 97%, respectively, under the dark condition. The lowest survival rate of cells treated with CdS was 28.5% confirmed that the high biotoxicity of CdS QDs. Compared with the group of treatment

with CdS QDs, the cell survival rate of the group of treatment with CdS-TiO₂ increased with the increase of the thickness of the TiO₂ shell. In the dark condition group corresponding to CdS-TiO₂ sample 1.0, the cell survival rate reached the highest 80%. These results indicated that it could significantly protect the cells under dark condition in the presence of TiO₂ shell and reduced the dark toxicity of CdS QDs. For CdS QDs, the release of Cd²⁺ was the main toxic mechanism [49]. The experimental results showed that the existence of TiO₂ shell could effectively protect the semiconductor QDs in solution and prevent corrosion [50]. In the dark toxicity test of this experiment, the existence of TiO₂ shell could effectively reduce the biological toxicity of CdS QDs by blocking their contact with the solution.

HL60 cells were treated with different concentrations of CdTe QDs and CdTe/TiO₂ nanocomposite (CdTe-based nanocomposite B); the relation between concentration and relative survival rate of cells is shown in Figure 8(b). When the concentration of CdTe/TiO₂ nanocomposite was lower than 20 µg/mL, the cytotoxicity was relatively lower. When the concentration was higher than 160 µg/mL, the cytotoxicity was relatively weakened. When the concentration was 80 µg/mL, the cytotoxicity to HL60 cells was the highest. It indicated that the cell uptake of nanoparticles had reached saturation in the concentration of 80 µg/mL. The dark toxicity of CdTe/TiO₂ was lower than that of CdTe QDs.

The effect of CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B) with different doping ratios of TiO₂ and CdSe on the activity of HL60 cells without irradiation is shown in Figure 8(c). When the concentration of nanoparticles was 4 µg/mL, the higher the doping ratio of CdSe and TiO₂, the lower the cell activity, and the dark toxicity of CdSe/TiO₂ with a doping ratio of two was the highest. The dark toxicity of CdSe/TiO₂ with a doping ratio of 0.5 and one was lower; it indicated that CdSe/TiO₂ nanocomposite had good biocompatibility.

3.3. Photodynamic Therapy of CdX (X=S, Te, Se)-Based Nanocomposites on HL60 Cells. The HL60 cells in the logarithmic growth phase were inoculated into two 96-well plates marked with A and B. Plate A was incubated for 12 hours without irradiation, next exposed to visible light for one hour, then incubated for 12 hours without irradiation. Plate B was incubated for 24 hours in the incubator without irradiation. Photodynamic inactivation efficiency (indicated by PDT_{efficiency}) was expressed by cell viability (OD value), which was calculated as follows:

$$\text{PDT}_{\text{efficiency}} = 1 - \frac{\text{OD}_{\text{Ir}}}{\text{OD}_{\text{Non-Ir}}} \cdot 100\%, \quad (4)$$

where the OD_{Ir} and OD_{Non-Ir} were the mean absorption values at 490 nm for the irradiation and no irradiation, respectively.

In Figure 9(a), compared with the without radiation group, the cell activity of the CdS-TiO₂ treatment group decreased significantly with radiation. The PDT inactivation efficiency of CdS/TiO₂ nanocomposite sample 0.6 reached the highest 95%. Even for the sample 1.0 with the lowest CdS con-

centration, the PDT inactivation efficiency reached 60%. These results indicated that the inactivation effect of CdS-TiO₂ nanocomposite (CdS-based nanocomposite A) on HL60 cells under irradiation was significantly enhanced due to the presence of CdS. Appropriate CdS QDs and TiO₂ coating can reduce dark toxicity and increase PDT efficiency.

Figure 9(b) shows the OD values of cells treated with different concentrations of TiO₂-CdTe (CdTe-based nanocomposite A) and cultured under irradiation and without irradiation, respectively. According to the above formula (4), the figure showed the PDT inactivation efficiency change tendency of TiO₂-CdTe mixed system with CdTe concentration of 0.25, 0.5, 0.75, and 1 µg/mL was 59.70%, 53.75%, 59.02%, and 71.54%, respectively. It indicated that the higher the CdTe concentration, the higher PDT efficiency and the lower dark toxicity in the low concentration range.

As shown in Figure 9(c), the relative survival rate of HL60 cells in the irradiation group treated with different concentrations of CdTe/TiO₂ nanocomposite (CdTe-based nanocomposite B) was significantly lower than that in the nonirradiation group. With the increasing concentration of CdTe/TiO₂ nanocomposite, the relative survival rate of HL60 cells decreased and showed a slowing trend after PDT. It indicated that PDT improved the inactivation efficiency of CdTe/TiO₂ nanocomposite on HL60 cells. In order to further explore the influence on PDT efficiency between the concentration of nanocomposite and irradiation time, the cells were irradiated with light at the dose of 18 J/cm² under the wavelength of 410 nm for 20, 40, and 60 minutes, respectively (Figure 9(d)). With the increase of irradiation time, the efficiency of PDT increased. Compared with the TiO₂ nanoparticle group, the PDT efficiency of the CdTe/TiO₂ nanocomposite group was significantly improved and showed an increasing trend with the increase of concentration. The experimental result showed that when the concentration of CdTe/TiO₂ nanocomposite was 320 µg/mL, the PDT efficiency reached the highest 87.7%. The photocatalytic activity of TiO₂ was enhanced by doping CdTe QDs under irradiation, which enhanced the PDT efficiency. However, when the concentration of CdTe/TiO₂ was higher than 80 µg/mL, the PDT efficiency did not increase significantly. The concentration of cell exposure has reached absorption saturation.

As shown in Figure 9(e), the PDT efficiency of the hydroxyacetic acid-coated CdTe QDs (CdTe-based nanocomposite C) and CdSe QDs (CdSe-based nanocomposite A) concentration of 1.0, 1.5, 2.0, and 2.5 µmol/L, respectively, under the optimal light dosage parameters (CdTe of 15 J/cm² and CdSe of 18 J/cm²) was shown. The inactivation efficiency of CdSe QDs was obviously higher than that of CdTe QDs. The PDT efficiency change tendency is shown in Figure 9(f), when the optimal concentration of both CdTe and CdSe QDs was 2.0 µmol/L, and the light dosages were 12, 15, 18, and 21 J/cm², respectively. Under the optimal light dosage parameters of 15 J/cm², the PDT inactivation efficiency of CdTe QDs with the concentration of 2.0 µmol/L on HL60 cells was 62.7%. Under the optimal light dosage of 18 J/cm², the PDT inactivation efficiency of HL60 cells with concentration of 2.0 µmol/L CdSe QDs was 83.0%.

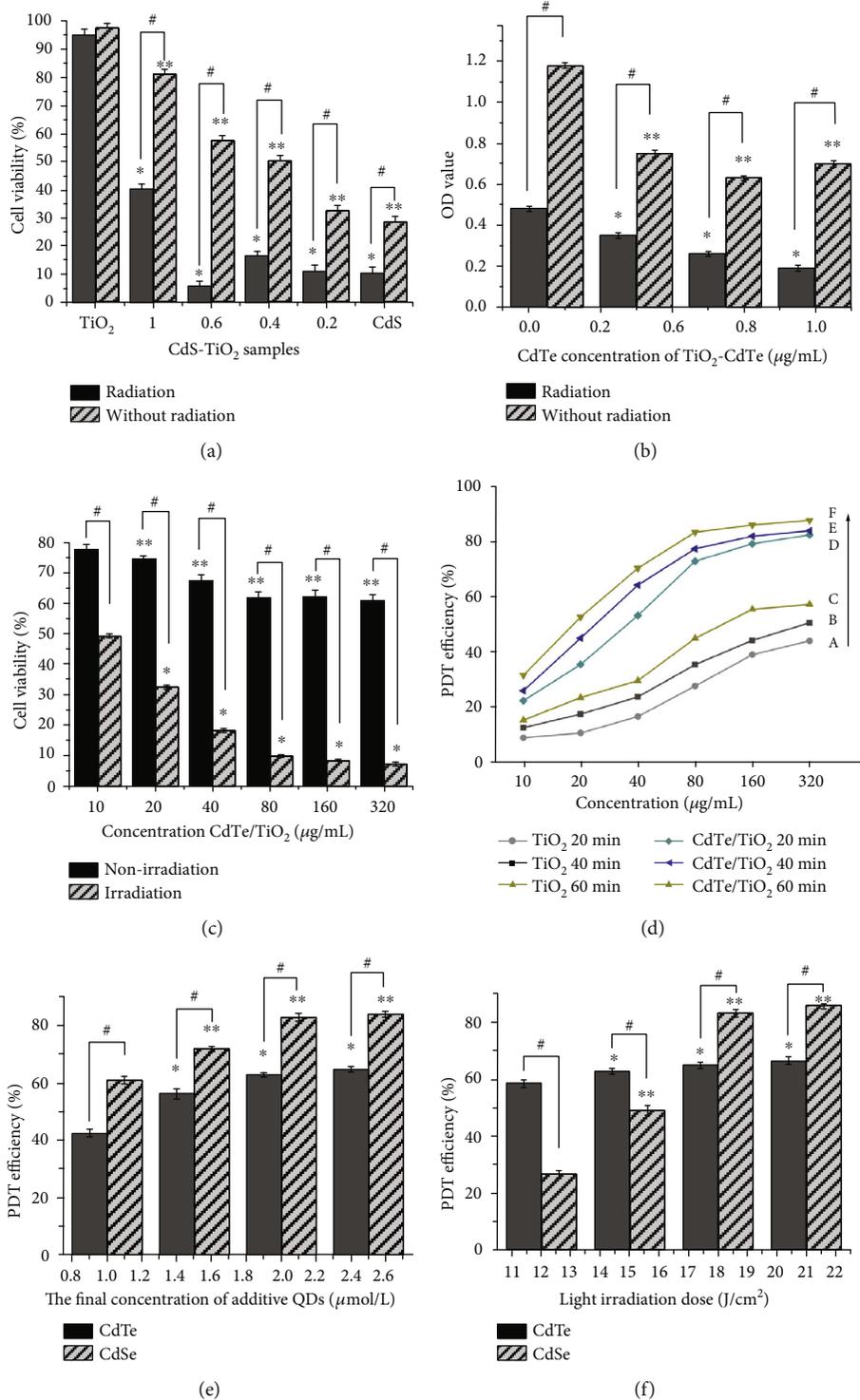


FIGURE 9: Continued.

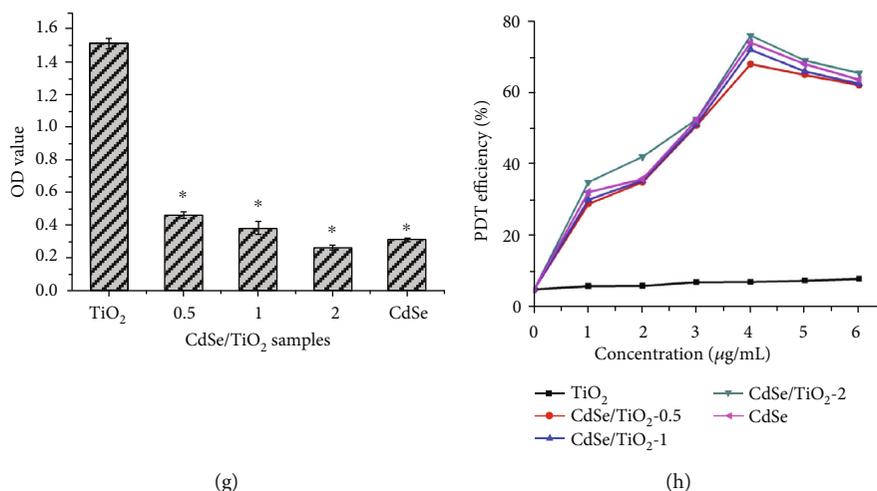


FIGURE 9: (a) Viability of cells treated with CdS-based nanocomposite A in different shell thickness at the concentration of 20 $\mu\text{g/mL}$. The group of irradiation was under 410 nm light exposure for 1 hour with a power density of 18 J/cm^2 . The without irradiation group was kept in dark condition for 24 hours. *t*-test was performed on the data of the irradiation group and the nonirradiation group. Each data point represents mean \pm SD ($n = 3$). * $P < 0.001$ indicated that there was a difference compared with groups of TiO₂ with irradiation, ** $P < 0.001$ indicated that there was a difference compared with groups of TiO₂ without irradiation, and # $P < 0.05$ compared with the two groups. (b) The influence of different CdTe concentrations of CdTe-based nanocomposite A to OD values of HL60 cells. Each data point represents mean \pm SD ($n = 3$). *t*-test was performed on the data of the irradiation group and the nonirradiation group. * $P < 0.001$ indicated that there was a difference compared with groups of irradiation in the different concentration, ** $P < 0.001$ indicated that there was a difference compared with groups of without irradiation in the different concentration, and # $P < 0.05$ indicated that there was a difference compared with two groups. (c) The effect of irradiation on the relative survival of HL60 cells with CdTe-based nanocomposite B. Each data point represents mean \pm SD ($n = 3$). *t*-test was performed on the data of the irradiation group and the nonirradiation group. * $P < 0.001$ indicated that there was a difference compared with groups of irradiation in the different concentration, ** $P < 0.001$ indicated that there was a difference compared with groups of without irradiation in the different concentration, and # $P < 0.05$ indicated that there was a difference compared with two groups. (d) The PDT efficiency of TiO₂ and CdTe-based nanocomposite B on HL60 with different irradiation times. Each data point represents mean \pm SD ($n = 3$). * $P < 0.05$ compared with groups of TiO₂. (e) The inactivation efficiency of hydroxyacetic acid-coated CdTe QDs (CdTe-based nanocomposite C) and CdSe QDs (CdSe-based nanocomposite A) under the best light dosage (CdTe: 15 J/cm^2 , CdSe: 18 J/cm^2). Each data point represents mean \pm SD ($n = 3$). *t*-test was performed on the data of the CdTe group and the CdSe group. * $P < 0.001$ indicated that there was a difference compared with groups of CdTe in different concentrations, ** $P < 0.001$ indicated that there was a difference compared with groups of CdSe in different concentrations, and # $P < 0.05$ indicated that there was a difference that CdTe compared with CdSe in the same concentration. (f) The inactivation efficiency of CdTe-based nanocomposite C and CdSe-based nanocomposite A under the best QD concentration (2.0 $\mu\text{mol/L}$). Each data point represents mean \pm SD ($n = 3$). *t*-test was performed on the data of the CdTe group and the CdSe group. * $P < 0.001$ indicated that there was a difference compared with groups of CdTe in different light irradiation doses, ** $P < 0.001$ indicated that there was a difference compared with groups of CdSe in different light irradiation doses, and # $P < 0.05$ indicated that there was a difference that CdTe compared with CdSe in the same light irradiation dose. (g) The viability of cells treated with TiO₂, CdSe, and CdSe-based nanocomposite B at the concentration of 4 $\mu\text{g/mL}$. The irradiation group was irradiated for 1 hour. Each data point represents mean \pm SD ($n = 3$). *t*-test was performed on the data of the CdSe/TiO₂ group and the TiO₂ group. * $P < 0.05$ compared with groups of TiO₂; (h) the PDT efficiency of TiO₂, CdSe, and CdSe-based nanocomposite B on HL60 cells at various concentrations. Each data point represents mean \pm SD ($n = 3$). * $P < 0.05$ compared with groups of TiO₂. All the data shown in the figure are representative.

The inactivation efficiency of CdSe QDs was higher than that of CdTe QDs at different concentrations of QDs under optimal light dosage parameters. The inactivation efficiency of CdSe was significantly higher than that of CdTe under the optimal concentration of QDs and different light dosages. In conclusion, the PDT efficiency of CdSe QDs was superior to that of CdTe QDs.

CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B) in different ratios of 0.5, 1, and 2 with final concentration of 1, 2, 3, 4, 5, and 6 $\mu\text{g/mL}$, CdSe and TiO₂ nanoparticle solutions were coincubated with HL60 cells for PDT experiment. The OD value of the irradiation group is shown in Figure 9(g). Compared with the nonirradiation group (Figure 8(c)), the OD value of cells treated with CdSe/TiO₂

nanocomposite under irradiation was significantly reduced. Besides, the OD value of cells treated with CdSe/TiO₂ nanocomposite was lower than that of pure TiO₂. The PDT efficiency of various nanoparticles at different concentrations is shown in Figure 9(h). The inactivation efficiency of HL60 cells induced by CdSe/TiO₂ was significantly higher than that of pure TiO₂. When the ratio of CdSe to TiO₂ equaled two and the concentration was 4 $\mu\text{g/mL}$, the PDT efficiency reached the highest 76%. The experimental result showed that the inactivation efficiency of CdSe/TiO₂ nanocomposite on HL60 cells was significantly enhanced under irradiation.

3.4. Ultrastructural Morphology of the Treated Cells. The samples of HL60 cells were collected by critical point drying

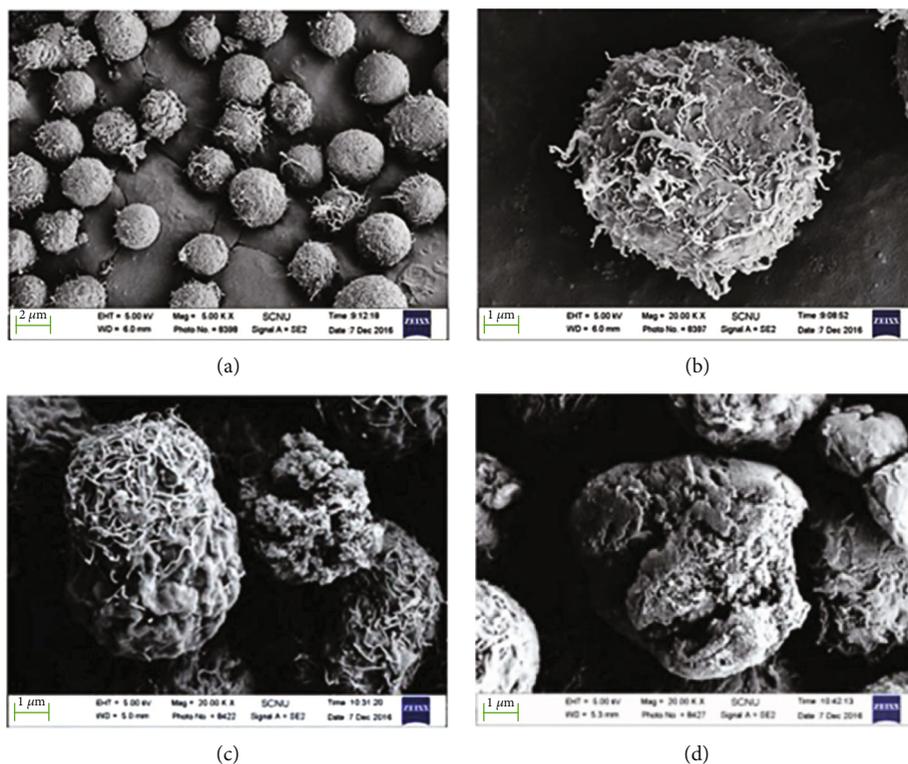


FIGURE 10: (a) The ultrastructural morphology of multiple normal HL60 cells; (b) the ultrastructural morphology of single normal HL60 cell; (c) the ultrastructural morphology of the HL60 cells after CdSe treatment; (d) the ultrastructural morphology of the HL60 cells after CdSe-based nanocomposite B treatment.

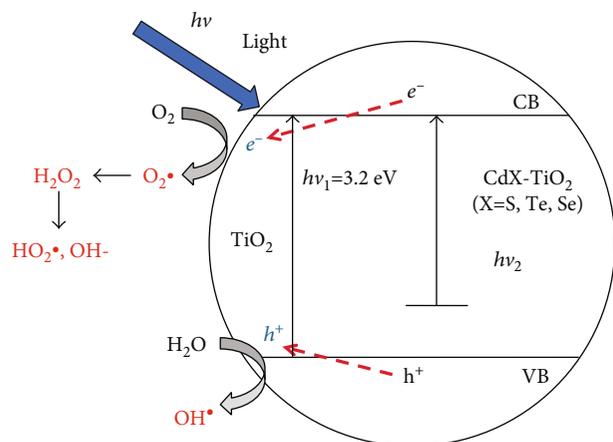


FIGURE 11: The possible mechanism of excitation and charge transfer process between CdX (X=S, Te, Se) and TiO₂ in the CdX (X=S, Te, Se)-based nanocomposites under irradiation.

method, which were normal HL60 cells in logarithmic growth phase and HL60 cells after PDT treatment, respectively. Firstly, the cells were fixed in 2.5% glutaraldehyde solution. Then, phosphate buffer solution (PBS) was used to rinse, and the upper layer of waste solution was discarded after centrifugation. It was dehydrated and fixed by gradient ethanol (50%, 70%, 80%, 90% once, and 100% twice, 10 minutes each time). Finally, it was fixed with ethanol isodiester twice, each time for three minutes. The cell samples

obtained by the above treatment were dried by a critical point dryer and then plated by a metal ion sputtering device and observed under a Zeiss Ultra-55 scanning electron microscope.

As shown in Figures 10(a) and 10(b), HL60 cells were not treated with nanoparticles. The diameter of the cells was about 10 μm , and the cells generally featured a regular spherical or spheroidal shape. Normal HL60 cells had the smooth surface, no flagellum, and plump cell type. The cell surface was not concave and convex, and the edge was relatively neat. As shown in Figures 10(c) and 10(d), HL60 cells were treated with CdSe/TiO₂ nanocomposite (CdSe-based nanocomposite B). The shape of the cells was changed, with obvious concavity, convexity, and cracks on the surface, showing a flake-like arrangement like fish scales. The cell membrane was ruptured, and small holes appeared on the surface of the membrane, proving that HL60 cells were in the stage of apoptosis.

3.5. Mechanism of Photocatalysis on CdX (X=S, Te, Se)-Based Nanocomposites. As shown in Figure 11, the valence band electrons are excited, and they migrate to the conduction band after the nanoparticles absorb the light energy greater than the band gap energy under the irradiation with appropriate wavelength, thus producing carriers. In this process, part of the electron-hole pairs is effectively separated, migrating to the surface of the particles and interacting with molecules such as water and oxygen in the cellular environment. Water molecules adsorbed on the surface of the

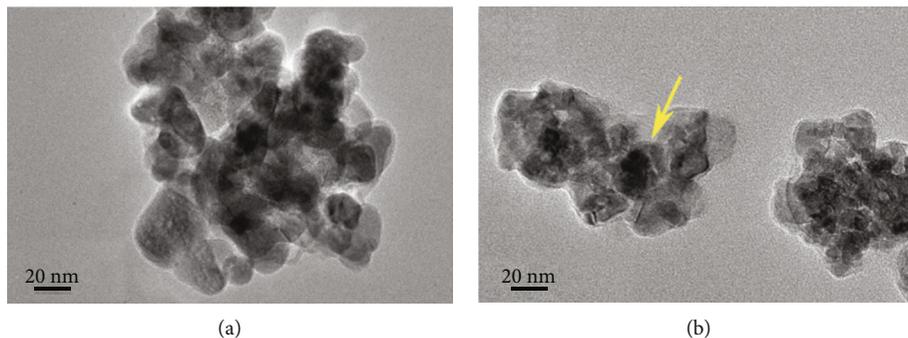


FIGURE 12: (a) The TEM images of CdSe-TiO₂; (b) the TEM images of CdSe-based nanocomposite C.

nanoparticles are oxidized by photogenerated holes, and oxygen molecules in the water are reduced by photogenerated electrons to generate active free radicals such as hydroxyl free radicals ($\cdot\text{OH}$) and superoxide ions ($\text{O}_2\cdot^-$).

It is well known that the UV-Vis absorption spectrum of TiO₂ has relatively high absorption in the ultraviolet region but weak absorption in visible light region, which is not conducive to being used as a visible light PDT drug. The absorption threshold of the UV-Vis absorption spectrum results can be calculated according to formula (2); the band gap energy of CdS, CdTe, and CdSe is 2.4, 2.36, and 1.95 eV, respectively. Since the CdX (X = S, Te, Se) band gap is narrow and the position of valence band is positive, the TiO₂ band gap energy (3.2 eV) is high and the position of conduction band is low. As X is changed from S, Te to Se, the more positive valence band position of CdSe QDs, the band gap of CdSe QDs is the narrowest. With the combination of CdX and TiO₂, the photogenerated electrons of CdX can be rapidly injected into the lower conduction band of TiO₂ due to the transfer of photogenerated carriers. The CdX (X = S, Te, Se)-TiO₂ band gap becomes narrow and photogenerated carriers are easier to separate. The CdSe-TiO₂ band gap is the narrowest, with the lowest recombination rate and highest separation efficiency of carriers.

The efficiency of carrier trapping, migration, and conversion of the nanocomposites after doping with CdX (X = S, Te, Se) QDs is changed by absorbing energy greater than the band gap energy. The lower the recombination rate of electrons and holes, the more photogenerated electrons and holes participate in redox reaction, which further enhances the photocatalytic activity of TiO₂ shell and the yield of reactive oxygen species and increases the PDT inactivation efficiency finally.

3.6. Targeting Ability of CdX (X = S, Se)-Based Nanocomposites Modified with Folic Acid. Studies show that folic acid is one of the most promising targeting ligands. Folate receptor has high sensitivity and specificity for folic acid and its analogs. Folate receptor mainly includes three subtypes: FR- α , FR- β , and FR- γ . FR- α and FR- β were overexpressed in cancer tissues but were rarely expressed in normal blood cells [34]. Folic acid modification with nanocomposite contributes to the targeted uptake of photosensitizers by cancer cells.

3.6.1. Characterization of Folic Acid Molecule

(1) *TEM Studies of FA-CdSe-TiO₂ (CdSe-Based Nanocomposite C).* Figure 12 is the TEM image of CdSe-TiO₂ nanocomposite prepared by the hydrolytic precipitation method and FA-CdSe-TiO₂ nanocomposite (CdSe-based nanocomposite C). CdSe-TiO₂ nanocomposite was spherical or spheroid, with certain dispersion, and size was about 30 nm shown in Figure 12(a). The dispersion of FA-CdSe-TiO₂ nanocomposite did not change significantly; a layer coated on the surface of nanoparticles could be observed (Figure 12(b) marked with an arrow). It proved that CdSe-TiO₂ nanocomposite modified with FA successfully, and nanocomposite size was about 40 nm.

(2) *Fourier Transform Infrared Spectra (FT-IR) of FA-CdS-TiO₂ (CdS-Based Nanocomposite B) and FA-CdSe-TiO₂ (CdSe-Based Nanocomposite C).* As shown in Figure 13(a), folic acid (curve A), CdS-TiO₂ (curve B), and FA-CdS-TiO₂ (curve C) were as shown by FT-IR. The absorption peaks of 3314, 3410, and 3535 cm⁻¹ in curve A were caused by the vibration of hydroxyl (-OH) and amino (NH-) in folic acid molecules. The absorption peaks of 726 cm⁻¹ in curve B corresponded to the vibration of O-Ti-O bond, and the absorption peaks located at 1605 and 3352 cm⁻¹ were caused by the vibration of hydroxyl (-OH) on the surface of anatase TiO₂. FT-IR (curve C) of FA-CdS-TiO₂ nanocomposite, the hydroxyl peaks at 1605 and 3352 cm⁻¹ disappeared or weakened, the absorption peaks corresponding to C-O and C=O bond vibration appeared at 1101 and 1638 cm⁻¹, and the characteristic peaks corresponding to the amino group (NH-) in folic acid molecules emerged at 3459 cm⁻¹. This demonstrated that the carboxyl group of folic acid was combined with the surface hydroxyl group of CdS-TiO₂ in the form of esterification.

As shown in Figure 13(b), FA-CdSe-TiO₂ (curve A), folic acid (curve B), and CdSe-TiO₂ (curve C) were shown by FT-IR. Absorption peaks of 3320 and 3540 cm⁻¹ in curve B were caused by the vibration of hydroxyl (-OH) and amino (NH-) in folic acid molecules. The absorption peaks locating at 807 cm⁻¹ in curve C corresponded to the vibration of O-Ti-O bond, and the peaks at 1645 and 3313 cm⁻¹ were caused by the vibration of hydroxyl (-OH) on the surface of TiO₂. In curve A, the absorption peaks near 1645 and 3313 cm⁻¹

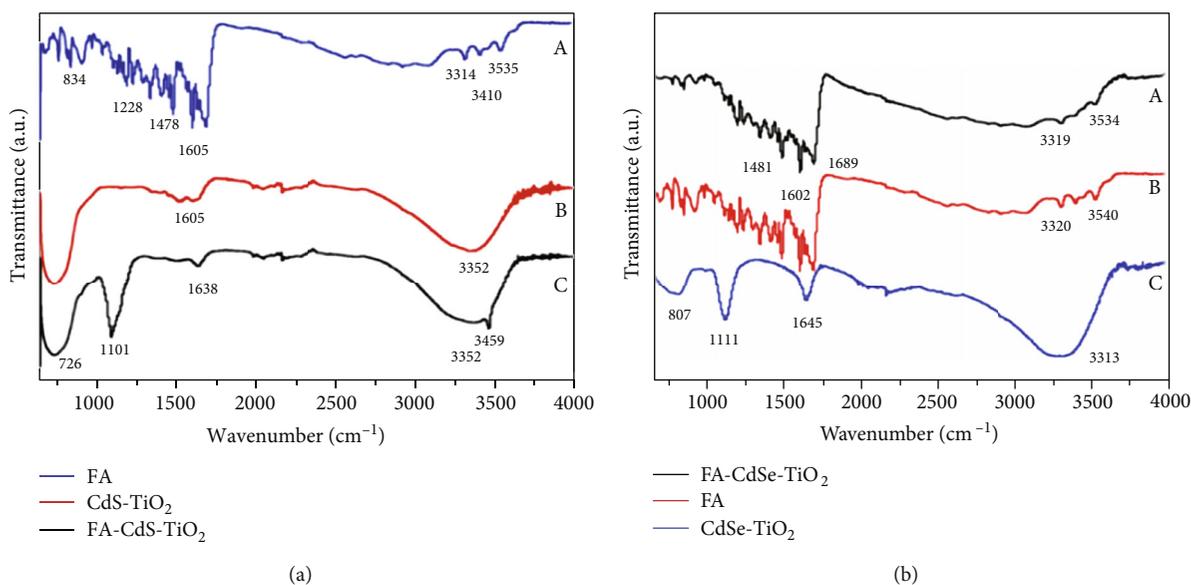


FIGURE 13: (a) The FT-IR spectra of folic acid (curve A), CdS-TiO₂ (curve B), and FA-CdS-TiO₂ (curve C) (CdS-based nanocomposite B); (b) the FT-IR spectra of FA-CdSe-TiO₂ (curve A) (CdSe-based nanocomposite C), folic acid (curve B), and CdSe-TiO₂ (Curve C).

disappeared or weakened, while the absorption peaks were similar to carboxylate (-COOM) appeared at 1481 cm⁻¹, the characteristic peak of the stretching vibration of the pteridine ring of folic acid appeared at 1689 cm⁻¹, and the absorption peaks corresponding to the vibration of the amino group (NH-) in folic acid molecules appeared around 3319 and 3534 cm⁻¹. This demonstrated that the carboxyl group of folic acid was combined with the surface hydroxyl group of CdSe-TiO₂ in the form of esterification.

(3) *FS Analysis of FA-CdS-TiO₂ (CdS-Based Nanocomposite B)*. To detect the fluorescence intensity of CdS-TiO₂ and FA-CdS-TiO₂ nanocomposite in solution and intracellular uptake of nanocomposites after 12 hours of incubation, respectively, it was found that the difference of fluorescence intensity between nanocomposite solution and after cellular uptake is smaller with FA modification. FA modification improved the cellular uptake efficiency of nanoparticles (Figure 14). FA-CdS-TiO₂ nanocomposite had good biocompatibility.

(4) *UV-Vis Spectroscopy of FA-CdSe-TiO₂ (CdSe-Based Nanocomposite C)*. As shown in Figure 15, absorption spectra of CdSe-TiO₂ and FA-CdSe-TiO₂ nanocomposites (CdSe-based nanocomposite C) red-shifted to the visible light region in different degrees compared with that of pure TiO₂, and the absorption intensity of FA-CdSe-TiO₂ slightly increased compared with CdSe-TiO₂ at 410 nm. It indicated that the doping of CdSe effectively enhanced the visible light absorption of TiO₂. After the modification of FA in proper proportion, it could still keep the effective visible light absorption of nanocomposite.

3.6.2. *Cytotoxicity of FA-CdS-TiO₂ (CdS-Based Nanocomposite B) and FA-CdSe-TiO₂ (CdSe-Based Nanocomposite C) on HL60 Cells*. As shown in Figure 16(a), the cell relative survival

rate of the cells treated with FA-CdS-TiO₂ nanocomposite was higher than 90% when the drug concentration was lower than 20 μg/mL. When the drug concentration was 20, 30, and 40 μg/mL, the survival rates of cells were 85%, 75%, and 70%, respectively, without irradiation. It indicated that the toxicity of FA-CdS-TiO₂ nanocomposite at appropriate concentration to HL60 cells was lower. Compared with the CdS-TiO₂ group, the dark toxicity of FA-CdS-TiO₂ nanocomposite decreased with the same concentration, and the decreasing trend increased with the increase of drug concentration. It indicated that FA-CdS-TiO₂ nanocomposite after FA modification had good biocompatibility in low concentration range.

The effect of CdSe-TiO₂ and FA-CdSe-TiO₂ nanocomposite modified with different FA ratios on the activity of HL60 cells without irradiation is shown in Figure 16(b). It could be seen that the relative survival rate of cells was maintained at above 80% treated with FA-CdSe-TiO₂ nanocomposite with the FA modified ratios of 0.2, 0.4, 1.0, and 2.0, respectively, without irradiation, when the mass concentration of the nanocomposite was less than 20 μg/mL. The relative survival rate of cells decreased when the drug concentration gradually increased. In addition, when the proportion of FA was less than 100%, the relative survival rate of cells decreased with the increase of the proportion of FA. It indicated that the targeting effect of folic acid molecules on HL60 cells improved the uptake efficiency of FA-CdSe-TiO₂ nanocomposite. When the proportion of FA was increased to 200%, cell viability increased compared with the nanocomposites with a lower proportion of FA without irradiation. This might be due to the large proportion of FA increasing, which reduced the proportion of CdSe in nanoparticles and thus reduced the toxicity of the drug itself.

As discussed above, it indicated that the modification of FA would not cause new cytotoxicity and might also reduce the toxicity of nanocomposites to a certain extent.

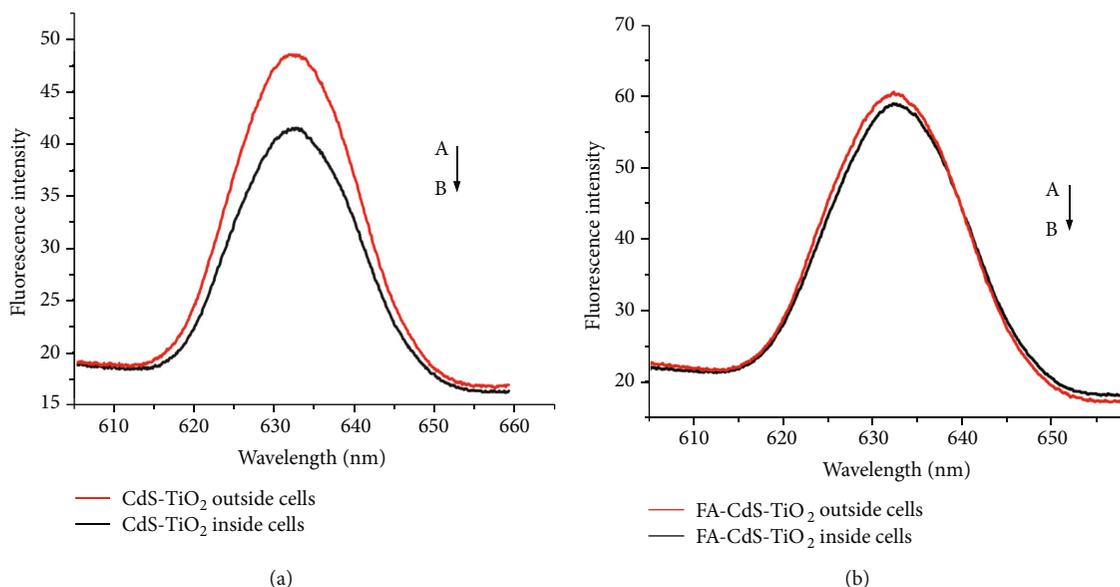


FIGURE 14: (a) The fluorescence emission spectrum of CdS-TiO₂ nanocomposite uptake by cells with the excitation wavelength of 410 nm. (b) The fluorescence emission spectrum of CdS-based nanocomposite B uptake by cells with the excitation wavelength of 410 nm.

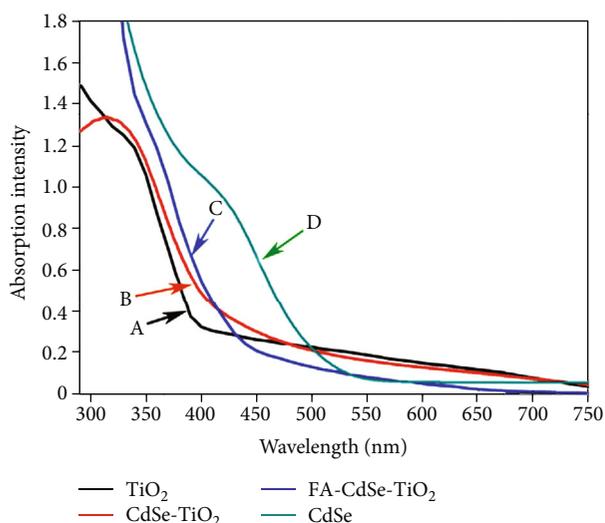


FIGURE 15: The UV-Vis absorption spectra of pure TiO₂, CdSe, CdSe-TiO₂ nanocomposite, and CdSe-based nanocomposite C.

3.6.3. Photodynamic Therapy of FA-CdS-TiO₂ (CdS-Based Nanocomposite B) and FA-CdSe-TiO₂ (CdSe-Based Nanocomposite C) on HL60 Cells. As shown in Figure 17(a), the PDT efficiency of the FA modified CdS-TiO₂ nanoparticles on HL60 cells was enhanced in comparison with that of CdS-TiO₂ in the increment of drug concentration. When the concentration of the nanoparticle drug was 40 $\mu\text{g}/\text{mL}$, the PDT inactivation efficiency of FA-CdS-TiO₂ nanocomposite reached the highest 80%, while the PDT inactivation efficiency of CdS-TiO₂ nanocomposite was only about 54%. According to the results of dark toxicity of nanocomposite on HL60 cells, it could be seen that the modification of FA not only reduced the dark toxicity of CdS-TiO₂ but also enhanced the PDT inactivation efficiency.

Figure 17(b) shows the PDT inactivation efficiency of CdSe-TiO₂ and FA-CdSe-TiO₂ nanocomposite with different FA ratios on HL60 cells under different light intensities. According to the result of the dark toxicity of FA-CdSe-TiO₂ nanocomposite to HL60 cells, the mass concentration of nanocomposite was chosen as 20 $\mu\text{g}/\text{mL}$, and the light dosages were 6, 12, and 18 J/cm^2 , respectively. As shown in the figure, the PDT inactivation efficiency of FA-CdSe-TiO₂ nanocomposite on HL60 cells was significantly improved compared with CdSe-TiO₂. With the increase of light intensity, PDT inactivation efficiency of different samples was improved. When the FA modification ratio was 1.0 and the light dosage was 18 J/cm^2 , the PDT inactivation efficiency reached the highest 84%. It was also found that when the ratio of FA was generally less than 1.0, the inactivation efficiency of FA-CdSe-TiO₂ nanocomposite enhanced with the increase of the ratio of FA. Because folic acid molecules on FA-CdSe-TiO₂ nanocomposite surface could combine with FR overexpressed on the cancer cell membrane, it could promote absorption of nanocomposites by HL60 cells. More nanocomposites involving PDT reaction result in higher inactivation efficiency of HL60 cell. However, when the modification ratio of FA was increased to 2.0, the PDT inactivation efficiency was significantly decreased, which might be due to the excessive modification of FA on the surface that hindered the excitation of CdSe-TiO₂ by visible light.

Figure 17(c) shows the PDT inactivation efficiency of different samples at the concentration of 20 $\mu\text{g}/\text{mL}$ and dosage of 18 J/cm^2 . Low concentration and appropriate light intensity were conducive to comparing PDT efficiency of different materials. High concentration of nanomaterials could increase PDT efficiency but also lead to increase dark toxicity [51]. CdS-TiO₂ (CdS-based nanocomposite A) when the ratio of CdS to TiO₂ was 1.0, the PDT inactivation efficiency reached 51%. FA-CdS-TiO₂ nanocomposite (CdS-based nanocomposite B) when the FA modification ratio

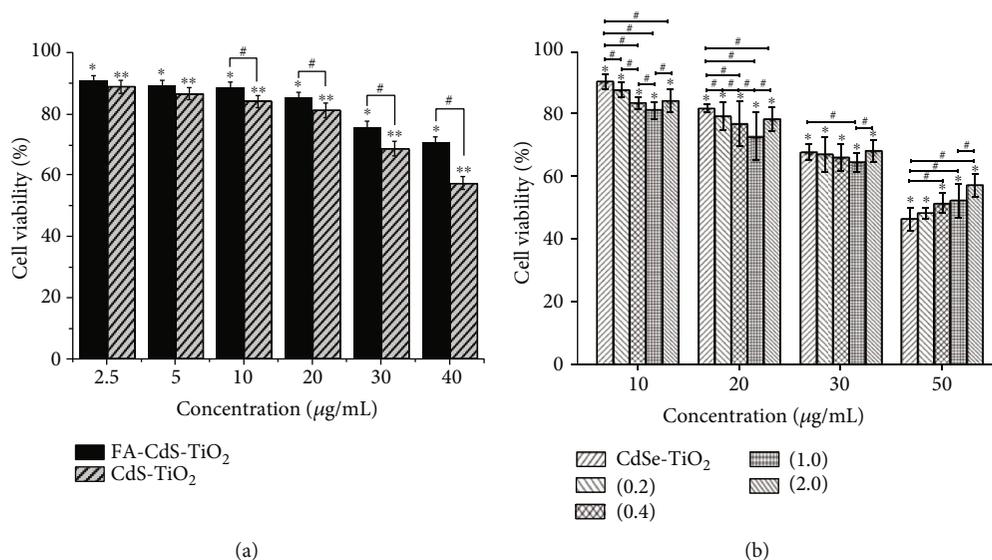


FIGURE 16: (a) Viability of HL60 cells treated with CdS-based nanocomposite B and CdS-TiO₂ without irradiation for 24 hours. Each data point represents mean \pm SD ($n = 3$). * $P < 0.001$ indicated that there was a difference compared untreated control cells with groups of FA-CdS-TiO₂ in different concentrations, ** $P < 0.001$ indicated that there was a difference compared untreated control cells with groups of CdS-TiO₂ in different concentrations, and # $P < 0.05$ compared with the two groups. (b) Viability of HL60 cells treated with CdSe-TiO₂ and CdSe-based nanocomposite C (with different folic acid ratios) in different concentrations without irradiation for 24 hours. Each data point represents mean \pm SD ($n = 3$). * $P < 0.001$ indicated that there was a difference compared with untreated control cells without irradiation, and # $P < 0.05$ compared with the two groups. All the data shown in the figure are representative.

was 1.0, the PDT inactivation efficiency reached 65.8%. The PDT inactivation efficiency of CdTe-TiO₂ (CdTe-based nanocomposite B) and the hydroxyacetic acid-coated CdTe QDs (CdTe-based nanocomposite C) were 53.8% and 42.3%, respectively. The PDT efficiency of the hydroxyacetic acid-coated CdSe QDs (CdSe-based nanocomposite A) was 60.2%. When the ratio of CdSe to TiO₂ was 1.0, the PDT inactivation efficiency of CdSe-TiO₂ reached 66.2%. FA-CdSe-TiO₂ nanocomposite (CdSe-based nanocomposite C) when the folic acid modification ratio was 1.0, the PDT inactivation efficiency reached the highest 84%. These results concluded that the PDT inactivation efficiency of FA-CdSe-TiO₂ nanocomposite (CdSe-based nanocomposite C) was the highest. Compared with the CdX-TiO₂ ($X = S, Te, Se$) nanocomposites, the PDT inactivation efficiency of CdSe-TiO₂ was the highest whereas CdS-TiO₂ was the lowest. The possible reason is that the band gap of CdSe QDs was the narrowest so the CdSe QDs coupling with TiO₂ promoted the photogenerated carrier separation of CdSe-TiO₂. The diameter of particles of CdTe-TiO₂ was larger resulting in low uptake of nanoparticles by HL60 cells. The PDT inactivation efficiency of CdX-TiO₂ ($X = Te, Se$) was higher than CdX QDs due to the CdX QD coupling with TiO₂ that could promote the photogenerated carrier separation and charge transfer efficiency. The advantages of using CdSe-TiO₂ nanocomposite for PDT in comparison with previous works about CdX ($X = S, Te, Se$) quantum dot modification were applied in different fields (see Table S1, Supporting Information). When the CdX-TiO₂ ($X = S, Se$) was modified with FA, the PDT inactivation efficiency was significantly increased. This was ascribed to a lot of folic acid molecules on FA-CdX-TiO₂ ($X = S, Se$) nanocomposite surface that could combine with

FR; it could promote absorption of nanoparticles by HL60 cells. The results clearly demonstrated that FA-CdSe-TiO₂ nanocomposite (CdSe-based nanocomposite C) was the best PDT nanomaterial.

3.6.4. Alteration of Reactive Oxygen Species in HL60 Cells after PDT. During photodynamic therapy, the photosensitizer transfers energy to the oxygen molecules in cells after being stimulated by visible light and produces a lot of highly oxidizing reactive oxygen species, resulting in the destruction of cell membranes or organelles, thereby killing cells [52, 53]. The level of intracellular reactive oxygen species (ROS) induced by irradiation is an important indicator for the evaluation of a photosensitizer. In this series of experiments, the fluorescence probe labeling technology was used to analyze the ROS levels in HL60 cells by FS.

The level of reactive oxygen species in the cells treated with FA-CdS-TiO₂ nanocomposite (CdS-based nanocomposite B) and CdS-TiO₂ nanoparticle before and after visible light irradiation was shown in Figure 18(a). When the concentration of nanoparticles was 20 µg/mL, the cells treated with FA-CdS-TiO₂ nanocomposite produced significant amounts of reactive oxygen species after 40 minutes of irradiation. Compared with the CdS-TiO₂ nanoparticle cell group under the same condition, the yield of ROS in the FA-CdS-TiO₂ group was significantly reduced without irradiation. However, the production of ROS of FA-CdS-TiO₂ nanocomposite was significantly increased under irradiation.

Figure 18(b) shows the intracellular ROS level of HL60 cells after being treated by CdSe-TiO₂ and FA-CdSe-TiO₂ nanocomposite (CdSe-based nanocomposite C) with 100% FA. The intracellular ROS level in the FA-CdSe-TiO₂-treated

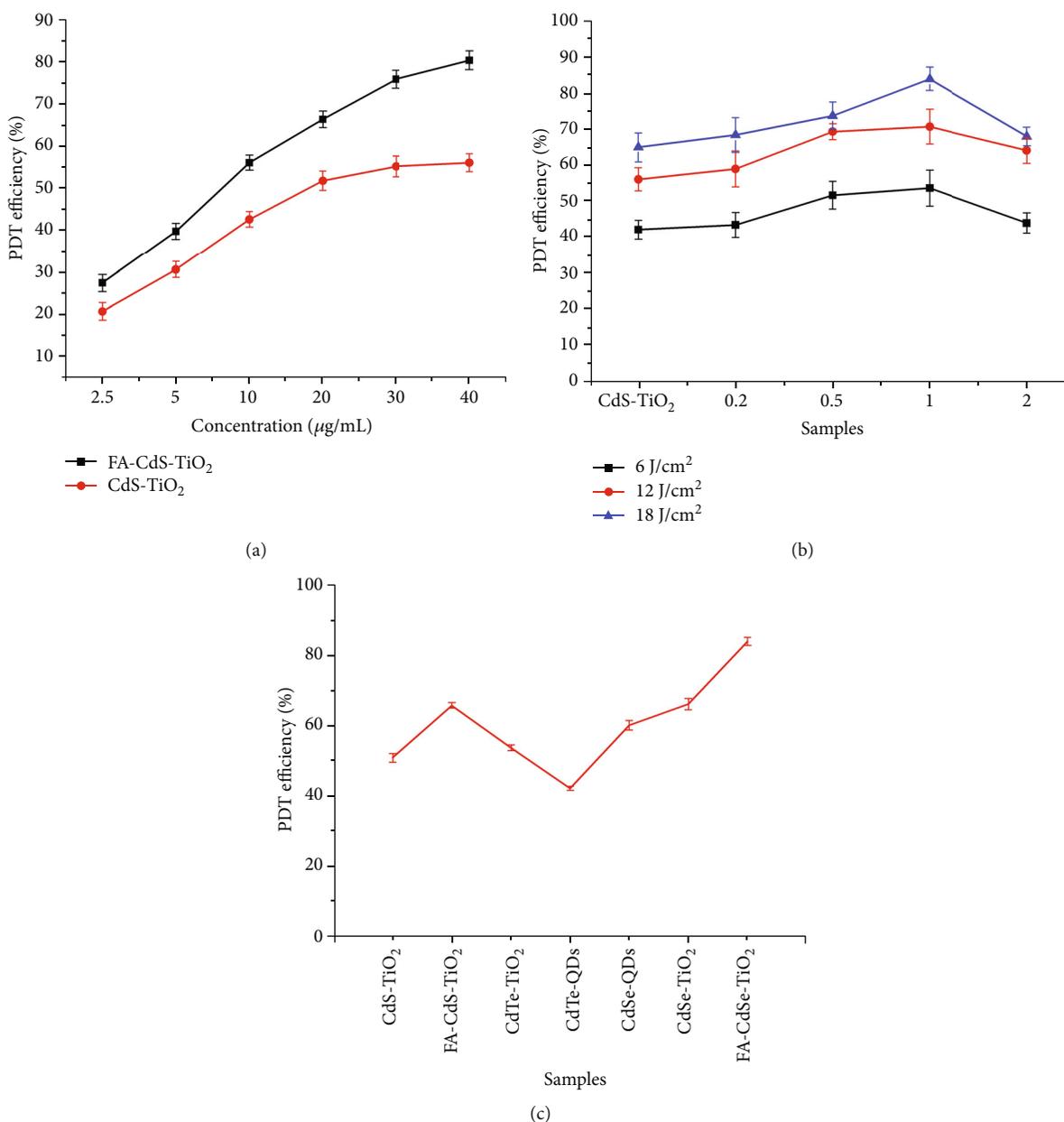


FIGURE 17: (a) The PDT efficiency of CdS-based nanocomposite B and CdS-TiO₂ on HL60 cells. Each data point represents mean \pm SD ($n = 3$). (b) The PDT efficiency of CdSe-TiO₂ and CdSe-based nanocomposite C modified with different folic acid ratios on HL60 cells at different light dosages. Each data point represents mean \pm SD ($n = 3$). (c) The PDT efficiency of different samples at the concentration of 20 $\mu\text{g/mL}$ and dosage of 18 J/cm^2 . Each data point represents mean \pm SD ($n = 3$). All the data shown in the figure are representative.

group was higher than that in the CdSe-TiO₂-treated group. The ROS level test result was consistent with the PDT efficiency test result described above (Figure 17(b)). These results indicated that FA modification could effectively improve the uptake of nanocomposites by HL60 cells, thus enhancing the photodynamic inactivation efficiency of HL60 cells.

3.6.5. Ultrastructural Morphology of the Treated Cells. Figure 19(a) shows normal HL60 cells with a cell diameter of about 8 μm and a complete cell structure with clear outline and villous. Figure 19(b) is the ultrastructural diagram

of HL60 cells after CdSe-TiO₂-mediated PDT. After coculture of CdSe-TiO₂ and HL60 cells with irradiation, the cell structure was damaged, and some cracks and pores were present on the cell membrane surface. The cells were incomplete and showed a tendency of lysis. It was supposed that CdSe-TiO₂ mainly accumulated on the surface of cell membrane and produced reactive oxygen species after visible light excitation, which caused damage to the cell membrane surface and eventually led to cell lysis and necrosis. Figures 19(c) and 19(d) are diagrams of multiple cells and a single cell after FA-CdSe-TiO₂ nanocomposite (CdSe-based nanocomposite C), and HL60 cells were incubated

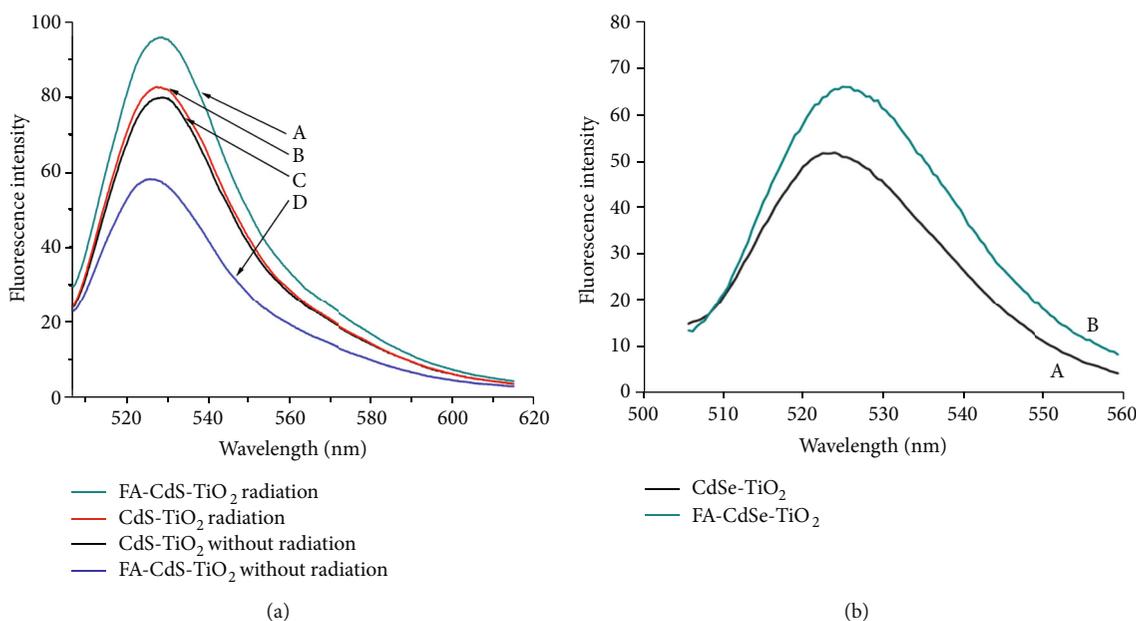


FIGURE 18: (a) The fluorescence spectra of reactive oxygen species probe in HL60 cells after PDT with the excitation wavelength of 485 nm (CdS-based nanocomposite B). (b) The fluorescence spectra of reactive oxygen species probe in HL60 cells after PDT with the excitation wavelength of 485 nm (CdSe-based nanocomposite C).

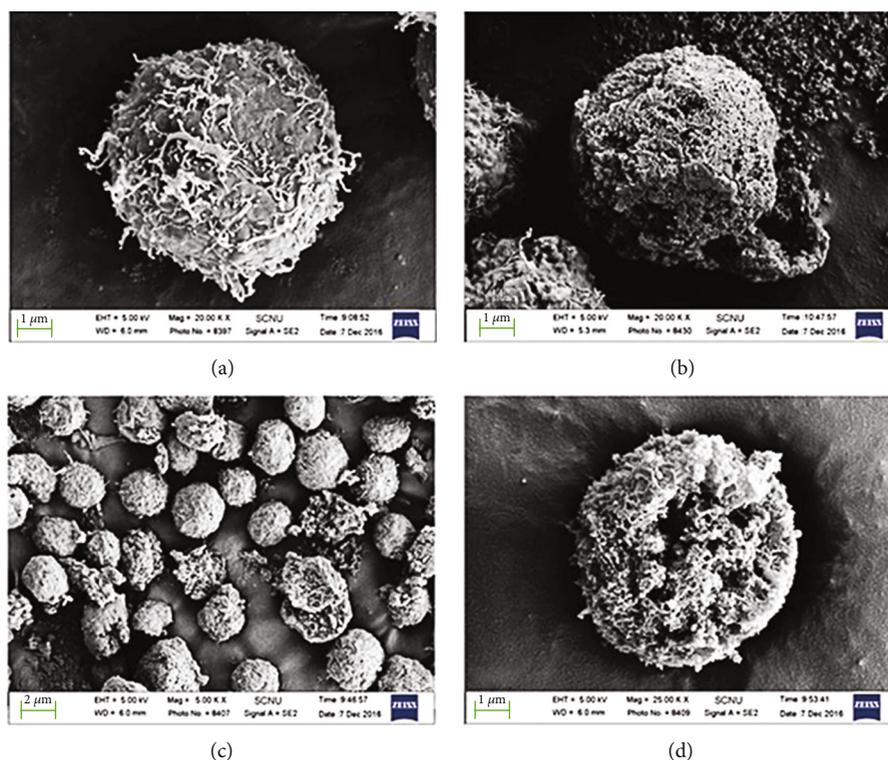


FIGURE 19: (a) The ultrastructural morphology of normal HL60 cell; (b) the ultrastructural morphology of the HL60 cell treated with CdSe-TiO₂ nanoparticles after PDT; (c) the ultrastructural morphology of the multiple HL60 cells treated with CdSe-based nanocomposite C after PDT; (d) the ultrastructural morphology of the single HL60 cell treated with CdSe-based nanocomposite C after PDT.

with irradiation, respectively. The cell structure was damaged seriously, and the cell membrane was severely ruptured and defective, and the organelle tissue was also damaged to a certain extent. The phenomenon may be related to FA-

CdSe-TiO₂ nanocomposite entering the cells through endocytosis and nanocomposite accumulated on the surface of cell membrane producing a large amount of ROS after light excitation. ROS reacted with the organelle and cell

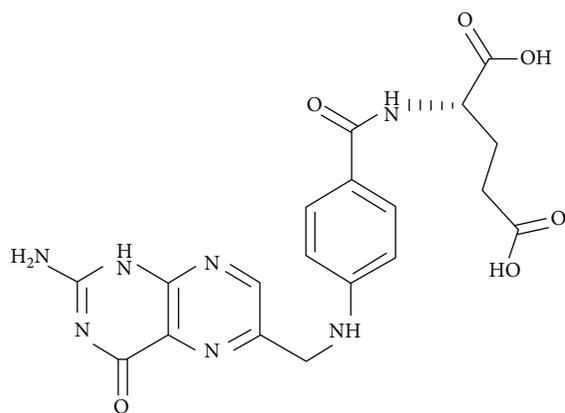


FIGURE 20: The molecular structure of folic acid.

membrane by oxidation, resulting in the destruction of the organelle and cell membrane, thus leading to the necrosis of HL60 cells.

3.6.6. Mechanism of Photocatalysis on CdX (X = S, Se)-Based Nanocomposites Modified with Folic Acid. Figure 20 shows the molecular structure of FA. It could be found that FA contained a lot of carboxyl groups, while the surface of TiO₂ contained a rich distribution of hydroxyl groups. It could be seen that carboxyl groups of folic acid molecules were combined with hydroxyl groups on the surface of CdS-TiO₂, CdSe-TiO₂ to form FA-CdS-TiO₂, and FA-CdSe-TiO₂ by esterification reaction, respectively. Because FA could selectively target the FR that are overexpressed on the cell membrane of cancer cells, cancer cells can absorb more of the nanocomposites. It could enhance the photocatalytic activity and the yield of ROS in the photodynamic process and finally increase the PDT inactivation efficiency.

Based on all of these analysis results, it was indicated that FA-CdSe-TiO₂ has the highest PDT efficiency and low dark toxicity. Due to CdSe QDs having narrower band gap, it combined with TiO₂ that can extend the visible response range of TiO₂. The separation efficiency of photogenerated carriers was improved. When FA is modified on the surface of CdSe-TiO₂, FA could specifically recognize FR to improve the efficiency of HL60 cell uptake of nanocomposites. As the present clinical experiment is only in vitro, it has not been tested in vivo. The next step is to conduct experiments on mice in vivo, to further explore the possibility of human leukemia HL60 cells being exposed to the concentration of nanocomposites whether there is a health risk to humans.

4. Conclusions

In this paper, titanium dioxide was modified with quantum dots such as CdS, CdTe, and CdSe, and the particle size and spectral response range of novel nanocomposites were explored by a transmission electron microscope (TEM), X-Ray Diffraction pattern (XRD), and UV-visible absorption spectrum (UV-Vis). The large particle size of CdTe-TiO₂ was not conducive to the uptake of nanoparticles by HL60

cells. Nevertheless, the particle size of CdSe-TiO₂ was smaller and had a larger specific surface area; it was beneficial to participate in photoreaction. CdSe has the longest absorption cutoff wavelength and narrowest band gap. The spectral response range could be red-shifted to the visible region due to the narrow band gap of the doped quantum dots. It was used to improve the visible light response range of TiO₂ under the visible light 410 nm. Fluorescence spectroscopy (FS) was used to analyze the capture, migration, and transformation efficiency of CdX-TiO₂ nanocomposites. The doping of QDs reduced the recombination rate of electrons and holes on the surface, improving the separation efficiency of photogenerated carriers and photocatalytic activity. In addition to exploring the physical and chemical properties of CdX-TiO₂ nanocomposites, it was also necessary to detect the biological properties such as low dark toxicity to cells or its PDT efficiency to human acute promyeloid leukemia cell (HL60 cells). CdSe-TiO₂ nanocomposite had lower dark toxicity and higher PDT inactivation efficiency. But CdX-TiO₂ NPs were nonspecific targeting; thus, it was limited in its biological applications. To achieve more efficient cancer targeting, FA is used to modify the prepared CdX-TiO₂ NPs. Results of Fourier transform infrared spectroscopy (FT-IR) indicated FA compounds combine with CdX-TiO₂ NPs in the form of esterification. The fluorescence probe labeling technology was used to analyze the higher ROS level in HL60 cells treated with FA-CdX-TiO₂ nanocomposite compared with CdX-TiO₂ after PDT by FS. Finally, serious damage to the ultrastructure of HL60 cells after treating with FA-CdSe-TiO₂ nanocomposites for photodynamic therapy inactivation could be observed by the scanning electron microscope (SEM) compared with normal cells. It could be proved that reactive oxygen species were produced under irradiation which oxidized cell membranes and caused damage. Optimum PDT efficiency of FA-CdSe-TiO₂ indicates that photocatalytic and targeting ability is much higher than pure TiO₂ and CdSe-TiO₂. Furthermore, the mechanisms of improving the PDT efficiency and targeting of photodynamic therapy simultaneously by CdX (X = S, Se)-based nanocomposites modified with FA are summarized. However, most QDs contain heavy metals, which would be toxic to normal cells and required further evaluation. As the present clinical experiment is only performed in vitro, the next step is to conduct experiments in vivo to further explore whether the FA-CdSe-TiO₂ nanocomposite is a superior photosensitizer for treating leukemia to human body.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this paper.

Acknowledgments

This work has been financially supported by the National Natural Science Foundation of China (61072029) and the Science and Technology Planning Project of Guangzhou City (2014J4100049).

Supplementary Materials

Figure S1: (a) Energy Dispersive Spectrometer (EDS) of CdS-TiO₂-0.2 (CdS-based nanocomposite A). (b) X-ray photoelectron spectroscopy (XPS) of CdTe-TiO₂ (CdTe-based nanocomposite B). (c) EDS of FA-CdSe-TiO₂ (CdSe-based nanocomposite C). Table S1: comparisons between recent reports of CdX (X=S, Te, Se) and our work. (*Supplementary Materials*)

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