

Review Article

Nanoplastics as an Invisible Threat to Humans and the Environment

Nenad Joksimovic¹, ¹ Dragica Selakovic¹, ² Nemanja Jovicic¹, ³ Nenad Jankovic¹, ⁴ Periyakaruppan Pradeepkumar, ⁵ Aziz Eftekhari⁶, ^{6,7} and Gvozden Rosic²

¹Department of Chemistry, Faculty of Science, University of Kragujevac, 34000 Kragujevac, Serbia

²Department of Physiology, Faculty of Medical Sciences, University of Kragujevac, 34000 Kragujevac, Serbia

³Department of Histology and Embryology, Faculty of Medical Sciences, University of Kragujevac, 34000 Kragujevac, Serbia

⁴Department of Sciences, Institute for Information Technologies Kragujevac, University of Kragujevac, 34000 Kragujevac, Serbia

⁵Department of Humanities and Science, MLR Institute of Technology (UGC-Autonomous), 500 043, Hyderabad, Telangana, India

⁶Research Center for Pharmaceutical Nanotechnology, Biomedicine Institute, Tabriz University of Medical Sciences, Tabriz, Iran

⁷Department of Pharmacology & Toxicology, Tabriz University of Medical Sciences, Tabriz, Iran

Correspondence should be addressed to Gvozden Rosic; grosic@medf.kg.ac.rs

Received 11 August 2022; Revised 13 September 2022; Accepted 22 September 2022; Published 13 October 2022

Academic Editor: Lakshmipathy R

Copyright © 2022 Nenad Joksimovic et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The consequence of the unstoppable growth of plastic production and inadequate waste disposal is plastic pollution, which is becoming a global challenge. Large quantities of discarded plastics in the environment are under the direct influence of biotic and abiotic factors, leading to the fragmentation of particles and the formation of tiny particles of plastics or nanoplastics. Nanoplastics are a fast-growing pollutant that, due to ubiquity in the environment, causes great public concern. It also attracts the attention of scientists in detecting harmful effects on health and the environment. This review is aimed at summarizing all adverse effects of nanoplastics on human health and the environment. Due to their toxic effects, it is necessary to reduce the disposal of plastics in the environment, develop or improve existing methods, and implement legislation that would reduce the release of nanoplastics into the environment. A possible ban or reasonable regulation of nanoplastics in cosmetics or food can be expected only after a critical mass of scientific objections has been created.

1. Introduction

Environmental pollution and its far-reaching consequences are two of the main problems facing today's modern society [1, 2]. Globalization, a dramatic increase in human population in the 20th century, and mass consumption of various products, followed by inadequate disposal of waste, have led to the worrying situation in which both land and water areas of our planet are affected today [3]. More importantly, such changes affect the animal communities that live in them. Marine pollution, as a part of this problem, is a subject of research by many scientists and societies for environmental protection, as many believe that environmental pollution is the most dangerous factor for human health [4–6]. The main component of environmental pollution is plastic [7], which is continuously disposed of, in substantial quantities, as a byproduct of human consumption. By plastics, we mean hundreds of different plastic polymers obtained from organic compounds. In today's society, plastic is an indispensable part of most everyday items. Since the creation of the first plastic polymers in the midtwentieth century, plastic production has increased dramatically, from 1.5 million tons in 1950 to 280 million tons of plastic produced in 2011, with an annual growth rate of 8.7 percent [8]. It is estimated that about 5.25 trillion pieces of plastic are currently circulating in the sea alone [9]. While a small portion of plastic waste ends up in the sea due to maritime activities, it is estimated that 80 percent comes from land-based sources [10]. Improperly



FIGURE 1: The most common forms of plastics for human consumption (a), structural formulas (b), and plastic pollutants are categorized by size (c).

disposed plastic waste accumulates in large quantities in the environment and comes under the influence of wind as well as river currents, which eventually reach the shores and enter the sea. The monitoring and "transport" of waste are proportionally connected and are inevitably associated with economic development, industrialization, and local infrastructure [11, 12].

The most common forms of plastics produced for human consumption and thus found in the sea are polymers polypropylene (PP), polyvinyl chloride (PVC), and polyethylene (PE) (Figure 1) [13]. The negative impact of larger pieces of plastic waste, better known as "macroplastics," on the marine environment is manifold. The effects on the human economy are most pronounced in many types of marine industries including fishing, aquaculture, and energy production, as well as maritime affairs, as plastics can greatly damage the equipment used in them. Another branch of the economy that is negatively affected by this problem is tourism, the main source of income for some countries [14]. However, the most severe and far-reaching consequences of marine pollution by plastics are borne by the ecosystem itself and all living organisms in it [15]. Some of the many consequences that plastic waste brings into the sea include injury and death of seabirds, mammals, and fish, occurring as a result of their interference with plastic parts, the introduction of plastic into the body by feeding, and transport of marine organisms from their natural habitat to new floating parts, as well as "choking" the seabed by preventing alteration gases due to deposits of plastic waste on it [16, 17]. Perhaps, the most shocking fact is that even if plastics were no longer disposed of in the sea, nanoplastic pollution would continue to grow as a result of the decomposition of preexisting plastic waste in the environment [18].

Nanoplastics can be defined as really small plastic parts, smaller than $1 \mu m$ in size. It includes a wide range of plastic particles, which vary in size, color, shape, and origin. The problem of nanoplastic pollution has become the subject of attention and concern of many scientists in recent years [19–21]. However, nanoplastics in the environment are not a completely new phenomenon. Namely, recent research has shown that nanoplastics have been actively accumulating in the seas and oceans around the world for the past 40 years. Due to their extremely small dimensions, nanoplastics are introduced into marine organisms through the food they eat and cause various blockages of the digestive and respiratory systems and other health problems [22, 23]. Nanoplastics can also be carriers of resistant organic pollutants which, due to the large ratio of surface to volume, as well as the hydrophobic surfaces of nanoplastics, are adsorbed on it. Intake of nanoplastics can thus also lead to the uptake of toxins into the food chain [24]. What worries scientists, even more, is the fact that marine organisms with accumulated nanoplastics in their bodies are being used in human nutrition. Thus, it is essential to study each aspect of nanoplastics individually.

2. The Invisible Spectrum of Plastic

Visible plastic can be removed from the environment, while micro- and nanoplastics remain an integral part of nature. Therefore, the toxic profile of micro- and nanoplastics is worse than that attributed to visible plastic [25]. Appropriate environmental clean-up actions relate to the removal of visible plastic, but not micro- and nanoplastics. Until recently, even microplastics were not the focus of the public debate, for the simple reason that they were not visible and detectable. Today, microplastics are a measurable quantity, a parameter of environmental plasticization that can be quantified [26]. Thus, in some ocean areas, there are more than a million microplastic fragments per square kilometer of the sea surface.

Just as microplastics are a bigger environmental problem than visible "bags and bottles," nanoplastics are, as already known, an additional and even greater risk to the environment and human health [27]. However, it is less known that microplastics can also be further converted into even smaller components. When the size of the resulting particles is less than one hundred nanometers, scientists talk about nanoplastics. Thus "small" (Greek: "micro") turns into "dwarf" (Greek: "nano"). However, until recently, that was just a guess. Apart from being invisible to the naked eye, like microplastics, nanoplastics are still difficult to access with analytical instruments [28]. Detection, quantification, and characterization of nanoplastic particles are too great of a methodological challenge for many scientists. Therefore, nanoplastics are stuck in the realm of the hypothesis, not laboratories.

2.1. Nanoplastics. Plastics appear in the environment in visible and invisible forms. The visible form refers to common scenes, such as bags "flying" on tree branches or PET bottles floating in rivers, lakes, and seas. The visible form of plastic in the environment is an aesthetic mockery and a reminder of the socially indiscriminate reception of all offers from the petrochemical industry. The invisible form of plastic is called nanoplastic.

Plastic particles of different sizes can be found in the natural environment, so plastics are divided into macroplastics (particles larger than 2.5 cm), mesoplastics (particles between 2.5 cm and 5 mm), microplastics, and nanoplastics (Figure 1) [29]. The upper limit of microplastic particle dimensions is usually 5 mm, and the lower limit has not yet been determined [30]. The European Commission defines the dimensions of microplastic particles between 5 mm and 100 nm, while the particle size of nanoplastics ranges between 1 and 100 nm [31].

Nanoplastic can be obtained from primary or secondary sources [32]. These are plastic fragments invisible to the human eye such as filaments, granules, and flakes. There is a so-called secondary nanoplastic that is formed by the decomposition of larger plastic pieces and packaging [33]. By the action of ultraviolet rays and chemical and biological processes, visible plastic is crushed into invisible nanoplastic. However, there are also the so-called primary or synthetic nanoplastics that are targeted in the polymer industry and inserted into various products, such as cosmetics or textiles [34].

For example, researchers at McGill University in Montreal recently published a study showing that nanoplastics are found in many cosmetic products, especially those filled with microplastic granules [35]. Cosmetic nanoplastics are formed during the production process of shampoos, shower gels, or facial care products. These are polyethylene and polystyrene microgranules that are inserted into cosmetic products, which also create nanoplastic by-products. Using special detection and isolation techniques, Canadian scientists have proven that nanoplastics are a regular cosmetic supplement.

2.2. Sources of Nanoplastics. In the scientific community, there is still a debate related to the classification of plastic waste in relation to the size of the particles and their chemical composition. However, we can say that nanoplastics are polymer-based particles in the nanometer size range. By primary particles of nanoplastic we mean particles of a defined size and composition that are intentionally produced for use in industry, while secondary particles are regarded as particles created by the decomposition of larger plastic waste. Nanoplastics can originate from several sources, but the most important are tires, synthetic textiles, marine coatings, road markings, personal care products, plastic pellets, city dust, and nanoplastic from industrial suppliers. To date, plastic pollutants have occurred in oceans [36, 37], lakes [38, 39], soils [40], and sediments [41, 42]. However, nanoplastics have even been found in isolated places with rare

or without human activity, such as the Arctic [43, 44] and the Antarctic [45]. Under the influence of external factors such as UV rays from sunlight, wind, and waves, the formation of nanoplastics could be formed if the mentioned pollutants have been found in the natural environment. Pollutants are subjected to mechanical fragmentation, and then, the small particles become even smaller and further transform into microplastic. At the moment of microplastic formation, the nanoplastic is also formed. This process cannot be stopped because it happens in nature [46]. It is a long and lasting process of the plastic lifecycle in the environment. Generally, poorly managed plastic matter leads to the formation of macro- and microplastic particles that further can be transformed into nanoplastic particles (e.g., polyethylene terephthalate (PET) or polystyrene) [47-50]. Compared to terrestrial ecosystems in the aquatic environment, plastic waste is significantly easier to fragment in the aquatic environment predominantly due to high salinity and the presence of microorganisms [51]. When microplastics are formed in environmental areas, the physicochemical factors affect further degradation of nanoplastics. The main factors are UV radiation, temperature, and pH. The presence of other contaminants (e.g., heavy metals and their oxides) can accelerate this process. Nonbiodegradation and biodegradation are key pathways in the mechanism of nanoplastic formation. The first path implies physical, thermal, oxidative, and photodegradation or even hydrolysis [52]. Thermal degradation is controlled by humans and cannot be reached in the environment. This process is followed by the oxidative transformation -C-C- bonds into -C=O and/or -C-O, thus significantly changing the hydrophobic properties of the plastics. Therefore, physical, oxidative, and photodegradation or hydrolysis of plastic matter have occurred in nature. Physical degradation is the first stage in the mechanism of nanoplastic formation. It implies the fragmentation of robust plastics into smaller pieces dependent on weathering conditions and sea waves. Different shapes are usually formed, e.g., flakes, discs, rectangles, cylinders, and spheres [53]. In this way, the contact surface is changed, which is important for the next steps in the degradation cycle. These formed species could be photodegraded via bond-breaking reactions that are dependent on UV rays, and consequently, the hydrolysis is processed. These associated abiotic processes convert a high to a low polymer. During this fragmentation process, significant changes will occur, such as structural differences, mechanical properties of the polymer, and increased surface area as the crucial property for their interaction with microbes [54].

Therefore, biodegradation is often a mechanistic path in the environment for converting plastic waste to nanoplastics [24, 55]. Biodegradation is slower compared to the nonbiodegradable path. Moreover, smaller pieces, as well as low polymers formed in nonbiodegradable paths, can be recognized by different microorganism populations that in combination with pH value, ions, oxygen, UV light, and temperature can produce nanoplastic particles.

2.3. Nanoplastics in the Environment. Nanoplastics are ubiquitous and are found all over the Earth: in the oceans, on the coasts, and inland [56], brought by wind from various sources, nanoplastic sediment on the ground, from where they easily travel through the soil, ending up in aquifers and ultimately contaminating drinking water. The path of nanoplastics in the environment is interconnected with all three components, and in each of these, their harmful effects are found [57]. However, the biggest problem is the organisms that live in aquatic systems.

In aquatic ecosystems, nanoplastics are found in all parts of the water column [58]. Due to their structure, polymers have a specific density $(0.9-2.3 \text{ g/cm}^3)$ that is lower than the water density (1.02 to 1.03 g/cm³), which leads to the floating of plastics and nanoplastics on the water surface [59]. However, on the surface of nanoplastics, organisms (e.g., algae) can colonize and increase the weight of particles, i.e., change the density, and thus reduce buoyancy, which allows immersion. Algae overgrow plastic waste that ends up in the sea, so much waste under the influence of sea currents transfers the species to places where they were originally absent [60]. Chemicals adsorbed on microplastics can have a toxic effect, and nanoplastic particles are small enough to pass through cell walls and membranes and reduce population growth and chlorophyll concentration, as studied in green algae Scenedesmus obliquus [30]. Fragmentation does not affect particle density because it does not change the chemical composition. Furthermore, the concentration of plastics, therefore nanoplastics, in the oceans depends on the period in which it is sampled. For example, if the coastal winds are low, the concentration is higher. Water that flows at a lower speed has lower energy, and the plastic that is denser than its environment will sediment. In areas of faster water flow, the plastic is easier to carry and resuspend. Furthermore, the areas of lower salinity show an increase in the plastic deposition as a result of an increase in plastic density [61].

Finally, the sum of the actions of coastal winds, the colonization of nanoplastic particles, and the exact distribution of nanoplastics in the water column remain unknown. Nanoplastics from cosmetic products, cleaning products, medicines, and mostly from washing machines that emit microfibers end up in municipal wastewater [35, 61]. Microfibers come from synthetic fabrics from which much of the garment is made, and it is estimated that 1900 synthetic microfibers are released during a washing cycle [61]. Furthermore, due to the very small size of the particles, they pass through filters in wastewater treatment facilities and end up in rivers or soil through direct discharge of treated water or using waste sludge, from where they travel to other parts of the environment. According to Mason et al., between 50000 and 15 million microplastic particles pass through wastewater treatment facilities daily [62].

2.4. Surface Toxicity. Just as microplastics are a bigger environmental problem than visible "bags and bottles," nanoplastics, according to scientists, are an additional and greater risk to the environment and human health. Nanomaterials, including nanoplastics, have a special property: although their size is on the scale of ordinary molecules, the (relative) surface area of nanomaterials is extremely large and active. Moreover, most of the biological, chemical, and technological properties of nanomaterials are attributed to this new dimension or parameter—the surface.

Nanoplastic size corresponds to the size of cell membranes, and due to hydrophobicity, it is easily absorbed into cells. This is sufficient for the mechanism of cellular toxicity (cytotoxicity) proven in nanoplastics [63, 64]. Unfortunately, the regulation of nanoplastics in cosmetics and food is unclear, which gives room to the industry to "hunt in the dark". In Table 1 are presented some *in vitro* and *in vivo* studies impaling the threats of nanosized plastics in the marine environment.

2.5. Cellular Responses to Nanoplastics. The toxicity of nanoplastics is a subject of discussion throughout the scientific community and has been attracting attention for many years. The cellular response depends on the way and route of absorption of nanoplastics and the location in the body where they are stacked (Figure 2). It is considered that gastrointestinal and airway cells are the most exposed places to nanoplastics.

Intra- and extracellular responses could be significantly different mainly because of the various physical properties of the nanoplastics particles. Effect of particle size and dose, the effect of particle charge, and time of exposition are the key factors that can induce different cellular responses if nanoplastics are found in the cell. Due to the enormous use of plastic in food packaging, the largest number of studies is focused on the effect of nanoplastics on the cells of the gastrointestinal tract [71]. The intestinal epithelial tissues primarily contain enterocytes, mucus-producing goblet cells, and microfold or M cells. There are two trafficking lines for polystyrene nanoparticles by Caco-2 cells. Primarily and the literally accepted mechanistic line was discovered and interpreted via micropinocytosis-mediated uptake [72]. Every few days (4-5), trapped nanoplastics might be excreted via epithelial cell shedding. A limited mechanistic path of the administration implies diffusion across the cell membrane. The process of elimination is followed by basal exocytosis. For instance, phagocytic M cells in the gut epithelia can significantly make nanoparticles transport better [73]. Furthermore, some types of adenocarcinoma cells are treated with nanoplastics sized from 44 to 100 nm. Experiments lasted one day while the tested concentrations were 1, 2, and 10 µg/mL [74]. During 1h exposure, any morphological changes in cells have not occurred. The important fact is that all sized particles during 1 h increased the level of interleukin-1 β , interleukin-6, and interleukin-8 proinflammatory cytokines. Going forwards, few studies showed low toxicity of nanosized plastics in Caco-2 intestinal cells. In that way, Cortés and coworkers demonstrated that after the treatment with 50-100 nm size particles, no increases of the ROS have been detected [75]. However, the authors have noticed the increase of mitochondrial membrane potential in a dosedependent manner (0-100 μ g/mL). In addition, the DNA cleavage or oxidative damage has not been approved after the treatment. Even though the gastrointestinal toxicity of nanoplastics of different sizes has not been proven in many studies, still, it is the subject of debate in the scientific community [76-79]. Knowing that the cell membrane has its

Study	Marine species	Potential toxicity/damage/threats
Wegner et al. [65]	Blue mussel (Mytilus edulis)	Extended exposure to nanoplastic can harm <i>Mytilus edulis</i> due to producing pseudofeces; it consumes energy, while reduced filtering activity can finally take to starvation.
Brandts et al. [66]	Mediterranean mussel (<i>Mytilus galloprovincialis</i>)	Significant alterations in the expression of genes associated with biotransformation, DNA repair, cell stress-response, and innate immunity have been discovered. Genotoxicity in hemocytes has been noticed.
Canesi et al. [67]	Mediterranean mussel (<i>Mytilus galloprovincialis</i>)	Increased cellular damage and ROS production.
Cole and Galloway [68]	Pacific oyster (Crassostrea gigas)	Exposure to nanoplastic concentrations greater than those meshured in the marine environment showed no measurable effects on the development or feeding capacity of the pacific oyster larvae.
González-Fernández et al. [69]	Pacific oyster (Crassostrea gigas)	The increase of relative cell size and ROS production was observed.
Besseling et al. [70]	Algae (Scenedesmus obliquus)	Nanoplastics inhibited the growth and development of this species. It also had an impact on increased mortality.

TABLE 1: Toxicity of nanoplastics to marine species and animals.



FIGURE 2: The most important routes of exposure, mechanisms of action, and effects of nanoplastics on the human organism.

own potential, it is very important to examine the effect of the nanoplastic surface charge on toxicity. For example, Walczak and collaborators investigated the effects of particle charge on the translocation efficiency across Caco-2/HT-29-MTX coculture. The highest translocation has been denoted for negatively charged particles [80, 81]. Mucin is a negatively charged glycoprotein with high molecular weight. Positively charged nanoplastics could smoothly bind to mucin resin *via* electrostatic forces and as consequence make changes within rheology [82, 83]. Similar conclusions have been obtained by Rieux and colleagues [84] who have investigated the effects of the absence of mucin. The study also showed that the transport of nanoparticles with amine resin has been higher in comparison with the carboxylated. Apoptosis was induced in the presence of catalase and nanoplastics with carboxylate resins. Thubagere and Reinhard suggested that this intracellular event was caused by nanoplastics particles and hydrogen-peroxide [85]. On the other hand, amine resins were capable to pass the mucin layer. Some investigations showed that after treatment with polystyrene nanoplastic beads, Caco-2 does not produce mucin, HT-29 produced a limited quantity of mucin, and LS174T produced abundant mucin cells [86]. In all treated cells, apoptosis was triggered after treatment with a high concentration ($100 \mu g$ /mL), whereas the lowest concentration ($20 \mu g$ / mL) caused higher cytotoxicity.

Besides, the inhaled nanoplastics can be responsible for plenty of respiratory problems such as irritation, dyspnea, decreased lung capacity, interstitial fibrosis, and granulomatous lesions [87, 88]. In one study [89], bronchial epithelial BEAS-2B cells were treated with different sizes of nanoplastics. Oxidative stress with significant ROS accumulation has been noticed upon 24 h. Cytokines interleukin-6 and interleukin-8 were also overexpressed. The nanoplastics with negatively charged resins and 60 nm size similarly generated oxidative stress in BEAS-2B cells [90]. Dose-dependent autophagy has also been provoked after the treatment. Namely, TCA cycle intermediate, pyruvate, and lactate were detected in nanoplastic-treated cells. All these facts implied that autophagy was caused by oxidative stress, due to the exposure of the cells to nanoplastics. The effects of different sizes of nanoplastics (25 and 70 nm) to A549 cells have also been investigated in the study by Xu and coworkers [91]. Namely, toxic effects and dose-dependent apoptosis were noted after 24 h for 25 (25–30 μ g/mL) and 70 nm (160–300 μ g/mL) particles. BEAS-2B cells were taken to investigate the effect of nanoparticles charge [92]. The aminated and carboxylated 60 nm particles have been used. After one day of incubation, the aminated particles caused the necrosis, most probably because of mitochondrial injury that is followed by fast ATP consumption. Higher toxicity of aminated compared to carboxylated nanoplastics has been discovered by Chiu and collaborators [93]. The higher toxicity has been proven in the *in vivo* study, where aminated nanoparticles caused up to 40-fold higher flux than carboxylated particles [94].

2.6. Nanoplastics in Food. Significant amounts of nanoplastics are found in aquatic species that are intended for human consumption, which allows nanoplastics to end up in the human body through the food chain. Nanoplastics are found in a variety of foods and beverages: seafood, beer, honey, sugar, salt, and drinking water [95]. 1000 particles of microplastics per person are introduced through sea salt. According to research by Orb Media on tap water samples from around the world, data show that a large proportion of drinking water is contaminated with microplastics, and the amount of microplastics ingested per person each year is 4,000 particles. Moreover, nanoplastic particles have been found in bottled drinking water, plastic, and glass bottles [96]. After being taken into the body, the nanoplastic particles are distributed into various tissues (testicles, liver, blood,

gills, and intestines). Numerous pieces of evidence are emerging indicating that nanoparticles affect the connection between the gastrointestinal and nervous system, gut-brain axis (Figure 3). Nanoparticles can cross the blood-brain barrier, a highly selective barrier necessary for neuronal functioning and crucial for maintaining necessary homeostasis and protecting the brain from toxins [97]. There are numerous ways of communication between the gut and the brain. The core of the gut-brain axis consists of the vagus nerve, which via afferent fibers sends information about the state of the internal organs to the brain and connects the CNS with the enteric nervous system. The sensitivity of the vagus nerve affects various responses in the brain, stimulating regions associated with feeding behavior and numerous psychological functions including emotions [97-99]. Efferent vagus activity affects the intestinal immune system and metabolism [97, 99, 100]. The CNS also interacts with the gastrointestinal tract through the hypothalamic-pituitaryadrenal axis (HPA axis) [99]. In response to hypothalamic and pituitary gland stimulation, adrenal glands produce the primary stress hormone, cortisol, which can modulate various processes in the gastrointestinal system [98]. The rare but consistent evidence shows that exposure to plastic nanoparticles can affect both the digestive and the nervous systems. The main effects include microbiota alterations, intestinal barrier permeability, oxidative stress, inflammation, neurotoxicity, and behavioral disturbances [97]. To supplement the information on the impact of nanoplastics on humans, it is important to summarize the potentially harmful effects of nanoplastics.

2.7. Side Effects of Nanoplastics. Numerous studies have shown that nanoplastic particles can lead to several side effects such as increased free radicals, activation of oxidative stress pathways, oxidative stress, DNA damage, lipid peroxidation, dysfunction of certain cellular organelles, and overactivation or inhibition of certain enzymes (Figure 4).

Our recent study investigated, in an animal model, the effects of oral uptake of polystyrene nanoparticles, a material that is widely present for human consumption. We demonstrated that after oral uptake, fluorescent polystyrene (PS) nanoparticles pass through the digestive system of mice, accumulate and aggregate in different organs, and induce functional changes in cells and organs. Apart from the increased accumulation in the inner ear, our results showed that PS effects were also gender-dependent. Male mice had significant accumulation in testicular tissue, which was associated with a decrease in testosterone levels. In addition, male mice showed a broad range of anxiogenic responses to PS nanoparticles, while hippocampal samples from treated females showed an increased expression of Bax and Nlrp3 genes, indicating a proapoptotic/proinflammatory effect of PS treatment [101].

A lysosome is a one-layer cytoplasmic organelle with high sensitivity to xenobiotics, including natural toxins. Consequently, lysosomal layer stability can be utilized as a biomarker to determine the potential effect of environmental contamination [102]. The lysosome capability disturbance was tracked down in blue mussel (*M. galloprovincialis*) [66,



FIGURE 3: The effects of nanoplastics exposure on the gut-brain axis.

103]. These toxicological studies proved significant genotoxicity after treatment with micro- and nanoplastics. Similar results were achieved after treatment of marine bivalve hemocytes with polystyrene nanoplastics in a physiological environment [104]. The presence of the mentioned nanoplastics increased cellular damage and reactive oxygen species (ROS) production compared to the control medium. Dysregulation of p38 mitogen-activated protein kinase (MAPK) signaling was also noted. Moreover, the connection between oxidative digestion and ROS generation and induced stress was established, including the harmful influence of nanomaterials on the lysosome function. Nanoplastics can prompt lysosomal harm following the following mechanism of action: (1) direct harm after ingestion of plastic particles in the cell through endocytosis or permeation (<50 nm) and attempts to process an unfamiliar body which might bring about lysosome interruption [105, 106]. The creation of ROS induced by the presence of nanoplastics impacts lysosomal membranes that are very sensitive to the oxidative effects of released ROS. However, the exact pathway within the mechanism of nanoplastics action on lysosome function has not yet been discovered and demands more toxicological studies (in vitro and in vivo).

Mitochondria are organelles in which most of the intracellular creation of ROS happens. Mitochondrial layer potential instability can lead to excessive generation of ROS through single-electron transporters (for example, cytochromes and iron-sulfur protein) and other oxidases [107].

From a mechanistic point of view, excessive exposure to oxidative stress results in a critical level of ROS, thus making the mitochondrial channels open. Consequently, mitochondrial membrane potential will collapse. Furthermore, the authors reported mitochondrial brokenness during the treatment of monogonont rotifer (Brachionus koreanus) with nanoplastics. During this process, the charge collapse has been denoted as a critical point [107, 108]. These results require further investigation as nanoplastics cannot directly accumulate in mitochondria. The mentioned studies in rotifer *B. koreanus* revealed that $0.5 \,\mu$ m polystyrene microbeads decreased the mitochondrial layer potential significantly. Furthermore, it was suggested that nanoplastics might influence the mitochondrial external layer (membrane) by implication through expanded ROS presence in cellular compartments beyond nearby mitochondria [107]. Exposure of mitochondria to unnecessary oxidative stress in the cytosol can trigger the activation of several mitochondrial Na/K



FIGURE 4: The intracellular side effects of nanoplastics.

transmembrane channels. Ion transition *via* an expanded membrane channel may further promote the breakdown of the mitochondrial layer potential and deliver ROS through a stimulated ROS scavenging mechanism [107].

Mitochondrial layer harm caused by nanoplastic presence is also related to the particle size. Nanoplastics could be less than 5 nm, implying that nanoplastics could force more extreme harm on mitochondria [108]. In that regard, nanoplastics activated mitochondria interruption in *D. magna* and an assortment of human cell lines [109, 110]. In addition, amine-functionalized polystyrene nanoplastics prompted essentially more extreme mitochondrial damage than the nonchanged ones. Therefore, this implies that the surface charge of nanoplastics takes on a significant role in causing adverse effects on mitochondrial function [110].

DNA strand damage might happen over the oxidative path. There is a current debate on whether nanoplastics are prompting DNA harm generally since just two articles to date have revealed that DNA damage happens in aquatic living models [103]. DNA strands could be broken down by polystyrene nanoparticles (20 nm) in the hemocytes of *S. plana*. Additionally, it was detected in mussels treated with polyethylene nanoparticles [111]. The two reports proposed that the noticed DNA harm could be related to the oxidative damage caused by nanoplastics.

Genotoxicity caused by nanoplastics, including damage to DNA strands, has also been under investigation. Nevertheless, an illustration from more extensive nanotoxicological research has shown that some nanoparticles (such as nanoplastics, titanium dioxide) can cause DNA damage through two systems: (I) direct interaction among nanoparticles and DNA and (II) aberrant DNA harm brought about by nanoparticle-produced ROS. An intracellular unique imaging study showed that cationic functionalized PS nanoplastics could result in a delayed G0/G1 work in the cell cycle during mitosis in NIH 3T3 cells [112]. The results demonstrated the potential for DNA harm. Paget and coworkers indicated that unmodified PS nanoplastics did not induce genotoxicity after $8.1 \,\mu g/cm$ discharge during one hour of exposure [113]. There is proof that nanoplastics can prompt DNA strands to break down predominately depending on the nanoplastics size and their surface charge.

Lipid peroxidation (LPO) is a self-sustaining reaction chain triggered by excessive ROS generation; the result is antioxidant damage to cell membranes and other fatty structures [114]. It has now become a topic of discussion as to whether the reaction and cell membranes in the exposure of nanoplastics are triggered on this human scale. In a study by Barboza and coworkers [115] in European countries, a significant increase in LPO in brain and muscle tissue has been noted to occur naturally after the release of nanoplastics in 0.69 mg/L doses within 24 hours. Another review showed that LPO was not essentially induced in hemocytes of the sea mussel Mytilus spp. after 7 days of nanoplastic exposure. Moreover, in S. plan, LPO levels in gills and stomach-related organs were either decreased or not substantially increased after 7 days of exposure to polystyrene nanoparticles. The authors proposed that such an outcome could be explained by an adequate limitation of antioxidant systems to defend the form of the organism against LPO damage. But it would be at least prudent to consider this molecular event as the main event induced by the nanoplastics [15]. This approach would be additionally upheld with nearly 10% of surveyed examinations showing that LPO and resulting harms were prompted by nanoplastic openness, subsequently proposing the likelihood that LPO might be a harm endpoint shared by the plastic particles in both the miniature and nanosize ranges.

2.8. Effects of Nanoplastic on Oxidative Stress. Oxidative pressure happens when there is unevenness between the creation of ROS and cancer prevention agent-based detoxification/balance. The antioxidant process is very complicated and includes different antioxidant compounds such as C, E, and D3 vitamins, respectively. Different enzymatic pathways are occupied by several manufacturers to remove oxidants, or antioxidants, as well as reactive nitrogen species and ROS. This goes past the extent of the previous framework review, and this translation is past the extent of the antioxidant system framework survey. Sies and colleagues explored oxidative pressure pathways after exposure to nanoplastics [116].

Antioxidant enzymes, such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx), play an important role in protecting tissues against ROS damage [117]. For example, a complex ROS scavenging framework in natural resistant cells uses SOD and CAT enzymatic systems to catalyze the conversion of superoxide anion radical (O_2^-) and hydrogen peroxide (H_2O_2) into water (H_2O) and oxygen (O_2) as metabolites, respectively [118]. CAT and SOD-based antioxidant mechanisms play a significant cellular role in the adverse effects of ROS that could be used as a sensor of xenobiotic-mediated oxidative stress [119, 120]. Exposure to nanoplastics could expand the grouping of ROS in the live form as we recently described, and the SOD and CAT antioxidative enzymes would respond to such a signal.

The determination of antioxidant properties related to nanoplastics was found in different animal models, such as water fleas (*Daphnia magna*) [121], rotifers (*B. koreanus*), mammals (*M. musculus*) [122], and fish (*D. rerio*) [122, 123]. This peculiarity proposes the presence of initial factors to start the disturbance of oxidative status induced by microplastic openness. Because of nanoplastic particle entry, an increase in oxidative stress parameter levels of approximately 50% was observed both in *in vitro* and *in vivo* studies. Those levels of oxidative imbalance have been reported to set the oxidative damage as one of the most decisive factors accompanied by adverse effects among all nanoplasticinduced poisonousness endpoints.

It was plainly shown that ROS can instigate or intervene in the enactment of the MAPK pathways [124]. However, definite instruments of MAPK downstream pathway guidelines remain unclear. Some studies revealed that activation of MAPK signaling could activate antioxidant response element-interceded quality articulation through the simultaneous component increase Nrf2 (nuclear factor erythroid 2related factor 2) activity, and apparently, MAPK enactment by ROS can be downregulated by concurrent expansion in Nrf2 action [125]. In contrast, the drawn-out initiation of MAPK framework with expanded ROS generation can activate different adverse pathways (such as autophagy), basically connected with the hindrance of extracellular-signalregulated kinase (ERK) and enactment of p38 MAPK parts. It was recently detailed that brominated fire-resistant prompted oxidative pressure in two copepod species enacted the MAPK pathway [126], and it was concluded that MAPK pathways assume a synergistic part with the Nrf2/Keap1 pathway in the oxidative pressure reaction in the scallops presented to benzo(a)pyrene [127].

In the following stage in the oxidative pressure occasion series, it has been seen that nanoplastic openness additionally enacts redox-sensitive signaling pathways such as MAPKs. MAPK downstream pathways were started postnanoplastic openness in the marine copepod *P. nana* and Mitten crab *E. sinensis* [27, 53]. Jeong and colleagues showed that the phosphorylation level increment of ERK and p38 kinase had a positive relationship with intracellular ROS age-level post-microplastic openness in *P. nana* [127]. The factor Nrf-2 was extended after openness to nanoplastics. As a consequence, nanoplastics set off breath bursts plausibly acted using ERK and p38 MAPK pathways activated by Nrf-2 [128].

The activation of the MAPK pathway is related to the size of a nanoparticle. Going forward, the MAPK pathway is activated after exposure to nanoplastics [66, 109, 129]. It was recommended that plastics with a size of 50 nm could create more serious oxidative pressure in comparison to 6μ m plastic, causing higher phosphorylation levels of p38 MAPKs in both *P. nana* and *B. koreanus* [130]. Furthermore, the surface charge of nanoplastic particles seems essential in the enactment of MAPK overflow, as plain/local polystyrene nanoparticles prompted the activation of p38 and c-Jun N-terminal kinase (JNK) alongside ROS enlistment, while adversely charged polystyrene nanoparticles activated p38 and JNK autonomously [109].

A free radical is a group of atoms with more than one unconnected electron. Generally, in the biological system, various radicals are synthesized from oxygen and are together referred to as ROS [131]. Several reports pointed out that the generation of ROS happens at release time, after exposure to the nanoplastics of different sizes and characteristics [108, 132, 133]. The remarkable nanoplastic process of ROS generation has two stages; intracellular and

extracellular. Electrocellular ROS generation induced by nanoplastics connects with the level of enduring (maturing) process during plastic polymers' openness to the components in the environment [134, 135]. Enduring cycles of nanoplastics include the concurrent or individual activity of photooxidation and warm oxidation, as well as UV radiation prompting synthetic changes on the outer layer of plastic polymers [136]. Photooxidation and/or UV light radiation can prompt the development of free radicals on nanoplastic surfaces as essential items through various pathways: deduction of a hydrogen atom from the macromolecular chain, or through cross-linking response (expansion to an unsaturated carbon chain bunch) [137]. Consequently, with the creation of free radicals in the polymer series chain, the second type of polymer chain is created due to appropriate conditions in the atmosphere [138]. This enduring prompted extracellular free radical release could be one of the potential mechanisms regarding the substantial increment of ROS amount observed after the cellular entry of the matured nanoplastics [67].

Nevertheless, the most primitive nanoplastics can trigger the abundant intracellular ROS production. This peculiarity was accounted for by utilizing a wide cluster of model frameworks, from mammalian cell lines to marine invertebrates and living fish models [134, 138]. Nanoplastics can be immersed by a cell through the processes of endocytosis or pinocytosis [132]. When inside a phagocyte, these nanoplastics seem to set off the natural insusceptible protection systems and are treated as unfamiliar substances [139]. Moreover, when a microplastic debases into nanosized particles, their surface-to-mass proportion increases and enables the nanoplastics to enter directly through the lipid layers. Changes in surface charge potential and mass/surface proportion in nanoplastics likewise empower simpler retention of free revolutionaries as well as more straightforward movement through the films, introducing areas of strength between the molecule size and ROS age potential: the more modest the molecule, the higher ROS age potential [135]. Size-subordinate contrasts in the harmfulness of nanoplastics are upheld by a few examinations [136].

3. Conclusions

In recent years, the problem of nanoplastics has gained importance both in the scientific community and in the general public. Numerous studies have revealed that nanoplastics are a ubiquitous pollutant of the human environment, which, in addition to environmental pollution, poses a serious threat to the organisms in them. Namely, numerous studies examining the organisms living in those ecosystems have revealed that nanoplastics can cause blockage of the digestive tract, inhibition of growth, reproduction disability, and ultimately cause death.

The presence of nanoplastics in fish and many other marine species has shown that nanoplastics can easily reach the human body through the food chain. The impact of nanoplastics, however, cannot be observed only at the level of a particular organism. Namely, nanoplastics can completely change the population structure of a particular species and ultimately lead to a change in the dynamics of the entire ecosystem. What is even more worrying is the potential impact of nanoplastics on humans themselves. By consuming marine organisms that contain accumulated nanoplastics, a human takes nanoplastic particles into his body. Depending on the concentration ingested, nanoplastics can have various negative consequences on human health as listed above. Many studies have shown that nanoplastics in the human body can cause an inflammatory response and disruption of the intestinal microbiome and cause many other side effects including oxidative stress.

Finally, it can be concluded that nanoplastics in the human environment have multiple negative impacts, and this problem should be approached with extreme seriousness and attention. A strategic plan should be made to solve it, including individuals, as well as local and global communities.

Conflicts of Interest

The authors confirm that this article content has no conflict of interest.

Authors' Contributions

Nenad Joksimovic and Dragica Selakovic contributed equally to this work.

Acknowledgments

This work was supported by the Faculty of Medical Sciences, University of Kragujevac, Serbia (JP 01/19), and the Serbian Ministry of Education, Science and Technological Development (No. 451-03-68/2020-14/200122).

References

- S. I. Zandalinas, F. B. Fritschi, and R. Mittler, "Global warming, climate change, and environmental pollution: recipe for a multifactorial stress combination disaster," *Trends in Plant Science*, vol. 26, no. 6, pp. 588–599, 2021.
- [2] F. O. Ajibade, B. Adelodun, K. H. Lasisi et al., "Environmental pollution and their socioeconomic impacts," in *Microbe Mediated Remediation of Environmental Contaminants, Chapter 25 - Environmental pollution and their socioeconomic impacts*, pp. 321–354, Woodhead Publishing, 2021.
- [3] L. Lebreton and A. Andrady, "Future scenarios of global plastic waste generation and disposal," *Palgrave Communications*, vol. 5, no. 1, 2019.
- [4] M. Ghayebzadeh, H. Aslani, H. Taghipour, and S. Mousavi, "Estimation of plastic waste inputs from land into the Caspian Sea: a significant unseen marine pollution," *Marine Pollution Bulletin*, vol. 151, p. 110871, 2020.
- [5] A. Turner, C. Wallerstein, R. Arnold, and D. Webb, "Marine pollution from pyroplastics," *Science of the Total Environment*, vol. 694, p. 133610, 2019.
- [6] N. Anbuselvan, D. Senthil Nathan, and M. Sridharan, "Heavy metal assessment in surface sediments off Coromandel Coast of India: implication on marine pollution," *Marine Pollution Bulletin*, vol. 131, pp. 712–726, 2018.

- [7] I. Ellen Napper and R. C. Thompson, "Plastic debris in the marine environment: history and future challenges," *Challenges*, vol. 4, no. 6, p. 1900081, 2020.
- [8] Plastics Europe 2012, "Plastics the Facts 2012: an analysis of European plastics production, demand and waste data for 2011," 2012, https://plasticseurope.org/fr/wp-content/ uploads/sites/2/2021/11/2012-Plastics-the-facts.pdf.
- [9] A. Kurtela and N. Antolović, "The problem of plastic waste and microplastics in the seas and oceans: impact on marine organisms," *Croatian Journal of Fisheries*, vol. 77, no. 1, pp. 51–56, 2019.
- [10] M. Smith, D. C. Love, C. M. Rochman, and R. A. Neff, "Microplastics in seafood and the implications for human health," *Current Environmental Health Reports*, vol. 5, no. 3, pp. 375–386, 2018.
- [11] M. F. G. Martinico-Perez, H. Schandl, T. Fishman, and H. Tanikawa, "The socio-economic metabolism of an emerging economy: monitoring progress of decoupling of economic growth and environmental pressures in the Philippines," *Ecological Economics*, vol. 147, pp. 155–166, 2018.
- [12] M. McCartney, "The China-Pakistan Economic Corridor (CPEC): infrastructure, social savings, spillovers, and economic growth in Pakistan," *Eurasian Geography and Economics*, vol. 63, no. 2, pp. 180–211, 2022.
- [13] Plastics Europe 2017, "Plastics the facts 2017: an analysis of European plastics production, Demand and Waste Data for 2017," 2018, https://plasticseurope.org/wp-content/uploads/ 2021/10/2017-Plastics-the-facts.pdf.
- [14] D. K. A. Barnes, F. Galgani, R. C. Thompson, and M. Barlaz, "Accumulation and fragmentation of plastic debris in global environments," *Philosophical Transactions of the Royal Society B*, vol. 364, no. 1526, pp. 1985–1998, 2009.
- [15] S. B. Kurniawan, S. R. Sheikh Abdullah, M. Fauzul Imron, and N. Ismail, "Current state of marine plastic pollution and its technology for more eminent evidence: a review," *Journal of Cleaner Production*, vol. 278, p. 123537, 2021.
- [16] M. R. Gregory, "Plastic 'scrubbers' in hand cleansers: a further (and minor) source for marine pollution identified," *Marine Pollution Bulletin*, vol. 32, no. 12, pp. 867–871, 1996.
- [17] C. J. Moore, "Synthetic polymers in the marine environment: a rapidly increasing, long-term threat," *Environmental Research*, vol. 108, no. 2, pp. 131–139, 2008.
- [18] L. A. Amaral-Zettler, E. R. Zettler, and T. J. Mincer, "Ecology of the plastisphere," *Nature Reviews Microbiology*, vol. 18, no. 3, pp. 139–151, 2020.
- [19] E. L. Ng, H. Esperanza, S. M. Lwanga et al., "An overview of microplastic and nanoplastic pollution in agroecosystems," *Science of the Total Environment*, vol. 627, pp. 1377–1388, 2018.
- [20] M. M. Allouzi, D. Y. Tang, K. W. Chew et al., "Micro (nano) plastic pollution: the ecological influence on soil-plant system and human health," *Science of the Total Environment*, vol. 788, p. 147815, 2021.
- [21] V. Menicagli, M. Ruffini Castiglione, E. Balestri et al., "Early evidence of the impacts of microplastic and nanoplastic pollution on the growth and physiology of the seagrass *Cymodocea nodosa*," *Science of the Total Environment*, vol. 838, Part 3, p. 156514, 2022.
- [22] I. Ferreira, C. Venâncio, I. Lopes, and M. Oliveira, "Nanoplastics and marine organisms: what has been studied?,"

Environmental Toxicology and Pharmacology, vol. 67, pp. 1–7, 2019.

- [23] S. Al-Thawadi, "Microplastics and nanoplastics in aquatic environments: challenges and threats to aquatic organisms," *Arabian Journal for Science and Engineering*, vol. 45, no. 6, pp. 4419–4440, 2020.
- [24] M. B. Paul, V. Stock, J. Cara-Carmona et al., "Micro- and nanoplastics – current state of knowledge with the focus on oral uptake and toxicity," *Advances*, vol. 2, no. 10, pp. 4350–4367, 2020.
- [25] C. Q. Yong, S. Valiyaveettil, and B. L. Tang, "Toxicity of microplastics and nanoplastics in mammalian systems," *International Journal of Environmental Research and Public Health*, vol. 17, no. 5, p. 1509, 2020.
- [26] K. Enders, A. Käppler, O. Biniasch et al., "Tracing microplastics in aquatic environments based on sediment analogies," *Scientific Reports*, vol. 9, no. 1, p. 15207, 2019.
- [27] I. Jakubowicz, J. Enebro, and N. Yarahmadi, "Challenges in the search for nanoplastics in the environment-a critical review from the polymer science perspective," *Polymer Testing*, vol. 93, p. 106953, 2021.
- [28] H. Cai, E. Genbo Xu, F. Du, R. Li, J. Liu, and H. Shi, "Analysis of environmental nanoplastics: progress and challenges," *Chemical Engineering Journal*, vol. 410, p. 128208, 2021.
- [29] M. Eriksen, L. C. Lebreton, H. S. Carson et al., "Plastic pollution in the world's oceans: more than 5 trillion plastic pieces weighing over 250,000 tons afloat at sea," *PLoS One*, vol. 9, no. 12, pp. 1–15, 2014.
- [30] I. L. Nerland, C. Halsband, I. Allan, and K. V. Thomas, *Microplastics in marine environments: occurrence, distribution and effects*, pp. 1–73, Report made for the Norwegian Environment Agency, 2014.
- [31] L. M. Rios Mendoza and M. Balcer, "Microplastics in freshwater environments: a review of quantification assessment," *Analytical Chemistry*, vol. 113, pp. 402–408, 2019.
- [32] A. Andrady, "Microplastics in the marine environment," *Marine Pollution Bulletin*, vol. 62, no. 8, pp. 1596–1605, 2011.
- [33] M. González-Pleiter, M. Tamayo-Belda, G. Pulido-Reyes et al., "Secondary nanoplastics released from a biodegradable microplastic severely impact freshwater environments," *Environmental Science: Nano*, vol. 6, pp. 1382– 1392, 2019.
- [34] L. P. Domínguez-Jaimes, E. I. Cedillo-González, E. Luévano-Hipólito, J. D. Acuña-Bedoya, and J. M. Hernández-López, "Degradation of primary nanoplastics by photocatalysis using different anodized TiO₂ structures," *Journal of Hazardous Materials*, vol. 413, p. 125452, 2021.
- [35] L. M. Hernandez, N. Yousefi, and N. Tufenkji, "Are there nanoplastics in your personal care products?," *Environmental Science & Technology Letters*, vol. 4, no. 7, pp. 280–285, 2017.
- [36] A. Keswani, D. M. Oliver, T. Gutierrez, and R. S. Quilliam, "Microbial hitchhikers on marine plastic debris: human exposure risks at bathing waters and beach environments," *Marine Environmental Research*, vol. 118, pp. 10–19, 2016.
- [37] L. M. R. Mendoza, H. Karapanagioti, and N. R. Alvarez, "Micro(nanoplastics) in themarine environment: current knowledge and gaps," *Current Opinion in Environmental Science & Health*, vol. 1, pp. 47–51, 2018.
- [38] X. Xiong, K. Zhang, X. Chen, H. Shi, Z. Luo, and C. Wu, "Sources and distribution of microplastics in China's largest

inland lake - Qinghai Lake," *Environmental Pollution*, vol. 235, pp. 899–906, 2018.

- [39] D. Materić, M. Peacock, J. Dean et al., "Presence of nanoplastics in rural and remote surface waters," *Environmental Research Letters*, vol. 17, no. 5, p. 54036, 2022.
- [40] A. Wahl, C. Le Juge, M. Davranche et al., "Nanoplastic occurrence in a soil amended with plastic debris," *Chemosphere*, vol. 262, p. 127784, 2021.
- [41] M. O. Rodrigues, N. Abrantes, F. J. M. Gonçalves, H. Nogueira, J. C. Marques, and A. M. M. Gonçalves, "Spatial and temporal distribution of microplastics in water and sediments of a freshwater system (Antua River, Portugal)," *Science of the Total Environment*, vol. 633, pp. 1549–1559, 2018.
- [42] X. Wen, C. Du, P. Xu et al., "Microplastic pollution in surface sediments of urban water areas in Changsha, China: abundance, composition, surface textures," *Marine Pollution Bulletin*, vol. 136, pp. 414–423, 2018.
- [43] A. Cozar, E. Martí, C. Duarte et al., "The Arctic Ocean as a dead end for floating plastics in the North Atlantic branch of the thermohaline circulation," *Science Advances*, vol. 3, no. 4, article e1600582, 2017.
- [44] I. Peeken, S. Primpke, B. Beyer et al., "Arctic sea ice is an important temporal sink and means of transport for microplastic," *Nature Communications*, vol. 9, no. 1, p. 1505, 2018.
- [45] D. Materić, H. AstridKjær, P. Vallelonga, J.-L. Tison, T. Röckmann, and R. Holzinger, "Nanoplastics measurements in Northern and Southern polar ice," *Environmental Research*, vol. 208, p. 112741, 2022.
- [46] K. Mattsson, L. A. Hansson, and T. Cedervall, "Nanoplastics in the aquatic environment," *Environmental Science: Processes & Impacts*, vol. 17, no. 10, pp. 1712– 1721, 2015.
- [47] S. Wagner and T. Reemtsma, "Things we know and don't know about nanoplastic in the environment," *Nature Nanotechnology*, vol. 14, no. 4, pp. 300-301, 2019.
- [48] J. R. Jambeck, R. Geyer, C. Wilcox et al., "Plastic waste inputs from land into the ocean," *Science*, vol. 347, no. 6223, pp. 768–771, 2015.
- [49] N. B. Hartmann, T. Hüffer, R. C. Thompson et al., "Are we speaking the same language? Recommendations for a definition and categorization framework for plastic debris," *Environmental Science & Technology*, vol. 53, no. 3, pp. 1039– 1047, 2019.
- [50] S. Lambert and M. Wagner, "Characterisation of nanoplastics during the degradation of polystyrene," *Chemosphere*, vol. 145, pp. 265–268, 2016.
- [51] D. L. Watters, M. M. Yoklavich, M. S. Love, and D. M. Schroeder, "Assessing marinedebris in deep seafloor habitats off California," *Marine Pollution Bulletin*, vol. 60, p. 131e138, 2010.
- [52] Z. Lin, T. Jin, T. Zou et al., "Current progress on plastic/ microplastic degradation: fact influences and mechanism," *Environmental Pollution*, vol. 304, p. 119159, 2022.
- [53] V. Hidalgo-Ruz, L. Gutow, R. C. Thompson, and M. Thiel, "Microplastics in the marine environment: a review of the methods used for identification and quantification," *Environmental Science & Technology*, vol. 46, no. 6, pp. 3060–3075, 2012.
- [54] N. Lucas, C. Bienaime, C. Belloy, M. Queneudec, F. Silvestre, and J.-E. Nava-Saucedo, "Polymer biodegradation: mecha-

nisms and estimation techniques - a review," *Chemosphere*, vol. 73, no. 4, pp. 429–442, 2008.

- [55] A. Trishul and D. Mukesh, "Biodegradation of aliphatic and aromatic polycarbonates," *Macromolecular Bioscience*, vol. 8, pp. 14–24, 2010.
- [56] J. Allan, S. Belz, A. Hoeveler et al., "Regulatory landscape of nanotechnology and nanoplastics from a global perspective," *Regulatory Toxicology and Pharmacology*, vol. 122, article 104885, 2020.
- [57] J. P. da Costa, "Nanoplastics in the environment," *Plastics* and the Environment, vol. 47, p. 82, 2018.
- [58] A. Koelmans, B. Albert, and W. J. Ellen, "Nanoplastics in the aquatic environment. critical review," *Marine Anthropogenic Litter*, pp. 325–340, 2015.
- [59] K. Pastor, B. Isić, M. Horvat et al., "Omnipresence of plastics: a review of the microplastic sources and detection methods," *Journal of Faculty of Civil Engineering*, vol. 39, pp. 29–43, 2021.
- [60] M. Rujnić-Sokele, "Plastični otpad-globalni ekološki problem," *Polimeri.*, vol. 36, pp. 1-2, 2015.
- [61] N. Welden, "Micro plastics: emerging contaminants requiring multilevel management," in *Waste (Second Edition)*, pp. 405–424, Academic Press, 2019.
- [62] S. A. Mason, D. Garneau, R. Sutton et al., "Microplastic pollution is widely detected in US municipal wastewater treatment plant effluent," *Environmental Pollution*, vol. 218, pp. 1045–1054, 2016.
- [63] M. Ban, R. Shimoda, and J. Chen, "Investigation of nanoplastic cytotoxicity using SH-SY5Y human neuroblastoma cells and polystyrene nanoparticles," *Toxicology In Vitro*, vol. 76, p. 105225, 2021.
- [64] Q. Shi, J. Tang, L. Wang, R. Liu, and J. P. Giesy, "Combined cytotoxicity of polystyrene nanoplastics and phthalate esters on human lung epithelial A549 cells and its mechanism," *Ecotoxicology and Environmental Safety*, vol. 213, p. 112041, 2021.
- [65] A. Wegner, E. Besseling, E. M. Foekema, P. Kamermans, and A. A. Koelmans, "Effects of nanopolystyrene on the feeding behavior of the blue mussel (Mytilus edulis L.)," *Environmental Toxicology and Chemistry*, vol. 31, no. 11, pp. 2490–2497, 2012.
- [66] I. Brandts, M. Teles, A. P. Gonçalves et al., "Effects of nanoplastics on *Mytilus galloprovincialis* after individual and combined exposure with carbamazepine," *Science of the Total Environment*, vol. 643, pp. 775–784, 2018.
- [67] L. Canesi, C. Ciacci, R. Fabbri et al., "Interactions of cationic polystyrene nanoparticles with marine bivalve hemocytes in a physiological environment: role of soluble hemolymph proteins," *Environmental Research*, vol. 150, pp. 73–81, 2016.
- [68] M. Cole and T. S. Galloway, "Ingestion of nanoplastics and microplastics by pacific oyster larvae," *Environmental Science* & *Technology*, vol. 49, no. 24, pp. 14625–14632, 2015.
- [69] C. González-Fernández, K. Tallec, N. Le Goïc et al., "Cellular responses of Pacific oyster (*Crassostrea gigas*) gametes exposed *in vitro* to polystyrene nanoparticles," *Chemosphere*, vol. 208, pp. 764–772, 2018.
- [70] E. Besseling, B. Wang, M. Lürling, and A. A. Koelmans, "Nanoplastic affects growth of S. obliquus and reproduction of D. magna," *Environmental Science & Technology*, vol. 48, no. 20, pp. 12336–12343, 2014.

- [71] A. Banerjee and W. L. Shelver, "Micro- and nanoplastic induced cellular toxicity in mammals: a review," *Science of the Total Environment*, vol. 755, Part 2, p. 142518, 2021.
- [72] J. Reinholz, C. Diesler, S. Schöttler et al., "Protein machineries defining pathways of nanocarrier exocytosis and transcytosis," *Acta Biomaterialia*, vol. 71, pp. 432–443, 2018.
- [73] A. Banerjee, J. Qi, R. Gogoi, J. Wong, and S. Mitragotri, "Role of nanoparticle size, shape and surface chemistry in oral drug delivery," *Journal of Controlled Release*, vol. 238, pp. 176–185, 2016.
- [74] M. Forte, G. Lachetta, M. Tussellino et al., "Polystyrene nanoparticles internalization in human gastric adenocarcinoma cells," *Toxicology In Vitro*, vol. 31, pp. 126–136, 2016.
- [75] C. Cortés, J. Domenech, M. Salazar, S. Pastor, R. Marcos, and A. Hernández, "Nanoplastics as a potential environmental health factor: effects of polystyrene nanoparticles on human intestinal epithelial Caco-2 cells," *Environmental Science*. *Nano*, vol. 7, no. 1, pp. 272–285, 2020.
- [76] V. Stock, L. Bohmert, E. Lisicki et al., "Uptake and effects of orally ingested polystyrene microplastic particles in vitro and in vivo," *Archives of Toxicology*, vol. 93, no. 7, pp. 1817–1833, 2019.
- [77] S. A. Kulkarni and S. S. Feng, "Effects of particle size and surface modification on cellular uptake and biodistribution of polymeric nanoparticles for drug delivery," *Pharmaceutical Research*, vol. 30, no. 10, pp. 2512–2522, 2013.
- [78] G. J. Mahler, M. B. Esch, E. Tako et al., "Oral exposure to polystyrene nanoparticles affects iron absorption," *Nature Nanotechnology*, vol. 7, no. 4, pp. 264–271, 2012.
- [79] B. Wu, X. Wu, S. Liu, Z. Wang, and L. Chen, "Size-dependent effects of polystyrene microplastics on cytotoxicity and efflux pump inhibition in human Caco-2 cells," *Chemosphere*, vol. 221, pp. 333–341, 2019.
- [80] A. P. Walczak, P. J. Hendriksen, R. A. Woutersen et al., "Bioavailability and biodistribution of differently charged polystyrene nanoparticles upon oral exposure in rats," *Journal of Nanoparticle Research*, vol. 17, no. 5, p. 231, 2015.
- [81] A. P. Walczak, E. Kramer, P. J. Hendriksen et al., "Translocation of differently sized and charged polystyrene nanoparticles in in vitro intestinal cell models of increasing complexity," *Nanotoxicology*, vol. 9, no. 4, pp. 453–461, 2015.
- [82] S. C. Baos, D. B. Phillips, L. Wildling, T. J. McMaster, and M. Berry, "Distribution of sialic acids on mucins and gels: a defense mechanism," *Biophysical Journal*, vol. 102, no. 1, pp. 176–184, 2012.
- [83] E. Y. Chen, Y. C. Wang, C. S. Chen, and W. C. Chin, "Functionalized positive nanoparticles reduce mucin swelling and dispersion," *PLoS One*, vol. 5, p. 15434, 2010.
- [84] A. des Rieux, E. G. Ragnarsson, E. Gullberg, V. Préat, Y. J. Schneider, and P. Artursson, "Transport of nanoparticles across an in vitro model of the human intestinal follicle associated epithelium," *European Journal of Pharmaceutical Sciences*, vol. 25, no. 4-5, pp. 455–465, 2005.
- [85] A. Thubagere and B. M. Reinhard, "Nanoparticle-induced apoptosis propagates through hydrogen-peroxide-mediated bystander killing: insights from a human intestinal epitheliumin vitromodel," ACS Nano, vol. 4, no. 7, pp. 3611–3622, 2010.
- [86] I. Inkielewicz-Stepniak, L. Tajber, G. Behan et al., "The role of mucin in the toxicological impact of polystyrene nanoparticles," *Materials*, vol. 11, no. 5, p. 724, 2018.

- [87] D. Allen, S. Allen, S. Abbasi et al., "Microplastics and nanoplastics in the marine-atmosphere environment," *Nature Reviews Earth & Environment*, vol. 3, no. 6, pp. 393–405, 2022.
- [88] S. L. Wright and F. J. Kelly, "Plastic and human health: a micro issue?," *Environmental Science and Technology*, vol. 51, no. 12, pp. 6634–6647, 2017.
- [89] C. D. Dong, C. W. Chen, Y. C. Chen, H. H. Chen, J. S. Lee, and C. H. Lin, "Polystyrene microplastic particles: in vitro pulmonary toxicity assessment," *Journal of Hazardous Materials*, vol. 385, p. 121575, 2020.
- [90] S. L. Lim, C. T. Ng, L. Zou et al., "Targeted metabolomics reveals differential biological effects of nanoplastics and nanoZnO in human lung cells," *Nanotoxicology*, vol. 13, no. 8, pp. 1117–1132, 2019.
- [91] M. Xu, G. Halimu, Q. Zhang et al., "Internalization and toxicity: a preliminary study of effects of nanoplastic particles on human lung epithelial cell," *Science of the Total Environment*, vol. 694, p. 133794, 2019.
- [92] T. Xia, M. Kovochich, M. Liong, J. I. Zink, and A. E. Nel, "Cationic polystyrene nanosphere toxicity depends on cellspecific endocytic and mitochondrial injury pathways," ACS Nano, vol. 2, no. 1, pp. 85–96, 2008.
- [93] H. W. Chiu, T. Xia, Y. H. Lee, C. W. Chen, J. C. Tsai, and Y. I. Wang, "Cationic polystyrene nanospheres induce autophagic cell death through the induction of endoplasmic reticulum stress," *Nanoscale*, vol. 7, no. 2, pp. 736– 746, 2015.
- [94] N. R. Yacobi, N. Malmstadt, F. Fazlollahi et al., "Mechanisms of alveolar epithelial translocation of a defined population of nanoparticles," *American Journal of Respiratory Cell and Molecular Biology*, vol. 42, no. 5, pp. 604–614, 2010.
- [95] B. Toussaint, B. Raffael, A. A. Loustau et al., "Review of micro- and nanoplastic contamination in the food chain," *Food Additives and Contaminants*, vol. 36, no. 5, pp. 639– 673, 2019.
- [96] Y. Huang, K. Ki Wong, W. Li et al., "Characteristics of nanoplastics in bottled drinking water," *Journal of Hazardous Materials*, vol. 15, p. 127404, 2022.
- [97] W. Grodzicki, K. Dziendzikowska, J. Gromadzka-Ostrowska, and M. Kruszewski, "Nanoplastic impact on the gut-brain axis: current knowledge and future directions," *International Journal of Molecular Sciences*, vol. 22, no. 23, p. 12795, 2021.
- [98] M. Carabotti, A. Scirocco, M. A. Maselli, and C. Severi, "The gut-brain axis: interactions between enteric microbiota, central and enteric nervous systems," *Annals of Gastroenterology*, vol. 28, no. 2, pp. 203–209, 2015.
- [99] J. F. Cryan, K. J. O'riordan, C. S. M. Cowan et al., "The microbiota-gut-brain axis," *Physiological Reviews*, vol. 99, no. 4, pp. 1877–2013, 2019.
- [100] L. H. Morais, H. L. Schreiber, and S. K. Mazmanian, "The gut microbiota-brain axis in behaviour and brain disorders," *Nature Reviews. Microbiology*, vol. 19, no. 4, pp. 241–255, 2021.
- [101] S. Nikolic, M. Gazdic-Jankovic, G. Rosic et al., "Orally administered fluorescent nanosized polystyrene particles affect cell viability, hormonal and inflammatory profile, and behavior in treated mice," *Environmental Pollution*, vol. 305, p. 119206, 2022.

- [102] M. N. Moore, J. Icarus Allen, and A. McVeigh, "Environmental prognostics: an integrated model supporting lysosomal stress responses as predictive biomarkers of animal health status," *Marine Environmental Research*, vol. 61, no. 3, pp. 278–304, 2006.
- [103] C. G. Avio, S. Gorbi, M. Milan et al., "Pollutants bioavailability and toxicological risk from microplastics to marine mussels," *Environmental Pollution*, vol. 198, pp. 211–222, 2015.
- [104] S. T. Stern, P. P. Adiseshaiah, and R. M. Crist, "Autophagy and lysosomal dysfunction as emerging mechanisms of nanomaterial toxicity," *Particle and Fibre Toxicology*, vol. 9, no. 1, p. 20, 2012.
- [105] N. von Moos, P. Burkhardt-Holm, and A. Kohler, "Uptake and effects of microplastics on cells and tissue of the blue mussel Mytilus edulis L. after an experimental exposure," *Environmental Science & Technology*, vol. 46, no. 20, pp. 11327–11335, 2012.
- [106] I. Fiorentino, R. Gualtieri, V. Barbato et al., "Energy independent uptake and release of polystyrene nanoparticles in primary mammalian cell cultures," *Experimental Cell Research*, vol. 330, no. 2, pp. 240–247, 2015.
- [107] D. B. Zorov, M. Juhaszova, and S. J. Sollott, "Mitochondrial ROS-induced ROS release: an update and review," *Biochimica et Biophysica Acta - Bioenergetics*, vol. 1757, no. 5-6, pp. 509–517, 2006.
- [108] C. Bum Jeong, E. Ji Won, H. Min Kang et al., "Microplastic size-dependent toxicity, oxidative stress induction, and p-JNK and p-p38 activation in the monogonont rotifer (Brachionus koreanus)," *Environmental Science & Technology*, vol. 50, no. 16, pp. 8849–8857, 2016.
- [109] W. Lin, R. Jiang, S. Hu et al., "Investigating the toxicities of different functionalized polystyrene nanoplastics on *Daphnia magna*," *Ecotoxicology and Environmental Safety*, vol. 180, pp. 509–516, 2019.
- [110] P. Ruenraroengsak and T. D. Tetley, "Differential bioreactivity of neutral, cationic and anionic polystyrene nanoparticles with cells from the human alveolar compartment: robust response of alveolar type 1 epithelial cells," *Particle and Fibre Toxicology*, vol. 12, no. 1, p. 19, 2015.
- [111] F. Ribeiro, A. R. Garcia, B. P. Pereira et al., "Microplastics effects in *Scrobicularia plana*," *Marine Pollution Bulletin*, vol. 122, no. 1-2, pp. 379–391, 2017.
- [112] Y. Liu, W. Li, F. Lao et al., "Intracellular dynamics of cationic and anionic polystyrene nanoparticles without direct interaction with mitotic spindle and chromosomes," *Biomaterials*, vol. 32, no. 32, pp. 8291–8303, 2011.
- [113] V. Paget, S. Dekali, T. Kortulewski et al., "Specific uptake and genotoxicity induced by polystyrene nanobeads with distinct surface chemistry on human lung epithelial cells and macrophages," *PLoS One*, vol. 10, no. 4, article e0123297, 2015.
- [114] T. G. Nam, "Lipid peroxidation and its toxicological implications," *Toxicology Research*, vol. 27, no. 1, pp. 1–6, 2011.
- [115] L. G. Barboza, L. R. Vieira, V. Branco et al., "Microplastics cause neurotoxicity, oxidative damage and energy-related changes and interact with the bioaccumulation of mercury in the European seabass, _Dicentrarchus labrax_ (Linnaeus, 1758)," Aquatic Toxicology, vol. 195, pp. 49–57, 2018.
- [116] H. Sies, C. Berndt, and D. P. Jones, "Oxidative stress," Annual Review of Biochemistry, vol. 86, no. 1, pp. 715–748, 2017.

- [117] D. R. Livingstone, "Contaminant-stimulated reactive oxygen species production and oxidative damage in aquatic organisms," *Marine Pollution Bulletin*, vol. 42, no. 8, pp. 656–666, 2001.
- [118] O. M. Ighodaro and O. A. Akinloye, "First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): their fundamental role in the entire antioxidant defence grid," *Alexandria Journal* of *Medicine*, vol. 54, no. 4, pp. 287–293, 2018.
- [119] R. T. Di Giulio, Indices of Oxidative Stress as Biomarkers for Environmental Contamination, M. A. Mayes and M. G. Barron, Eds., ASTM International, West Conshohocken, PA, 1991.
- [120] H. K. Imhof, J. Rusek, M. Thiel, J. Wolinska, and C. Laforsch, "Do microplastic particles affect Daphnia magna at the morphological, life history and molecular level?," *PLoS One*, vol. 12, no. 11, article e0187590, 2017.
- [121] Y. Deng, Y. Zhang, B. Lemos, and H. Ren, "Tissue accumulation of microplastics in mice and biomarker responses suggest widespread health risks of exposure," *Scientific Reports*, vol. 7, no. 1, p. 46687, 2017.
- [122] Y. Lu, Y. Zhang, Y. Deng et al., "Uptake and accumulation of polystyrene microplastics in zebrafish (Danio rerio) and toxic effects in liver," *Environmental Science & Technology*, vol. 50, no. 7, pp. 4054–4060, 2016.
- [123] J. A. McCubrey, M. M. LaHair, and R. A. Franklin, "Reactive oxygen species-induced activation of the MAP kinase signaling pathways," *Antioxidants and Redox Signaling*, vol. 8, no. 9–10, pp. 1775–1789, 2006.
- [124] X. Shi and B. Zhou, "The role of Nrf2 and MAPK pathways in PFOS-induced oxidative stress in zebrafish embryos," *Toxico-logical Sciences*, vol. 115, no. 2, pp. 391–400, 2010.
- [125] J. C. Park, J. Han, M. C. Lee et al., "Adverse effects of BDE-47 on life cycle parameters, antioxidant system, and activation of MAPK signaling pathway in the rotifer *Brachionus koreanus*," *Aquatic Toxicology*, vol. 186, pp. 105– 112, 2017.
- [126] H. Wang, L. Pan, R. Xu, L. Si, and X. Zhang, "The molecular mechanism of Nrf2-Keap1 signaling pathway in the antioxidant defense response induced by BaP in the scallop *Chlamys farreri*," *Fish & Shellfish Immunology*, vol. 92, pp. 489–499, 2019.
- [127] J. Chang-Bum, K. Hye-Min, L. Min-Chul et al., "Adverse effects of microplastics and oxidative stress-induced MAPK/ Nrf2 pathway-mediated defense mechanisms in the marine copepod *Paracyclopina nana*," *Scientific Reports*, vol. 7, no. 1, pp. 41323–41323, 2017.
- [128] J. Chang-Bum, K. Hye-Min, L. Min-Chul et al., "Accumulation of polystyrene microplastics in juvenile *Eriocheir sinensis* and oxidative stress effects in the liver," *Aquatic Toxicology*, vol. 200, pp. 28–36, 2018.
- [129] A. A. de Souza Machado, W. Kloas, C. Zarfl, S. Hempel, and M. C. Rillig, "Microplastics as an emerging threat to terrestrial ecosystems," *Global Change Biology*, vol. 24, no. 4, pp. 1405–1416, 2018.
- [130] M. C. Celina, "Review of polymer oxidation and its relationship with materials performance and lifetime prediction," *Polymer Degradation and Stability*, vol. 98, no. 12, pp. 2419–2429, 2013.
- [131] A. Jahnke, H. Peter, H. Arp et al., "Reducing uncertainty and confronting ignorance about the possible impacts of

weathering plastic in the marine environment," *Environmen*tal Science & Technology Letters, vol. 4, no. 3, pp. 85–90, 2017.

- [132] G. F. Schirinzi, I. Pérez-Pomeda, J. Sanchís, C. Rossini, M. Farré, and D. Barceló, "Cytotoxic effects of commonly used nanomaterials and microplastics on cerebral and epithelial human cells," *Environmental Research*, vol. 159, pp. 579– 587, 2017.
- [133] I. Pont, C. Lacroix, C. G. Fernández et al., "Exposure of marine mussels *Mytilus* spp. to polystyrene microplastics: toxicity and influence on fluoranthene bioaccumulation," *Environmental Pollution*, vol. 216, pp. 724–737, 2016.
- [134] E. Yousif and R. Haddad, "Photodegradation and photostabilization of polymers, especially polystyrene: review," *Springer Plus*, vol. 2, no. 1, pp. 398–398, 2013.
- [135] X. Chen, M. Song, B. Zhang, and Y. Zhang, "Reactive oxygen species in the immune system," *International Reviews of Immunology*, vol. 32, no. 3, pp. 249–270, 2013.
- [136] F. Yu, C. Yang, Z. Zhu, X. Bai, and J. Ma, "Adsorption behavior of organic pollutants and metals on micro/nanoplastics in the aquatic environment," *Science of the Total Environment*, vol. 694, p. 133643, 2019.
- [137] Comprehensive Polymer Science and Supplements, Comprehensive Polymer Science and Supplements-Online, Elsevier Science, 1996.
- [138] P. Pannetier, J. Cachot, C. Clérandeau et al., "Toxicity assessment of pollutants sorbed on environmental sample microplastics collected on beaches: part I-adverse effects on fish cell line," *Environmental Pollution*, vol. 248, pp. 1088–1097, 2019.
- [139] L. Lei, M. Liu, Y. Song et al., "Polystyrene (nano) microplastics cause size-dependent neurotoxicity, oxidative damage and other adverse effects in Caenorhabditis elegans," *Environmental Science: Nano*, vol. 5, no. 8, pp. 2009–2020, 2018.