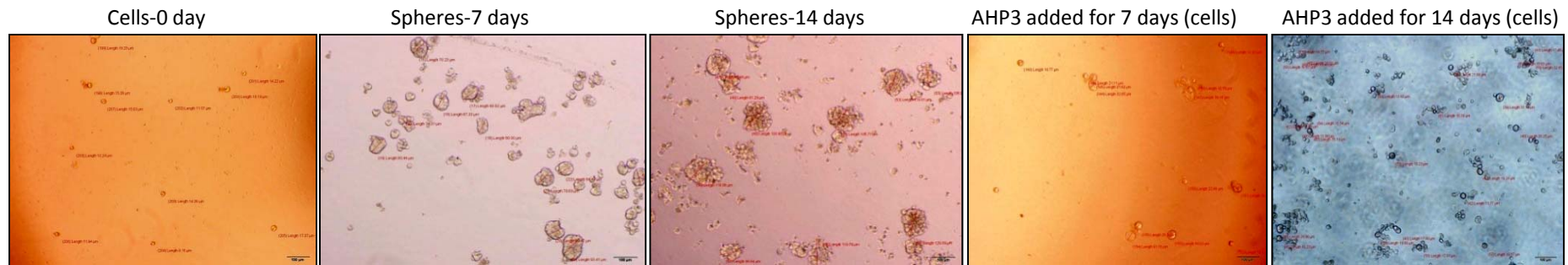
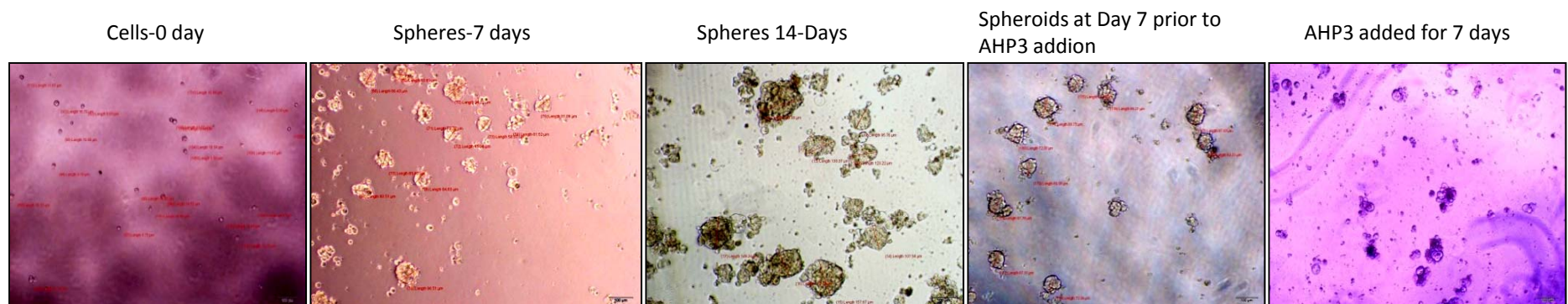
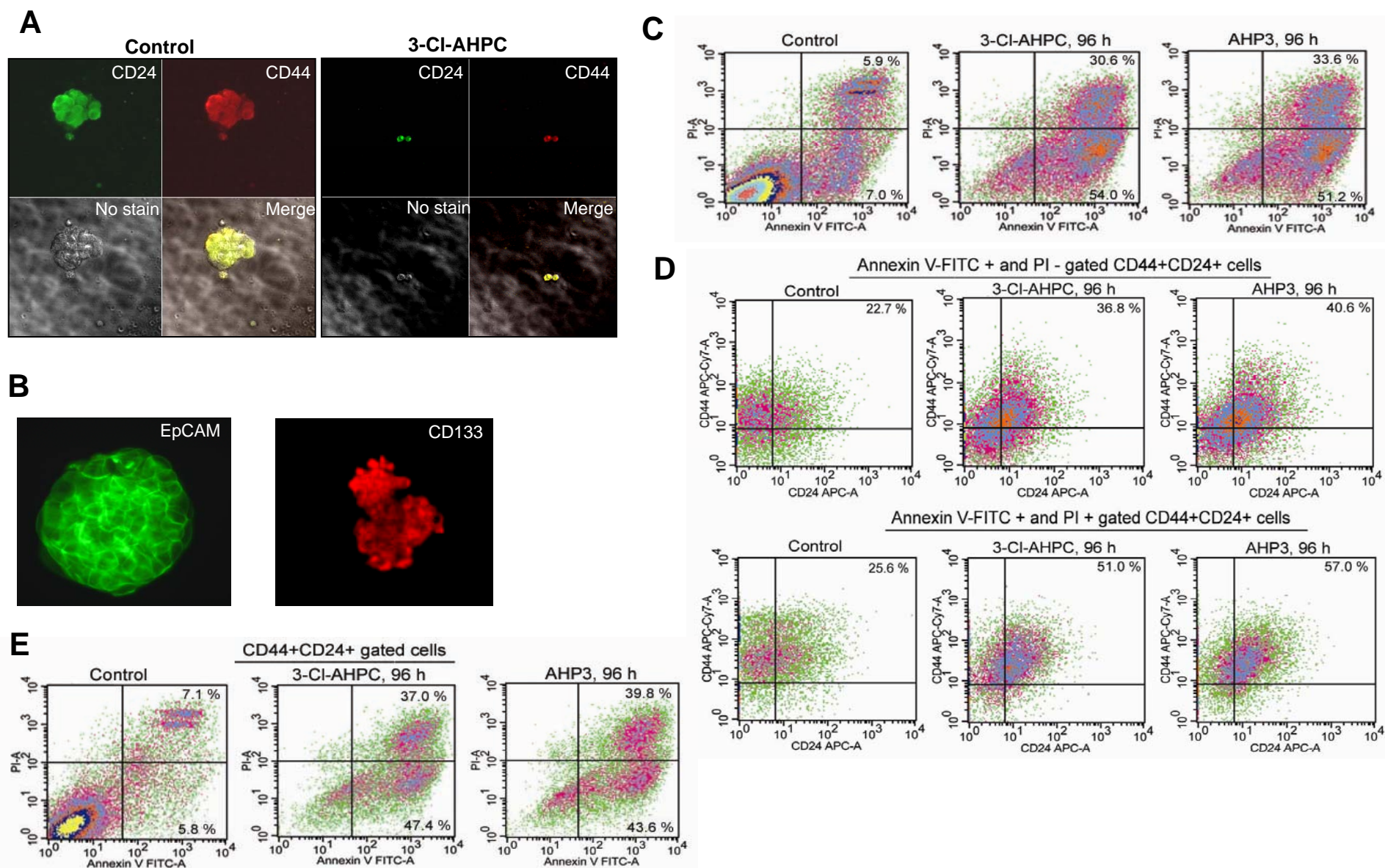
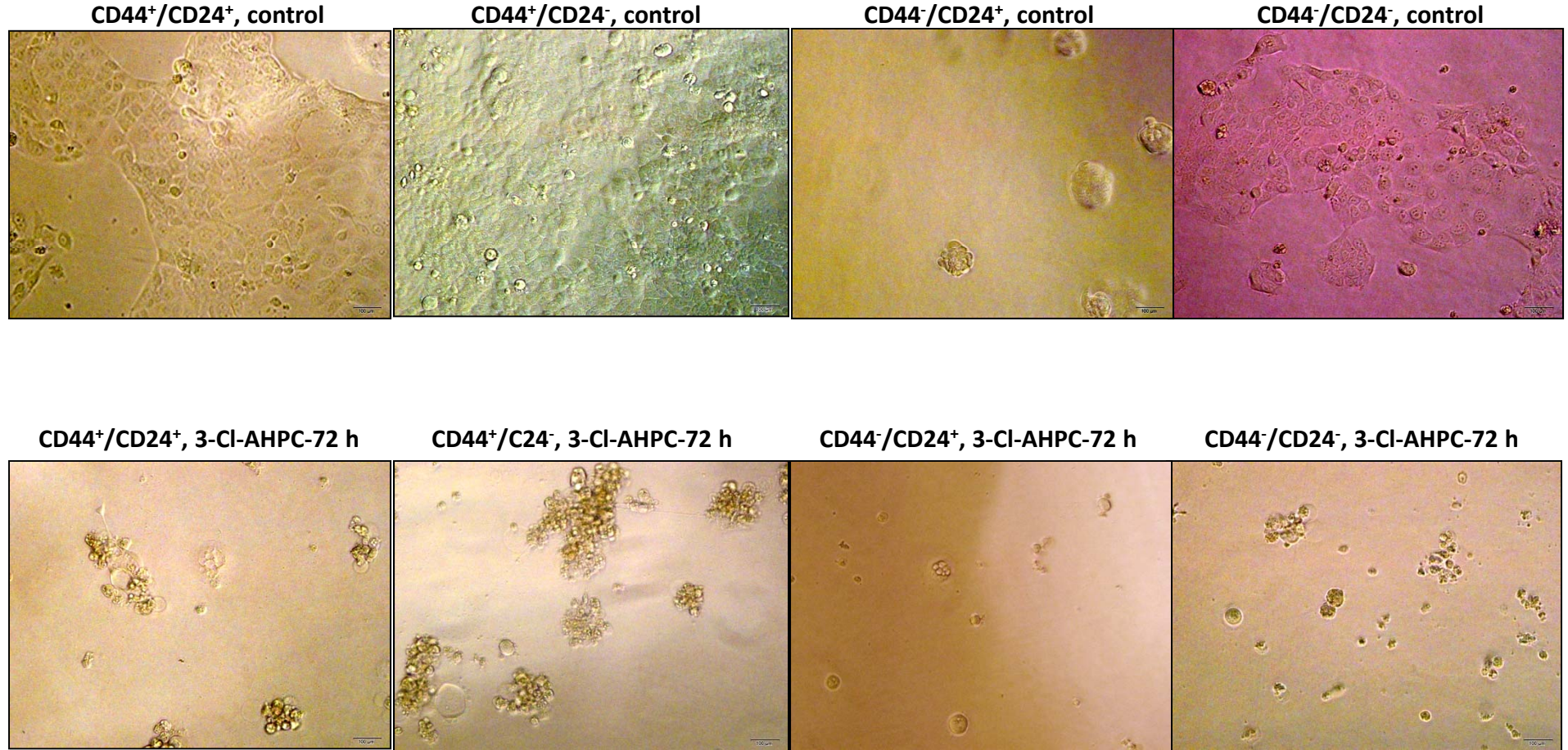


A**AHP3 added the day after seeding the cells****B****AHP3 added after spheres formation**

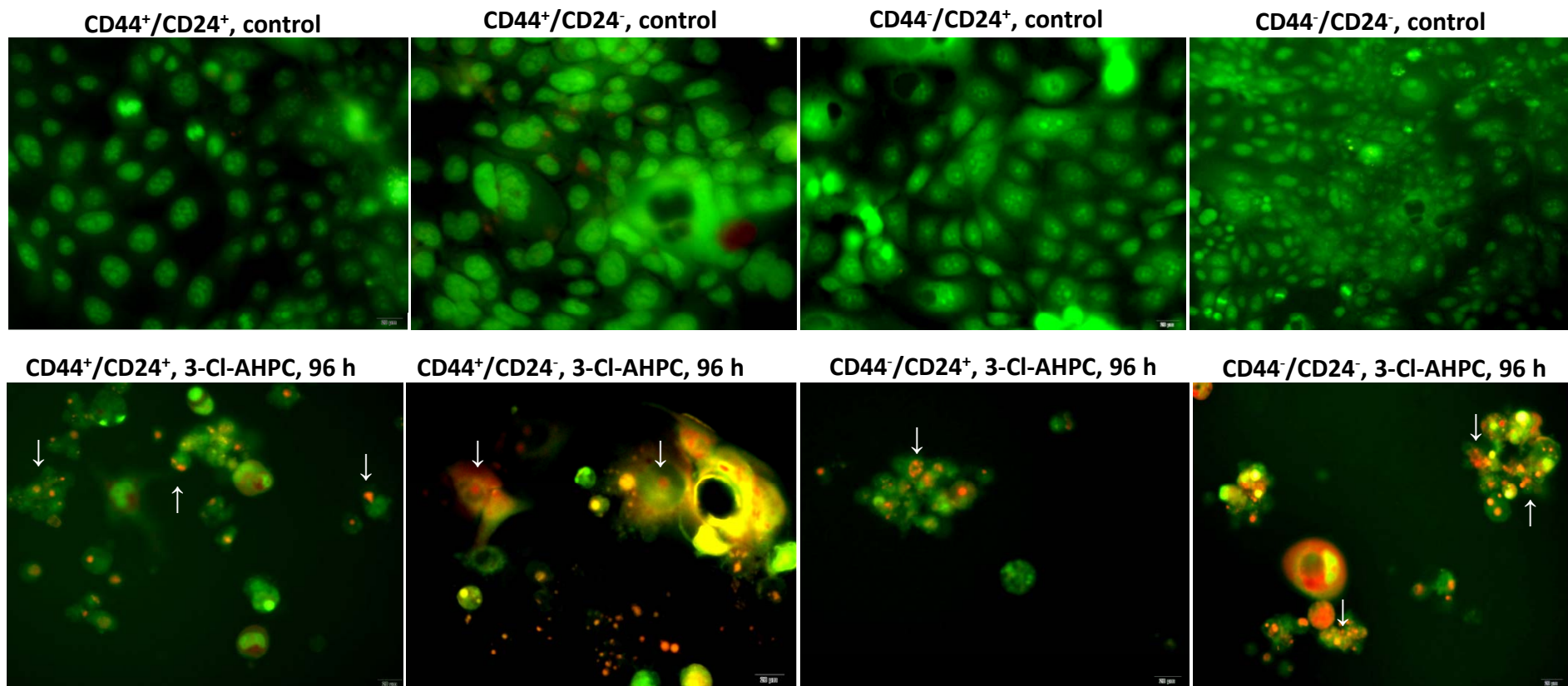
Supplementary Figure S1: CD44⁺/CD24⁺ cell sphere formation and AHP3-mediated inhibition and degradation of spheres derived from PANC-1 cells (A) AHP3 inhibited CD44⁺/CD24⁺ sphere formation in PANC-1 cells. (B) AHP3 exposure resulted in inhibition of growth and degradation of the CD44⁺/CD24⁺ derived spheres in PANC-1 cells.



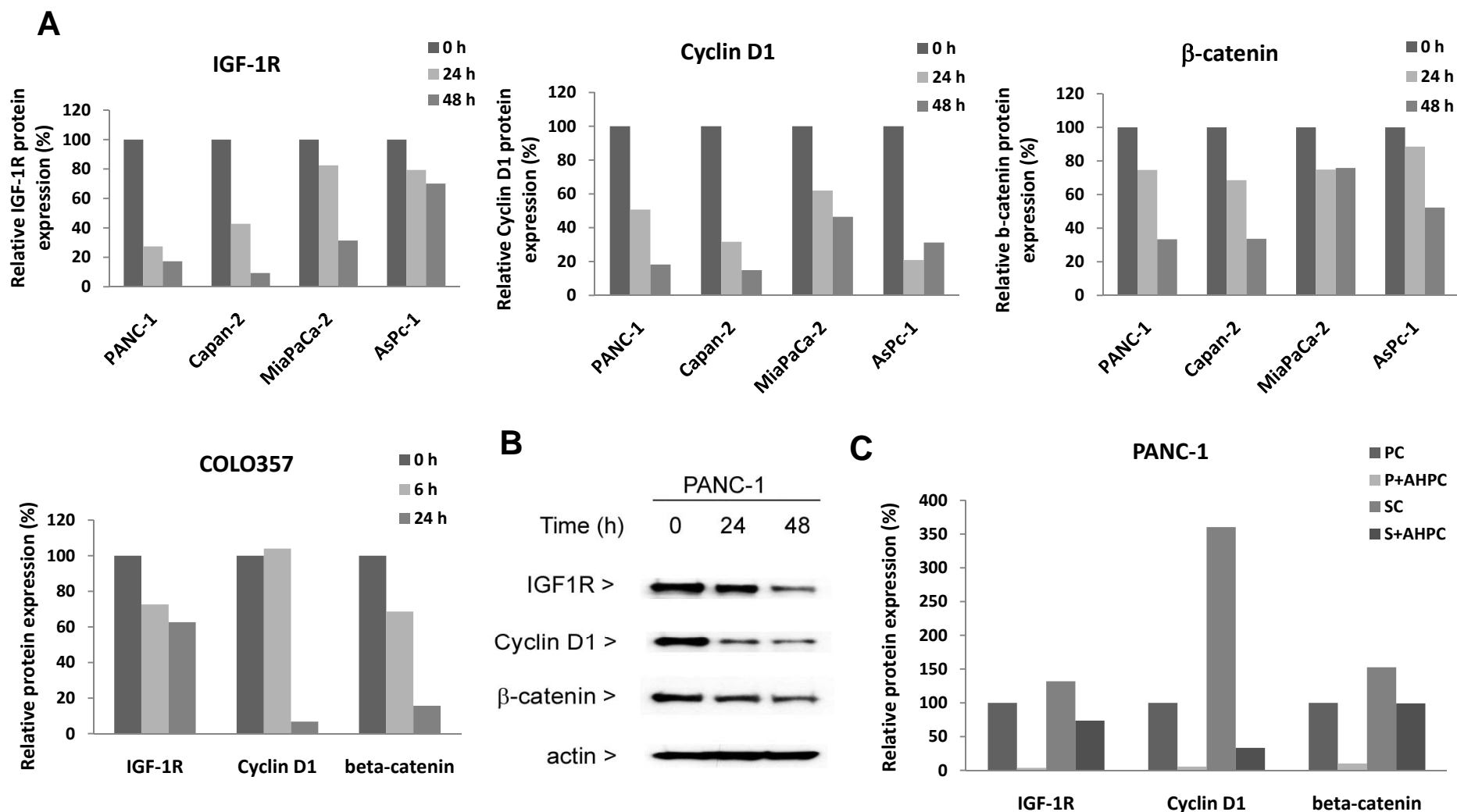
Supplementary Figure S2: Pancreatic spheres are composed of CD44⁺/CD24⁺/CD133⁺/EpCAM⁺ in cells and 3-Cl-AHPC (1 μ M) mediated apoptosis of CD44/CD24 PANC-1 cells. (A) CD44/CD24 spheres were stained with CD44, CD24. Cells in vehicle and 3-Cl-AHPC (1 μ M) treated spheres for 7 days were stained with either CD44 or CD24. Images were obtained using confocal microscope and the images merged. (B) CD44/CD24 spheres expressed EpCAM and CD133 positivity by confocal and fluorescence microscope. (C) ARR-mediated total apoptotic cells (D) early apoptotic CD44⁺/CD24⁺ cells (upper panel) and early & late apoptotic CD44⁺/CD24⁺ cells (lower panel), and (E) total CD44⁺/CD24⁺ apoptotic cells. PANC-1 cells were analysed in flow cytometry.



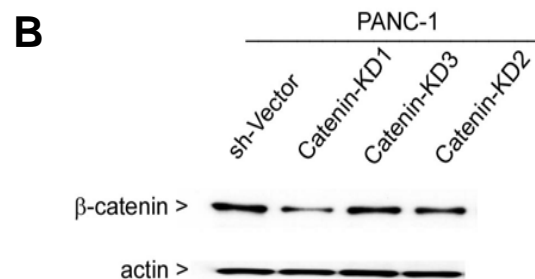
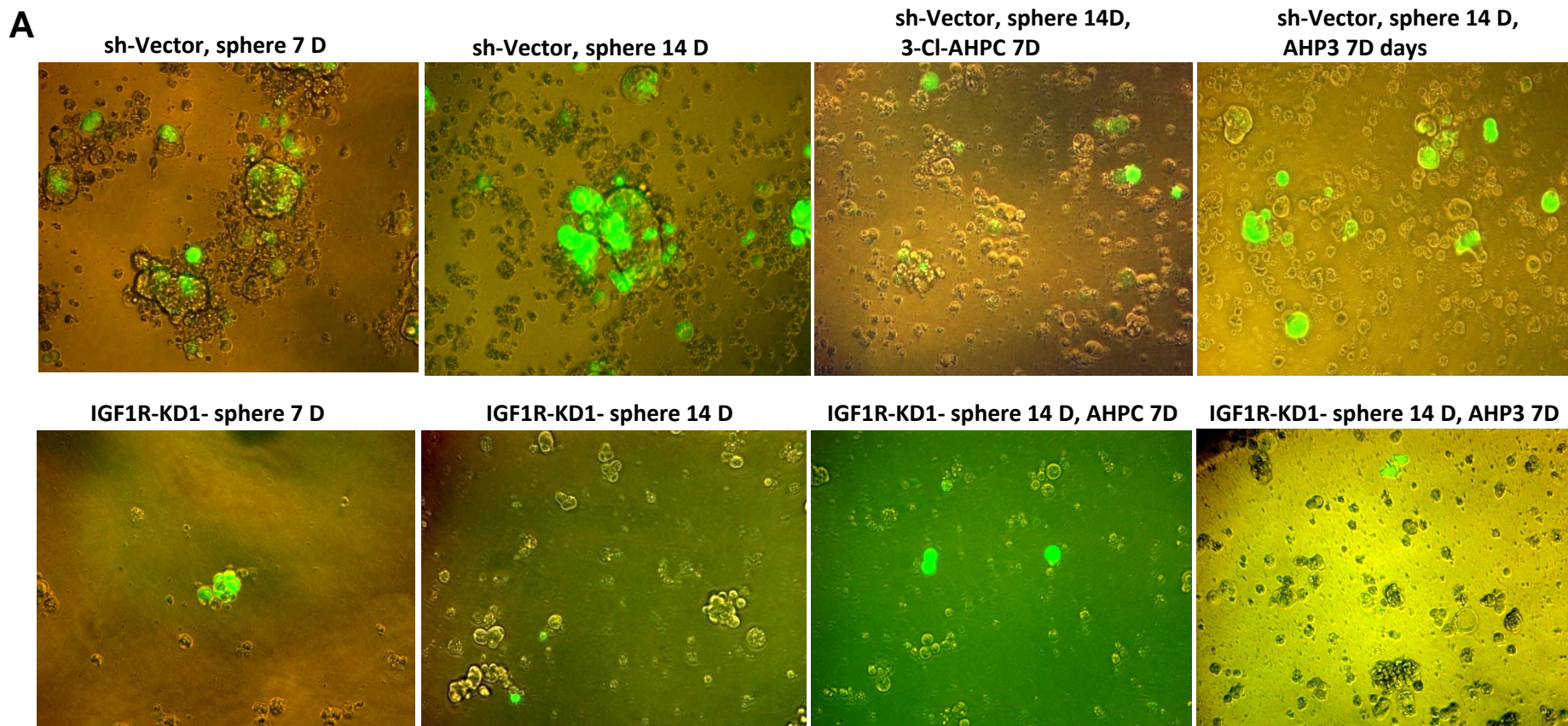
Supplementary Figure S3: 3-Cl-AHPC (1 μ M) mediated growth inhibition and degradation of CD44/CD24 PANC-1 cells. The four populations of CD44/CD24 cells were sorted by flow cytometry and grown with B27 medium without growth factors in 24 well plates and 1.0 μ M 3-Cl-AHPC added the day after seeding. The cells were photographed using Olympus microscope digital camera and DP2-BSW software.



Supplementary Figure S4: 3-Cl-AHPC -mediated apoptosis of PANC-1 CD44/CD24 cell populations. The four types of CD44/CD24 cell populations were sorted by flow cytometry and were seeded with B27 medium without growth factors in 24-well plates. 1.0 μ M 3-Cl-AHPC was added the day after seeding. Cells were stained with acridine orange/ethidium bromide. Arrow indicates fragmentation of DNA in apoptotic cells. The cells were photographed using Olympus microscope digital camera and DP2-BSW software.



Supplementary Figure S5: (A) Densitometric analysis of protein expression of IGF-1R, cyclin D1 and β -catenin in pancreatic cancer cell lines. Cells were incubated with 1.0 μ M 3-Cl-AHPC for times indicated and western blots performed as described in Materials and Methods section. (B) IGF-1R, cyclin D1 and β -catenin expression decreased in cells following exposure of AHP3. (C) Densitometric analysis of expression of IGF-1R, cyclin D1 and β -catenin in PANC-1 parental and spheroid cells. PC, parental control; SC, spheroid control; AHPC, 3-Cl-AHPC ; P+AHPC, parental cells 3-Cl-AHPC treated; S+AHPC, spheroid 3-Cl-AHPC treated. Parental cells and spheroids were treated for 96 h and 7 days, respectively.



Supplementary Figure S6. (A) Knock down of IGF-1R expression inhibits PANC-1 sphere formation. Spheres were formed at indicated times (D, day) from stably transfected GFP-tagged sh-vector and GFP-tagged -IGF-1R-KD PANC-1 cells. Spheres were grown as described in Materials and Methods. Vehicle or 1 μ M 3-Cl-AHPC or AHP3 were added after 7 days of sphere formation and the spheres incubated an addition 7 days. Spheres were visualized and photographed with fluorescence microscope (100 μ m scale and magnification 400 X) after 14 days 3-Cl-AHPC treatment. (B) Knock down of β -catenin in stably transfected PANC-1 cell lines.