

Review Article

Pulmonary Salivary Gland Tumor, Mucoepidermoid Carcinoma: A Literature Review

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Received 27 March 2022; Revised 10 May 2022; Accepted 31 August 2022; Published 2 November 2022

Academic Editor: Goo Lee

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Pulmonary mucoepidermoid carcinoma (PMEC) is the most common malignant salivary gland tumor in the lungs and accounts for 0.1–0.2% of all lung malignancies in adults. It has no specific epidemiological or clinical characteristics. Correct diagnosis requires the combined examinations of images, laboratories, pathology, and immunohistochemistry (IHC) as well as molecular characteristics. PMEC tumors are characterized by squamous, intermediate, and mucus-secreting cells. Currently, histological appearance, mitotic frequency, cellular atypia, and necrocytosis allow the classification of PMEC into low grade or high grade. Molecular changes are crucial to pathological diagnosis. The driver of PMEC seems to be the fusion protein MECT1-MAML2 that is generated from a genetic mutation in *t*(11; 19) (*q*21; *p*13), while other gene mutations are also reported. However, no treatment of PMEC exists so far; surgical excision is still the primary treatment, while the efficacies of chemotherapy or radiotherapy are undefined. Tyrosine kinase inhibitor (TKI) therapy and immunotherapy showed to have significant therapeutic effects but require more investigation and better understanding. This review focuses on the clinical characteristics, imaging and pathological features, immunohistochemical examination, mutation analysis, differential diagnosis, prognosis, and treatment of PMEC.

1. Introduction

In 1952, Smetana et al. [1] first reported pulmonary mucoepidermoid carcinoma (PMEC) as a scarce malignant neoplasm in the lungs that accounted for 0.1–0.2% of all lung malignancies in adults [2–6]. PMEC, the most common primary salivary gland carcinoma (SGC) in the lung, originates from the minor salivary glands in the submucosa of large airways [7]. PMEC occurs in any age group; some studies reported its occurrence primarily in younger adults under 50 [8]. The clinical symptoms and epidemiological characteristics of PMEC are not specific and representative. Accordingly, correct diagnosis of PMEC requires the

combination of clinical characteristics, histopathological examination, immunohistochemistry (IHC), and molecular mutational analysis. Tumors caused by PMEC are characterized by the histopathology of squamous epithelial, mucous, and intermediate cells. The classification of PMEC into high grade or low grade depends on histological appearances, mitotic frequencies, cellular atypia, and necrocytosis [2, 8, 9].

The most common genetic change is *t*(11; 19) (*q*21; *p*13), which generates the fusion protein MECT1-MAML2. Since MECT1-MAML2 fusion was demonstrated to be present in more than 66% of all PMEC cases [8, 10, 11], it was proposed to drive PMEC progress [7]. Several other gene mutations

were also reported. For example, a mutation in the gene for the epidermal growth factor receptor (EGFR) was observed in patients with P MEC [12]. These genomic alterations are potential for selecting therapy.

Up to now, there is still no consolidated strategy for the therapy of P MEC, and the complete surgical resection is recognized as the primary therapeutic method. The effects of chemotherapy or radiotherapy as valid therapies have not been shown yet [13]. Several case reports show that EGFR-inhibiting agents (gefitinib and erlotinib) have efficacies in patients with P MEC [14–16]. An immunotherapy approach for P MEC is limited so far and needs to be explored in-depth; therefore, advanced research currently prioritizes targeted therapy and immunotherapy.

2. Clinical Characteristics

From an epidemiological perspective, P MEC is a rare pulmonary tumor that appears in a wide range of ages. Generally, the age of onset ranges from 7 to 87 years, and the mean age is approximately 50–60 years. The incidence of P MEC in patients over 75 years is rare, and Abdalla et al. [17] reported a rare case of an 81-year-old male with P MEC. In terms of gender, although some studies suggested that the incidence in males is higher than in females [2, 18, 19], most reports demonstrated that the incidence between males and females possesses a similar distribution [8, 20, 21]. Interestingly, only a few patients stated they were smoking. Hence, there does not seem to be a correlation between P MEC onset and smoking, and this needs to be confirmed in advanced studies.

P MEC is not accompanied by any specific clinical symptom, while the most common symptom is cough. Other symptoms include blood-tinged or whitish sputum, fever, hemoptysis, chest tightness, chest pain, hoarseness, and dyspnea; yet some patients have no obvious symptoms and are only diagnosed during a physical examination. Three patients were even diagnosed with P MEC upon hospital admission because of a cough [17, 22, 23]. Interestingly, the clinical symptom of a P MEC patient, as well as its frequency and extent, depends on the position of the lesion. Tumors located in the central bronchus will appear as obstructive airway symptoms and primarily manifest as cough, dyspnea, or asthma. On the other hand, 85% of P MEC appears in the peripheral lung. These tumors may manifest as cough, chest pain, and pulmonitis [24, 25] or can be asymptomatic and only be found in physical examinations. These results suggest that P MEC has no obvious symptoms, while it can be easily misdiagnosed or overlooked. Hence, due to the challenge of correctly identifying this benign disease, it is necessary to raise awareness of P MEC to decrease the rate of misdiagnosis.

3. Imaging Examination

Computed tomography (CT) is a vital and necessary approach for the diagnosis and differential diagnosis of P MEC. CT is a noninvasive and convenient technique that can be adopted to explore suspected trachea and lung lesions. Most

studies based on CT describe P MEC as a well-defined mass characterized as the central or hilar type, oval or round shape, with smooth margins and marked enhancement [26–28]. Wang et al. [29] used CT images to distinguish between low-grade and high-grade P MEC. Low-grade P MEC usually manifests as a central bronchial mass with marked homogeneous enhancement. In contrast, high-grade P MEC tends to be in the periphery and manifests as a lobular and heterogeneous mass with poorly defined margins and minor enhancement. Additionally, Cheng et al. [27] used multisession-computed tomography (MSCT) to reveal an oval or lobulated, mildly enhanced mass with calcification and mucus lakes that may indicate P MEC. Similarly, Park et al. [30] applied ^{18}F -FDG PET/CT to predict the pathologic grade and prognosis of P MEC. The authors concluded that patients with SUV_{max} greater than 6.5 tend to have high-grade P MECs, lymph node metastasis, and recurrences. The size of tumors varies approximately from 1 cm to 10 cm. The majority of literature reports indicate tumor sizes of 2.0–3.0 cm.

The preferential location of P MEC distribution is rather varied as shown by several studies. For example, Zhang et al. [31] and Salem et al. [10] showed that tumors often occur in the left lower lobe and right upper lobe. Qiu et al. [20] also observed that P MEC commonly occurs in the upper lobe and lower lobe rather than in the bronchus. However, Cheng et al. examined 43 patients with P MEC and observed that P MECs more likely occur in the right lower lobe and left upper lobe [27]. In addition, Huo et al. [8] investigated 26 patients with P MEC and found that tumors were rather located in the segmental bronchus and lobe. Hence, it can be concluded that P MEC tumors can occur in any lobe of the lung with no preference to its location.

4. Pathology and Immunohistochemistry Examinations

Grossly, P MEC tumors are tan or light brown polypoid mass. The central bronchus may be present as an exophytic tumor and nearly completely occlude the bronchial lumen [32]. P MEC are histopathologically defined by a combination of squamous epithelial, mucous, and intermediate cells with defects in keratinization. The standard classifies P MEC into low-grade and high-grade tumors, depending on histological appearance, mitotic frequency, cellular atypia, and necrocytosis [2, 8, 9]. Low-grade P MECs are combinations of all three cell types without any specific differentiation, comprised of cystic changes prevailing. Mitotic figures, nuclear atypia, and necrosis are rarely observed. Microscopic invasion into pulmonary parenchyma is unusual [33]. High-grade P MEC primarily consists of squamous epithelial and intermediate cells with a small number of mucous cells, with a presentation of predominantly solid pattern growth, continual necrosis, mitoses (more than 4/10 high-power fields), or distinct atypia [8, 10, 27, 29, 34]. Chin et al. [18] observed that high-grade tumors had a higher proportion of squamous epithelium. Besides, invasion into the adjacent pulmonary parenchyma and regional lymph node involvement are more frequent in high-grade P MEC

TABLE 1: IHC results of the literature review.

Ref.	P63+	CK7+	Muc5Ac+	P40+	CK5/ 6+	TTF-1-	Napsin A-	HER2-	Ki-67
[8]	26/ 26	26/26	26/26	26/26	NM	26/26	NM	26/26	Median 4.1% (low-grade) Median 22.4% (high-grade)
[11]	25/ 25	NM	NM	23/25	NM	25/25	25/25	NM	NM
[17]	1/1	NM	NM	1/1	NM	NM	NM	NM	NM
[19]	5/5	6/6	NM	NM	2/5	6/6	2/2	NM	<10% (4/6 low-grade) ≥20% (2/6 high-grade)
[22]	1/1	NM	NM	1/1	NM	1/1	NM	NM	NM
[23]	NM	1/1	NM	1/1	1/1	1/1	1/1	NM	40% (high-grade)
Total.	58/ 58 100%	33/33 100%	26/26 100%	52/54 96.3%	3/6 50%	59/59 100%	28/28 100%	26/26 100%	

NM: No mention.

[9, 21, 33, 35]. In the majority of the series, the authors concluded that the pathological classification of PMEC is significant for diagnosis, treatment, and prognosis. Patients with low-grade PMEC have a better survival outcome compared with those with high-grade PMEC [8, 9, 13, 19, 25, 29, 36, 37]. In addition, Wang et al. [29] compared the association between pathologic grade and predilection sites of PMEC and observed that low-grade PMEC usually appears in the central lung, whereas high-grade PMEC often occurs in the peripheral lung.

The IHC characteristics of PMEC are retrospectively analyzed and summarized in Table 1. Here, the positive percentage of p63, CK7, Muc5Ac, p40, and CK5/6 was found to be 58/58 (100%), 33/33 (100%), 26/26 (100%), 52/54 (96.3%), and 3/6 (50%), respectively. Napsin A, TTF-1, and human epidermal growth factor receptor 2 (HER2) were all negative. Most of the studies reported that p63 and p40 are expressed, while TTF-1 and Napsin A are negative in PMEC. However, Zhang et al. [31] reported that some cases were positive for TTF-1 and napsin A, which is inconsistent with the results of the majority of reports. In low-grade cases, the Ki-67 labeling index was less than 10%, while in cases of high-grade PMEC, the index was more than 20% [8, 19, 23]. Hence, the Ki-67 labeling index potentially be used as an auxiliary index for differentiating high-grade from low-grade PMEC.

P63 commonly supports the diagnosis of squamous cells. Therefore, p63 is confirmed to be positive in PMEC. It is yet generally positive in adenosquamous carcinoma and squamous cell carcinoma, so it may lead to misdiagnosis [8, 38]. P63 could be adopted to distinguish PMEC from other salivary gland tumors, especially acinic cell carcinoma, since p63 is generally negative in acinic cell carcinoma [39]. P40 is another IHC marker adopted to diagnose PMEC. Roden et al. [11] observed that the expression pattern between p40 and p63 was semblable in most cases, while the p40 expression score was lower than p63 in nearly one-quarter of PMEC. A few p40-negative cases have focal p63 expression. Consequently, they realized p63 could be a more sensitive marker. Despite, it has recently been proposed to be more specific than p63 for squamous differentiation [38]. Since

TTF-1 is always negative in PMEC, it is conducive to distinguishing PMEC from primary pulmonary adenosquamous carcinoma and adenocarcinoma [8, 11, 13].

HER2 gene and protein change are molecular basics for target therapy in cancer. As a whole, it is reported HER2 gene amplification in 1.0%–14.3% by fluorescence in situ hybridization (FISH) and protein overexpression in 4.3%–38% by IHC of salivary mucoepidermoid carcinoma (MEC) [40–43]. In addition, both HER2 amplification and protein overexpression were also reported to be associated with high-grade tumors [40, 42]. One patient with metastatic MEC expressed HER2 positive achieved therapeutic response to trastuzumab [44]. Therefore, Clauditz et al. [40] suggested that IHC and FISH analyses of HER2 should be applied in the cases of recrudescence and/or metastatic disease. Until now, only a few studies have investigated the expression of HER2 in PMEC, and in Table 1, HER2 is negative in all cases (0/26). However, the detection and analysis of HER2 should be explored in larger samples of PMEC.

In summary, the combined detection of p63, p40, CK5/6, CK7, Ki-67 labeling index, the absence of TTF-1, and Napsin A may be an auxiliary diagnostic index of PMEC. HER2 detection (protein overexpression and gene amplification) could be a necessary complement.

5. Molecular Characteristics

The mutation *t* (11; 19) (*q21*; *p13*) generating the MECT1-MAML2 fusion protein has been demonstrated to be the specific genetic event for PMEC onset [45]. The rearrangement is fused by mucoepidermoid carcinoma translocated 1 (MECT1) at 19p13 and mastermind-like 2 (MAML2) at 11q21 [46]. Tonon et al. [47] observed that MECT1-MAML2 activated HES1 transcription to disrupt Notch signaling. Wu et al. [48] also found that this fusion protein activates CREB and thus mimics the constitutive activation of cAMP signaling. Therefore, presence of the MECT1-MAML2 rearrangement can support PMEC diagnosis as this genetic change is found in 66% to 100% of PMEC. Simultaneously, some studies proposed that the

MECT1-MAML2 rearrangement is more common in low-grade than in high-grade PMEC. For example, Salem et al. [10] showed that 88% (8/9) of PMEC contained the MECT1-MAML2 fusion protein, of which all had a low-grade morphologic tumor. A study by Huo et al. [8] showed similar results; 83.3% (10/12) of low-grade PMEC contained the MAML2 rearrangement, while only 33.3% (2/6) of high-grade PMEC did. Also, Roden et al. [11] detected the MAML2 rearrangement by FISH and confirmed all 24 cases (3 low, 19 intermediate, and 2 high grade) to be positive.

Little is known about the genomic background of PMEC in addition to MECT1-MAML2 translocations. Wang et al. [49] employed a comprehensive genomic profiling to investigate salivary mucoepidermoid carcinomas (3 high-grade PMECs) and revealed the appearance of diverse genomic alterations. These may bring new targets for an immunological therapy approach. Although the detailed genomic change of PMEC was not reported separately, the authors concluded that the majority of patients had at least one genomic alteration and that the most common genomic alterations were in *CDKN2A* and *TP53*. They also indicated that the frequency of both *PIK3CA* alterations and PI3K pathway activation in high-grade tumors is higher than their frequencies in low-grade tumors. Consequently, more potentially actionable genomic alterations have been observed now that can influence therapy selection.

Overexpression of the EGFR protein was common in most cases of PMEC. On the contrary, amplification or mutation within the tyrosine kinase domain of the EGFR gene has been barely reported [8, 50]. However, Yu et al. [12] unveiled 5 cases (25%) in 20 PMEC patients with an uncommon EGFR mutation (exon 21 L861Q heterozygous mutation). This study proved the appearance of EGFR mutations in PMEC and the L861Q mutation to be the predominant EGFR mutation. Yamamoto et al. [51] reported that in two out of nine (22.2%) patients, EGFR gene abnormalities (exon 21) were detected by the IHC method, and in one (11.1%) patient, the EGFR mutation (exon 21 L858R mutation) was observed by the cycleave method.

6. Differential Diagnosis

Distinguishing high-grade PMEC from adenosquamous carcinoma (ASC) is rather challenging due to only minor differences in their IHC and histopathological patterns. A study by Huo et al. [8] misdiagnosed 2 ASCs as PMEC based on the presence of mucous cells, solid nests, and the consistency of IHC-positive results. Only by considering the keratinization and positivity of TTF-1 could the diagnosis be modified. Similarly, Chenevert et al. [52] decided to reclassify their ASC cases due to the presence of dysplastic and/or *in situ* carcinoma in the mucosa and extensive keratinization. Therefore, although the differences between PMEC and ASC are only minor, PMEC rarely shows an expression of keratinization and *in situ* carcinoma and a complete absence of TTF-1.

The incidence rate of primary adenoid cystic carcinoma (PACC) is lower than the rate of PMEC in adults [7]. However, no remarkable difference in the clinical

manifestation between PMEC and PACC exists [3]. Comparing the epidemiological characteristics with PACC, patients with PMEC are often of younger age at tumor onset, have smaller tumors, less lymph nodes, or distant metastases, and are more likely to be in the early stage of the disease [3, 53]. There are significant distinctions in the predilection site and features shown in CT. PACC occurs more frequently in the central type (located in the main bronchus or trachea) and appears more often as a lobulated mass. Homogeneous or heterogeneous thickening owing to infiltration of the luminal wall is common. PMEC manifests commonly as the hilar type, concomitant by distant bronchial dilatation with mucoid obstruction. CT findings are more likely to suggest an obstructive airway disease. PMEC is more frequently present as an obvious enhancement than PACC [28, 54]. Kumar et al. [25] observed a similar result; PACC usually occurs in the central airways and main bronchial tube, while PMEC was more frequently located in the lungs. As for the results of the immunohistochemical examination, PACC expresses CD117(c-kit protein) and myoepithelial markers, including pancytokeratin, p63, and CK7. *MYB-NFIB* fusion carcinogens generated by tumor-specific t(6;9)(q22-23;p23-24) translocation are considered to be specific to PACC [7, 55].

The most challenging distinction may be with hyalinizing clear cell carcinoma. Both carcinomas show a similar presence of mucin pools, intracytoplasmic mucin, and hyalinized stroma and immunohistochemically squamous differentiation [56–58]. Takamatsu et al. [57] observed mucin production, yet no mucin-secreting cells were present in hyalinizing clear cell carcinoma. On the contrary, mucin-secreting cells are one of the essential components in PMEC. Furthermore, there is no significant difference in immunohistochemical findings between PMEC and hyalinizing clear cell carcinoma. Similar to PMEC, CK7, CK 5/6, p63, and p40, cytokeratin cocktail is usually positive in hyalinizing clear cell carcinoma, which reveals squamous differentiation. TTF-1, napsin A, CK20, chromogranin, synaptophysin, SMA, HMB45, and melan A, on the other hand, are negative [56, 57]. Ki-67 labeling was ranged from 3 to 10% [57]. Interestingly, molecular analysis can be instrumentally adapted to distinguish PMEC from hyalinizing clear cell carcinoma. *EWSR1-ATF1* fusion is confirmed to be specific in hyalinizing clear cell carcinoma [56–58]. Chapman et al. [58] reported three cases initially diagnosed as PMEC and then demonstrated *EWSR1-CREM* fusion to sustain a diagnosis of hyalinizing clear cell carcinoma. Hence, one can conclude that performing essential cytogenetic and molecular analysis supports a correct differential diagnosis. The pathological diagnostic flowchart of PMEC is shown in Figure 1.

7. Treatment

At present, there is no consolidated standard to treat PMEC. However, the principles to treat PMEC are consistent in the domestic and foreign literature. Complete surgical resection is recognized as the predominant therapeutic strategy, which even implies a better survival outcome, especially for stage

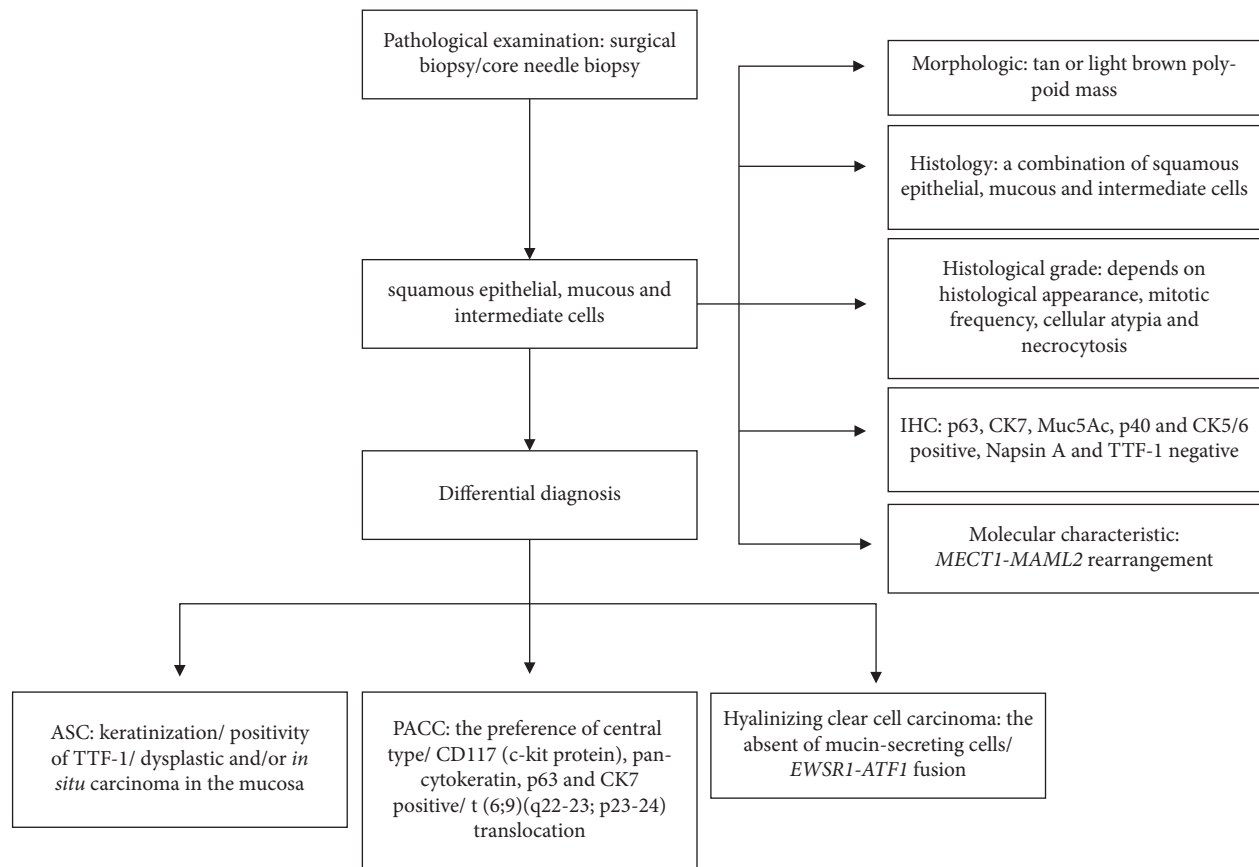


FIGURE 1: The pathological diagnostic flowchart of PMEC.

I–II PMEC [8, 20, 31]. Zhang et al. [31] summarized the median overall survival (OS) of surgery (57/87), radiotherapy (5/87), and others (25/87) as 61 months, 60 months, and 42 months, respectively. It is suggested that the prognosis of surgery is better than that of nonsurgery. Qiu et al. [20] analyzed survival outcomes of treatments in patients with TNM stage I–II and stage III–IV PMEC separately. They concluded that surgical treatments had the highest cancer-specific survival (CSS) compared to other therapies (radiation and/or chemotherapy and surgery plus radiation and/or chemotherapy). Zhang et al. [31] observed that the median OS of surgery (30) and nonsurgery (15) is 60 months and 52 months in stage I–II patients, respectively ($P = 0.013$). Qiu et al. [20] found that local surgical resection should be avoided since patients with local tumor excision had the worst OS and CCS. Besides, concerning PMEC patients with high uptake on PET/CT imaging, Park et al. [30] suggested that mediastinal lymph node dissection and adjuvant therapies were feasible. Consequently, the majority of the studies agree that complete surgical resection and systematic lymph node dissection are essential and necessary for patients who consider surgical treatment.

The efficacy of chemotherapy or radiotherapy remains controversial [13]. However, adjuvant chemotherapy or radiotherapy is probably feasible in patients with high-grade PMEC, especially in cases of extrathoracic invasion. Yan et al. [23] reported a case of comprehensive therapy

combining apatinib with fractionated stereotactic radiotherapy. This approach had a therapeutic effect on high-grade PMEC with limited brain metastases, which inspiringly improved brain edema and OS. Sonobe et al. [59] reported a case of a 59-year-old man with high-grade PMEC responding to carboplatin and paclitaxel and suggested that this combination therapy of carboplatin and paclitaxel provided an option for PMEC treatment.

EGFR-tyrosine kinase inhibitor (EGFR-TKI) is the most effective therapy for terminal patients harboring EGFR mutation. Han et al. [15] as well as Rossi et al. [16] reported PMEC patients whose tumor neither showed EGFR protein overexpression nor had EGFR genomic variations by FISH and mutational analysis. However, these patients had a certain response to gefitinib. According to these findings, O'Neill [60] suggested that the tumor-specific $t(11; 19)(q21; p13)$ gene mutation producing the MECT1-MAML2 fusion protein may be an effective target for EGFR-TKI therapy. Lee et al. [14] reported a case of metastatic PMEC with a response to EGFR-TKI erlotinib. It provides a possibility for PMEC treatment. Chen et al. [61] suggested that the MECT1-MAML2 fusion protein upregulates the expression of EGFR ligand amphiregulin (AREG) by binding directly to transcription factor CREB. In the next step, AREG activates EGFR signaling to support the growth and survival of tumor cells, which is why anti-EGFR agents in PMEC-targeted therapy are highly efficient. Additionally, Clauditz et al. [40]

TABLE 2: The expression of PD-1, PD-L1, and PD-L2 in MEC.

Ref.	Number of cases	Positions	PD-1 (%)	PD-L1 (%)	PD-L2 (%)
[63]	41	Lungs	63.4	0	
[64]	27	Salivary gland	81.5	25.9	
[65]	9	Salivary gland	NM	55.6	
[66]	7	Salivary gland	NM	57.1	
[67]	34	Salivary gland	NM	9	

Low-grade: cytomembrane of squamous cells and intermediate cells shows medium or higher intensity (N = 8).
High-grade: cytomembrane of squamous cells and intermediate cells shows focal positive (N = 33)

observed that the HER2 positive group is characterized by high-level gene amplification; thus, trastuzumab may have a response. According to the immunohistochemical results of Wang et al. [49], PI3K/mTOR inhibitors may have a therapeutic effect, resulting from 52% high-grade tumors were observed gene alternations in PI3K/mTOR pathway. Recently, a multicenter phase 2 study has looked at the effect of nintedanib in patients with recurrent or metastatic salivary gland cancer of the head and neck [62]. This work revealed a promising clinical efficacy and achieved a 75% disease-control rate (15/20) of nintedanib to treat this condition.

The knowledge on immunotherapy for P MEC is limited so far. Undoubtedly, programmed death-1, programmed death ligand-1, and programmed death ligand-2 (PD-1, PD-L1, and PD-L2) are the most studied immune pathway targets in various carcinomas. Some of the literature about salivary MEC could show a directional effect. The expression rates of PD-1, PD-L1, and PD-L2 are summarized in Table 2. In studies with small sample sizes, the positive rate of PD-L1 is approximately 50%–60%, whereas the rate of PD-L2 is rather low. However, contrary results of PD-L1 are reported in large sample studies. Liu et al. [63] found that the intensity of PD-L2 expression had a positive relation with the histological grade. Besides, PD-L2 is likely to be associated with tumor recurrence [63, 64]. It can be predicted that P MEC is a tumor with low expression of PD-L1, whose evasion mechanisms are likely related to PD-L2.

The KEYNOTE-028 phase IB trial measured pembrolizumab against advanced PD-L1-positive SGCs. Among SGC patients (3 MECs), 26 PD-L1 were positively treated with pembrolizumab and the overall response rate (ORR) was 12%. The trial reported that only three patients had partial responses (PRs) [68]. Tumor mutational burden (TMB) can estimate tumor neoantigen load [69]; therefore, cancer with high TMB has strong immunogenicity [70]. The MyPathway trial observed that one high-TMB MEC patient achieved PR with the treatment of atezolizumab [71]. PD-L1 and TMB are reliable biomarkers to evaluate the curative effects of immunotherapy [69], what yet has low-to-moderate immunogenicity in the prevalent MEC. Consequently, its potential as the target for current immunotherapy seems not to be remarkable [72], whereas immunotherapy aimed at PD-L2 is a potential strategy.

8. Prognosis

P MEC is a relatively inert tumor whose prognosis is usually considered optimistic. The 5-year OS of P MEC is approximately 45% to 70% in general [2, 8, 31, 73], although it is strongly influenced by the TMN stage and the pathological grade [2, 20, 27, 31]. The survival outcome of patients with P MEC seems to be better than that of patients with small-cell lung cancer (SCLC) and non-small-cell lung cancer (NSCLC) [20, 74, 75]. In the study by Cheng et al. [27], low-grade tumors are much more common in the younger age group, while high-grade ones are common in the older group. Huo et al. [8] concluded that age <50 years old, central/endobronchial growth pattern, tumor size <3 cm,

low-grade tumor, Ki-67 labeling index <10%, and complete resection indicated a better OS and prognosis. Qiu et al. [20] drew a similar conclusion. They primarily employed CCS to predict survival curves, and their multivariate Cox analysis revealed that age >60 years, poor differentiation, tumor sizes >30 mm, lymph node metastases, and distant metastases were independent factors of a poor prognosis. In the study by Hsieh et al. [2], the tumor pathological grade influenced neither disease-free survival (DFS) nor OS, differing from previous studies. Park et al. [30] proposed this to be a factor of adverse prognosis. Tumors with greater than 6.5 SUV_{max} are more likely to be high-grade tumors, appeared more often in lymph nodes and distant metastasis, and deduced worse survival outcomes. Press et al. [41] showed that *HER2/neu* immunostaining and amplification were predictors of a poor prognosis independent of pathological grades, tumor sizes, and lymph node metastasis. Mukaigawa et al. [67] observed that the prognosis of PD-L1-positive patients was significantly worse. Moreover, PD-L1 expression is associated with poor DFS.

MECT1-MAML2 rearrangement is more common in low-grade P MEC. Similarly, the survival rate of patients with low-grade P MEC is significantly higher than that of patients with high-grade P MEC. Hence, MECT1-MAML2 rearrangement seems to indicate a better prognosis. However, some studies suggested that not only MECT1-MAML2 rearrangements were correlated with P MEC pathological grading, but also the translocation status was irrelevant to prognosis [8, 76].

In summary, age \geq 50–60 years, tumor size \geq 3 cm, poor pathological differentiation, Ki-67 labeling index \geq 10%, $SUV_{max} >$ 6.5, HER-2/*neu* immunostaining and amplification, PD-L1-positive, lymph node metastases, and distant metastases are associated with poor prognostic factors in P MEC.

9. Conclusion

P MEC, first reported by Smetana in 1952, is a rare primary pulmonary malignant neoplasm. Detailed results of clinical characteristics, epidemiological features, treatment, and prognosis are summarized in Supplementary Table 1. As the most common malignant salivary gland tumor, the survival outcomes of patients with P MEC seem to be better than those of patients with NSCLC and SCLC; however, accurate and early diagnosis plays critical roles. There is no clear diagnostic clinical symptom that has been related to P MEC. Most patients with P MEC present symptoms of bronchial obstruction, while several asymptomatic patients were only diagnosed with P MEC during a physical examination. CT is a necessary approach for the diagnosis and differential diagnosis: a tumor with well-defined mass and oval or round shape and a smooth margin, central type or hilar type, and marked enhancement more possibly diagnosed as P MEC. P MEC is histopathologically characterized by three cell types: squamous, intermediate, and mucus-secreting cells, classified into low-grade and high-grade histological appearance, mitotic frequency, cellular atypia, and necrocytosis. Immunohistochemical findings show that p63, p40,

CK5/6, and CK7 are usually positive, while TTF-1 and napsin A are negative. A gene mutation in *t* (11; 19) (*q21; p13*) generates the fusion protein MECT1-MAML2, which is proposed to drive P MEC onset. Therefore, we can identify some diseases by using IHC and molecular examination. In the meantime, an EGFR mutation (exon 21 L861Q heterozygous mutation) is also certified to exist in P MEC. It has been demonstrated to be the molecular interpretation of EGFR signal activation that the MECT1-MAML2 fusion protein upregulates expression of AREG by direct binding to CREB. It provides clinical evidence for the effectiveness of TKI therapy. Some other gene mutations may lead to custom treatment options. It may provide new directions for future studies. There is temporarily no consolidated standard for treating P MEC, and surgical resection is the mainstay of treatment for low-grade P MEC. The effects of chemotherapy or radiotherapy are undefined. They could be used for patients with high-grade tumors with extrathoracic invasion. TKI therapy such as gefitinib and erlotinib had therapeutic responses. An immunotherapy approach for P MEC has powerful potential and needs to be explored in depth. The adverse prognostic factors are age ≥ 50 –60, tumor size ≥ 3 cm, poor differentiation, Ki-67 labeling index $\geq 10\%$, $SUV_{max} > 6.5$, HER-2/neu immunostaining and amplification, PD-L1-positive, lymph node metastases, and distant metastases.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Shumin Hu and Jiali Gong authors contributed equally to this work. Shumin Hu and Jiali Gong are co-first author.

Acknowledgments

This study was supported by the Wu Jieping Medical Foundation (320.6750.2021-01-50).

Supplementary Materials

Supplementary Table 1: detailed results of clinical characteristics, epidemiological features, diagnostic modality, the follow-up time, treatment, and prognosis of P MEC in the literature. (*Supplementary Materials*)

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