Review Article

The Role of Praziquantel in the Prevention and Treatment of Fibrosis Associated with Schistosomiasis: A Review

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Schistosomiasis remains a major global public health concern. Currently, the control of this neglected tropical disease still depends on chemotherapy to reduce the prevalence and intensity of the parasite infection. It has been widely accepted that praziquantel is highly effective against all species of Schistosoma, and this agent is virtually the only drug of choice for the treatment of human schistosomiasis. Mass drug administration (MDA) with praziquantel has been shown to be effective in greatly reducing the prevalence and morbidity due to schistosomiasis worldwide. In addition to antischistosomal activity, a large number of experiential and clinical evidence has demonstrated the action of praziquantel against fibrosis caused by S. mansoni and S. japonicum infections through decreasing the expression of fibrotic biomarkers such as α-smooth muscle actin (α-SMA), collagen, matrix metalloproteinase (MMP), and tissue inhibitor of metalloproteinase (TIMP), and inhibiting the expression of proinflammatory cytokines such as interleukin (IL)-6, tumor necrosis factor (TNF)-α, and transforming growth factor (TGF)-β, as well as chemokines, and similar anti-fibrotic activity was observed in mouse models of fibrosis induced by carbon tetrachloride (CCI4) and concanavalin A (Con-A). In this review, we discuss the role of praziquantel in the prevention and treatment of fibrosis associated with schistosomiasis and the possible mechanisms. We call for randomized, controlled clinical trials to evaluate the efficacy and safety of praziquantel in the treatment of schistosomiasis-induced hepatic fibrosis, and further studies to investigate the potential of praziquantel against fibrosis associated with alcohol consumption, viruses, and toxins seem justified.

1. Introduction

Schistosomiasis, a neglected tropical disease caused by the blood fluke of the genus Schistosoma, remains a global public health concern that ranks second to malaria among human parasitic diseases in terms of the number of people infected and at risk of infection [1]. Worldwide, it is estimated that over 140 million people are thought to have the disease, with a further 779 million at risk of infection [2]. Although the elimination of this tropical parasitic disease requires a multidisciplinary integrated approach [3–5], the control of schistosomiasis still relies on chemotherapy, which has been proven to be effective in reducing the prevalence and intensity of the parasitic infection [6–8].

Hepatosplenic schistosomiasis is characterized by typical pathological changes at chronic and advanced stages, including egg granulomas, fibrosis, and tissue damage within the liver and other host tissues [1]. Hepatic fibrosis, which is characterized by excessive deposition of collagen and other extracellular matrix (ECM) components resulting from granulomatous responses triggered by soluble egg antigens (SEA) secreted by parasite eggs, is the main pathological mechanism and the major lesion of hepatosplenic schistosomiasis [9]. Hepatic stellate cells (HSCs), hepatic macrophages, immune cells, cytokines (interleukin (IL)-13, transformation growth factor (TGF)-β1, interferon (IFN)-γ), and microRNAs (miRNAs) have been found to contribute to the pathogenesis of schistosomiasis-induced hepatic fibrosis [10–17]. Portal hypertension and ascites associated with hepatic fibrosis have been identified as the main causes of mortality among patients with chronic hepatosplenic schistosomiasis [18], and currently, there is no cure for hepatic fibrosis except liver transplantation [19]. However, liver transplantation suffers from problems of lack
of donors, high costs, and use of immunosuppressive agents, which limits its clinical applications [20]. A search for novel treatments for hepatic fibrosis is therefore given a high priority.

Since praziquantel, a broad-spectrum schistosomicide, was developed in 1970s [21], it has replaced other anti-schistosomal agents to become the only drug of choice for treatment of human schistosomiasis due to high efficacy, low toxicity, easy administration, and low cost [22–24]. The agent is found to be active against all species of *Schistosoma*, notably, adult stages of the parasite [25]. As a consequence, the introduction of praziquantel led to the global schistosomiasis control strategy shifting from disease control to morbidity control [26]. Mass drug administration (MDA) with praziquantel has been shown to be effective in reducing the prevalence and morbidity due to schistosomiasis [27–29]. Furthermore, an antifibrotic activity of praziquantel was reported in both animal models and patients infected with schistosomiasis [30,31]. This review article aims to discuss the role of praziquantel in the prevention and control of fibrosis associated with schistosomiasis and the possible mechanisms.

2. Literature Search Strategy

A joint search was performed in international and national electronic databases, including Web of Knowledge, PubMed, Scopus, Google Scholar, Wanfang Data (https://www.wanfangdata.com.cn/), CNKI (https://www.cnki.net) and VIP (https://qikan.cnki.net) using the terms “schistosomiasis,” “fibrosis,” and “praziquantel” to retrieve publications concerning the action of praziquantel against fibrosis associated with schistosomiasis during the period from January 1st, 1970 to December 31st, 2021. Inclusion criteria involved: (1) studies reporting the activity of praziquantel against fibrosis associated with schistosomiasis; and (2) animal studies or clinical reports. Publications that met the following exclusion criteria were: (1) review articles; or (2) the full text was unavailable.

2.1. Experimental Evidence. In a murine model of *S. mansoni*-induced hepatic fibrosis, administration of praziquantel at a dose of 250 mg/kg modestly diminished liver fibrosis as compared to untreated controls 10 weeks post-treatment and suppressed fibrosis and reduced liver collagen content to normal levels 20 weeks post-treatment [32]. In Swiss albino mice experimentally infected with *S. mansoni*, praziquantel treatment resulted in a reduction in total collagen content and a recovery of the type III (Col3)/I collagen (Col1) ratio to normal limits [33]. In Syrian golden hamsters infected with 100 *S. mansoni* cercariae each, a significant reduction in hepatic and splenic granulomas, fibrosis, and circulating cathodic antigen (CCA), and circulating anodic antigen (CAA) was seen following praziquantel treatment [34]. Following praziquantel administration, amelioration of hepatic granulomas and reduction of Col1, Col3, and Col4 gene expression were observed 6 and 12 months post-treatment, and 71.4% resorption of hepatic fibrous tissues was found 12 months post-treatment in a CBA/J mouse model of *S. mansoni* infections [35]. In Swiss Webster outbred mice infected with *S. mansoni*, treatment with praziquantel resulted in elimination of egg granulomas or collagen fibrils, and reduced the expression of signal transducer and activator of transcription 1 (STAT1), STAT4, IFN-γ, T-BET, IL-4, C-C motif chemokine ligand 12 (CCL12), and CCL22 [36]. In addition, treatment with praziquantel at a dose of 80 mg/kg 50 days postinfection for 5 consecutive days was reported to result in a significant reduction in the volume density of hepatocytes, sinusoids, and hepatic fibrosis in mice infected with *S. mansoni* and fed either a low-protein diet (8% protein) or standard chow (22% protein) [37]. In a recent study, however, treatment with praziquantel at a dose of 400 mg/kg twice daily 12 weeks postinfection for a week followed by praziquantel treatment at a dose of 400 mg/kg twice daily for 4 weeks was found to achieve comparable collagen deposition, hydroxyproline level, and granuloma areas relative to untreated controls in the murine model of chronic experimental schistosomiasis mansoni [38] (Table 1).

In addition, combined therapy of praziquantel with alpha lipoic acid [39], silymarin [40], AT1 receptor antagonist losartan [41], was reported to achieve a greater activity against hepatic fibrosis induced by *S. mansoni* infection than praziquantel monotherapy. Since stem cell therapy has shown a potential value in treatment of schistosomiasis-associated fibrosis [42–44], the antifibrotic activity of human Wharton’s jelly-derived mesenchymal stem cells in combination with praziquantel was evaluated in *S. mansoni*-infected mice, and the combination was found to achieve a higher beneficial efficacy against *S. mansoni*-induced liver fibrosis than monotherapy with mesenchymal stem cells or praziquantel alone, as revealed by histopathological, morphometric, and gelatin zymographic results as well as reduction of alpha smooth muscle actin (α-SMA), Col1, and IL-13 expression [45].

In animal models of fibrosis induced by *S. japonicum*, praziquantel treatment was found to reduce serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) concentrations, the areas of egg granulomas, areas of collagen deposition and hepatic hydroxyproline contents, alleviate fibrotic proliferation, inflammatory infiltration, hepatocyte degeneration and necrosis, improve liver size and texture, decrease the levels of fibrosis-related parameters including α-SMA, SPET4, matrix metalloproteinase (MMP)-9, tissue inhibitor of metalloproteinase (TIMP)-1, Col1α1, and Col3α1 in relative to infected but untreated controls, and inhibit the expression of proinflammatory cytokines tumor necrosis factor (TNF)α and TGF-β1 [31, 46–52] (Table 2). Combined treatment with praziquantel and IFN-γ was reported to result in a greater decline in hepatic fibrosis score and area of collagen deposition, higher reduction of Col1 and Col3 expression, more down-regulation of Smad2 expression and upregulation of Smad7 expression as compared to treatment with praziquantel alone [53, 54], and the combination of praziquantel and the *Amusium pleuronectes* polysaccharide presented a more inhibitory effect on hepatic fibrosis than treatment with
praaziquantel or the polysaccharide alone in *S. japonicum*-infected mice, as revealed by lower Col1, Col2, and Col4 levels, lower parasite egg burdens and higher IL-12 and IFN-γ productions [55]. In addition, combining praziquantel and extracts from traditional Chinese medicines achieved synergistic activity in the treatment of *S. japonicum*-induced hepatic fibrosis [56–58].

The results of these experimental studies demonstrate the effectiveness of praziquantel against hepatic fibrosis induced by *S. mansoni* or *S. japonicum* infection, which encourages the randomized, controlled clinical trials to confirm the antifibrotic activity of praziquantel.

2.2. Exciting Findings from Human Trials. To date, there are no randomized, controlled trials to examine the role of praziquantel in prevention and control of fibrosis associated with schistosomiasis; however, available data have shown the antifibrotic value of praziquantel in treatment of patients with schistosomiasis-induced fibrosis [30, 59]. In *S. mansoni*-infected subjects, administration of praziquantel at a total dose of 40 or 50 mg/kg resulted in improvements in the ultrasonographic parameters of fibrosis [60–64] (Table 3). Among *S. japonicum*-infected patients, praziquantel treatment at a total dose of 40 or 60 mg/kg was found to improve the ultrasonographic and biochemical parameters in patients with mild fibrosis but not in patients with severe fibrosis [30, 65–68] (Table 4); however, Fabre and colleagues reported an elevated prevalence of hepatic fibrosis in *S. japonicum*-infected Filipinos following one-year treatment of praziquantel at a total dose of 60 mg/kg given in a split dose [69]. Furthermore, multicenter, randomized, double-blind, and controlled clinical trials are required to validate the antifibrotic efficacy of praziquantel.

<table>
<thead>
<tr>
<th>Animal model</th>
<th>Pattern of fibrosis</th>
<th>Treatment regimen</th>
<th>Effect on schistosomiasis-induced fibrosis</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mouse</td>
<td>Hepatic fibrosis</td>
<td>Administration of praziquantel at a dose of 250 mg/kg 8 weeks postinfection</td>
<td>A prompt reduction in liver parasite egg load with no viable eggs, and a moderate decrease in liver fibrosis 10 weeks post-treatment; and arrest of fibrosis and reduction of liver collagen content to normal levels by 20 weeks post-treatment</td>
<td>32</td>
</tr>
<tr>
<td>Swiss albino mouse</td>
<td>Hepatic fibrosis</td>
<td>Administration of praziquantel at a dose of 250 mg/kg 8, 12, and 20 weeks postinfection</td>
<td>A reduction of total collagen content and recovery of type III/I collagen ratio to normal limits following treatment given 8 weeks postinfection</td>
<td>33</td>
</tr>
<tr>
<td>Syrian golden hamster</td>
<td>Hepatic fibrosis and splenic fibrosis</td>
<td>Administration of praziquantel at a dose of 100 mg/kg 12, 13, 14, and 15 weeks postinfection</td>
<td>A significant reduction of granulomas, CAA and CCA in hepatic specimens and a clear-cut reduction of fibrosis, granulomas, CAA and CCA in splenic specimens</td>
<td>34</td>
</tr>
<tr>
<td>CBA/J mouse</td>
<td>Hepatic fibrosis</td>
<td>Administration of praziquantel at a dose of 300 mg/kg for 2 days 8 weeks postinfection</td>
<td>Amelioration of hepatic granulomatous pathology, reduction of collagen I, III, and IV gene expression at 6 and 12 months post-treatment, and resorption of liver fibrous tissue by 71.4% 12 months post-treatment</td>
<td>35</td>
</tr>
<tr>
<td>Swiss webster mice</td>
<td>Hepatic fibrosis</td>
<td>Administration of praziquantel at a dose of 250 mg/kg 32 days postinfection for 4 consecutive days</td>
<td>Elimination of egg granulomas or collagen fibrils, and a reduction in the expression of STAT1, STAT4, IFN-γ, Tbet, IL-4, CCL12, and CCL22</td>
<td>36</td>
</tr>
<tr>
<td>Swiss webster mice</td>
<td>Hepatic fibrosis</td>
<td>Treatment with praziquantel at a dose of 80 mg/kg 50 days postinfection for 5 consecutive days</td>
<td>A significant reduction in the volume density of hepatocytes, sinusoids, and hepatic fibrosis</td>
<td>37</td>
</tr>
<tr>
<td>Balb/C mouse</td>
<td>Hepatic fibrosis</td>
<td>Treatment with praziquantel at a dose of 400 mg/kg twice daily 12 weeks postinfection for a week followed by praziquantel treatment at a dose of 400 mg/kg twice daily for 4 weeks</td>
<td>No significant reduction in collagen deposition, hydroxyproline level, or granuloma areas</td>
<td>38</td>
</tr>
</tbody>
</table>

CAA, circulating anodic antigen; CCA, circulating cathodic antigen; STAT, signal transducer and activator of transcription; IFN, interferon, IL, interleukin; CCL, C–C motif chemokine ligand.
expression at both translational and transcriptional levels [48], and increased plasminogen activator activity [71], which was considered to contribute to the reversal of schistosomiasis-induced fibrosis. Since multiple factors are involved in schistosomiasis-induced fibrosis, the exact mechanisms underlying the action of praziquantel against schistosomiasis-induced fibrosis require further investigations.

### Table 2: Activity of praziquantel monotherapy against hepatic and splenic fibrosis associated with schistosomiasis japonica.

<table>
<thead>
<tr>
<th>Animal model</th>
<th>Pattern of fibrosis</th>
<th>Treatment regimen</th>
<th>Effect on schistosomal fibrosis</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>BABL/c mouse</td>
<td>Hepatic fibrosis</td>
<td>Praziquantel at a dose of 300 mg/kg twice daily for 30 days administered 8 and 15 weeks postinfection</td>
<td>A significant reduction in the areas of sirius red-stained liver, liver hydroxyproline contents, spleen weight, spleen index, and levels of Col1α1, Col3α1, α-SMA, MMP9, and TIMP1 as compared to infected but untreated controls</td>
<td>31</td>
</tr>
<tr>
<td>Rabbit</td>
<td>Hepatic fibrosis</td>
<td>Oral administration of praziquantel at a single dose of 100 mg/kg 6, 12 or 24 weeks postinfection</td>
<td>A significant decrease in portal vein pressure, number and size of egg granulomas, and liver collagen content, and improvement of echogenic bands and nodules</td>
<td>46</td>
</tr>
<tr>
<td>Kunming mouse</td>
<td>Hepatic fibrosis</td>
<td>Administration of praziquantel at a daily dose of 500 mg/kg for 2 days, followed by praziquantel treatment twice a week for 8 weeks</td>
<td>Significantly reduced egg granulomas area, type I and type III collagen levels, and ALT and AST activities, remarkable improvements in liver size and texture, and significantly reduced hepatic fibrosis degree as compared to infected but untreated controls</td>
<td>47</td>
</tr>
<tr>
<td>ICR mouse</td>
<td>Hepatic fibrosis</td>
<td>Praziquantel given at a daily dose of 250 mg/kg for 3 days 6 weeks postinfection</td>
<td>A clear-cut decline in diameters of egg granulomas, areas of collagen deposition and α-SMA expression, inhibition of TNF-α and IL-6 mRNA expression, and reduced SEPT4 expression at transcriptional and translation levels as compared to infected but untreated controls</td>
<td>48</td>
</tr>
<tr>
<td>Kunming mouse</td>
<td>Hepatic fibrosis</td>
<td>Administration of praziquantel at a daily dose of 500 mg/kg for 2 weeks postinfection</td>
<td>Alleviated fibrotic proliferation and inflammatory infiltration, significantly reduced egg granulomas and hepatocyte degeneration and necrosis, and significantly decreased serum NO, hepatic inducible nitric oxide synthase (iNOS), TGF-β1, type I and type III collagen, and TNF-α levels, and significantly elevated Bcl-2 and INF-γ levels as compared to infected but untreated controls</td>
<td>49–52</td>
</tr>
</tbody>
</table>

Col1α1, collagen Iα1; Col3α1, collagen IIIα1; α-SMA, α-smooth muscle actin; MMP9, matrix metalloproteinase 9; TIMP1, metalloproteinase-1; TNF-α, tumor necrosis factor α; TGF-β1, transforming growth factor β1; IL-6, interleukin-6; INF-γ, interferon-γ; ALT, alanine transaminase; AST, aspartate transaminase.

### Table 3: Clinical efficacy of praziquantel in treatment of patients with schistosomiasis mansoni-induced fibrosis.

<table>
<thead>
<tr>
<th>Pattern of fibrosis</th>
<th>Treatment regimen</th>
<th>Clinical outcomes of fibrosis</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hepatic periportal fibrosis</td>
<td>Praziquantel at a total dose of 40 mg/kg</td>
<td>Improvement of periportal thickening/fibrosis in 17.6% of the cohorts, total resolution in 34.7% 26-months post-treatment</td>
<td>60</td>
</tr>
<tr>
<td>Hepatic periportal fibrosis</td>
<td>Praziquantel at a total dose of 40 mg/kg</td>
<td>Reduction of hepatic periportal thickening from 46% at baseline to 32% one-year post-treatment and 35% three-year post-treatment</td>
<td>61</td>
</tr>
<tr>
<td>Hepatic periportal fibrosis</td>
<td>Praziquantel at a total dose of 50 mg/kg</td>
<td>Significant reduction in the mean values for longitudinal and anteroposterior measurements of liver (left and right lobes) as well as portal vein diameter, a considerable reduction in moderate fibrosis and IL-10 level one-year post-treatment</td>
<td>62</td>
</tr>
<tr>
<td>Hepatic periportal fibrosis</td>
<td>Praziquantel at a total dose of 40 mg/kg</td>
<td>Complete reversal of periportal lesions seen in 28.2% subjects; a reduction in periportal thickening followed by a decrease in the size of the left hepatic lobe 3 to 5 years post-treatment</td>
<td>63</td>
</tr>
<tr>
<td>Hepatic periportal fibrosis</td>
<td>Praziquantel at a total dose of 40 mg/kg</td>
<td>Improvement in the sonomorphological abnormalities of periportal fibrosis and organomorphometry of livers and spleens one-year post-treatment</td>
<td>64</td>
</tr>
</tbody>
</table>
3. Conclusions and Perspectives

Currently, there are no effective treatments for hepatic fibrosis except liver transplantation [19]. Early diagnosis and interventions are of great importance to attenuate hepatic fibrosis; however, there are still challenges for early diagnosis of hepatic fibrosis [72]. Nevertheless, artificial intelligence (AI) opens the new era of precision medicine in hepatology, which facilitates precise identification and prediction of disease severity and progression, the presence of complications, and outcomes of hepatic fibrosis [73–75]. In addition, stem cell therapy has shown great potential in the treatment of hepatic fibrosis, which provides a new hope for the treatment of hepatic fibrosis [76–78]; however, further prospective clinical trials to examine the efficacy and safety of stem cell therapy for hepatic fibrosis associated with schistosomiasis are required.

It has been proven that praziquantel is a highly effective and mildly toxic schistosomicide [79], and praziquantel-based chemotherapy has been widely implemented in the national schistosomiasis control program around the world and plays a great role in the transmission of schistosomiasis [80]. In addition to antischistosomal action, both experimental studies and clinical trials have revealed the antifibrotic activity of praziquantel against hepatic fibrosis induced by *S. japonicum* and *S. mansoni* infection [47–52, 61–68]. Moreover, oral administration of praziquantel at a dose of 300 mg/kg given twice daily for 30 days resulted in a significant reduction of the collagen areas, content of hepatic hydroxyproline, and serum ALT and AST levels in a CCl₄-induced mouse model of hepatic fibrosis [30]. In the concanavalin A (Con A)-induced model of hepatic fibrosis, praziquantel was also found to improve the morphological and biochemical parameters associated with hepatic fibrosis [81]. Since hepatic fibrosis may be induced by virus, alcohol and toxin [82–84], and there are millions of subjects suffering from hepatic fibrosis due to viral hepatitis, parasitic diseases, toxic chemicals, and alcohol consumption, further studies to investigate the potential of praziquantel against alcoholic, viral, and toxin-induced fibrosis seem justified.

In summary, praziquantel, an old schistosomicide, is a promising antifibrotic drug. Although randomized, controlled clinical trials are required to validate the antifibrotic action, and there are still a large number of challenges ahead, the antifibrotic activity of praziquantel may benefit thousands of patients, due to its low cost, easy administration, and low toxicity. In addition, the timing, dosing, optimal regimens, and mechanisms of action of praziquantel for treatment of fibrosis require further investigation.

### Data Availability

All data presented in this study are available from the corresponding author upon request.

### Conflicts of Interest

The authors declare that they have no conflicts of interest.

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