Research Article
MiR-6869-5p Induces M2 Polarization by Regulating PTPRO in Gestational Diabetes Mellitus

Pingping Wang,1 Zhenzhi Ma,2 Zengyan Wang,3 Ximei Wang,1 Guifeng Zhao4, and Zengfang Wang1

1Department of Gynecology and Obstetrics, the Affiliated Hospital of Maternal and Child Health, Weifang Medical University, Weifang 261000, China
2Department of Pharmacy, the First Affiliated Hospital of Weifang Medical University, Weifang 261000, China
3Operating Room, Zhucheng People’s Hospital, Zhucheng 262200, China
4Department of Antenatal Diagnosis, the First Affiliated Hospital of Maternal and Child Health, Weifang Medical University, Weifang 261000, China

Correspondence should be addressed to Guifeng Zhao; wfrmzhaogf@126.com and Zengfang Wang; wangzf123@yeah.net

Received 17 December 2020; Revised 10 March 2021; Accepted 1 April 2021; Published 3 May 2021

Academic Editor: Jie Chen

The role of microRNA (miRNA) in gestational diabetes mellitus has been widely investigated during the last decade. However, the altering effect of miR-6869-5p on immunity and placental microenvironment in gestational diabetes mellitus is largely unknown. In our study, the expression of miR-6869-5p was documented to be significantly decreased in placenta-derived mononuclear macrophages, which was also negatively related to PTPRO. Besides, PTPRO was negatively regulated by miR-6869-5p in placenta-derived mononuclear macrophages. In vitro, miR-6869-5p inhibited macrophage proliferation demonstrated by EdU and CCK-8 experiments. The inflammatory response in macrophages was also significantly inhibited by miR-6869-5p, which could regulate PTPRO as a target documented by luciferase reporter assay. Moreover, miR-6869-5p promoted M2 macrophage polarization and thus restrain inflammation. Accordingly, miR-6869-5p is involved in maintaining placental microenvironment balance by preventing from inflammation and inducing M2 macrophages in gestational diabetes mellitus.

1. Introduction

Gestational diabetes mellitus is a common complication in pregnant females, who usually have normal glucose metabolism or potential abnormal glucose tolerance [1]. The incidence of gestational diabetes mellitus has been increasing during the past few decades. Most patients with gestational diabetes mellitus can return to normal glucose metabolism postpartum, but they are at an elevated risk of developing type II diabetes in the future. The pathogenesis of gestational diabetes mellitus is unclear yet. Most importantly, abnormal glucose metabolism during gestation can cause pregnancy failure, dystocia, stillbirth, fetal death, and fetal macrosomia increased owing to complicated factors [2]. Therefore, identifying useful way for gestational diabetes mellitus prevention and treatment is urgent.

Mononuclear macrophages are key cells involved in regulating placental immunity and homeostasis. Macrophages can be induced to differentiate into classically activated M1 cells and alternatively activated M2 cells under specific regulators and microenvironment in placenta [3, 4]. Type 2 macrophages (M2) play a critical role in maintaining placental microenvironment balance [5]. Accumulated studies have suggested that microRNAs (miRNAs) participate in the pathogenesis of gestational diabetes mellitus by regulating β cell development, insulin sensitivity, and resistance [6, 7]. It has been well documented that some miRNAs can affect macrophages differentiation and polarization, such as miR-657, miR-145, and miR-221-3p [8–11]. We have previously found miR-657 could induce macrophages to M1 in gestational diabetes mellitus, which is a promising target for disease diagnosis and treatment [8, 12]. MiR-6869-5p has been
firstly found as a cancer suppressor [13]. There is no evidence showing that miR-6869-5p is attributed to gestational diabetes mellitus pathogenesis. We have previously found that the expression of miR-6869-5p was significantly decreased in placenta-derived mononuclear macrophages, which was also negatively related to PTPRO. The bioinformatics analysis has suggested that PTPRO is a potential target of miR-6869-5p. However, little is known about the influence of miR-6869-5p in placental macrophages differentiation and function. In this study, we aim to elucidate the altering effect and potential molecular mechanism of miR-6869-5p on immunity and placental microenvironment in gestational diabetes mellitus, which will provide insight into the disease pathogenesis and gaining promising strategies for disease prevention and therapy.

2. Material and Methods

2.1. Participants and Samples Preparation. Participants with gestational diabetes mellitus (26 cases) and normal pregnancies (23 controls) were enrolled. Inclusion and exclusion criteria are as follows: pregnant women with GDM having full-term cesarean section are enrolled as the case group, while healthy women having full-term pregnancy are regarded as the control group. Those pregnancies with premature or overdue births are all excluded. Table 1 lists the characteristics of participants. The study is approved by the hospital’s Institutional Ethics Committee of Affiliated Hospital of Maternal and Child Health, Weifang Medical University. Participants have signed the informed consent. Macrophages are freshly separated from placental tissues, which are divided into small pieces before separation. Placental tissue suspension is used for isolating cells by density gradient centrifugation. Then, we used CD14 positive microbeads (Biologend, USA) for macrophage purification.

2.2. Cell Line. Human THP-1 cells are cultured in RPMI 1640 with 10-20% fetal bovine serum (Gibco, USA). PMA (100 nM, Sigma, USA) is used to stimulated cells for 48 h to make them differentiate into macrophages. miR-6869-5p mimics and miR-6869-5p mimic control are used to coculture with macrophages, which are purchased from Genechem Company (Shanghai, China). Lentivirus plasmids with or without overexpression of miR-6869-5p in placental macrophages were used to coculture and make them differentiate.

2.3. CCK-8 and EdU. CCK-8 kit (Vazyme Biotech Nanjing, China) is used to assess cell proliferation. In brief, macrophages (5 × 10^4/ml per well) with or without overexpression of PTPRO are seeded into 96-well plate for 12 h and, then, are treated with miR-6869-5p mimics and miR-6869-5p mimic control for 24 h and 48 h. 10 μl CCK-8 reagent solution is finally detected.

2.4. Real-Time PCR. Trizol reagent (Invitrogen, USA) is used to isolate RNAs from placental tissue macrophages and

### Table 1: Characteristics of patients and controls.

<table>
<thead>
<tr>
<th></th>
<th>Patients</th>
<th>Controls</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>n</td>
<td>26</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>Age (y)</td>
<td>30.6 ± 4.4</td>
<td>29.2 ± 3.5</td>
<td>0.808</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>70.8 ± 5.5</td>
<td>64.6 ± 4.9</td>
<td>0.409</td>
</tr>
<tr>
<td>Infant weight (g)</td>
<td>3708 ± 110.2</td>
<td>3528 ± 124.5</td>
<td>0.283</td>
</tr>
<tr>
<td>Gestational weeks</td>
<td>38.1 ± 1.2</td>
<td>39.4 ± 1.2</td>
<td>0.449</td>
</tr>
<tr>
<td>Blood pressure</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>119.3 ± 11.1</td>
<td>102.4 ± 10.3</td>
<td>0.274</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>71.9 ± 7.2</td>
<td>65.2 ± 8.1</td>
<td>0.538</td>
</tr>
<tr>
<td>Glucose metabolism</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fasting glucose (mmol/L)</td>
<td>5.1 ± 1.0</td>
<td>4.2 ± 0.8</td>
<td>0.043</td>
</tr>
<tr>
<td>1 h glucose (mmol/L)</td>
<td>10.8 ± 1.7</td>
<td>6.09 ± 0.72</td>
<td>0.019</td>
</tr>
<tr>
<td>2 h glucose (mmol/L)</td>
<td>8.5 ± 1.4</td>
<td>5.3 ± 1.1</td>
<td>0.034</td>
</tr>
<tr>
<td>Fasting insulin (mIU/L)</td>
<td>9.6 ± 1.1</td>
<td>7.9 ± 1.3</td>
<td>0.320</td>
</tr>
</tbody>
</table>

THP-1 cells according to the protocol. RNA (0.5 μg) is used as the template for cDNA synthesis based on the protocol of PrimeScriptTM RT Kit (Takara, Beijing, China). cDNA is applied for PCR amplification by use of Takara SYBR Premix. TaqMan PCR assay kit (ThermoFisher Scientific, USA) is used to determine the expression of miR-6869-5p in cells. Primers are as follows: Human PTPRO, F, TATTGTGAGCCTCCGT GTGT; R, GCCAAGGCTTCTTCACTGACA. Human IL-1β, F, ACATGACCACTGTACGATCA; R, TCATTCAACAC GCAGGACAGC.

Human TNF-α, F, CCGTGAAAACACCCCTCAGA; R, AAGAGGCTGAGGAACAAGCA. Human GAPDH, F, CCCTGAAAACAACCCTCAGA; R, TCCACCACCTGT TGCTGTA.

2.5. Enzyme-Linked Immune Sorbent Assay (ELISA). As previously described [12], THP-1 macrophages with or without overexpression of PTPRO are incubated in serum-free medium for 12 h. Subsequently, macrophages are transfected by use of Takara SYBR Premix. TaqMan PCR assay kit (ThermoFisher Scientific, USA) is used to determine the expression of miR-6869-5p in cells. Primers are as follows: Human PTPRO, F, TATTGTGAGCCTCCGT GTGT; R, GCCAAGGCTTCTTCACTGACA. Human IL-1β, F, ACATGACCACTGTACGATCA; R, TCATTCAACAC GCAGGACAGC.

Human TNF-α, F, CCGTGAAAACACCCCTCAGA; R, AAGAGGCTGAGGAACAAGCA. Human GAPDH, F, CCCTGAAAACACCCCTCAGA; R, TCCACCACCTGT TGCTGTA.

2.6. Luciferase Reporter Assay. pGL3 vectors carrying the luciferase reporter gene are used to clone the 3’ untranslated region (3’UTR) of PTPRO (wild and mutant types). The luciferase activity is estimated using the system of dual luciferase reporter assay. Details have been presented in our previous study [8].

2.7. Flow Cytometry. Macrophages from placenta tissues of gestational diabetes mellitus patients with high or low expression of miR-6869-5p are incubated with FITC-conjugated CD14 Ab, and PE-HLA-DR-conjugated Ab, or PE-CD206-conjugated Ab (Biolegend, San Diego, CA, USA) at room temperature for 30 min. Cells are then centrifuged and harvested for detection by flow cytometry. THP-1 macrophages (5 × 10^4/ml) with or without overexpression
of PTPRO are seeded into 24-well plate overnight. Then, cells
by miR-6869-5p mimics or mimics control for another 24 h.
After incubating with PE-HLA-DR-conjuncted Ab or PE-
CD206-conjuncted Ab (Biolegend, San Diego, CA, USA) at
room temperature for 30 min, we harvest cells and apply flow
cytometry for determination.

2.8. Statistical Analysis. Mean ± SEM is used for data calcula-
tion. All results are normally distributed. The GraphPad Soft-
ware and SPSS Software are used. Di-
ferences between two
groups are statistically analyzed by use of independent sam-
ple Student’s T-test for parametric data, while differences
among more than three groups are estimated by ANOVA.

P < 0.05 is considered to be significant.

3. Results

3.1. MiR-6869-5p Was Significantly Decreased in Macrophages
from Placenta and Associated with M2 Macrophages
Polarization. The expression of miR-6869-5p in placenta-
derived macrophages from gestational diabetes mellitus
patients was significantly reduced when comparing with that
in placenta-derived macrophages from normal pregnancies
(Figure 1(a)). Reversely, increased expression of PTPRO was
found in placenta-derived macrophages from patients with
gestational diabetes mellitus (Figure 1(b)). Negative associa-
tion between miR-6869-5p and PTPRO was demonstrated
regarding their expression in placental tissue-derived macro-
phages (Figure 1(c)). We also detected the expression of
CD206+ macrophages and HLA-DR+ macrophages in pla-
cental tissues derived macrophages. Interestingly, CD206+
macrophages but not HLA-DR+ macrophages were signifi-
cantly increased in those patients with high expression of
miR-6869-5p in placental tissue-derived macrophages, while
CD206+ macrophages but not HLA-DR+ macrophages were
significantly decreased in those patients with low expression
of miR-6869-5p in placental tissue-derived macrophages
(Figure 2). Accordingly, we hypothesize that miR-6869-5p
might regulate macrophage polarization in placental immune
microenvironment and induces macrophages towards M2
polarization in gestational diabetes mellitus.

3.2. MiR-6869-5p Inhibited Macrophage Proliferation by
Targeting PTPRO. To assure whether miR-6869-5p could
regulate PTPRO as a target, we screened the Targetscan database and found that miR-6869-5p could recognize the 3’UTR sequence of PTPRO. The predicted consequent pairing of target region (top) and miRNA (bottom) was shown in Figure 3(a). The luciferase reporter assay further demonstrated our hypothesis that PTPRO was a target gene of miR-6869-5p in THP-1 macrophages (Figure 3(b)). In the following experiments, the altering effects of miR-6869-5p on macrophage proliferation were estimated. As evidenced by CCK-8 (Figure 3(c)) and EdU (Figure 3(d)), the proliferation of THP-1 macrophages could be effectively enhanced when PTPRO was overexpressed in macrophages, while miR-6869-5p mimics could successfully rescue its effect because the proliferation of THP-1 macrophages was significantly inhibited in miR-6869-5p treated group compared with miR-6869-5p mimics control group.

3.3. MiR-6869-5p Prevented from Inflammation in Macrophages by Inducing M2 Macrophages. To estimate the effect of miR-6869-5p on macrophage, THP-1 macrophages were treated by miR-6869-5p mimics and simultaneously stimulated by LPS. As shown in Figures 4(a)–4(d), PTPRO overexpression could induce high expression of TNF-α and IL-1β at both levels of mRNA and protein in macrophages. Besides, reduced rate of CD206+ macrophages and elevated rate of HLA-DR+ macrophages was observed when PTPRO was overexpressed in macrophages (Figure 5). However, decreased expression of TNF-α and IL-1β in macrophages was observed when PTPRO-overexpressed cells were treated by miR-6869-5p mimics (Figures 4(a)–4(d)). Moreover, miR-6869-5p mimic-treated macrophages are more likely to differentiate into CD206+ macrophages (Figure 5). Accordingly, miR-6869-5p could rescue the inflammatory response in macrophages induced by PTPRO. Taken together, miR-6869-5p is capable of preventing from inflammation in macrophages by inducing macrophages towards M2.

4. Discussion

Gestational diabetes mellitus is a complication of pregnancy, which poses high risks for both the mother and the fetus. The role of noncoding RNAs in gestational diabetes mellitus has drawing more and more attention in the past few years, such as long noncoding RNA and miRNA [14–16]. Although they do not encode active proteins or polypeptides, many noncoding RNAs are involved in the pathogenesis of gestational diabetes mellitus by RNA-RNA or RNA-protein interactions. Some circulating noncoding RNAs can serve as useful disease biomarkers for the onset and progression of gestational diabetes mellitus [17].

miRNA is a small noncoding RNA, which functions by targeting specific mRNAs. A number of miRNAs have been demonstrated to serve crucial roles in gestational diabetes mellitus by protecting pancreatic β-cell function, affecting insulin resistance, insulin sensitivity as well as liver gluconeogenesis, for instance, miR-143, miR-351, and miR-96 [18–

Figure 2: Ratio of CD206+ macrophages and HLA-DR+ macrophages in placenta of GDM patients compared with normal pregnancies.
The study by Yan et al. has reported that miR-6869-5p was dysregulated in colorectal cancer and contributed to cancer cell proliferation, invasion, and migration by negatively regulating TLR4/NF-κB signaling pathway [13]. Exosome-encapsulated miR-6869-5p has also been demonstrated to participate in cancer [21]. However, the modifying effect of miR-6869-5p in macrophages mediated gestational disorders has not been evaluated. In this study, we aim to investigate the miR-6869-5p involvement in gestational diabetes mellitus. MiR-6869-5p is significantly downregulated in placenta derived macrophages from gestational diabetes mellitus patients. It is involved in regulating placental immune microenvironment and inducing macrophages towards M2. MiR-6869-5p prevents from macrophage proliferation and inflammation by targeting PTPRO and promoting macrophages polarization to M2 cells. Accordingly, miR-6869-5p can serve as a suppressor in macrophages mediated inflammatory and immune responses in gestational diabetes mellitus. However, the precise molecular mechanism regarding miR-6869-5p regulation in macrophages proliferation and polarization warrants to be elucidated by more future studies.

Mounting data have implicated that macrophages mediated local immunity plays an important role in maintain the balance of immune microenvironment in placenta [22–24]. Also, macrophages play a critical role in adipose tissue inflammation and immunity [3, 25, 26]. To the best of our knowledge, macrophages can be divided into two common cell types, namely, classically activated M1 and alternatively activated M2 [3, 27]. M1 cells usually possess proinflammatory activity, while M2 cells exert anti-inflammatory effects. The classic markers for M1 macrophages are CD11c, HLA-DR, and TNF-α. The typical markers for M2 macrophages are CD206, CD163, and IL-10. An anti-inflammatory M2 phenotype macrophages is essential for controlling gestational diabetes mellitus [28]. Previously, we have found M1/M2 balance is critical for the maintenance of the maternal-fetal interface immune balance [8]. M1/M2 imbalance would lead to sustained inflammation and immune disorders in the microenvironment of maternal-fetal interface, which may cause premature birth, stillbirth, and so on. Accumulating data have suggested miRNAs participate in the regulating of macrophages proliferation, differentiation, and

![Diagram](image_url)
Figure 4: MiR-6869-5p prevented from the production of TNF-α and IL-1β in macrophages (compared with the control group, **P < 0.01, ***P < 0.001; compared with the PTPRO (+) group, *P < 0.05; **P < 0.01; ***P < 0.001). (a) mRNA expression of TNF-α. (b) mRNA expression of IL-1β. (c) TNF-α in the supernatant of macrophages. (d) IL-1β in the supernatant of macrophages.

Figure 5: MiR-6869-5p promoted macrophages polarization towards M2.
polarization, which thus contributes to diabetes mellitus [29–31]. Nonetheless, the effect of miRNAs on macrophages polarization in gestational diabetes mellitus is largely unknown. In our study, miR-6869-5p is firstly documented to be positively associated with M2 polarization and protect normal pregnancy in patients with gestational diabetes mellitus. MiR-6869-5p is a promising marker for gestational diabetes mellitus.

PTPRO belongs to protein tyrosine phosphatase family. A number of studies have implicated that PTPRO participates in the regulation of macrophage-mediated inflammatory response, hepatic ischemia reperfusion injury, and tumor immunity [32–35]. In our previous study, PTPRO has been demonstrated to be significantly upregulated in preeclampsia patients, which is also found to be involved in regulating macrophage inflammation in preeclampsia [36]. The current study has firstly suggested that PTPRO is upregulated in placenta-derived macrophages from gestational diabetes mellitus patients. Therefore, we hypothesize that PTPRO may affect the differentiation and function of macrophages and, thus, participate in regulating local immune balance in the placenta. Findings of in vitro study have implicated that PTPRO can promote the expression of inflammatory cytokines TNF-α and IL-1β at both levels of mRNA and protein in macrophages. In addition, PTPRO is capable of inducing macrophages polarization towards M1 and enhancing the inflammatory response. As suggested by the bioinformatics analysis, PTPRO is a targeted gene of miR-6869-5p. MiR-6869-5p might negatively regulate PTPRO at the posttranscriptional level in macrophages. Interestingly, miR-6869-5p can prevent from inflammation by inducing higher expression of CD206 and Arg-1 but lower HLA-DR and CD11c in macrophages. As demonstrated by luciferase reporter assay, the well-established target of miR-6869-5p is PTPRO, a key factor in regulating macrophage-mediated inflammation and immune disorders. In general, miR-6869-5p possesses anti-inflammation activity by targetedly regulating PTPRO and inducing M2 macrophages ultimately. Nevertheless, whether miR-6869-5p could exert the same effect in vivo needs to be investigated in future studies.

In summary, the present study has identified a miR-6869-5p signature involvement in gestational diabetes mellitus, which may contribute to maintain the balance of placental immune microenvironment by targeting PTPRO and inducing macrophages polarization towards M2.

Data Availability
Data can be available from upon requesting for the corresponding author.

Conflicts of Interest
All authors have declared no conflict of interest.

Authors’ Contributions
Pingping Wang, Zhenzhi Ma, and Zengyan Wang are cofirst authors.

Acknowledgments
This work is supported by grants from Medical Health Science and Technology Development Program of Weifang (wfwsjk2019-031 and 2020YX048).

References


