

***In vivo* and *in vitro* Development of Human Mesencephalic Dopaminergic Neurons**

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Development of mesencephalic dopaminergic cells is well characterized in rodents and partially defined in primates. In the human fetus, tyrosine hydroxylase (TH) immunoreactivity was first observed at 7 weeks of gestation.

Migration and neuritic elongation of TH⁺ cells were monitored from 7 to 12 weeks of gestational age. Intact fetal mesencephalon was post-fixed and sections obtained to demonstrate dopaminergic cell anatomical location. At 8 weeks, TH⁺ neurons were found in a crescentic band occupying the middle third of the ventral mesencephalon. TH⁺ cell migration was confirmed in the medial third of the ventral mesencephalon at 9 weeks and by 11 weeks a large number of dopaminergic cells had elaborated neural processes. These data complement the observations of Freeman *et al.* /3/ that similarly described early dopaminergic cellular development.

Growth potentialities of the dopaminergic cells were evaluated in primary cultures. Neuronal cells were identified for non-specific markers (Neuron Specific Enolase). TH⁺ cells represented 1 to 2.5% of the total neuronal population, strictly dependent on precise dissection. One peculiar feature of TH⁺ neurons is the formation of long neurite (axons) projections in the presence and absence of the co-cultured striatal-specific target. Treatment of these cultures with fibroblast growth factors (FGF) significantly increased the number of surviving TH⁺ neurons at 8 d *in vitro* and induced proliferation of glial cells. Dopaminergic cells showed specific tropism for astroglial GFAP⁺ cells. Previous studies demonstrated that mesencephalic glia specifically support the survival of dopamin-

ergic neurons and induce dendritic outgrowth /2,4/. Some of the GFAP⁺ cells were shown to express NGF-low affinity receptors (NGF-R) on the membrane surface. NGF-R were identified with double staining for GFAP and NGF-R, using a confocal microscopy technique (monoclonal antibody against NGF-R obtained from clone ME 20-4 and 8211).

TH, the rate-limiting enzyme in the biosynthesis of catecholamines, was biochemically assayed as a quantitative index of dopaminergic cell number and viability. Enzymatic activity was determined using the TH microassay described by Bostwick *et al.* /1/. This microradiometric assay can detect the production of 5 pmol of ¹⁴CO₂ (coupled non-enzymatic decarboxylation of Dopa). 10⁵ cells were plated in a 96 microwell plate and the TH activity assayed at days 6 to 10. Linearity of the TH enzyme reaction with time and protein concentration was demonstrated. TH activity was shown to increase by culture day 6 and then to decline.

Characterization of developmental events in the human mesencephalic dopaminergic neurons provides significant pre-clinical information to define the optimal age characteristics for brain grafting. Percentage of dopaminergic neurons and number of proliferating glial cells may be controlled after precise anatomical dissection. A TH microassay was applied to the human dopaminergic cells *in vitro* and FGF was shown to affect TH⁺ neuronal cell development. Further investigations on glial-mediated processes in the human nigra are necessary to define the direct/indirect effects of FGF on dopaminergic neurons.

REFERENCES

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