Hindawi Oxidative Medicine and Cellular Longevity Volume 2019, Article ID 6512485, 12 pages https://doi.org/10.1155/2019/6512485



Research Article

Effects of Angiotensin-(1-7) and Angiotensin II on Acetylcholine-Induced Vascular Relaxation in Spontaneously Hypertensive Rats

Feng Zhang,^{1,2} Yu Xu,¹ Yan Pan,¹ Shuo Sun,¹ Aidong Chen,¹ Peng Li,³ Changlei Bao,⁴ Jian Wang,⁵ Haiyang Tang,⁰,^{4,5} and Ying Han,⁰

Correspondence should be addressed to Haiyang Tang; tanghy2008@yahoo.com and Ying Han; yhancn@njmu.edu.cn

Received 25 April 2019; Revised 22 July 2019; Accepted 12 August 2019; Published 20 November 2019

Academic Editor: Colin Murdoch

Copyright © 2019 Feng Zhang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Endothelial dysfunction of small arteries occurs in patients with hypertension and in various hypertensive models. Endothelial function is usually evaluated by the degree of acetylcholine- (ACh-) induced vascular relaxation. Our previous study has found that compared to Wistar-Kyoto rats (WKY), ACh-induced vasodilatation was attenuated significantly in the mesenteric artery (MA), coronary artery (CA), and pulmonary artery (PA) of spontaneously hypertensive rats (SHR). This study investigated the influence of angiotensin- (Ang-) (1-7) and Ang II on blood pressure and ACh-induced vascular relaxation, as well as their interactive roles and downstream signal pathways in SHR and WKY. Intravenous injection of Ang II significantly increased, while Ang-(1-7) decreased the mean arterial pressure (MAP) in SHR. Ang-(1-7) improved ACh-induced relaxation in the MA, CA, and PA of SHR, while Ang II further attenuated it, which were inhibited by pretreatment with Mas receptor antagonist A-779 or AT₁ receptor antagonist losartan, respectively. Ang-(1-7) decreased the basal arterial tension, and Ang II induced great vasoconstriction in SHR. Pretreatment with Ang-(1-7) inhibited the Ang II-induced pressor response, vasoconstriction, and the effects on ACh-induced relaxation in SHR. AT₁ receptor expression was higher, while nitric oxide (NO), cGMP, and protein kinase G (PKG) levels of arteries were lower in SHR than in WKY. Ang II decreased, while Ang-(1-7) increased, the levels of NO, cGMP, and PKG of arteries. In addition, pretreatment with Ang-(1-7) inhibited the Ang II-induced reduction of NO, cGMP, and PKG in SHR. These results indicate that the activation of the Mas receptor by Ang-(1-7) can improve endothelial function and decrease MAP in SHR and inhibit the deteriorative effect of Ang II on endothelial function through the NO-cGMP-PKG pathway.

1. Introduction

The integrity of vascular endothelial function plays an important role in maintaining equilibrium and homeostasis of vascular tension in a normal physiological state [1]. Vascular endothelial cells (VECs) release vasoconstrictor factors, such as endothelin-1. More importantly, VECs can also

release several vasodilators, such as nitric oxide (NO), endothelium-derived hyperpolarizing factor (EDHF), and prostaglandin (PGI₂). Among them, NO plays the most important roles in vasodilatation [2]. Endothelial nitric oxide synthase (eNOS) catalyzes the release of NO, which then diffuses into adjacent vascular smooth muscle cells (VSMCs), resulting in relaxation of smooth muscle through activation

¹Key Laboratory of Targeted Intervention of Cardiovascular Disease, Collaborative Innovation Center of Translational Medicine for Cardiovascular Disease, Department of Physiology, Nanjing Medical University, Nanjing, Jiangsu 211166, China

²Department of Geriatrics, Shandong Provincial Hospital Affiliated to Shandong First Medical University, Jinan, Shandong 250021, China

³Department of Cardiology, The First Affiliated Hospital of Nanjing Medical University, Nanjing, Jiangsu 210029, China

⁴College of Veterinary Medicine, Northwest A&F University, Yangling, Shaanxi 712100, China

⁵State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, Guangzhou Institute of Respiratory Health, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong 510120, China

of the intracellular guanylate cyclase- (GC-) cGMP-protein kinase G (PKG) signal pathway [3]. Acetylcholine (ACh) stimulates endothelial cells to release NO and induce vasodilatation; therefore, ACh-induced vasodilatation is usually used to evaluate endothelial function [1, 4].

Some factors, such as hypoxia, increased oxidative stress, increased inflammatory factors, and vascular mechanical dystonia, hurt artery endothelial cells and cause endothelial dysfunction [5]. The major cause of endothelial dysfunction is the imbalance of the release of vasoconstrictor factors and vasorelaxation factors, resulting in the decline of vascular diastolic function and the enhancement of vascular contraction function [6, 7]. Patients with essential hypertension [8, 9] or secondary hypertension [10, 11] have been reported to show endothelial dysfunction in small arteries. This phenotype is also typical in multiple hypertensive models [12]. Impaired NO release caused by endothelial dysfunction of small arteries and subsequent impaired vascular relaxation are implicated in the development and progression of hypertension [13] and involved in the further progression of organ damage in hypertension [14]. Our previous study has also found that compared to Wistar-Kyoto rats (WKY), ACh-induced vasodilatation was attenuated significantly in the mesenteric artery (MA), coronary artery (CA), and pulmonary artery (PA) of spontaneously hypertensive rats (SHR) [15], but the mechanisms involved in it are still not very clear.

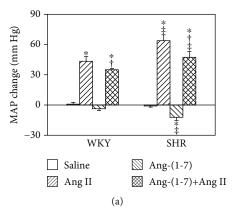
Biologically active peptide angiotensin- (Ang-) (1-7) is one important member of the renin-angiotensin system (RAS) family which regulates cardiovascular activity by activation of the Mas receptors [16-19]. The Mas receptor is massively expressed in the VECs and selectively blocked by its specific antagonist D-alanine-Ang-(1-7) (A-779) [20–22]. It has been reported that Ang-(1-7) in a peripheral circulation system prevents the development of chronic hypertension and end-organ damage in SHR [19, 23, 24], but the specific mechanism for this antihypertensive effect of Ang-(1-7) is still unknown. A study has shown that Ang-(1-7) can improve endothelial function and delays the development of cardiac remodeling and heart failure in rats with myocardial infarction [25]. Ang-(1-7) induces the mesenteric arterial relaxation of normal rats [26, 27], and intravenous injection of Ang-(1-7) decreases blood pressure [28]. In addition, some studies have shown that the Ang-(1-7)/Mas receptor axis plays opposite roles to Ang II and acts as a counterregulator of the classic Ang II/AT₁mediated effects in some peripheral tissues [29-32]. Activation of the Mas receptor blocks Ang II-induced vasoconstriction, and interfering with the expression of the Mas receptor significantly enhances the effect of Ang II on vascular tension [33]. Ang-(1-7) plays an antagonistic role in an Ang II-induced pressor effect [34] and inhibits Ang II-induced phosphorylation of ERK, p38, and JNK [35]. Ang-(1-7) inhibits myocardial hypertrophy induced by Ang II by activating Mas receptors [36]. Given the current body of data, Ang-(1-7) is a relevant area of study for mitigating negative endothelial functions and involved roles of Ang II. Specific effects of Ang-(1-7) and Ang II on endothelial dysfunction and the interaction between Ang-(1-7) and Ang II in hypertension are still unclear and warrant further investigation.

In this study, we used the MA, CA, and PA to determine the effects of Ang-(1-7) and Ang II on ACh-induced arterial relaxation, the influence of Ang-(1-7) on Ang II-induced responses, and their signal molecular mechanisms in SHR and WKY.

2. Materials and Methods

All procedures were approved by Nanjing Medical University Experimental Animal Care and complied with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH publication, 8th edition, 2011). Thirteen-week-old male WKY and SHR rats (Vital River Laboratory Animal Technology Co. Ltd., Beijing, China) were kept in a temperature-controlled room on a 12 h-12 h light-dark cycle with free access to standard chow and water. The major methods used in this study were done as described in a previous report [15].

- 2.1. Intravenous Injection. Intravenous injection of $100\,\mu\text{L}$ of either Ang-(1-7) or Ang II was performed through an external jugular vein catheter; injection was performed at a controlled rate for 10 minutes. A dual-channel microdialysis infusion syringe pump (53101V, Stoelting Co., Illinois, USA) was used to control the injection rate.
- 2.2. Mean Arterial Pressure (MAP) and Heart Rate (HR) Recording. Rats were anesthetized with urethane (800 mg/kg, intraperitoneal). A cannula was inserted into the right carotid artery and connected to a pressure transducer (MLT0380, ADInstruments, Australia). Arterial blood pressure, MAP, and HR were measured continuously.
- 2.3. Systolic Blood Pressure (SBP) Measurements. The SBP of the tail artery of conscious rats was measured with a noninvasive computerized tail-cuff system (NIBP, ADInstruments, Australia) as indicated in our previous report [37]. The SBP was obtained by averaging 10 measurements.
- 2.4. Isometric Tension Measurements in Arteries. In order to evaluate vascular function, the isometric tension of arteries was measured as previously described [15, 38]. Arterial rings (1.0 to 1.2 mm in length) from the third-order coronary arteries, pulmonary arteries, or mesenteric arteries were isolated from rats. The tissue was mounted onto four-chambered myographs (620M, DMT, Denmark) and set at a resting tension of 0.1 g. In order to evaluate the functionality of each arterial ring, the tissue was contracted by a high K⁺ solution (as previously reported [15]). Then, prostaglandin $F2\alpha$ (PGF 2α) was used to induce arterial ring contraction, followed by 6 doses of ACh (10⁻⁹~10⁻⁴ mol/L) administrated in a dose-dependent manner to induce vasodilatation. The degree of relaxation is shown as a percentage of PGF 2αinduced contraction. To determine the effects of Ang-(1-7) or Ang II on ACh-induced vasodilatation, the chemicals were added 20 minutes before the contraction induced by PGF 2α .
- 2.5. Artery Sample Preparation. The MA, CA, or PA were isolated from rats and were flash-frozen in liquid nitrogen and stored at -70°C. Arterial tissues were homogenized and



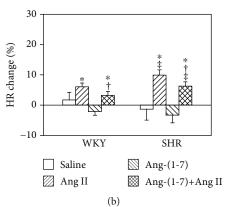


FIGURE 1: Effects of intravenous injection of saline, Ang II (10^{-4} mol/L), Ang-(1-7) (10^{-4} mol/L), and Ang-(1-7)+Ang II on mean arterial pressure (MAP) and heart rate (HR) in WKY and SHR. Values are the mean \pm SE. *P < 0.05 compared with saline. $^{\dagger}P < 0.05$ compared with Ang II alone. $^{\ddagger}P < 0.05$ compared with WKY. n = 6 for each group.

centrifuged; the supernatant was collected to extract total protein. Total protein concentration was measured using a commercial protein assay kit (BCA, Pierce Chemical, USA). The protein was then used for the detection of AT₁ receptor protein expression by western blotting or immunohistochemistry. To determine the effects of Ang-(1-7) or Ang II on NO, cGMP, and PKG levels of arteries, the MA, CA, or PA were isolated from rats and incubated in Krebs-Henseleit solution (as previously described [15]) in combination with Ang-(1-7) or Ang II for 20 minutes. The arteries were then flash-frozen in liquid nitrogen and stored at -70°C.

- 2.6. Western Blotting. The AT₁ receptor protein expressions in the arteries were determined by western blotting as described in our previous reports [39, 40]. Briefly, proteins purified from the supernatant of artery tissue homogenates were transferred to a nitrocellulose membrane after being run on a gel. An antibody specific for the AT₁ receptor (diluted 1:500; Abcam, Cambridge, MA, USA) was used, followed by blotting with horseradish peroxidase-conjugated goat anti-rabbit IgG (diluted 1:5000; Immunology Consultants Lab, Portland, OR, USA). An enhanced chemiluminescence ECL system (Pierce Chemical, Rockford, IL, USA) was used to visualize the blots. Protein loading was normalized by probing blots with the house-keeping gene, glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (1:5000; Bioworld Technology Inc., Louis, MN, USA); the total protein amount of the AT₁ receptor was derived after normalization to GAPDH.
- 2.7. Measurement of NO, cGMP, and PKG Levels. NO production of arteries was detected by a Nitrate/Nitrite Colorimetric Assay Kit (Cayman Chemical Co., Ann Arbor, MI, USA), to measure the concentration of stable nitrate and nitrite metabolites. In order to quantify the levels of cGMP and PKG in the arteries, a commercially available enzyme immunoassay kit was used (Cayman Chemical Co., Ann Arbor, MI, USA; Yi Fei Xue Biotechnology, Nanjing, China).
- 2.8. Immunohistochemistry. AT₁ receptor immunohistochemistry of the arteries was performed with a commercially available immunohistochemistry kit (Abcam, MA, USA).

Table 1: Body weight, SBP, MAP, and HR in one representative group of WKY and SHR.

| | WKY | SHR |
|-----------------|------------------|-------------------|
| Body weight (g) | 322.4 ± 4.7 | 320.5 ± 3.9 |
| SBP (mm Hg) | 114.7 ± 4.8 | $195.1 \pm 4.9^*$ |
| MAP (mm Hg) | 89.6 ± 4.2 | $135.4 \pm 4.2^*$ |
| HR (beats/min) | 384.3 ± 11.3 | 379.4 ± 12.6 |

SBP: systolic blood pressure; MAP: mean arterial pressure; HR: heart rate. Values are expressed as the mean \pm SE. *P < 0.05 compared with the WKY rats. n = 6 for each group.

Table 2: The high K^+ -induced contraction (mg/mm) in MA, CA, and PA in WKY and SHR.

| | WKY | SHR |
|----|-------------------|--------------------|
| MA | 428.5 ± 52.9 | $876.9 \pm 60.6^*$ |
| CA | 100.31 ± 12.3 | $202.5 \pm 18.6^*$ |
| PA | 115.4 ± 14.3 | $181.0 \pm 12.4^*$ |

Values are the mean \pm SE. *P < 0.05 compared with WKY. n = 6 for each group.

Coronal sections (5 μ m) were made from the isolated arteries and incubated overnight at 4°C with an antibody specific for the AT₁ receptor (diluted 1:500; Abcam, Cambridge, MA, USA) protein. The arteries were then incubated with a secondary antibody (biotinylated goat anti-rabbit IgG) for one hour, followed by staining with DAB in accordance with the manufacturer's instructions. AT₁ receptor immunoreactivity was observed using light microscopy (DP70, Olympus, Tokyo, Japan) after covering the section slides with mounting media.

2.9. Chemicals. All chemicals were dissolved in normal saline to create working solutions at stated concentrations. Ang-(1-7) and D-alanine-Ang-(1-7) (A-779, an antagonist of Mas receptors) were obtained from Bachem (Bubendorf, Switzerland). Ang II, losartan, prostaglandin F2 α (PGF 2α),

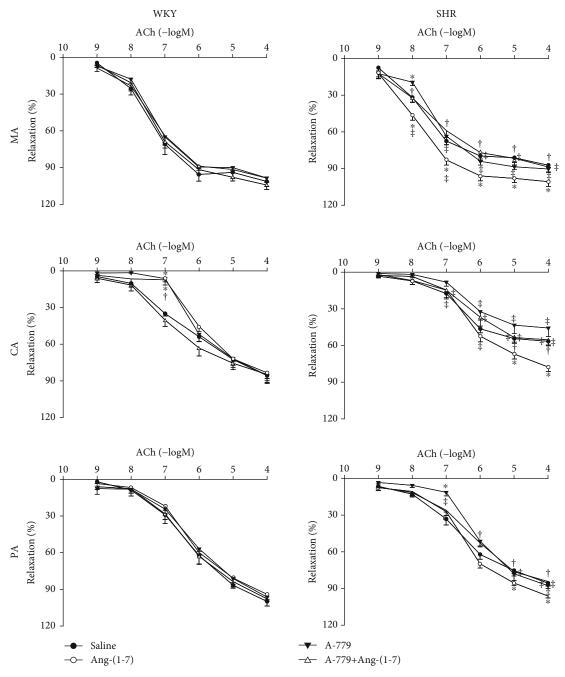


FIGURE 2: Effects of saline, Ang-(1-7) (10^{-6} mol/L), A-779 (10^{-5} mol/L), and A-779+Ang-(1-7) on ACh-induced dose-dependent relaxation in the MA, CA, and PA in WKY and SHR. Values are the mean \pm SE. *P < 0.05 compared with saline. †P < 0.05 compared with Ang-(1-7) alone. †P < 0.05 compared with WKY. P = 0.05 for each group.

and acetylcholine (ACh) were purchased from Sigma Chemical Co. (St. Louis, MO, USA).

2.10. Statistical Analysis. Data are expressed as the mean \pm SE. One-way or two-way ANOVA was used, followed by the Bonferroni test, for post hoc analysis when multiple comparisons were made. P < 0.05 was considered statistically significant.

3. Results

3.1. Effects of Ang-(1-7) and Ang II on Blood Pressure and Heart Rate. Intravenous injection of Ang II significantly increased MAP (Figure 1(a)) and HR (Figure 1(b)) in both WKY and SHR. Ang-(1-7) decreased MAP but had no significant effect on HR in SHR. Ang-(1-7) pretreatment inhibited the Ang II-induced elevation of blood pressure and HR in both WKY and SHR (Figure 1).

| | | Saline | Ang-(1-7) | Ang II | Ang-(1-7)+Ang II | A-779 | A-779+Ang-(1-7) | Losartan | Losartan+Ang II |
|-----|----|----------------|---------------------|---------------------------|-----------------------------|---------------|--------------------|----------------|----------------------------|
| WKY | MA | -0.9 ± 3.0 | -10.2 ± 2.4* | 155.2 ± 16.3* | $6.3 \pm 3.0^{\ddagger}$ | 6.3 ± 3.6 | $5.6 \pm 2.8^{\#}$ | 0.1 ± 1.1 | $6.2 \pm 1.1^{\ddagger}$ |
| | CA | 0.5 ± 2.7 | -2.9 ± 2.5 | $46.5 \pm 7.9^*$ | $13.2 \pm 2.3^{*\ddagger}$ | 4.6 ± 2.0 | 5.4 ± 2.0 | 0.1 ± 1.2 | $-1.1 \pm 2.8^{\ddagger}$ |
| | PA | 3.3 ± 1.9 | -0.6 ± 3.1 | $201.0 \pm 14.1^*$ | $19.4 \pm 4.7^{*\ddagger}$ | 8.1 ± 3.2 | 3.4 ± 2.3 | 5.3 ± 2.3 | $12.2 \pm 3.0^{*\ddagger}$ |
| SHR | MA | 2.7 ± 2.4 | -15.7 ± 2.5* | 137.4 ± 18.3* | $5.6 \pm 3.2^{\ddagger}$ | 8.6 ± 2.5 | $4.2 \pm 2.9^{\#}$ | 1.4 ± 3.9 | $-0.7 \pm 3.7^{\ddagger}$ |
| | CA | 3.0 ± 1.9 | $-9.1 \pm 1.7^{*}$ | $76.3 \pm 7.4^{*\dagger}$ | $15.8 \pm 3.5^{*\ddagger}$ | 3.0 ± 1.5 | $4.1 \pm 3.0^{\#}$ | -2.3 ± 2.1 | $8.1 \pm 2.1^{\ddagger}$ |
| | PA | -2.1 ± 2.0 | $-12.4 \pm 3.0^{*}$ | $230.6 \pm 17.5^*$ | $13.5 \pm 4.6^{* \ddagger}$ | 3.5 ± 3.4 | $5.4 \pm 2.0^{\#}$ | 1.0 ± 1.7 | $3.0 \pm 3.5^{\ddagger}$ |

Table 3: Influence of saline, Ang-(1-7), Ang II, Ang-(1-7)+Ang II, A-779, A-779+Ang-(1-7), losartan, and losartan+Ang II on the basal vascular tension (mg/mm) in WKY and SHR.

Data showed the changes (mg/mm) of vascular tension from the values before chemical intervention. Values are expressed as the mean \pm SE. *P < 0.05 vs. the saline. †P < 0.05 vs. WKY. *P < 0.05 compared with Ang II alone. *P < 0.05 compared with Ang-(1-7) alone. n = 6 for each group.

- 3.2. Vasoconstriction and Vasodilatation Function in WKY and SHR. There were no significant differences in body weight or HR between SHR and WKY, while the SBP and MAP of SHR were significantly higher than those observed in WKY (Table 1). The constriction of the MA, CA, and PA induced by high K⁺ solution was enhanced significantly in SHR (Table 2), while ACh-induced dose-dependent relaxations in the MA, CA, and PA were attenuated significantly in SHR compared with WKY, which was consistent with our previous finding [15] (Figure 2).
- 3.3. Effects of Ang-(1-7) and Ang II on Vascular Tension. Ang-(1-7) decreased basal vascular tension of the MA, CA, and PA in SHR, which was blocked by Mas receptor antagonist A-779. Ang II induced significant vasoconstriction in the MA, CA, and PA in both WKY and SHR models, which was blocked by the AT₁ receptor antagonist losartan. Pretreatment with Ang-(1-7) on the arteries inhibited the Ang II-induced vasoconstriction in both WKY and SHR. Neither A-779 nor losartan had a significant effect on the basal vascular tension of the MA, CA, and PA in both WKY and SHR (Table 3).
- 3.4. Effects of Ang-(1-7) on ACh-Induced Vascular Relaxation. ACh-induced dose-dependent relaxation of the MA, CA, and PA was improved by treatment with Ang-(1-7) in SHR but not in WKY. This effect of Ang-(1-7) was blocked by A-779 pretreatment. A-779 had no significant effect on ACh-induced relaxations in both WKY and SHR (Figure 2).
- 3.5. Effects of Ang II on ACh-Induced Vascular Relaxation. Ang II attenuated ACh-induced relaxations in the MA, CA, and PA, and this phenomenon was more significant in SHR compared with WKY. Furthermore, this effect was blocked by the AT₁ receptor antagonist losartan pretreatment. Losartan had no significant effect on ACh-induced relaxations in both WKY and SHR (Figure 3).
- 3.6. Effects of Ang-(1-7) on ACh-Induced Vascular Relaxation Response to Ang II. Pretreatment with Ang-(1-7) significantly inhibited the effects of Ang II on ACh-induced vascular relaxation in the MA, CA, and PA in both WKY and SHR (Figure 4).

- $3.7.\,AT_1$ Receptor Protein Expression of Arteries. Western blotting (Figure 5(a)) and immunohistochemistry (Figure 5(b)) revealed that AT_1 receptor protein expressions in the MA, CA, and PA of SHR arteries were significantly higher than the expression levels measured in WKY arteries.
- 3.8. The NO, cGMP, and PKG Levels of MA, CA, and PA. Compared with WKY, the NO, cGMP, and PKG levels of the MA, CA, and PA in SHR were decreased. Ang II further decreased, while Ang-(1-7) increased or normalized, NO, cGMP, and PKG levels of the MA, CA, and PA in SHR. In addition, the effect of Ang II on NO, cGMP, and PKG levels was inhibited by pretreatment with Ang-(1-7) on the arteries in both WKY and SHR (Figure 6).

4. Discussion

Endothelial dysfunction is a hallmark of hypertension and involved in further progression and deterioration. Our recent study has found that ACh-induced endothelium-dependent relaxation in the MA, CA, and PA of SHR was impaired significantly [15]. As we had talked about in a previous study [15], the MA, CA, and PA have been shown to play a major role in the damaging effects of hypertension; we therefore continue choosing these particular vessels for this study to investigate the effects of Ang (1-7) and Ang II on endothelial function. The present study demonstrates the following new findings: (1) Ang-(1-7) decreased blood pressure, while Ang II elevated it; (2) Ang-(1-7) enhanced ACh-induced vascular relaxation, while Ang II further attenuated it in the MA, CA, and PA of SHR. They were inhibited by pretreatment with Mas receptor antagonist A-779 and AT₁ receptor antagonist losartan, respectively; (3) Ang-(1-7) decreased the basal tension of the MA, CA, and PA, while Ang II induced vasoconstriction in SHR; (4) pretreatment with Ang-(1-7) on the arteries inhibited the Ang II-induced pressor response, vasoconstriction, and the effects on ACh-induced relaxation in SHR; (5) compared with WKY, NO, cGMP, and PKG levels of the MA, CA, and PA were decreased, but the AT₁ receptor expression of arteries was increased in SHR; (6) Ang II decreased but Ang-(1-7) increased NO, cGMP, and PKG levels of arteries; in addition, Ang-(1-7) pretreatment inhibited the Ang II-induced reduction of NO, cGMP, and PKG levels of arteries in SHR. These results indicate

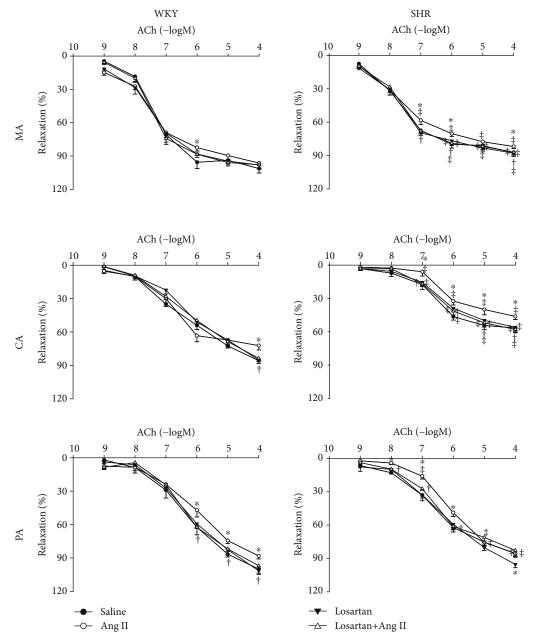


FIGURE 3: Effects of saline, Ang II (10^{-6} mol/L), losartan (10^{-5} mol/L), and losartan+Ang II on Ach-induced dose-dependent relaxation in the MA, CA, and PA in WKY and SHR. Values are the mean \pm SE. *P < 0.05 compared with saline. $^{\dagger}P < 0.05$ compared with Ang II alone. $^{\ddagger}P < 0.05$ compared with WKY. n = 6 for each group.

that activation of the Mas receptor by Ang-(1-7) restores endothelial dysfunction, decreases blood pressure, and inhibits the deteriorative effects of Ang II on vascular tension and Ang II induced pressor response through the NO-cGMP-PKG pathway.

In this study, SHR models had a significant increase in SBP and MAP as compared to WKY models. Additionally, when exposed to a high K⁺ solution to induce arterial constriction, SHR had a drastically enhanced response as compared to the WKY. SHR models also showed an impaired endothelium relaxation in their MA, CA, and PA after ACh induction. These results indicated that endothelial dysfunction of small

arteries and consequential enhanced vasoconstriction and attenuated vasodilatation occurred in SHR.

Dense Mas receptors are found in the VECs [41] and VSMCs [42]. Ang-(1-7) is found playing important roles in the regulation of cardiovascular activity [43, 44], and more interestingly, the regulation roles of Ang-(1-7) are different in different sites of the body. Our recent studies have found that microinjection of Ang-(1-7) into either the paraventricular nucleus (PVN) or the rostral ventrolateral medulla (RVLM) increases renal sympathetic nerve activity and MAP in renovascular hypertensive rats [39, 40, 45]. Ang-(1-7) in the PVN or RVLM increases NAD(P)H

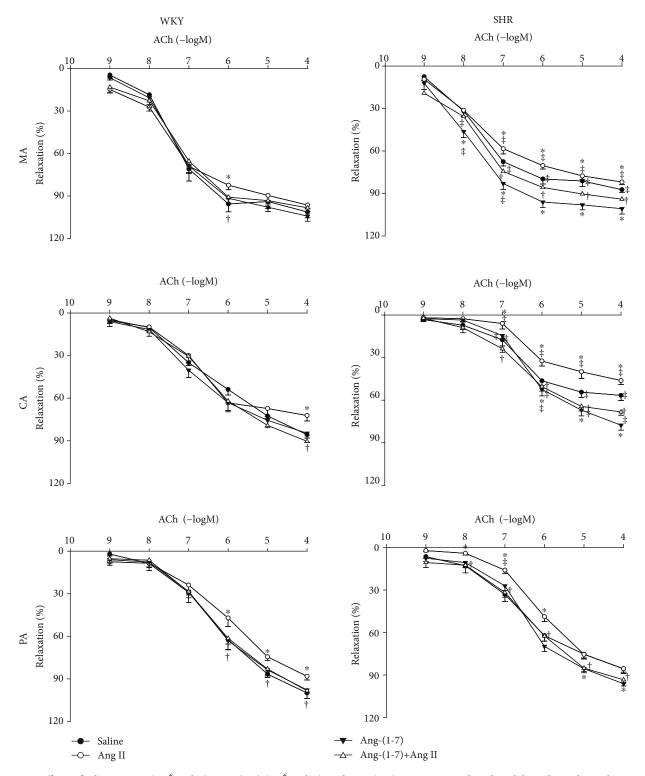


FIGURE 4: Effects of saline, Ang II (10^{-6} mol/L), Ang-(1-7) (10^{-6} mol/L), and Ang-(1-7)+Ang II on ACh-induced dose-dependent relaxation in the MA, CA, and PA in WKY and SHR. Values are the mean \pm SE. *P < 0.05 compared with saline. $^{\dagger}P$ < 0.05 compared with Ang II alone. $^{\ddagger}P$ < 0.05 compared with WKY. n = 6 for each group.

oxidase activity and superoxide anion level, and superoxide anions but not NO modulate the effects of Ang-(1-7) in the PVN or RVLM on sympathetic activity and blood pressure in normal rats or chronic heart failure rats [46, 47]. How-

ever, in regard to cardiovascular activity, Ang-(1-7) plays opposite regulating roles in peripheral tissues. Ang-(1-7) has been shown to decrease blood pressure [48] and induce mesenteric arterial relaxation in normotensive rat models

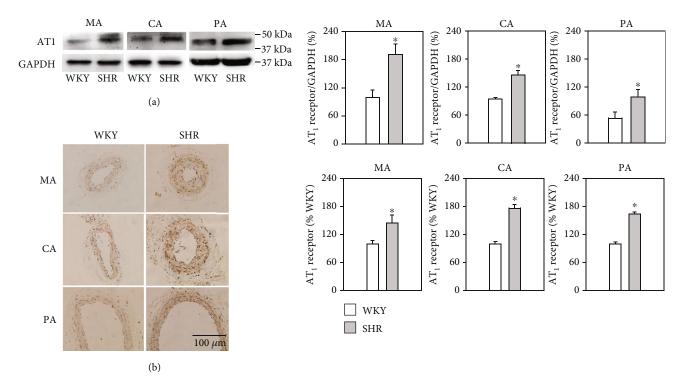


FIGURE 5: AT₁ receptor protein expressions of the MA, CA, and PA in WKY and SHR detected by western blotting (a) and immunohistochemistry (b). Values are the mean \pm SE. *P < 0.05 compared with WKY. n = 6 for each group.

[49, 50]. Ang-(1-7) has also been shown to improve endothelial function and delay the development of cardiac remodeling, even heart failure, in rats with myocardial infarction [51]. In the present study, we found that Ang-(1-7) enhanced ACh-induced vascular relaxation and decreased blood pressure and the basal tension of the MA, CA, and PA in SHR, which were blocked by pretreatment with Mas receptor antagonist A-779. These findings indicated that Ang-(1-7) has a beneficial effect to improve endothelial function and decrease blood pressure and vascular tension by activation of the Mas receptor in the arteries in a hypertensive state. In addition, a study has reported that Ang-(1-7)-dependent vasorelaxation of the renal artery was sensitive to antagonists against not only Mas but also AT₁, AT₂, and bradykinin receptor subtypes [52], which we will study in the future.

It has been reported that the Ang-(1-7)/Mas receptor axis presumably acts as a counterregulator of classic Ang II/AT₁-mediated effects in heart failure [53], emotional stress [54], or inflammatory conditions [55]. Ang-(1-7) plays an antagonistic role in the Ang II-induced pressor effect [34] and inhibits Ang II-induced phosphorylation of ERK, p38, and JNK [35]. Activation of the Mas receptor blocks Ang II-induced vasoconstriction [33]. In the present study, we found that Ang II elevated blood pressure significantly, attenuated ACh-induced vascular relaxation, and induced the constriction of the MA, CA, and PA in SHR, which were inhibited by pretreatment with the AT₁ receptor antagonist losartan, suggesting the worse effects of activity of the Ang II/AT₁ receptor on endothelial dysfunction and hypertension. It is important to note that pretreatment with Ang-(1-7) on the

arteries inhibited the Ang II-induced pressor response and vasoconstriction, as well as the effects on ACh-induced vascular relaxation in SHR. The results indicate that Ang-(1-7) not only improves the impaired endothelial function by activation of the Mas receptor directly but also protects endothelial function through antagonizing the adverse effects of Ang II indirectly, which emphasized the important roles of Ang-(1-7) in restoring endothelial dysfunction in hypertension.

In addition, we previously found that the Mas receptor protein expressions in the MA, CA, and PA of SHR were decreased [15], while in this study, AT₁ receptor protein expressions of the arteries of SHR were increased significantly compared with WKY. From these results, we speculated that the activity of the Ang-(1-7)/Mas receptor was impaired and the activity of Ang II/AT₁ was enhanced in SHR, which might be an important reason for inducing endothelial dysfunction and subsequent enhanced vasoconstriction and attenuated vasodilatation in a hypertensive state. Improvement of the activity of the Ang-(1-7)/Mas receptor would be beneficial for inhibition of the development and progression of endothelial dysfunction in hypertension.

Some studies have shown that NO mediates effects of Ang-(1-7) in peripheral tissues. Ang-(1-7) induces canine middle cerebral artery relaxation through stimulating NO release from endothelial cells [56]. Ang-(1-7) activates eNOS and increases the production of NO in human aortic endothelial cell, Mas-transfected Chinese hamster ovary cells [57], and cardiomyocytes [58]. As we know, NO released from endothelial cells induces VSMC relaxation through activation of the intracellular cGMP-PKG signal pathway.

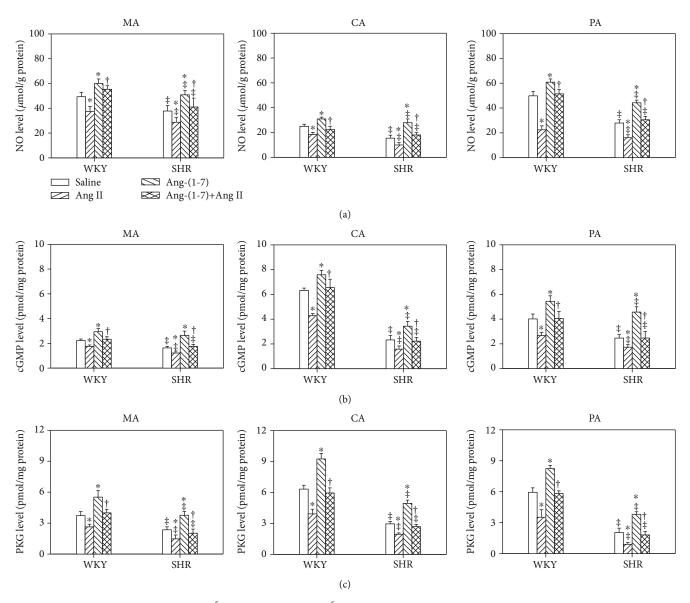


FIGURE 6: Effects of the saline, Ang II (10^{-6} mol/L), Ang-(1-7) (10^{-6} mol/L), and Ang-(1-7)+Ang II on the NO (a), cGMP (b), and PKG (c) levels of the MA, CA, and PA in WKY and SHR. Values are the mean \pm SE. *P < 0.05 compared with saline. $^{\dagger}P < 0.05$ compared with Ang II alone. $^{\ddagger}P < 0.05$ compared with WKY. n = 6 for each group.

The present study found that NO, cGMP, and PKG levels of the MA, CA, and PA in SHR were much lower than that in WKY, which further suggests the presence of endothelial dysfunction in SHR and that the NO-cGMP-PKG signal pathway plays important roles during endothelial dysfunction of SHR. Ang-(1-7) increased or normalized, while Ang II further decreased, NO, cGMP, and PKG levels of the MA, CA, and PA in SHR. More importantly, pretreatment with Ang-(1-7) on the arteries inhibited the Ang II-induced reduction of NO, cGMP, and PKG levels in SHR. These results suggest that the NO-cGMP-PKG signal pathway might be an important intracellular signaling mechanism mediating the effects of Ang-(1-7) or Ang II on vascular endothelial function. Activation of Mas receptors on the endothelial cell membrane by Ang-(1-7) might stimulate

NO generation and then NO relaxed the VSMCs through the cGMP-PKG signal pathway, while activation of ${\rm AT_1}$ receptors on the endothelial cell membrane by Ang II decreased NO generation and subsequent activation of the cGMP-PKG signal pathway. As to the mechanisms of the inhibitory influence of Ang-(1-7) on Ang II effects, we speculated that it might be some molecule mechanisms of activation or inactivation of eNOS inside of the endothelial cell, which need to be studied in the future.

In conclusion, the activity of the Ang-(1-7)/Mas receptor in the arteries was impaired and the activity of the Ang II/AT₁ receptor was enhanced in SHR. Activation of the Mas receptor by Ang-(1-7) decreased blood pressure, improves endothelial function, and inhibits the deteriorative effects of Ang II on endothelial function, vascular tension,

and hypertension through the NO-cGMP-PKG pathway in SHR. Improvement of Ang-(1-7)/Mas receptor activity in the arteries might be developed as a therapeutic strategy to ameliorate endothelial dysfunction and inhibition of hypertension.

Data Availability

All data supporting the findings of this study are available within the main manuscript or from the corresponding authors upon reasonable request.

Conflicts of Interest

The authors declare no conflict of interest.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (81470538, 31571168, 81770059, and 81970052), Qing Lan Project of Jiangsu Province, China, and the Open Project of the State Key Laboratory of Respiratory Disease (SKLRD-OP-201911).

References

- [1] E. L. Schiffrin and R. M. Touyz, "From bedside to bench to bedside: role of renin-angiotensin-aldosterone system in remodeling of resistance arteries in hypertension," *American Journal of Physiology Heart and Circulatory Physiology*, vol. 287, no. 2, pp. H435–H446, 2004.
- [2] A. Deussen, V. Ohanyan, A. Jannasch, L. Yin, and W. Chilian, "Mechanisms of metabolic coronary flow regulation," *Journal of Molecular and Cellular Cardiology*, vol. 52, no. 4, pp. 794–801, 2012.
- [3] U. Forstermann, E. I. Closs, J. S. Pollock et al., "Nitric oxide synthase isozymes. Characterization, purification, molecular cloning, and functions," *Hypertension*, vol. 23, 6, Part 2, pp. 1121–1131, 1994.
- [4] R. F. Furchgott and J. V. Zawadzki, "The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine," *Nature*, vol. 288, no. 5789, pp. 373–376, 1980.
- [5] H. A. Hadi, C. S. Carr, and J. Al Suwaidi, "Endothelial dysfunction: cardiovascular risk factors, therapy, and outcome," Vascular Health and Risk Management, vol. 1, pp. 183–198, 2005.
- [6] C. M. Sena, A. M. Pereira, and R. Seica, "Endothelial dysfunction a major mediator of diabetic vascular disease," *Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease*, vol. 1832, no. 12, pp. 2216–2231, 2013.
- [7] M. A. Incalza, R. D'Oria, A. Natalicchio, S. Perrini, L. Laviola, and F. Giorgino, "Oxidative stress and reactive oxygen species in endothelial dysfunction associated with cardiovascular and metabolic diseases," *Vascular Pharmacology*, vol. 100, pp. 1–19, 2018.
- [8] C. De Ciuceis, E. Porteri, D. Rizzoni et al., "Structural alterations of subcutaneous small-resistance arteries may predict major cardiovascular events in patients with hypertension," American Journal of Hypertension, vol. 20, no. 8, pp. 846–852, 2007
- [9] A. Virdis, A. Bacca, R. Colucci et al., "Endothelial dysfunction in small arteries of essential hypertensive patients: role of

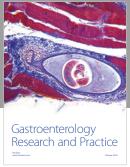
- cyclooxygenase-2 in oxidative stress generation," *Hypertension*, vol. 62, no. 2, pp. 337–344, 2013.
- [10] Y. Higashi, S. Sasaki, K. Nakagawa, H. Matsuura, T. Oshima, and K. Chayama, "Endothelial function and oxidative stress in renovascular hypertension," *The New England Journal of Medicine*, vol. 346, no. 25, pp. 1954–1962, 2002.
- [11] A. C. P. T. Henriques, F. H. C. Carvalho, H. N. Feitosa, R. H. M. Macena, R. M. S. Mota, and J. C. G. Alencar, "Endothelial dysfunction after pregnancy-induced hypertension," *International Journal of Gynaecology & Obstetrics*, vol. 124, no. 3, pp. 230–234, 2014.
- [12] P. Y. Chen and P. W. Sanders, "Role of nitric oxide synthesis in salt-sensitive hypertension in Dahl/Rapp rats," *Hypertension*, vol. 22, no. 6, pp. 812–818, 1993.
- [13] S. Kerr, M. J. Brosnan, M. McIntyre, J. L. Reid, A. F. Dominiczak, and C. A. Hamilton, "Superoxide anion production is increased in a model of genetic hypertension: role of the endothelium," *Hypertension*, vol. 33, no. 6, pp. 1353–1358, 1999.
- [14] U. Landmesser and H. Drexler, "Endothelial function and hypertension," *Current Opinion in Cardiology*, vol. 22, no. 4, pp. 316–320, 2007.
- [15] F. Zhang, H. Tang, S. Sun et al., "Angiotensin-(1-7) induced vascular relaxation in spontaneously hypertensive rats," *Nitric Oxide: Biology and Chemistry*, vol. 88, pp. 1–9, 2019.
- [16] S. Keidar, M. Kaplan, and A. Gamliel-Lazarovich, "ACE2 of the heart: from angiotensin I to angiotensin (1–7)," *Cardiovas-cular Research*, vol. 73, pp. 463–469, 2007.
- [17] C. Mercure, A. Yogi, G. E. Callera et al., "Angiotensin(1-7) blunts hypertensive cardiac remodeling by a direct effect on the heart," *Circulation Research*, vol. 103, no. 11, pp. 1319–1326, 2008.
- [18] R. A. S. Santos, A. C. S. e Silva, C. Maric et al., "Angiotensin-(1–7) is an endogenous ligand for the G protein-coupled receptor Mas," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 14, pp. 8258–8263, 2003.
- [19] I. F. Benter, D. I. Diz, and C. M. Ferrario, "Cardiovascular actions of angiotensin(1-7)," *Peptides*, vol. 14, no. 4, pp. 679–684, 1993.
- [20] R. A. S. Santos, M. J. Campagnole-Santos, N. C. V. Baracho et al., "Characterization of a new angiotensin antagonist selective for angiotensin-(1-7): Evidence that the actions of angiotensin-(1-7) are mediated by specific angiotensin receptors," *Brain Research Bulletin*, vol. 35, no. 4, pp. 293–298, 1994.
- [21] R. A. S. Santos, A. S. Haibara, M. J.: Campagnole-Santos et al., "Characterization of a new selective antagonist for angiotensin-(1–7), d -Pro 7 -angiotensin-(1–7)," *Hypertension*, vol. 41, no. 3, pp. 737–743, 2003.
- [22] R. Faria-Silva, F. V. Duarte, and R. A. Santos, "Short-term angiotensin(1-7) receptor MAS stimulation improves endothelial function in normotensive rats," *Hypertension*, vol. 46, no. 4, pp. 948–952, 2005.
- [23] I. F. Benter, C. M. Ferrario, M. Morris, and D. I. Diz, "Antihypertensive actions of angiotensin-(1-7) in spontaneously hypertensive rats," *The American Journal of Physiology*, vol. 269, pp. H313–H319, 1995.
- [24] I. F. Benter, M. H. M. Yousif, J. T. Anim, C. Cojocel, and D. I. Diz, "Angiotensin-(1–7) prevents development of severe hypertension and end-organ damage in spontaneously hypertensive rats treated with L-NAME," American Journal of Physiology Heart and Circulatory Physiology, vol. 290, no. 2, pp. H684–H691, 2006.

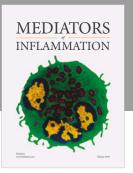
- [25] M. Durik, R. van Veghel, A. Kuipers et al., "The effect of the thioether-bridged, stabilized angiotensin-(1-7) analogue cyclic ang-(1-7) on cardiac remodeling and endothelial function in rats with myocardial infarction," *International Journal of Hypertensiont*, vol. 2012, article 536426, 8 pages, 2012.
- [26] R. A. Marangoni, A. K. Carmona, R. C. A. T. Passaglia, D. Nigro, Z. B. Fortes, and M. H. C. de Carvalho, "Role of the kallikrein–kinin system in Ang-(1-7)-induced vasodilation in mesenteric arterioles of Wistar rats studied in vivo–in situ," *Peptides*, vol. 27, no. 7, pp. 1770–1775, 2006.
- [27] J. A. Grace, S. Klein, C. B. Herath et al., "Activation of the MAS receptor by angiotensin-(1-7) in the renin-angiotensin system mediates mesenteric vasodilatation in cirrhosis," *Gastroenterology*, vol. 145, no. 4, pp. 874–884.e5, 2013, e875.
- [28] Y. le Tran and C. Forster, "Angiotensin-(1-7) and the rat aorta: modulation by the endothelium," *Journal of Cardiovascular Pharmacology*, vol. 30, no. 5, pp. 676–682, 1997.
- [29] L. S. A. Capettini, F. Montecucco, F. Mach, N. Stergiopulos, R. A. S. Santos, and R. F. da Silva, "Role of renin-angiotensin system in inflammation, immunity and aging," *Current Pharmaceutical Design*, vol. 18, no. 7, pp. 963–970, 2012.
- [30] T. Kanda and H. Itoh, "The ACE2/Ang(1-7)/Mas receptor axis in cardiovascular and renal diseases," *Nihon Rinsho*, vol. 70, pp. 1487–1491, 2012.
- [31] E. Mendoza-Torres, A. Oyarzun, D. Mondaca-Ruff et al., "ACE2 and vasoactive peptides: novel players in cardiovascular/renal remodeling and hypertension," *Therapeutic Advances in Cardiovascular Disease*, vol. 9, no. 4, pp. 217–237, 2015.
- [32] L. M. Kangussu, P. S. Guimaraes, A. P. Nadu, M. B. Melo, R. A. Santos, and M. J. Campagnole-Santos, "Activation of angiotensin-(1–7)/Mas axis in the brain lowers blood pressure and attenuates cardiac remodeling in hypertensive transgenic (mRen2)27 rats," *Neuropharmacology*, vol. 97, pp. 58–66, 2015.
- [33] E. Kostenis, G. Milligan, A. Christopoulos et al., "G-protein-coupled receptor Mas is a physiological antagonist of the angiotensin II type 1 receptor," *Circulation*, vol. 111, no. 14, pp. 1806–1813, 2005.
- [34] P. I. Ehlers, L. Nurmi, A. M. Turpeinen, R. Korpela, and H. Vapaatalo, "Casein-derived tripeptide Ile-Pro-Pro improves angiotensin-(1-7)- and bradykinin-induced rat mesenteric artery relaxation," *Life Sciences*, vol. 88, no. 5-6, pp. 206–211, 2011
- [35] Z. Su, J. Zimpelmann, and K. D. Burns, "Angiotensin-(1-7) inhibits angiotensin II-stimulated phosphorylation of MAP kinases in proximal tubular cells," *Kidney International*, vol. 69, no. 12, pp. 2212–2218, 2006.
- [36] E. A. Tallant, C. M. Ferrario, and P. E. Gallagher, "Angiotensin-(1-7) inhibits growth of cardiac myocytes through activation of the mas receptor," American Journal of Physiology Heart and Circulatory Physiology, vol. 289, no. 4, pp. H1560– H1566, 2005.
- [37] P. Li, H. J. Sun, B. P. Cui, Y. B. Zhou, and Y. Han, "Angiotensin-(1-7) in the rostral ventrolateral medulla modulates enhanced cardiac sympathetic afferent reflex and sympathetic activation in renovascular hypertensive rats," *Hypertension*, vol. 61, no. 4, pp. 820–827, 2013.
- [38] Y. Han, Y. E. Cho, R. Ayon et al., "SGLT inhibitors attenuate NO-dependent vascular relaxation in the pulmonary artery but not in the coronary artery," *American Journal of Physiology*

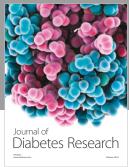
- Lung Cellular and Molecular Physiology, vol. 309, no. 9, pp. L1027–L1036, 2015.
- [39] P. Li, F. Zhang, H. J. Sun, F. Zhang, and Y. Han, "Angiotensin-(1-7) enhances the effects of angiotensin II on the cardiac sympathetic afferent reflex and sympathetic activity in rostral ventrolateral medulla in renovascular hypertensive rats," *Journal of the American Society of Hypertension*, vol. 9, no. 11, pp. 865–877, 2015.
- [40] H. J. Sun, P. Li, W. W. Chen, X. Q. Xiong, and Y. Han, "Angiotensin II and angiotensin-(1-7) in paraventricular nucleus modulate cardiac sympathetic afferent reflex in renovascular hypertensive rats," *PLoS One*, vol. 7, no. 12, article e52557, 2012
- [41] N. Alenina, P. Xu, B. Rentzsch, E. L. Patkin, and M. Bader, "Genetically altered animal models for Mas and angiotensin-(1-7)," *Experimental Physiology*, vol. 93, no. 5, pp. 528–537, 2008.
- [42] F. Zhang, X. Ren, M. Zhao, B. Zhou, and Y. Han, "Angiotensin-(1-7) abrogates angiotensin II-induced proliferation, migration and inflammation in VSMCs through inactivation of ROS-mediated PI3K/Akt and MAPK/ERK signaling pathways," *Scientific Reports*, vol. 6, no. 1, article 34621, 2016.
- [43] J. C. Q. Velez, K. J. Ryan, C. E. Harbeson et al., "Angiotensin I is largely converted to angiotensin (1-7) and angiotensin (2-10) by isolated rat glomeruli," *Hypertension*, vol. 53, no. 5, pp. 790–797, 2009.
- [44] A. Q. Gomes da Silva, C. H. Xavier, M. J. Campagnole-Santos et al., "Cardiovascular responses evoked by activation or blockade of GABA_A receptors in the hypothalamic PVN are attenuated in transgenic rats with low brain angiotensinogen," *Brain Research*, vol. 1448, pp. 101–110, 2012.
- [45] Y. Han, H. J. Sun, P. Li et al., "Angiotensin-(1-7) in paraventricular nucleus modulates sympathetic activity and cardiac sympathetic afferent reflex in renovascular hypertensive rats," *PLoS One*, vol. 7, no. 11, article e48966, 2012.
- [46] P. Li, F. Zhang, Y. B. Zhou, B. P. Cui, and Y. Han, "Superoxide anions modulate the effects of angiotensin-(1-7) in the rostral ventrolateral medulla on cardiac sympathetic afferent reflex and sympathetic activity in rats," *Neuroscience*, vol. 223, pp. 388–398, 2012.
- [47] X. Ren, F. Zhang, M. Zhao et al., "Angiotensin-(1-7) in paraventricular nucleus contributes to the enhanced cardiac sympathetic afferent reflex and sympathetic activity in chronic heart failure rats," Cellular Physiology and Biochemistry: International Journal of Experimental Cellular Physiology, Biochemistry, and Pharmacology, vol. 42, no. 6, pp. 2523–2539, 2017.
- [48] A. N. G. Braga, M. da Silva Lemos, J. R. da Silva, W. R. P. Fontes, and R. Augusto Souza Dos Santos, "Effects of angiotensins on day-night fluctuations and stress-induced changes in blood pressure," *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, vol. 282, no. 6, pp. R1663–R1671, 2002.
- [49] G. Raffai, M. J. Durand, and J. H. Lombard, "Acute and chronic angiotensin-(1-7) restores vasodilation and reduces oxidative stress in mesenteric arteries of salt-fed rats," *American Journal of Physiology Heart and Circulatory Physiology*, vol. 301, no. 4, pp. H1341-H1352, 2011.
- [50] W. O. Sampaio, A. A. S. Nascimento, and R. A. S. Santos, "Systemic and regional hemodynamic effects of angiotensin-(1-7) in rats," *American Journal of Physiology Heart* and Circulatory Physiology, vol. 284, no. 6, pp. H1985– H1994, 2003.

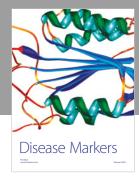
- [51] J. Wang, W. He, L. Guo et al., "The ACE2-Ang (1-7)-Mas receptor axis attenuates cardiac remodeling and fibrosis in post-myocardial infarction," *Molecular Medicine Reports*, vol. 16, no. 2, pp. 1973–1981, 2017.
- [52] M. H. M. Yousif, I. F. Benter, D. I. Diz, and M. C. Chappell, "Angiotensin-(1-7)-dependent vasorelaxation of the renal artery exhibits unique angiotensin and bradykinin receptor selectivity," *Peptides*, vol. 90, pp. 10–16, 2017.
- [53] V. B. Patel, J. C. Zhong, M. B. Grant, and G. Y. Oudit, "Role of the ACE2/angiotensin 1-7 axis of the renin-angiotensin system in heart failure," *Circulation Research*, vol. 118, no. 8, pp. 1313–1326, 2016.
- [54] M. A. P. Fontes, A. Martins Lima, and R. A. S. Santos, "Brain angiotensin-(1–7)/Mas axis: a new target to reduce the cardio-vascular risk to emotional stress," *Neuropeptides*, vol. 56, pp. 9–17, 2016.
- [55] R. R. Gaddam, S. Chambers, and M. Bhatia, "ACE and ACE2 in inflammation: a tale of two enzymes," *Inflammation & Allergy-Drug Targets*, vol. 13, no. 4, pp. 224–234, 2014.
- [56] K. Feterik, L. Smith, and Z. S. Katusic, "Angiotensin-(1-7) causes endothelium-dependent relaxation in canine middle cerebral artery," *Brain Research*, vol. 873, no. 1, pp. 75–82, 2000.
- [57] W. O. Sampaio, R. A. Souza dos Santos, R. Faria-Silva, L. T. da Mata Machado, E. L. Schiffrin, and R. M. Touyz, "Angiotensin-(1-7) through receptor Mas mediates endothelial nitric oxide synthase activation via Akt-dependent pathways," *Hypertension*, vol. 49, no. 1, pp. 185–192, 2007.
- [58] M. F. Dias-Peixoto, R. A. S. Santos, E. R. M. Gomes et al., "Molecular mechanisms involved in the angiotensin-(1-7)/Mas signaling pathway in cardiomyocytes," *Hypertension*, vol. 52, no. 3, pp. 542–548, 2008.

















Submit your manuscripts at www.hindawi.com













Journal of Obesity

