

SUPPORTING INFORMATION

to

Products of sulfide/selenite interaction possess antioxidant properties, scavenge superoxide-derived radicals, react with DNA, modulate blood pressure and tension of isolated thoracic aorta

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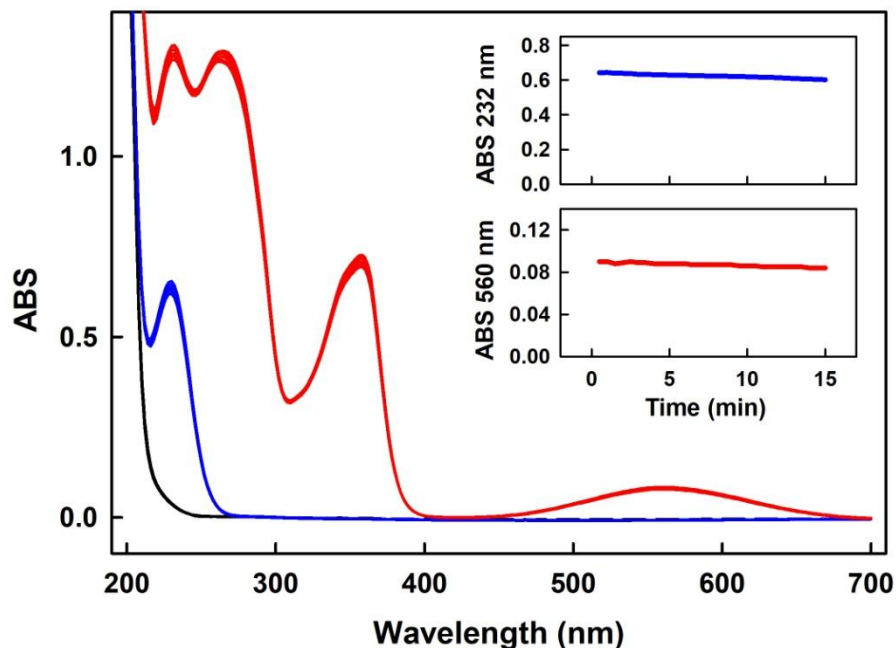
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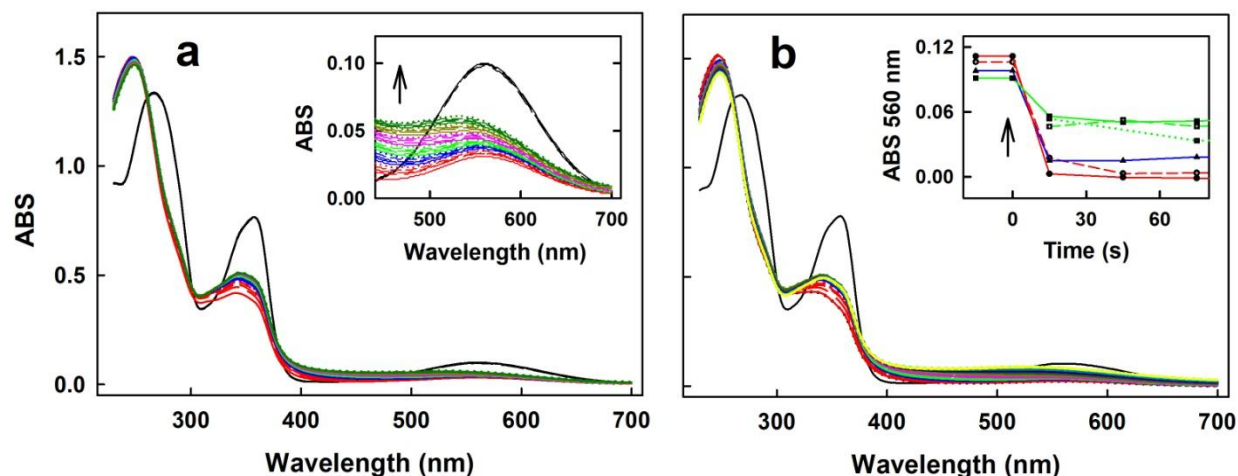
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Supplementary Figure 1: Time-resolved UV-VIS spectra of Na_2SeO_4 , H_2S and their mixture with $\bullet\text{cPTIO}$.

UV-VIS spectra of $100 \mu\text{mol L}^{-1}$ Na_2SeO_4 (repeated 3 x 30 s, black), subsequent addition of $100 \mu\text{mol L}^{-1}$ H_2S (30 x 30 s, blue) followed by $100 \mu\text{mol L}^{-1}$ $\bullet\text{cPTIO}$ (30 x 30 s, red). Upper insert: Kinetic trace of ABS at 232 nm (corresponding to HS^-) in the mixture of $100 \mu\text{mol L}^{-1}$ Na_2SeO_4 + $100 \mu\text{mol L}^{-1}$ H_2S (blue). Lower insert: Kinetic trace of $100 \mu\text{mol L}^{-1}$ $\bullet\text{cPTIO}$ reduction in the mixture of $100 \mu\text{mol L}^{-1}$ Na_2SeO_4 + $100 \mu\text{mol L}^{-1}$ H_2S (red) measured as change of ABS at 560 nm. Buffer: 100 mmol L^{-1} sodium phosphate buffer, $100 \mu\text{mol L}^{-1}$ DTPA, pH 7.4 at 37°C .

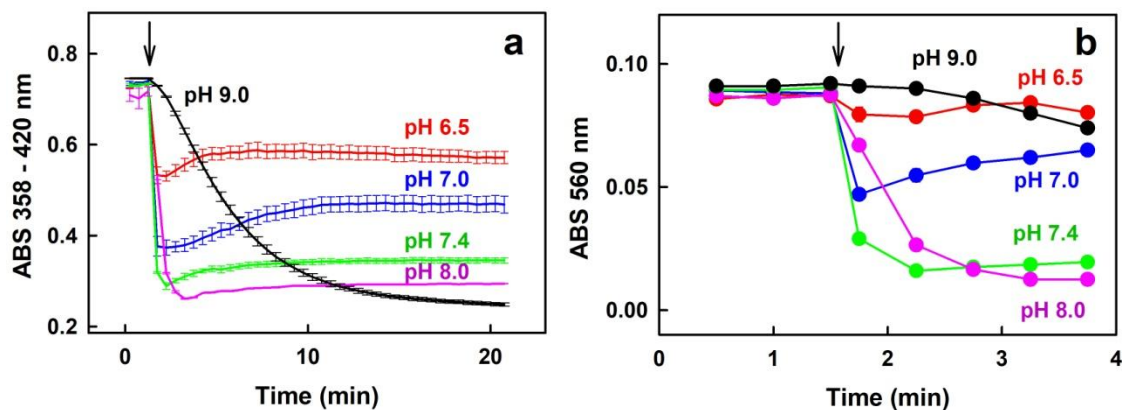


Supplementary Figure 2: Time resolved UV-VIS spectra of studied compounds.

(a) Time resolved UV-VIS spectra of \bullet cPTIO, SeCl_4 and their mixture with H_2S . UV-VIS spectra of $100 \mu\text{mol L}^{-1}$ \bullet cPTIO with $100 \mu\text{mol L}^{-1}$ SeCl_4 (repeated 3 times every 30 s, black). Subsequent addition of $100 \mu\text{mol L}^{-1}$ H_2S measured every 30 s for 15 min. Inset represents detail of ABS at 440-710 nm. The arrow indicates the increase of absorbance at around 420 nm.

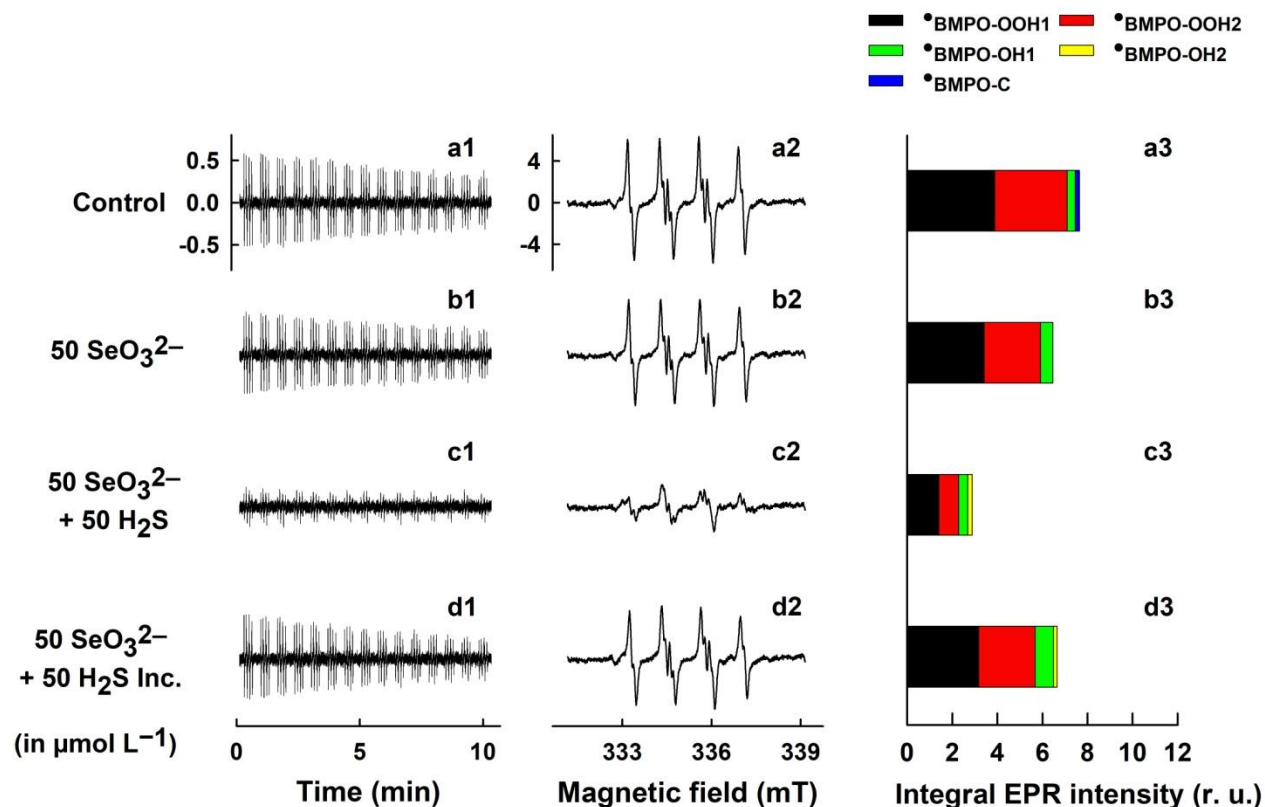
(b) Time resolved UV-VIS spectra of the interaction of \bullet cPTIO/ SeO_3^{2-} with H_2S in argon de-aerated solutions. UV-VIS spectra of $100 \mu\text{mol L}^{-1}$ \bullet cPTIO with $100 \mu\text{mol L}^{-1}$ SeO_3^{2-} (black) and subsequent addition of $100 \mu\text{mol L}^{-1}$ H_2S (spectra were collected every 30 s for 30 min, the first spectrum, indicated by the red line, was measured in 15th second after addition of H_2S). Inset: The reduction of the \bullet cPTIO radical was detected as the decrease of ABS at 560 nm. Red lines: $110 \mu\text{mol L}^{-1}$ H_2S was added to \bullet cPTIO/ SeO_3^{2-} (110/110 in $\mu\text{mol L}^{-1}$); Blue lines: $100 \mu\text{mol L}^{-1}$ H_2S was added to \bullet cPTIO/ SeO_3^{2-} (100/100 in $\mu\text{mol L}^{-1}$) and green lines: $60 \mu\text{mol L}^{-1}$ H_2S was added to \bullet cPTIO/ SeO_3^{2-} (100/100 in $\mu\text{mol L}^{-1}$). Arrow marks H_2S addition.

The solid red line of UV-VIS spectra indicates the first spectrum after addition of H_2S into SeCl_4 or SeO_3^{2-} , which is followed each 30 s by: long dash red, medium dash red, short dash red, dotted red, solid blue line, long dash blue, medium dash blue, etc. Buffer: 100 mmol L^{-1} sodium phosphate, $100 \mu\text{mol L}^{-1}$ DTPA, pH 7.4, 37°C .



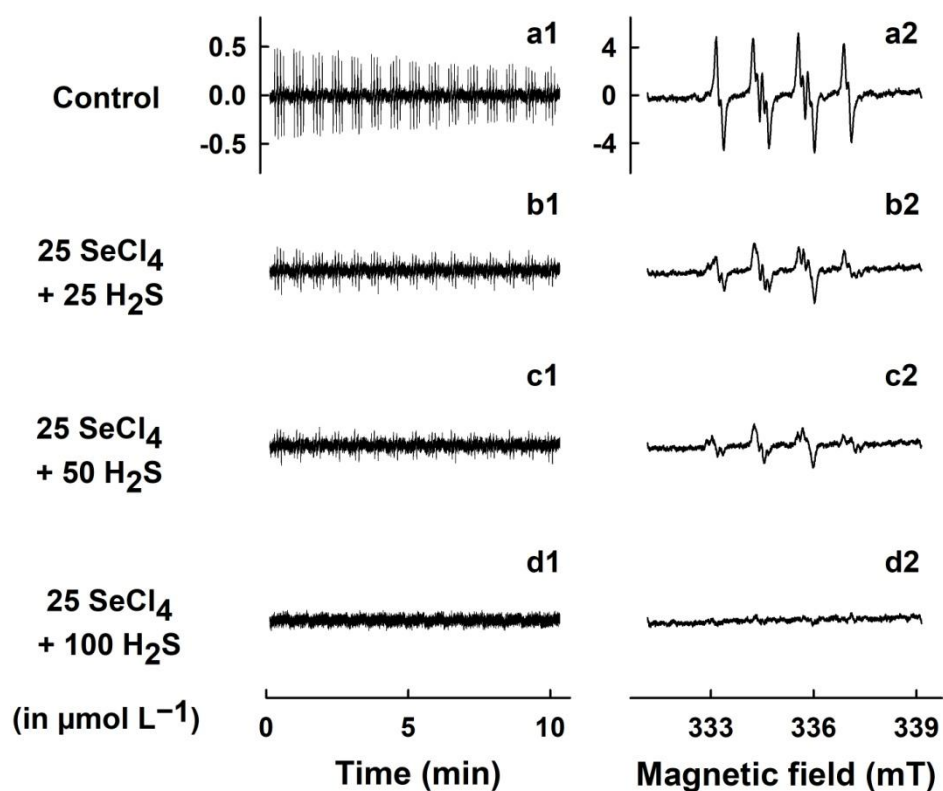
Supplementary Figure 3: Effect of pH on the kinetics of \bullet cPTIO reduction induced by H_2S in the presence of SeO_3^{2-} .

The reduction of the \bullet cPTIO radical was detected as decrease of ABS at 358 nm minus ABS at 420 nm (a) or as a decrease of the absorbance at 560 nm (b). Buffer: 100 mmol L^{-1} sodium phosphate, 100 $\mu\text{mol L}^{-1}$ DTPA, pH 6.5, 7.0, 7.4, 8.0, and 9.0 at 37 $^\circ\text{C}$. Arrow indicates addition of H_2S (100 $\mu\text{mol L}^{-1}$) into \bullet cPTIO/ SeO_3^{2-} (100/50 in $\mu\text{mol L}^{-1}$) solution. In the case of experiments at pH 6.5 and 7.0, UV-VIS spectra of $\text{H}_2\text{S}/\text{SeO}_3^{2-}$ were subtracted from the \bullet cPTIO/ $\text{H}_2\text{S}/\text{SeO}_3^{2-}$ spectra. Values are the means \pm S.E.M., $n = 2-3$.



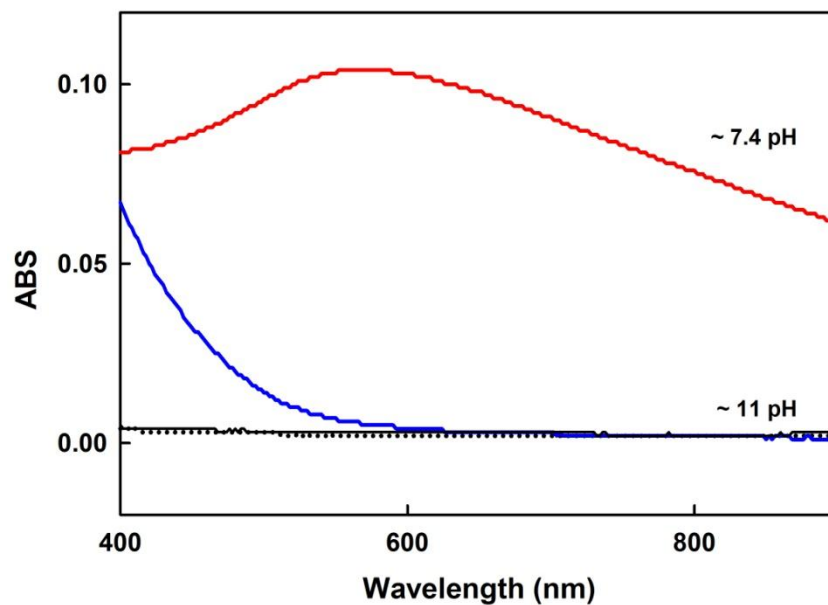
Supplementary Figure 4: EPR spectra of \bullet BMPO in the presence of $\text{O}_2^{\bullet-}$ and modulated by $\text{H}_2\text{S}/\text{SeO}_3^{2-}$.

Representative EPR spectra of the \bullet BMPO-adducts monitored in 10% v/v saturated KO_2/DMSO solution in 50 mmol L^{-1} sodium phosphate buffer, 0.1 mmol L^{-1} DTPA, pH 7.4, 37°C in the presence of studied compounds and 20 mmol L^{-1} BMPO. The sets of individual EPR spectra of the \bullet BMPO-adducts monitored upon 15 sequential scans, each 42 s (a1-d1), starting acquisition 2 min after sample preparation in: control 10% v/v KO_2/DMSO in the buffer (a1), KO_2/DMSO in the presence of $50 \mu\text{mol L}^{-1}$ SeO_3^{2-} (b1), 50/50 in $\mu\text{mol L}^{-1}$ $\text{H}_2\text{S}/\text{SeO}_3^{2-}$ (c1) and 50/50 in $\mu\text{mol L}^{-1}$ $\text{H}_2\text{S}/\text{SeO}_3^{2-}$ preincubated 5 min before KO_2/DMSO addition (d1). The details of accumulated 15 spectra (a2-d2). The intensities of the time-resolved EPR spectra (a1-d1) and detailed spectra (a2-d2) are comparable, as they were measured at the identical EPR settings. EPR 0.1 mT modulation amplitude was used. (a3-d3) comparison of integral EPR spectra intensities of the individual components of simulated $\text{BMPO} + \text{O}_2^{\bullet-}$ without (control) and with chalcogen species showed in a1-d1. The first five EPR spectra were accumulated and used for simulation. The data represent the means of $n = 2$, standard error was $\leq 10\%$ of the mean value. The \bullet BMPO-OOH and \bullet BMPO-OH were simulated based on two conformers. The means of used hyperfine coupling constants were: \bullet BMPO-OOH1 $a_N = 13.30 \pm 0.03 \text{ G}$; $a_H = 11.7 \pm 0.1 \text{ G}$; \bullet BMPO-OOH2 $a_N = 13.24 \pm 0.03 \text{ G}$; $a_H = 9.4 \pm 0.1 \text{ G}$; \bullet BMPO-OH1 $a_N = 13.7 \pm 0.3 \text{ G}$; $a_H = 12.3 \pm 0.4 \text{ G}$; \bullet BMPO-OH2 $a_N = 13.6 \pm 0.2 \text{ G}$; $a_H = 15.3 \pm 0.1 \text{ G}$. For \bullet BMPO-C $a_N = 15.2 \pm 0.1 \text{ G}$; $a_H = 21.5 \pm 0.1 \text{ G}$.



Supplementary Figure 5: EPR spectra of \bullet BMPO in the presence of $\text{O}_2^{\bullet-}$ modulated by $\text{H}_2\text{S}/\text{SeCl}_4$.

Representative EPR spectra of the \bullet BMPO-adducts monitored in 10% v/v saturated KO_2/DMSO solution in 50 mmol L^{-1} sodium phosphate buffer, 0.1 mmol L^{-1} DTPA, pH 7.4, 37°C in the presence of studied compounds and 20 mmol L^{-1} BMPO. The sets of individual EPR spectra of the \bullet BMPO-adducts monitored upon 15 sequential scans, each 42 s (a1-d1), starting acquisition 2 min after sample preparation in: control 10% v/v KO_2/DMSO in the buffer (a1), KO_2/DMSO in the presence of 25/25 in $\mu\text{mol L}^{-1}$ $\text{H}_2\text{S}/\text{SeCl}_4$ (b1), 50/25 in $\mu\text{mol L}^{-1}$ $\text{H}_2\text{S}/\text{SeCl}_4$ (c1) and 100/25 in $\mu\text{mol L}^{-1}$ $\text{H}_2\text{S}/\text{SeCl}_4$ (d1). The detail of accumulated 15 spectra (a2-d2). The intensities of the time-resolved EPR spectra (a1-d1) and detailed spectra (a2-d2) are comparable, as they were measured at the identical EPR settings. EPR modulation amplitude was 0.1 mT.



Supplementary Figure 6: VIS spectra of H₂S, SeO₃²⁻ and their mixture.

H₂S (100 μmol L⁻¹; black full), SeO₃²⁻ (50 μmol L⁻¹; dotted black) and their mixture (100/50 in μmol L⁻¹) prepared in 0.9 % NaCl pH ~ 11 (blue) and in phosphate buffer of pH ~ 7.4 (red). Sample with phosphate buffer was prepared as follows: 123.5 μL of 100 mmol L⁻¹ phosphate buffer, 100 μmol L⁻¹ DTPA, pH 7.4, 37°C; + 14 μL of 1 mol L⁻¹ HCl; + 62.5 μL of 40 mmol L⁻¹ SeO₃²⁻ prepared in 0.9 % NaCl and + 50 μL of 100 mol L⁻¹ H₂S in H₂O. Final concentrations in the stock mixture of 250 μL volume were 20 mmol L⁻¹ Na₂S and 10 mmol L⁻¹ SeO₃²⁻. pH was measured by paper pH indicator.