

Supplemental Materials

Redox states of protein cysteines in pathways of protein turnover and cytoskeleton dynamics are changed with aging and reversed by Slc7a11 restoration in mouse lung fibroblasts

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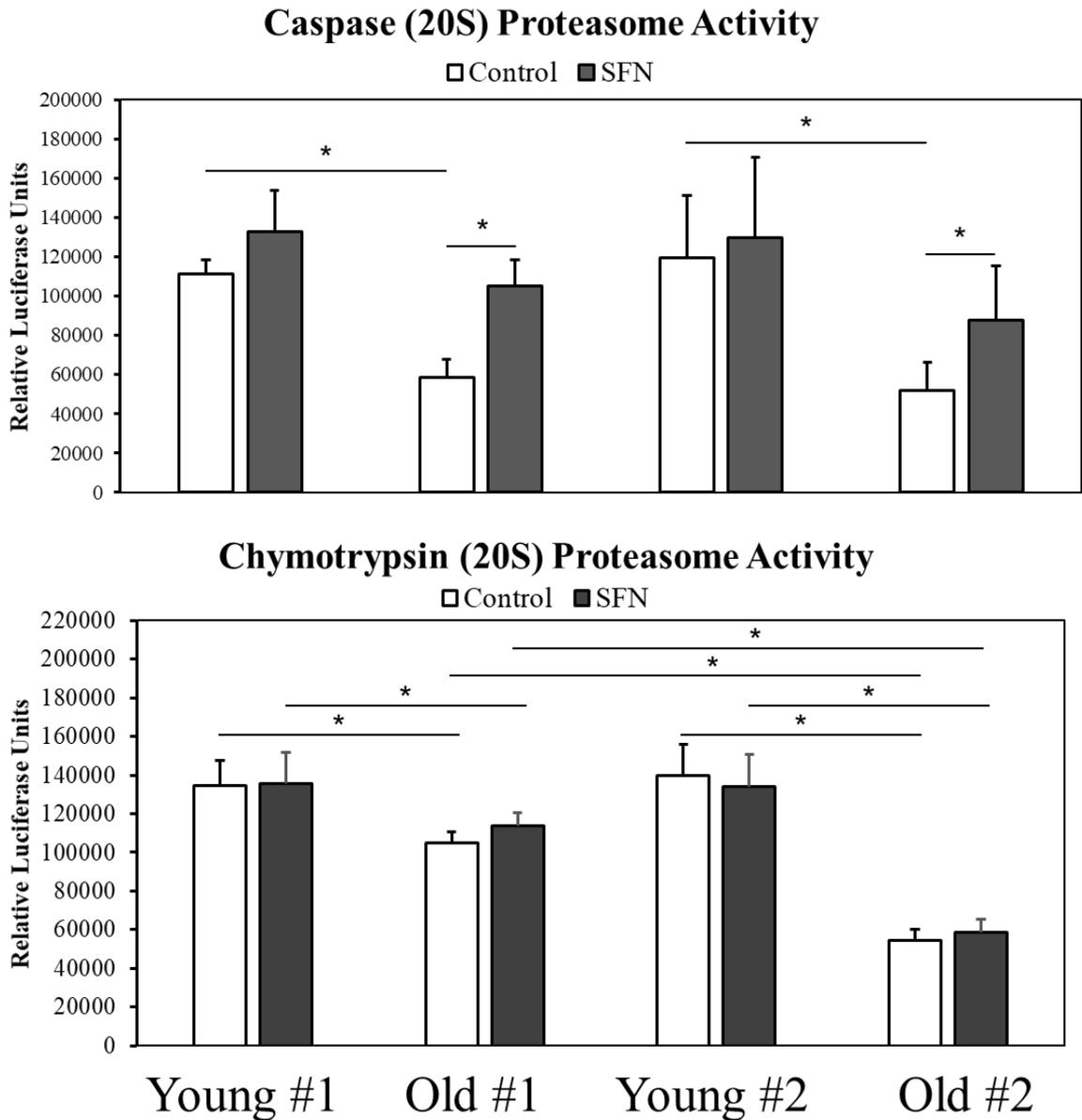
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Supplemental Methods

Proteasomal activity assay. Proteasomal activity was measured using the Proteasome-Glo Chymotrypsin-Like and Caspase-Like Cell-Based Assays from Promega (#G8660, #G8860, Madison, WI) according to the manufacturer's instructions. Briefly, primary lung fibroblasts (15,000 cells/well) from young or old female C57BL/6 mice were plated onto a 96 well (white walled) tissue culture plate and incubated in DMEM supplemented with 10% fetal bovine serum (FBS) and 1% antibiotic-antimycotic solution in a humidified incubator with 5% CO₂. After attachment, cells were treated with or without the addition of 5 μ M sulforaphane for 6 hours. Afterwards, some fibroblasts were incubated in the presence of a proteasome inhibitor lactacystin (10 μ M, Cayman Chemical, Ann Arbor, MI) for 2 hours to identify nonspecific protease activity not related to the proteasome. The Proteasome-Glo Cell-Based Reagent (100 μ l) was then added to each sample and mixed for 2 min at 700 rpm using a plate shaker. Samples were incubated at room temperature for 10 minutes prior to the measurement of luminescence using a Luminoskan Ascent Luminometer (Labsystems, Helsinki, Finland). Samples were normalized to total protein measured using protein assay dye (Bio-Rad, Hercules, CA) and values were reported as relative luciferase units.

Supplemental Figure 1 shows proteasomal activity is lower in old cells, and sulforaphane treatment restores caspase-like, but not chymotrypsin-like, proteasomal activity. This figure is related to Table 1 and described in the 4th paragraph of the Discussion section.



SUPPLEMENTAL FIGURE 1: Caspase-like (top) and chymotrypsin-like (bottom) proteasomal activity. Fibroblasts were isolated from the lungs of 2 Young (#1 and #2) and 2 Old (#1 and #2) female C57BL/6J mice. Caspase-like and chymotrypsin-like proteasomal activity were measured before and after induction of Slc7a11 activity with sulforaphane as described in Supplemental Methods. Both types of proteasomal activity were lower in old cells, but sulforaphane treatment restored only caspase-like proteasomal activity. Fibroblasts from 2 different young mice had similar caspase- and chymotrypsin-like activities. Fibroblasts from both old mice had lower activities than either young mouse, but one biological replicate had even lower chymotrypsin-like activity than the other. Six wells were plated for each condition. * - $p < 0.05$ by 2-way ANOVA and Tukey's post-hoc test.

Supplemental Table 2 provides IPA analysis of proteins whose redox state changes with Slc7a11 overexpression in old fibroblasts regardless of their redox states in young fibroblasts. Cysteine redox states were compared between old fibroblasts and old Slc7a11-overexpressing fibroblasts. This table is described in the Results section 3.2.

SUPPLEMENTAL TABLE 2: IPA analysis of proteins whose redox states were changed by Slc7a11 overexpression in old fibroblasts. Fisher's exact test *p*-values for all the items shown were less than 0.05.

Name	p-value	Overlap
Regulation of eIF4 and p70S6K Signaling	2.09E-09	6.4 % 10/157
EIF2 Signaling	6.26E-08	4.5 % 10/224
mTOR Signaling	4.53E-06	3.8 % 8/210
Antiproliferative Role of Somatostatin Receptor 2	2.41E-05	6.5 % 5/77
Relaxin Signaling	5.68E-05	4.0 % 6/150

Supplemental Table 3 provides IPA analysis of age-dependent proteins whose redox states were not rescued by Slc7a11 overexpression in old fibroblasts. Those age-dependent Slc7a11-nonresponsive protein cysteines indicate there are Slc7a11-independent regulation of protein redox signaling. This table is described in the Results section 3.3.

SUPPLEMENTAL TABLE 3: IPA analysis of age-dependent proteins whose redox states were not rescued by Slc7a11 overexpression in old fibroblasts. Fisher's exact test *p*-values for all the items shown were less than 0.05.

Name	p-value	Overlap
Hepatic Fibrosis / Hepatic Stellate Cell Activation	3.27E-05	2.7 % 5/186
Protein Ubiquitination Pathway	2.00E-04	1.8 % 5/273
Epithelial Adherens Junction Signaling	2.32E-04	2.6 % 4/152
Actin Cytoskeleton Signaling	9.04E-04	1.8 % 4/218
EIF2 Signaling	1.00E-03	1.8 % 4/224

Supplemental Table 4 provides GO-based biological process analysis of proteins whose redox states were not rescued by Slc7a11 overexpression in old fibroblasts. Those age-dependent Slc7a11-nonresponsive protein cysteines indicate there are Slc7a11-independent regulation of protein redox signaling. This table is described in the Results section 3.3.

SUPPLEMENTAL TABLE 4: GO-based biological process analysis of proteins whose redox states were not rescued by Slc7a11 overexpression in old fibroblasts. Fisher's exact test *p*-values for all the items shown were less than 0.05.

Term	Count	p-value	-log10 (p-value)
protein folding	6	<0.001	4.845
wound healing	4	0.002	2.811
actomyosin structure organization	3	0.002	2.709
mitotic cytokinesis	3	0.002	2.618
translation	6	0.003	2.554
skin development	3	0.009	2.057
IRES-dependent viral translational initiation	2	0.017	1.772
platelet degranulation	2	0.019	1.714
heart development	4	0.025	1.594
cytoskeleton organization	3	0.026	1.577
single organismal cell-cell adhesion	3	0.028	1.547
cerebellar Purkinje cell layer development	2	0.038	1.417
actin filament-based movement	2	0.041	1.392
actin filament capping	2	0.043	1.367