

Research Article

Single-Cell Integration Analysis of Heterotopic Ossification and Fibrocartilage Developmental Lineage: Endoplasmic Reticulum Stress Effector Xbp1 Transcriptionally Regulates the Notch Signaling Pathway to Mediate Fibrocartilage Differentiation

Yisheng Chen ^{1,2}, Yaying Sun,² Yuzhen Xu ³, Wei-Wei Lin ⁴, Zhiwen Luo,² Zhihua Han,¹ Shaohua Liu,² Beijie Qi,² Chenyu Sun,⁵ Ken Go,⁶ x.-R. Kang,⁷ and Jiwu Chen ¹

¹Department of Orthopedics, Shanghai General Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai Jiao Tong University, Shanghai 200080, China

²Department of Sports Medicine, Huashan Hospital, Fudan University, Shanghai, China

³Department of Rehabilitation, The Second Affiliated Hospital of Shandong First Medical University, Taian, Shandong Province 271000, China

⁴Department of Neurosurgery, Second Affiliated Hospital of Zhejiang University School of Medicine, Zhejiang University, 88 Jiefang Road, Hangzhou, 310009 Zhejiang, China

⁵Internal Medicine, AMITA Health Saint Joseph Hospital Chicago, 2900 N. Lake Shore Drive, Chicago, 60657 Illinois, USA

⁶Department of Clinical Training Centre, St. Marianna Hospital, Tokyo, Japan

⁷Shanghai Jiao Tong University, Shanghai 200080, China

Correspondence should be addressed to Jiwu Chen; jeevechen@gmail.com

Received 12 July 2021; Revised 21 September 2021; Accepted 1 October 2021; Published 26 October 2021

Academic Editor: Wen-Jun Tu

Copyright © 2021 Yisheng Chen et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction. Regeneration of fibrochondrocytes is essential for the healing of the tendon-bone interface (TBI), which is similar to the formation of neurogenic heterotopic ossification (HO). Through single-cell integrative analysis, this study explored the homogeneity of HO cells and fibrochondrocytes. **Methods.** This study integrated six datasets, namely, GSE94683, GSE144306, GSE168153, GSE138515, GSE102929, and GSE110993. The differentiation trajectory and key transcription factors (TFs) for HO occurrence were systematically analyzed by integrating single-cell RNA (scRNA) sequencing, bulk RNA sequencing, and assay of transposase accessible chromatin seq. The differential expression and enrichment pathways of TFs in heterotopically ossified tissues were identified. **Results.** HO that mimicked pathological cells was classified into HO1 and HO2 cell subsets. Results of the pseudo-temporal sequence analysis suggested that HO2 is a differentiated precursor cell of HO1. The analysis of integrated scRNA data revealed that ectopically ossified cells have similar transcriptional characteristics to cells in the fibrocartilaginous zone of tendons. The modified SCENIC method was used to identify specific transcriptional regulators associated with ectopic ossification. Xbp1 was defined as a common key transcriptional regulator of ectopically ossified tissues and the fibrocartilaginous zone of tendons. Subsequently, the CellPhoneDB database was completed for the cellular ligand-receptor analysis. With further pathway screening, this study is the first to propose that Xbp1 may upregulate the Notch signaling pathway through Jag1 transcription. Twenty-four microRNAs were screened and were found to be potentially associated with upregulation of XBPI expression after acute ischemic stroke. **Conclusion.** A systematic analysis of the differentiation landscape and cellular homogeneity facilitated a molecular understanding of the phenotypic similarities between cells in the fibrocartilaginous region of tendon and HO cells. Furthermore, by identifying Xbp1 as a hub regulator and by conducting a ligand-receptor analysis, we propose a potential Xbp1/Jag1/Notch signaling pathway.

1. Introduction

Tendon to bone healing is a major concern in sports medicine. Regardless of the advances in rotator cuff repair and anterior, healing in the tendon-bone interface (TBI) is the chief problem that has attracted significant attention [1–4].

The TBI contains four coherent layers with gradual composition, structure, and mechanical properties. Histologically, the TBI is composed of bone, calcified fibrocartilage, noncalcified fibrocartilage, and tendon, which ensures the smooth transmission of force from the muscle to bone [5–8]. However, following rotator cuff repair or anterior cruciate ligament reconstruction, large amounts of fibrovascular tissue accumulate in the TBI, hindering healing and weakening the biomechanical properties of the joint [6–12]. Accumulating evidence has demonstrated that fibrochondrocytes localized within the TBI are essential for tendon to bone healing [13]. Rui et al. found that the tendon and bone healed well in rabbits with more fibrocartilage cells, and the promotion of fibrocartilage cell proliferation could enhance tendon-bone healing [8]. Therefore, the study of the origin and characteristics of fibrochondrocytes could help improve tendon to bone healing [13].

Neurogenic heterotopic ossification is a common complication of acute ischemic stroke; however, its exact mechanism remains unknown. Heterotopic ossification (HO) is the appearance of osteoblasts in the soft tissues and the formation of bone tissue [14, 15]. Interestingly, HO demonstrates several characteristics similar to the healing process of the TBI. First, the tissue in HO is histologically similar to the TBI [14–16]. As an important feature of the tissue in HO and an essential process for fibrocartilage development, mineralization is observed in both HO and TBI formation [6, 16, 17]. Second, mechanical stimulation is pivotal for the development of both TBI and HO [16, 18]. Third, bone marrow-derived mesenchymal stem cells (BM-MSCs) and tendon-derived stem cells, the cellular sources of fibrocartilage and HO, have been reported to possess a multidifferentiation potential [19–25]. Fourth, numerous studies have demonstrated that the Notch and Hedgehog (Hh) signaling pathways could drive ectopic ossification and tendon to bone healing by mediating associated transcription factors, although the exact underlying mechanisms remain unclear [20, 26–30]. Fifth, some genes, such as COL2A1 and MKX, were found to be associated with the upregulation of the Notch and Hh signaling pathways and have the potential to promote tendon to bone healing [27, 30–33]. Moreover, the transcript levels of these genes (including COL2A1 and MKX) were found to be upregulated in ectopic ossified tissues [27, 34, 35]. Therefore, considering the homogeneity between HO and the TBI, we hypothesized that research on HO might serve as the reference for a better understanding of tendon to bone healing. Exploring the key regulators and pathways associated with the development of HO might provide novel insights into fibrocartilage restoration in the TBI.

The single-cell technique was used in this study to investigate the homogeneity between HO and the TBI. The single-cell transcriptome, which is a recent, novel research

method, enables comprehensive exploration of tissue microstates and thus the characterization and changes in the cellular condition [36–39]. The scRNA-seq transcriptome sample integration analysis has been used to assess cellular homology across different sequencing platforms at different centers [40, 41]. We speculate that the homogeneity between the two cells can be explored by integrating the analysis of HO tissue and fibrocartilage cells by single-cell techniques.

Transcription factors are targets for the regulation of cell differentiation [42]. In recent years, significant progress was achieved in the construction of gene regulatory networks (GRNs) based on single-cell transcriptome expression data. This data would help us understand the transcriptional regulatory networks of cells and uncover key TFs [43, 44]. On the other hand, transcription factor-mediated changes in cellular communication and pathway signaling are important manifestations of cellular homogeneity [45]. Cellular communication is defined as the transmission of information from one cell to another through a medium to produce a corresponding response. Ligand-receptor complex-mediated cell-cell communication is essential for coordinating various biological processes, such as development, differentiation, and inflammation [46]. CellPhoneDB serves as an integrated algorithm for identifying biologically relevant ligand-receptor interactions from the scRNA-seq data, which could be used to investigate new targets for disease treatment, while ligand proteins in biological signaling are regulated via transcription [47]. Numerous studies have demonstrated that ligand manipulation therapy is a viable method of targeted therapy [31]. Therefore, after constructing GRNs based on single-cell transcriptome expression data, we propose the use of that technique for ligand-receptor analysis, which may contribute to the identification of key transcription factors for cellular developmental targets in the HO and fibrocartilage regions, thus providing a theoretical foundation for future ligand therapy.

In this study, three single-cell sequencing datasets (GSE168153, GSE138515, and GSE102929) were available for the cells in the tendon fibrochondral region and HO tissue [46–48]. After the GRNs were constructed, ligand-receptor analysis was performed to identify the core transcription factors. We aimed to identify common intercellular differentiation and transcriptional regulation pathways associated with fibrocartilage development in the TBI and HO.

2. Materials and Methods

2.1. Data Retrieval. All data were obtained from the Gene Expression Omnibus database, which mainly includes the GSE94683, GSE144306, GSE168153, GSE138515, GSE102929, and GSE110993 datasets (Table 1). GSE94683 includes transcriptional profiles of mesenchymal stromal cells in ectopically ossified tissues from nine healthy donors and seven patients with HO. GSE168153 is the only single-cell sequencing dataset available for the fibrocartilaginous region of mouse tendons. GSE144306 and GSE168153 are datasets of the same study, and GSE144306 is an assay for transposase-accessible chromatin with high-throughput

TABLE 1: Raw data working sheet.

Mesenchymal stromal cells in ectopic ossified tissues	GSE94683 from GEO database
A single-cell sequencing dataset available from the fibrocartilaginous region of mouse tendons	GSE144306 from GEO database
A ATAC-seq dataset available from the fibrocartilaginous region of mouse tendons	GSE168153 from GEO database
A scRNA-seq dataset from the Achilles tendon of 6-week-old male C57Bl/6 mice	GSE138515 from GEO database
A scRNA-seq dataset from 4-week-old mouse tail tendon cells	GSE102929 from GEO database
A dataset reflecting microRNA changes after acute ischemic stroke	GSE110993 from GEO database
Datasets used to screen for differentially expressed TF in HO tissue	The TTRUST dataset

sequencing (ATAC-seq) dataset. GSE138515 contains single-cell transcriptome data from the Achilles tendon of 6-week-old male C57Bl/6 mice. GSE102929 reveals single-cell RNA transcriptome data from 4-week-old mouse tail tendon cells under physiological and simulated HO pathological conditions. Lactating rat tail tendon has less extracellular matrix, more number of cells per tissue volume, and strong cell growth and differentiation ability. Primary and passaged tendon cells can be obtained in large quantities relatively quickly, facilitating subsequent experiments. Therefore, GSE102929 can be used to reveal the differentiation trajectory characteristics and heterogeneity of tendon-derived cells under pathological HO conditions. GSE110993 is a dataset reflecting microRNA changes following acute ischemic stroke. This dataset was used to screen for microRNAs with altered expression in the serum after acute ischemic stroke. The target mRNAs of the miRNAs were predicted using the miRWalk database (<http://mirwalk.umm.uni-heidelberg.de/>).

2.2. Differential Expression Analysis of TFs. The TTRUST dataset is a database that records the regulatory relationships of TFs and contains not only the target genes corresponding to TFs but also the regulatory relationships among TFs [49]. It was used to screen for differentially expressed TFs in HO tissues. Differential analysis was performed by the R software limma package, and p values less than 0.05 were defined as significantly different. Differentially expressed genes were ranked according to $|\log FC|$, and 70 genes that were highly expressed in HO tissues were obtained.

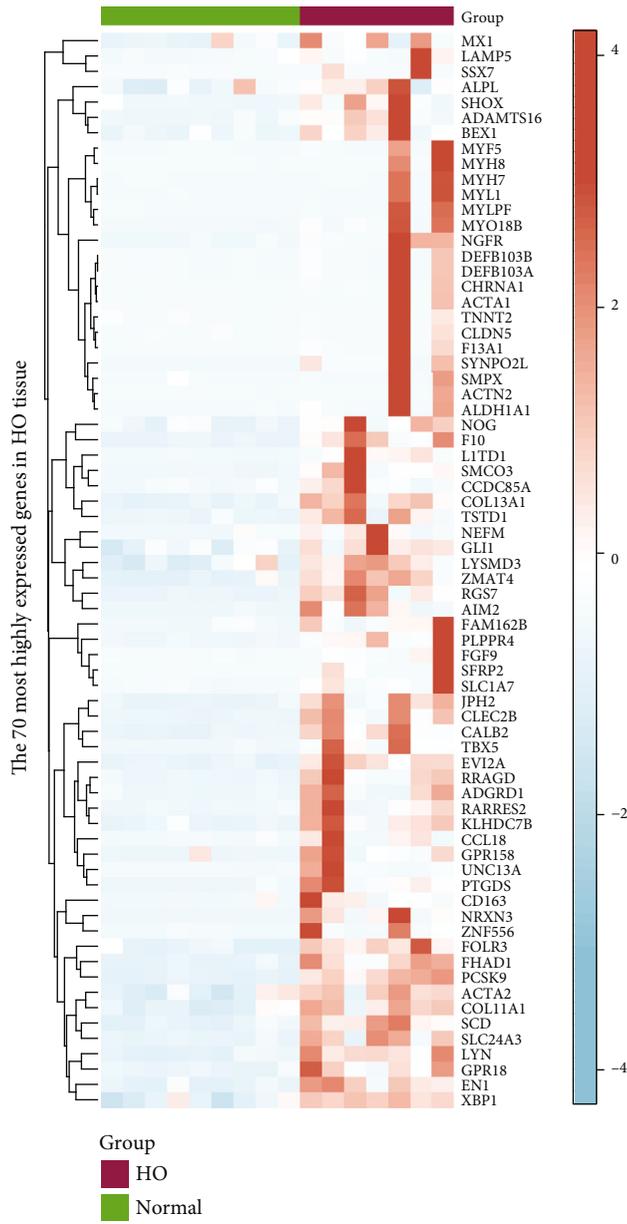
2.3. Single-Cell RNA Sequencing (RNA-seq) Analysis. RNA-seq analysis was performed using the Seurat package based on the R software (version 4.0.2) [50]. Canonical correlation analysis (CCA) was also used for dataset integration. Principal component analysis (PCA) and uniform manifold approximation and projection/ t -distributed stochastic neighbor embedding (UMAP/TSNE) methods were used for the scRNA-seq dimensionality reduction analysis. Cell annotation of GSE168153 and GSE138515 was completed according to the information provided in the original article, and the CellMarker database and SingleR package were used for further correction [51, 52]. The “FindAllMarkers” function in Seurat was further used for the differential expression analysis, as described in the previous analysis. Monocle 2 and Monocle 3 were used to perform pseudo-temporal trajectory analysis [53]. Cellular ligand-receptor analysis was performed according to the instruction manual using the Cell-

PhoneDB database and software [54]. Raw data of all analysis processes were saved.

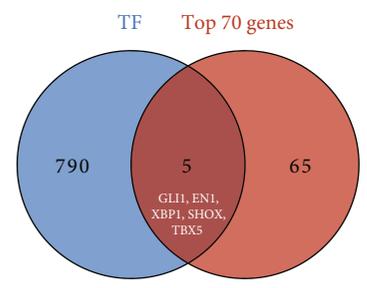
2.4. Hub TF Analysis. To identify specific transcriptional regulators associated with ectopic ossification, a modified version of the SCENIC method was used to construct a regulatory network between HO-associated targets and TFs [55, 56]. Thus, each TF has its corresponding downstream target gene. Currently, SCENIC only supports transcriptional positive regulation analysis. Based on the above results, the regulon activity score was calculated for each cell, and the method was repeated three times to confirm the stability of the regulatory relationships [57]. Then, by performing Pearson correlation coefficient analysis for each pair of regulatory relationships, the similarity of the overall regulatory activity of different TFs was assessed, and this was used to quantify the regulatory correlations among cell types. The top 5 genes in each cell were screened for correlation analysis.

2.5. ATAC-seq Analysis. The Fastq sequencing data for the ATAC-seq dataset of the tendon-bone union region were obtained from GSE144306. Filtering and quality control were performed with the trim_galore software, and bowtie2 was used for comparison and calculation of statistical comparison rates. After finding peaks using MACS2, the length of the insert fragments, fraction of reads in peak values, and irreproducibility discovery rate were used to calculate duplicates. The MEME tool was then used to annotate motifs and verify potential transcriptional regulatory relationships [58].

2.6. Enrichment Analyses. Four main methods of enrichment analysis were used in this study, namely, Gene Set Enrichment Analysis (GSEA), Gene Set Variation Analysis (GSVA), pathway enrichment analysis, and Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analysis: (1) GSEA was conducted using the Molecular Signatures Database (MSigDB) Collection gene set database, and the database for the reference genes was MSigDB Collections (c2.cp.v7.2. symbols.gmt) [59, 60]. (2) The GSVA for microarray and RNA-Seq dataset uses the reference gene database MSigDB Collections (2.cp.kegg.v6.2.symbols) [61]. False discovery rate < 0.25 and $p.adjust < 0.05$ were considered significantly enriched. (3) The pathway enrichment analysis of single cells was performed using the Single-Cell Signature Explorer, and the expression abundance of the target pathway is calculated.



(a)



(b)

FIGURE 1: Continued.

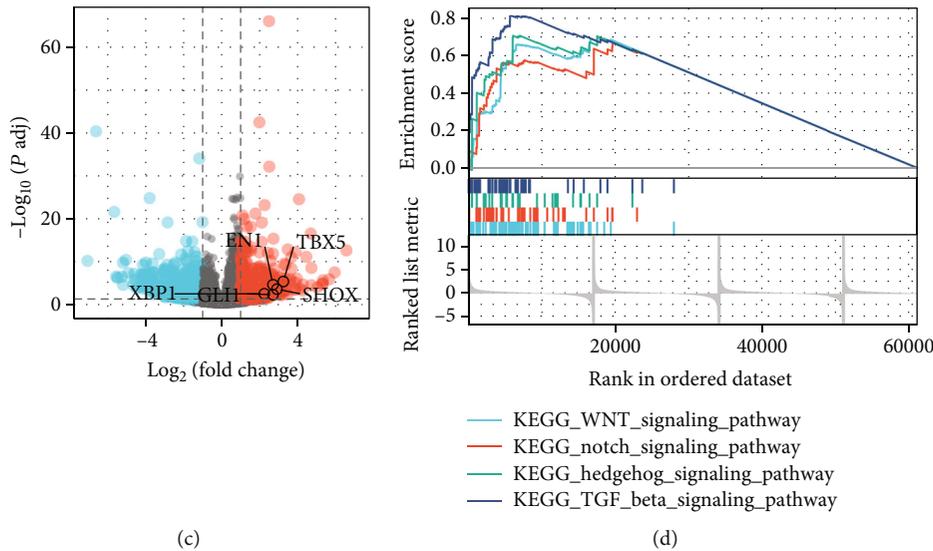


FIGURE 1: Characterization of the differential expression and enrichment pathways of transcription factors in ectopically ossified tissues. (a) Heat map of the top 70 genes expressed in heterotopic ossification (HO) tissues. (b) Intersection of the top 70 genes and 795 genes from the TTRUST database. (c) Volcano plot of five differential transcription factor expression levels based on GSE94683 data. (d) Visualization of four HO-related KEGG pathways based on GSEA results. GSEA: Gene Set Enrichment Analysis; KEGG: Kyoto Encyclopedia of Genes and Genomes.

(4) GO and KEGG enrichment analyses are widely used research methods; they are conducted using the clusterProfiler package (version 3.14.3), and the org.Mm.eg.db package (version 3.10.0) was used for ID transformation [59, 62, 63].

2.7. Statistical Analysis and Graphing. Statistical graphs were created using the R software (version 4.0.2, R Foundation for Statistical Computing, Vienna, Austria). Comparisons between two groups were performed using a two-tailed, unpaired Student's *t*-test, and the results are presented as standard deviation \pm mean. Analysis of variance was used to compare multiple groups.

3. Results

3.1. Differential Expression and Enrichment Pathways of TFs in Ectopically Ossified Tissues. Differential analysis was performed on 17076 genes from GSE94683 (including nine healthy donor samples and seven samples of ectopically ossified tissues from patients with HO). Among them, 1203 genes satisfied the threshold of $|\log_2(\text{FC})| > 1$ and $p.\text{adj} < 0.05$, 456 genes were highly expressed in HO tissues, and 747 genes were lowly expressed in HO tissues. Then, 70 genes with the highest expression in HO tissues were screened, and the differential expressions of these genes were visualized in different samples by drawing a heat map (Figure 1(a)). Moreover, 795 TFs identified were obtained from the TTRUST database. By plotting Venn diagrams, five genes, namely, GLI1, EN1, XBP1, SHOX, and TBX5, were found to be the intersection of the 795 TFs and 70 genes highly expressed in HO tissues (Figure 1(b)). The characteristics of these five TFs, namely, GLI1, EN1, XBP1, SHOX, and TBX5, differentially expressed in GSE94683 are shown

in Figure 1(c). Thus, to explore the biological pathways differentially expressed in HO tissues, we performed GSEA. The analysis suggested that the WNT, Notch, Hh, and transforming growth factor (TGF) beta signaling pathways in the KEGG pathway were highly expressed in HO tissues ($p < 0.05$).

3.2. Heterogeneity and Proposed Temporal Differentiation Trajectory of an Ectopic Ossification Cell Model. At present, the differentiation trajectory of ectopically ossified cells remains unknown, and the associated single-cell transcriptome has not been fully characterized. Silencing *Mkx* is shown as a viable method of constructing ectopic ossification models [15, 64, 65]. GSE102929 reveals scRNA transcriptomic data from 4-week-old mouse tail tendon cells under physiological and simulated HO pathological conditions, all filtered, normalized, and clustered. UMAP clustering analysis of single-cell RNA-seq revealed that single cells of mouse tail tendon cells could be distinguished into four clusters under physiological and simulated HO pathological conditions (Figure 2(a)). These cell clusters were found to correlate with the cell model groups. Among them, clusters 0 and 3 belong to the scRNA-seq data of mouse tail tendon cells under physiological conditions, while clusters 1 and 2 belong to the scRNA-seq data of mouse tail tendon cells under pathological conditions (Figure 2(b)). Furthermore, GSVA was used to quantify pathway activity within every single cell, all differential pathways ($p < 0.05$) were extracted, and a pathway heat map (Supplementary Figure 1A) was drawn according to the pathway activity of each sample. Cluster 2 has a more pronounced degree of HO differentiation than cluster 1. Therefore, the pathway activities of clusters 2 and 0 were compared to further identify the pathways that were

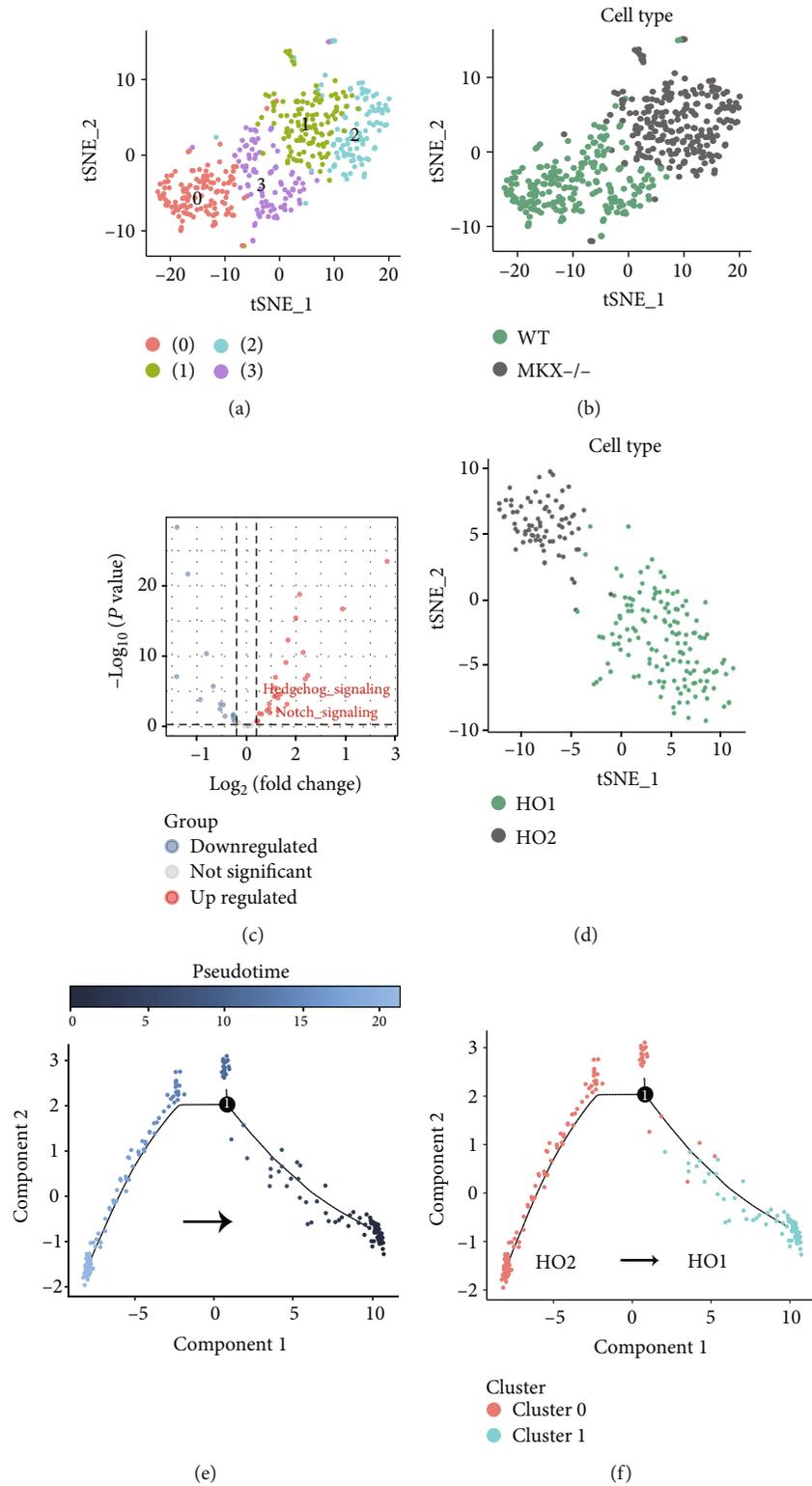


FIGURE 2: Continued.

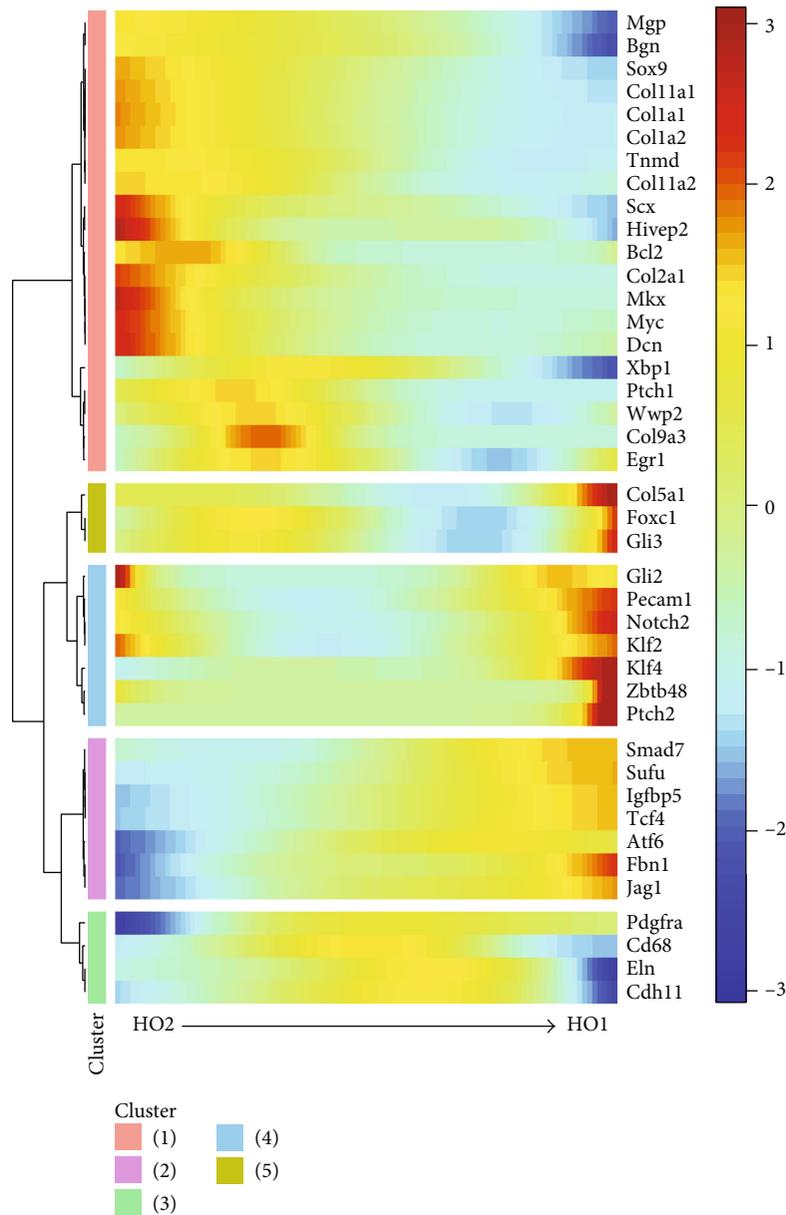


FIGURE 2: Characterization of the heterogeneity and pseudo-temporal differentiation trajectories in heterotopic ossification (HO) cell models. (a) tSNE subcluster clustering maps of mouse tail tendon cells under physiological and simulated HO pathological conditions, with different clusters indicated by colors. (b) tSNE subcluster clustering maps of mouse tail tendon cells under physiological and simulated HO pathological conditions, with green and gray indicating tail tendon cells under physiological conditions and simulated HO pathological conditions, respectively. (c) Volcano map of the pathway differential expression in mouse tail tendon cells under physiological and simulated HO pathological conditions. (d) Uniform manifold approximation and projection (UMAP) clustering of mouse tail tendon cells under simulated HO pathological conditions, with HO1 class cells (cluster 0) defined in green and HO2 class cells (cluster 1) defined in gray. (e, f) Pseudo-temporal sequence of HO cell differentiation in mice. (g) Expression of HO-related genes in mouse tail tendon cells under simulated HO pathological conditions with the change of differentiation chronology.

significantly upregulated/downregulated in ectopic ossified tissues. Compared with the control group (cluster 0), the myogenic MYOGENESIS and MYC TARGETS pathways were lowly expressed in the ectopic ossification model group (cluster 2), and KRAS SIGNALING DN was also lowly expressed in cluster 2 (Supplementary Figure 1B-1D). By contrast, the ectopic ossification-related pathways, namely,

Hh and Notch signaling pathways, were highly expressed, and the oxidative phosphorylation pathway was also highly expressed in cluster 2 (Supplementary Figure 1E-1G). Mlx was significantly upregulated in cluster 4 (Supplementary Figure 2A, 2B). Then, heat maps were plotted to further demonstrate the degree of differential expression between the Notch and the Hh signaling pathways (Figure 2(c)).

Ectopic ossification mimicking pathological cells were reclustered and defined as HO1 (corresponding to Supplementary Figure 2C, cluster 1) and HO2 (corresponding to Supplementary Figure 2C, cluster 0) (Figure 2(d) and Supplementary Figure 2C). The results of the mimetic timing analysis suggested that HO2 was a precursor cell for HO1 differentiation (Figures 2(e) and 2(f)). To further characterize the pathological differentiation of ectopically ossified cells, the proposed chronological expression profile of genes associated with the ectopic ossification phenotype was revealed (Figure 2(g)). The gene expression profiles of *Xbp1*, *E2f7*, *Fgfr1*, *Icam1*, *Jag1*, and *Ncam1* are shown in Supplementary Figure 2D. *Mgp*, *Bgn*, *Sox9*, *Col11a1*, *Col1a1*, *Col1a2*, *Tnmd*, *Col11a2*, *Scx*, *Hivep2*, *Bcl2*, *Col2a1*, *Mkx*, *Myc*, and *Dcn* were enriched in HO2-like cells. *Col5a1*, *Foxc1*, *Gli3*, *Gli2*, *Pecam1*, *Klf2*, *Klf4*, *Zbtb48*, *Ptch2*, *Smad7*, *Sufu*, *Igfbp5*, *Tcf*, *Fbn1*, and *Jag1* were enriched mainly in HO1-like cells (Figure 2(g)). Among them, *Gli3*, *Cli2*, *Pecam1*, and *Ptch2* are the star genes of the Hh signaling pathway, and *Notch2* and *Jag1* are the star genes of the Notch signaling pathway, which suggested that the activities of the Notch and Hh signaling pathways may be upregulated in HO1-like cells. According to the expression characteristics of these genes, HO1-like cells are more similar to chondrocyte-like cells, and HO2-like cells are more similar to tenocyte-like cells. Therefore, HO2 cells were defined as “tenocyte-like cells.”

3.3. Endoplasmic Reticulum (ER) Stress Effector XBP1 Was Found to Be a Key Transcriptional Regulator in Ectopically Ossified Tissues. In this study, key TFs in HO1-like cells (chondrocyte-like cells) and HO2-like cells (tenocyte-like cells) were identified using comprehensive network analysis (Supplementary Figure 3). The network analysis identified *Ebf1*, *Usf2*, *Klf3*, *Zmiz1*, and *Sin3a* as key transcriptional regulators in HO1-like cells, and *Ep300*, *Smarcc*, *Foxj2*, *Xbp1*, and *E2f5* are key transcriptional regulators in HO2-like cells. To systematically compare the transcriptional profiles in different cells, we compared the expression correlation of each TF based on the connection specificity index (CSI) (Figure 3(a)). In HO1-like cells (chondrocyte-like cells) and HO2-like cells (tenocyte-like cells), *Xbp1* and *Klf3* were found to be key transcriptional regulators of MH1 module cells, and TF *Ebf1* was found to be a key transcriptional regulator of MH2 module cells. In addition, the MH1 module was found to be more active in HO2-like (tenocyte-like cells) cells, while the MH2 module was more active in HO1-like (chondrocyte-like cells) cells (Figures 3(b) and 3(c)). This also demonstrates that *Xbp1* and *Klf3* are key transcriptional regulators in HO2 (tenocyte-like cells), while *Ebf1* is a key transcriptional regulator in HO1 (chondrocyte-like cells). The Venn diagram reveals that *Xbp1* is a common key TF for HO tissue bulk-RNA-seq and scRNA-seq analyses (Figure 3(d)).

3.4. Ectopically Ossified Cells Have Similar Transcriptional Characteristics to Cells in the Fibrocartilaginous Zone of Tendons. Based on the proposed temporal analysis characteristics of the gene expression in Figure 2(g), combined with

the results of previous studies, the gene expression characteristics of ectopic ossification demonstrated strong similarities to those of the fibrocartilaginous region of the tendon-bone union [8, 66–68]. Therefore, we hypothesized that ectopically ossified cells have similar transcriptional characteristics to cells in the fibrocartilaginous zone of the tendon-bone junction region. Then, we integrated scRNA-seq transcriptome data from mouse Achilles tendon, scRNA-seq transcriptome data from mouse tendon junction region, and scRNA-seq transcriptome data from mouse tail tendon cells under simulated HO pathological conditions for scRNA-seq integration analysis.

First, the cell types of single-cell transcriptome data were annotated for GSE138515 and GSE168153. GSE138515 contains single-cell transcriptome data from the Achilles tendon of 6-week-old male C57Bl/6 mice, and PCA grouping was performed after quality control of these cells (Supplementary Figure 4A, 4C). With the UMAP method, GSE138515 was found to have 12 clusters of cell types (Figure 4(a) and Supplementary Figure 4B). The cell differentiation trajectory of GSE138515 is shown in Supplementary Figure 4D. A preliminary annotation of these cells was performed based on SingleR (Supplementary Figure 4E). Among the cell clusters, clusters 0, 7, and 11 were initially classified as endothelial cells; cluster 2 was initially classified as erythrocytes; clusters 1, 3, 4, 8, 9, 10, and 12 were initially classified as fibroblasts; cluster 5 was tentatively classified as macrophages; cluster 6 was tentatively classified as oligodendrocytes. In addition, these cells were further annotated regarding the results of previous studies (Figure 4(a)) [69]. As shown above, the quality of scRNA-seq data of GSE168153 was controlled and grouped into nine clusters (Supplementary Figure 5A, 5D). The cell differentiation trajectory of GSE168153 is shown in Supplementary Figure 5B. Moreover, these cells were further annotated based on the results of previous studies (Figure 4(b)) [66]. Accordingly, the proposed temporal expression profile of genes associated with the phenotypic cartilaginous region of tendon fibers is shown in Figure 4(c). The cell annotation method combines the results of the pseudo-temporal analysis to classify cells into tenocyte, tenocyte-like attachment cell, attachment cell, chondrocyte-like attachment cell, and chondrocyte, which are the five major cell clusters (Figure 4(c) and Supplementary Figure 5C). The trajectory of cell differentiation is presented in the following order: tenocytes, tenocyte-like attachment cells, attachment cells, chondrocyte-like attachment cells, and chondrocytes (Supplementary Figure 5C). To the best of our knowledge, this study provided the most detailed subcellular classification of the tendon junctional area to date. Moreover, this classification demonstrates the spatial distribution characteristics of cells in the fibrocartilage layer. Previous analyses identified *Xbp1* as a common key TF for HO tissue bulk-RNA-seq and scRNA-seq analyses. In the present study, the results of the pseudo-temporal analysis suggested that *Xbp1* was enriched in tenocytes, tenocyte-like attachment cells, and attachment cells. “Chondrocyte-like attachment cells” and “Chondrocytes” were less expressed (Figures 4(d) and 4(e)).

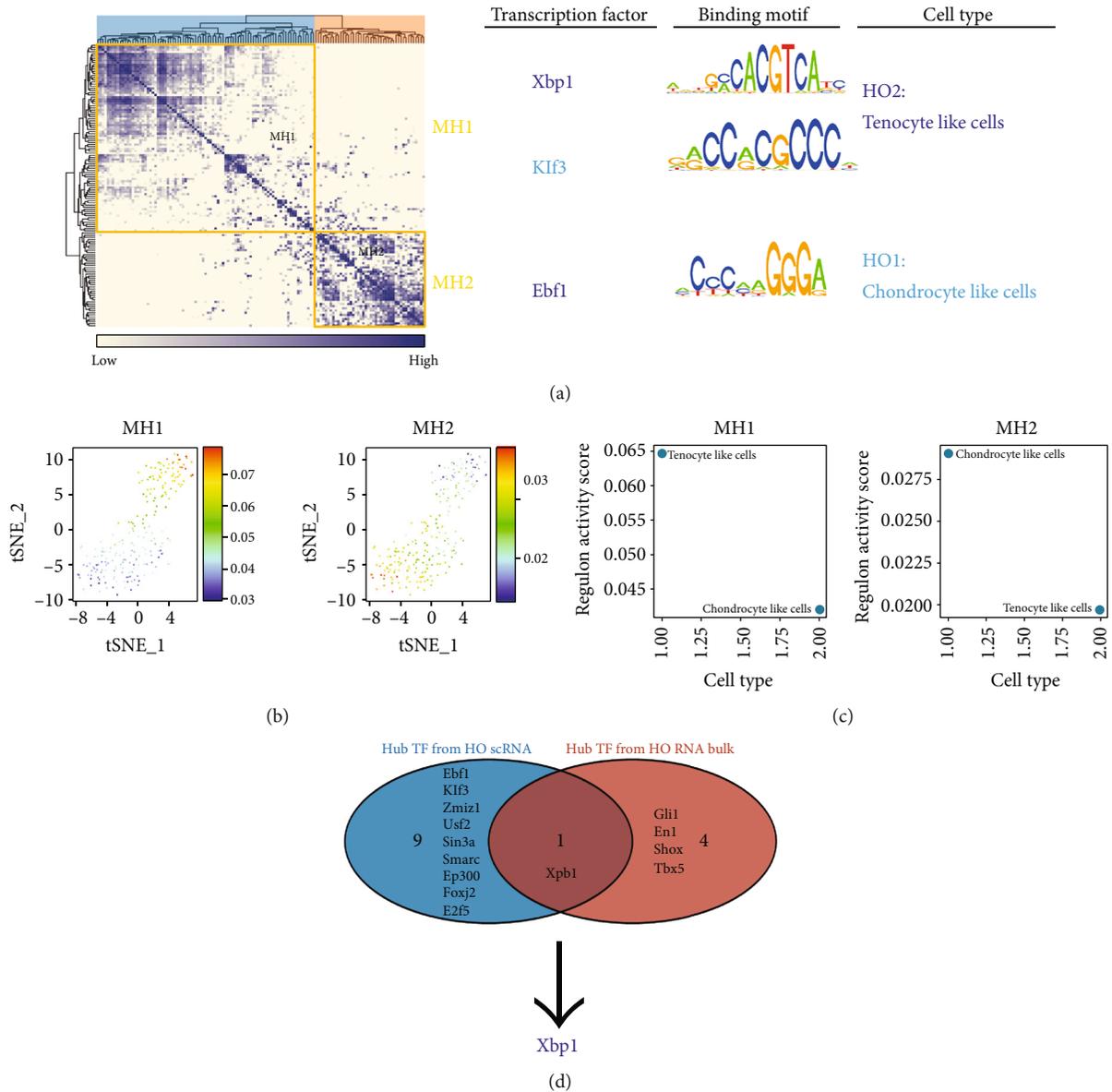


FIGURE 3: Endoplasmic reticulum stress effector XBP1 was found to be a key transcriptional regulator in ectopic ossified tissues. (a) Regulatory modules were identified based on the regulon connection specificity index matrix, and the core transcription factors, binding motifs, and corresponding cell types were extracted. (b) The activity of each module in different cells is represented by tSNE plots. (c) Corresponding cell types of different modules. (d) The Venn diagram reveals that Xbp1 is a common key transcription factor for heterotopic ossification bulk-RNA-seq and scRNA-seq analyses.

CCA was also used for dataset integration. Three datasets (including GSE168153, GSE138515, and GSE102929 under simulated HO pathological conditions) were integrated using the CCA approach. These datasets were then grouped into nine cell clusters (Supplementary Figure 6A, 6B). The top 5 marker genes in each cell cluster were flagged (Figure 4(f) and Supplementary Figure 6B). In addition, the UMAP graph was used to show all cell clusters, and the original annotations of all cells were also identified (Supplementary Figure 6C). Figure 4(g) shows the distribution of datasets in the UMAP plot. Before reannotating the cells, HO2 was found to be enriched in attachment cell cluster 1 and tendon fibroblasts, while HO1 was enriched near chondrocyte-like

attachment cell 3 (Figure 4(g) and Supplementary Figure 6C). After reannotation of the cells, HO2 was found to be enriched in tenocyte-like attachment cell 1, while HO1 was enriched in chondrocyte-like attachment cell 1 (Figure 4(h)). This suggests that ectopically ossified cells have similar transcriptional characteristics to cells in the fibrocartilaginous zone.

3.5. CCA Integration of Pseudo-Temporal Differentiation Trajectories and Gene Expression Signatures in scRNA-seq. The cell differentiation trajectory of mouse tail tendon cells under simulated HO pathological conditions started from HO2 to HO1 (Figures 2(e) and 2(f)). To verify that the cell

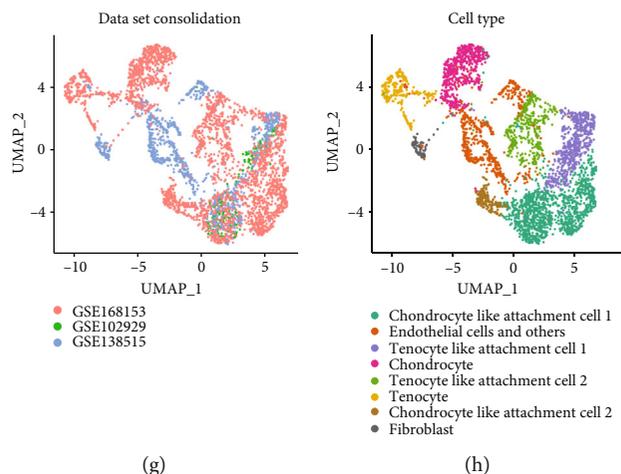


FIGURE 4: Ectopically ossified cells have similar transcriptional characteristics to cells in the tendon fibrocartilaginous region. (a) For GSE138515, cell clustering and annotation are shown on the uniform manifold approximation and projection (UMAP) plots. (b) For GSE168153, cell clustering and annotation are presented on the UMAP plots. (c) For GSE168153, the key gene expression features of ectopic ossification and the tendon fibrocartilaginous zone are presented with a pseudo-temporal analysis. (d) Expression features of *Xbp1* on pseudo-temporal trajectories. (e) Expression characteristics of *Xbp1* in different cells in GSE168153 shown on the UMAP plots. (f) Differential gene expression analysis of the top 5 genes showing upregulated expression in all eight clusters in the integrated dataset (including GSE168153, GSE138515, and GSE102929 under simulated HO pathological conditions). (g) UMAP plots showing the distribution of the different datasets. (h) Cellular annotations for the integrated datasets are shown on the UMAP plots, with different colors representing cell types.

differentiation trajectory of the CCA-integrated single-cell dataset is similar to that of mouse tail tendon cells under simulated HO pathological conditions, a pseudo-temporal analysis of the CCA-integrated single-cell dataset was performed based on Monocle 3 (Figure 5(a)). In the CCA-integrated single-cell dataset, the differentiation trajectory of cells started from tenocyte-like attachment cell 1 to chondrocyte-like attachment cell 1 (Figures 4(h) and 5(b)). In this study, the genes associated with ectopic ossification and the cartilage region of tendon fibers (*Atf4*, *Bgn*, *Cdh11*, *Col11a1*, *Col1a1*, *Col1a2*, *Col5a1*, *Dcn*, *Eln*, *Ep300*, *Fbn1*, *Gli3*, *Hes1*, *Hivep2*, *Klf2*, *Klf3*, *Klf4*, *Mgp*, *Notch2*, *Pdgfra*, *Smad4*, *Smad7*, *Smo*, *Tbx5*, *Tcf4*, *Tnmd*, *Usf2*, *Xbp1*, *Zmiz1*, etc.) were found in tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1. “Chondrocyte-like attachment cell 1” was significantly enriched in “tenocyte-like attachment cell 1” and “chondrocyte-like attachment cell 1.” The expression profiles of the key genes of ectopic ossification and tendon fibrochondral regions in the CCA-integrated single-cell dataset were presented by the pseudo-temporal analysis. *Atf4*, *Bcl2*, *Bgn*, *Cdh11*, *Col11a1*, *Col11a2*, *Col2a1*, *Col5a1*, *Dcn*, *Foxc1*, *Hes1*, *Hivep2*, *Klf3*, *Klf4*, *Mgp*, *Mkx*, *Smad4*, *Sox9*, *Tnmd*, *Xbp1*, and *Zmiz1* had high expression in the early stages of the pseudo-temporal distribution and low expression in the late stages.

3.6. Heterogeneity Analysis of Tenocyte-Like Attachment Cell 1 and Chondrocyte-Like Attachment Cell 1. The CCA-integrated pseudo-temporal differentiation trajectory and gene expression characterization of the single-cell dataset presented above revealed that HO cells were mainly concentrated in tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1. To demonstrate that ectopically ossi-

fied cells have similar transcriptional characteristics to cells in the fibrocartilaginous zone of the tendon-bone union, it is necessary to further confirm the transcriptional characteristics of cells from different samples in tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1. Therefore, the cells belonging to tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1 were extracted in each sample separately, a new PCA clustering analysis was performed, and five PCs were finally identified (Supplementary Figure 7A and Figure 6(a)). The first 10 marker genes of each cell cluster were marked (Supplementary Figure 7B). Marker genes of PC1 and PC2 are shown in Supplementary Figure 7C. Based on the cell annotation map of UMAP, the cell clusters were divided into the following five clusters: tenocyte-like attachment cells and chondrocyte-like attachment cells 1-4 (Figure 6(b)). Moreover, the distribution of cells from different sources is shown in Supplementary Figure 7D. The correspondence of these cells with the annotated subpopulation of cells in Figure 4(h) is also shown in Figure 6(c). Most of the cells in the tenocyte-like attachment cell 1 group in Figure 4(h) belonged to the tenocyte-like attachment cell 1 group in Figure 6(b) annotation, but a few cells belonged to chondrocyte-like attachment cell 2 cluster. The pseudo-temporal analysis of this single-cell dataset is shown in Figures 6(d) and 6(e), which suggests a cell differentiation trajectory from tenocyte-like attachment cell to chondrocyte-like attachment cell 2 and finally to chondrocyte-like attachment cell 1. This finding also suggests that the differentiation process of tenocyte-like attachment cell to chondrocyte-like attachment cell 1 is gradually progressive. The chondrocyte-like attachment cell 2 stage is the intermediate stage of the differentiation process.

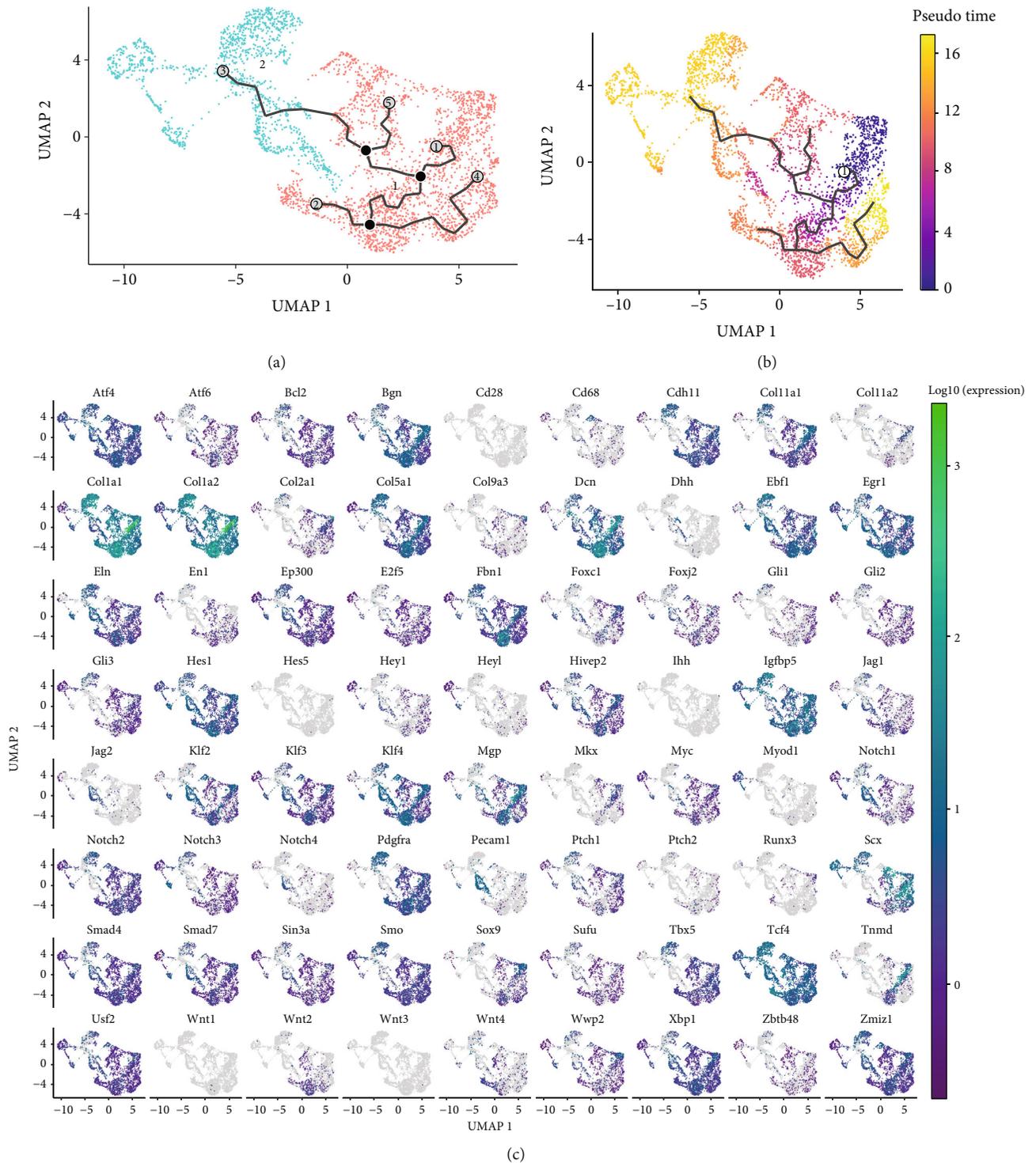


FIGURE 5: Pseudo-temporal differentiation trajectories and gene expression characteristics of the canonical correlation analysis- (CCA-) integrated single-cell datasets. (a) Pseudo-temporal series analysis of the CCA-integrated single-cell dataset based on Monocle 3. (b) Different pseudo-temporal sequences are represented by different colors in the CCA-integrated single-cell dataset. (c) Based on single-cell gene expression profiles, genes related to ectopic ossification and tendon fibrochondral region that were significant in both tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1 were significantly enriched.

For the review of cell fractionation, we found the relative proportions of the cell subpopulations, and the average cell numbers in each tissue are shown in Figure 6(f). Tenocyte-like attachment cells and chondrocyte-like attachment cells

1 and 2 were found to be the major cell types in tendon tissues and mouse tail tendon cells under simulated HO pathological conditions. The core marker genes and sample preferences for each cluster are shown in Figure 6(g). *Anxa2*

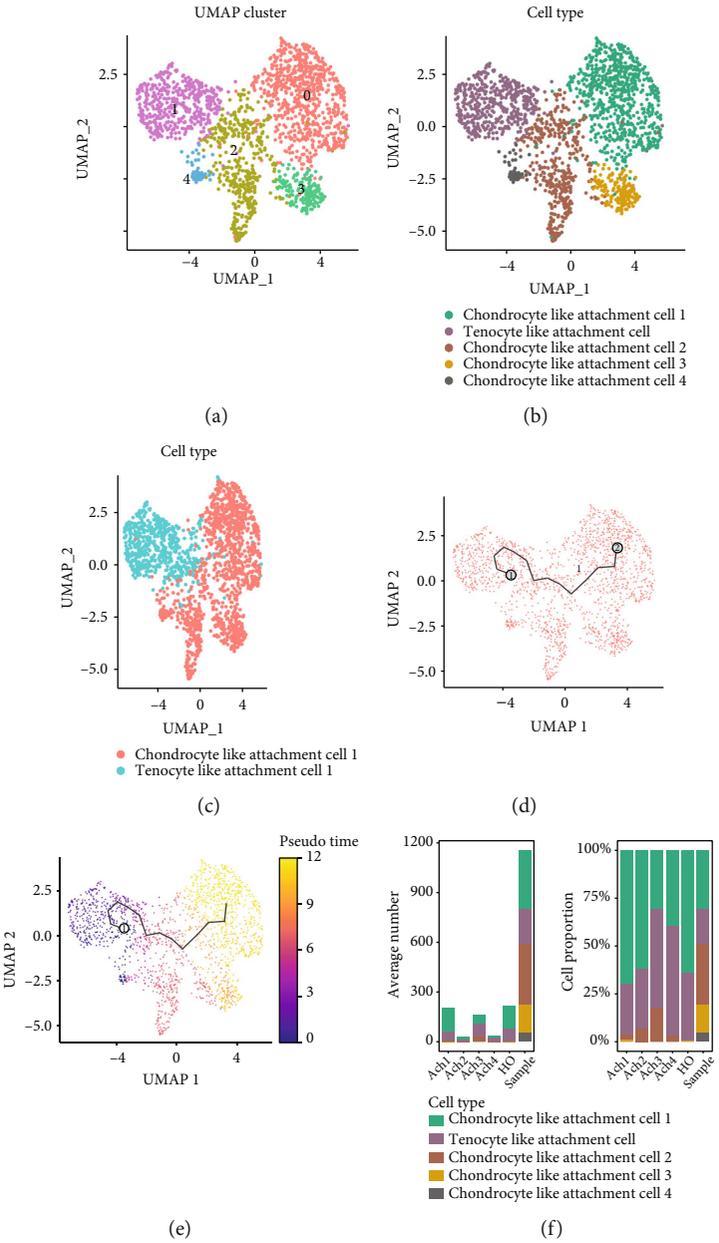


FIGURE 6: Continued.

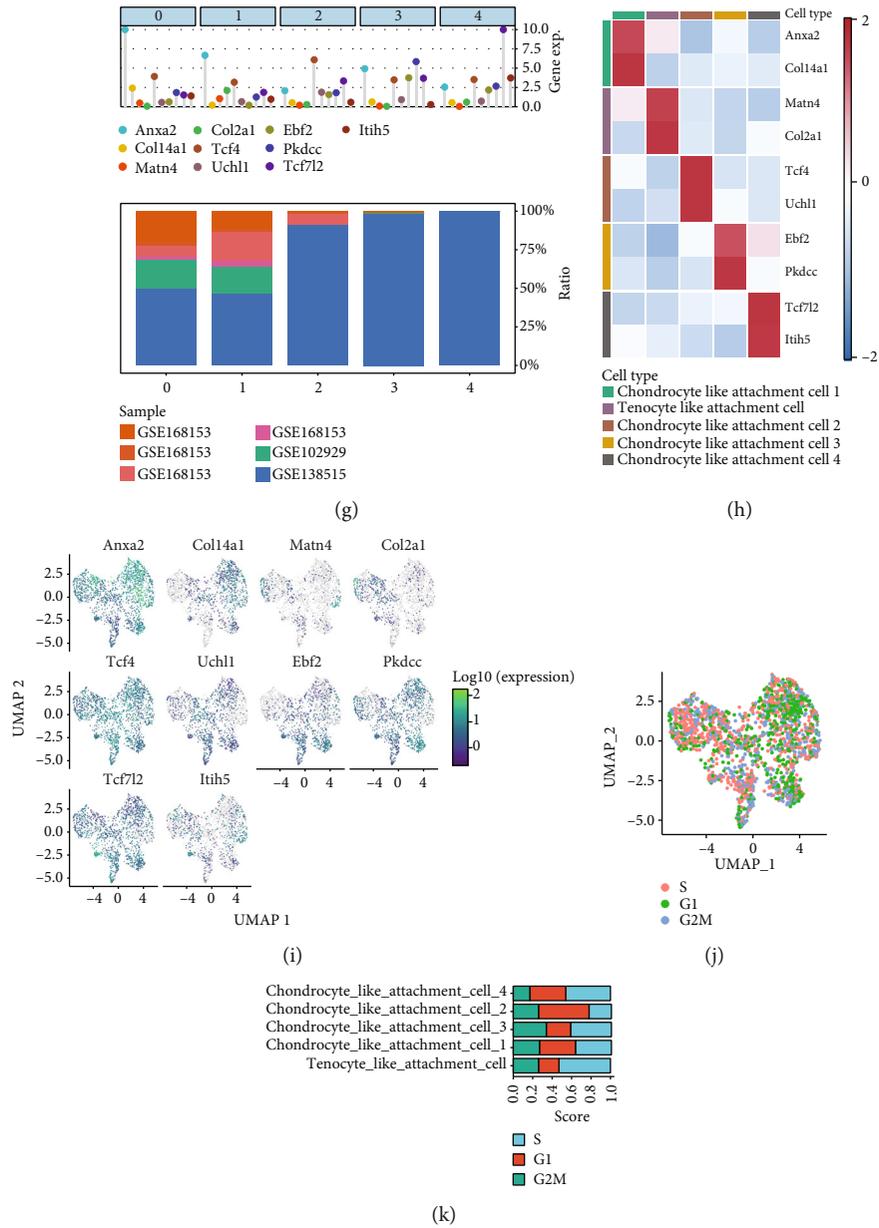


FIGURE 6: Heterogeneity analysis of tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1. (a) Uniform manifold approximation and projection- (UMAP-) based analysis of tenocyte-like attachment cell 1 and chondrocyte-like attachment cell 1 that were classified into five clusters. (b) Cell annotation map based on UMAP-classified cell clusters, namely, tenocyte-like attachment cell and chondrocyte-like attachment cells 1–4. (c) A UMAP diagram shows the correspondence of these cells with the annotated subgroups of Figure 4(h) cells. Blue indicates tenocyte-like attachment cell 1, and red indicates chondrocyte-like attachment cell 1. (d) Pseudo-temporal series analysis of this single-cell dataset based on Monocle 3. (e) Different degrees of pseudo-temporal differentiation sequences in this dataset are indicated by different colors. (f) Relative proportions and average cell counts of cell subpopulations in each tissue. (g) Core marker genes and sample preferences for each cluster. (h) Heat map of the relative expression characteristics of marker genes associated with each cell subpopulation. (i) UMAP plots of the expression characteristics of marker genes associated with each cell subpopulation. (j) The cell division cycle of all cells was explored with the Seruat package. Note: red indicates the S phase, green indicates the G1 phase, and blue indicates the G2/M phase. (k) Bar graphs of the heterogeneity of cell mitosis in each cell subpopulation.

and *Col14a1* are marker genes for chondrocyte-like attachment cell 1, *Matn4* and *Col2a1* are marker genes for tenocyte-like attachment cell, *Tcf4* and *Uchl1* are marker genes for chondrocyte-like attachment cell 2, *Ebf2* and *Pkdcc* are marker genes of chondrocyte-like attachment cell 3, and *Tcf712* and *Itih5* are marker genes of chondrocyte-like

attachment cell 4. The expression characteristics of these cell subpopulation-associated marker genes are shown in Figure 6(i). Then, the cell division cycles of all cells were explored using the Seruat package, and more significant heterogeneity in the mitotic cell divisions of each cell subpopulation was found (Figures 6(j) and 6(k)). These series of

single-cell gene expression profiles were also provided, systematically demonstrating the gene expression profile of the fibrocartilaginous region of ectopically ossified tissues and tendons (Supplementary Figure 8A). Information on the altered expression of HO-related genes with pseudo-temporal changes is shown in Supplementary Figure 8B. These cell differentiation trajectories further validate that ectopically ossified cells have similar transcriptional characteristics to cells in the fibrocartilaginous region of tendons.

3.7. XBP1 Is a Key Cotranscriptional Regulator in the Tendon-Bone Union Region and Ectopically Ossified Cells. TFs largely determine the characteristics of cell differentiation trajectories, so finding key TFs can help us explore the core mechanisms of ectopic ossification and differentiation. First, key TFs were identified in five clusters (tenocyte-like attachment cells, chondrocyte-like attachment cells 1-4) using a comprehensive network analysis combined with a modified SCENIC approach (Figure 7(a)). The key TFs in the tenocyte-like attachment cell cluster include *Mef2c*, *Foxo3*, *Xbp1*, *Sp4*, and *Klf11*. The key TFs in chondrocyte-like attachment cell 1 were *Stat1*, *Reta*, *Cebpb*, *Nfkb1*, and *Zfp704*. The key TFs in chondrocyte-like attachment cell 2 were *Sox4*, *Tbx5*, *Bmyc*, *Smarcc1*, and *Ybx1*. The key TFs in chondrocyte-like attachment cell 3 include *Hoxa3*, *Hoxa4*, *Hoxa5*, *Pbx1*, and *Meis*. The key TFs in chondrocyte-like attachment cell 4 were *Gli1*, *Tcf3*, *Tcf712*, *Hoxb5*, and *Maf*. The expression characteristics of some core TFs for each cell group are shown in Figure 7(b). The regulatory modules were identified based on the regulon CSI matrix, and the core TFs, binding motifs, and corresponding cell types were extracted (Figure 7(c)). The M1 module mainly included the two cell types: attachment cells and chondrocyte-like attachment cell 1 (Figures 7(d) and 7(e)). This reflects the similarity in the transcriptional regulation between tenocyte-like attachment cell and chondrocyte-like attachment cell 1. The M2 and M3 modules mainly included chondrocyte-like attachment cell 4, chondrocyte-like attachment cell 3, and chondrocyte-like attachment cell 2. The M2 and M3 modules mainly included chondrocyte-like attachment cell clusters 3 and 4 (Figures 7(d) and 7(e)). By contrast, chondrocyte-like attachment cell 2 did not exhibit module specificity. The three core TFs of the M1 module (i.e., *Foxo3*, *Mef2c*, and *Xbp1*) are tenocyte-like attachment cell. *Stat1* is the core TF of chondrocyte-like attachment cell 1, and *Tbx5* is the core TF of chondrocyte-like attachment cell 2. Intersection analysis of these key TFs with previously studied TFs revealed that TF *Xbp1* was again found to be a common key TF for HO bulk-RNA-seq, HO scRNA-seq, and the integrated scRNA data.

To explore the potential regulatory function of *Xbp1* in the onset of ectopic ossification, the Venn diagram was used to screen 160 genes that collectively serve as downstream regulatory targets of XBP1 in the integrated scRNA data and HO scRNA data (Figure 8(a)). These 160 genes were subjected to GO and KEGG enrichment analysis (Figure 8(b)). GO enrichment analysis included three modules, namely, biological process (BP), cell component (CC),

and molecular function (MF). The Golgi vesicle transport, protein localization to the ER, and ER to Golgi vesicle-mediated transport were enriched in BP; the Golgi-associated vesicle membrane, COPI-coated vesicle, and Golgi apparatus part were enriched in CC, and the glucocorticoid receptor binding, primary active transmembrane transporter activity, and P-P-bond-hydrolysis-driven transmembrane transporter activity were enriched in MF. The pathways enriched in the KEGG were the circadian rhythm, protein export, and protein processing in the ER. In summary, genes that were downstream regulatory targets of *Xbp1* were found to play important roles mainly in Golgi vesicle transport, ER protein processing, cellular communication, and transport. To visualize the interaction relationships of *Xbp1* downstream regulatory target genes, string sites were used to map the protein-protein interaction (PPI) network analysis of XBP1 downstream regulatory genes.

3.8. Ligand-Receptor and ATAC-seq Analyses Reveal That Xbp1 Upregulate the Notch Signaling Pathway by Promoting Jag1 Transcription. Cellular communication is an important pathway for inducing cell differentiation. We hypothesized that XBP1 may affect intercellular communication by regulating ligand transcription. To investigate the commonalities and differences in cellular signaling exchange for each cell subtype, the CellPhoneDB was used to infer intercellular communication. Based on the results of the CellPhoneDB analysis, five ligand genes (i.e., *Fgfr1*, *Grin2b*, *Icam1*, *Jag1*, and *Ncam1*) were identified as potential target genes of *Xbp1* (Figure 9(a)). The Sankey plot was used to demonstrate the potential receptor genes (including *AREG*, *BDNF*, *FGF23*, *FGF3*, *FGF5*, *FGF7*, *FGF8*, *FGF9*, and *FGFR2*) for the five ligand genes (i.e., *Fgfr1*, *Grin2b*, *Icam1*, *Jag1*, and *Ncam1*), *GDNF*, *IL16*, *KL*, *NCAM1*, *NOTCH2*, and *NOTCH3*. Accordingly, the XBP1 functional pattern was mapped (Figure 9(c)). This suggests that XBP1 mediates the regulation of tenocyte-like attachment cell, chondrocyte-like attachment cell 2, and chondrocyte-like attachment cell 1 phenotype by promoting the transcriptional regulation of these five ligand genes, namely, *Fgfr1*, *Grin2b*, *Icam1*, *Jag1*, and *Ncam1*, by receptor genes. The Pearson correlation analysis based on the GTEX database also revealed that XBP1 was significantly and positively correlated ($p < 0.05$) with the expression of *JAG1* (A), *NOTCH2* (B), and *NOTCH3* (C) (Supplementary Figure 9A-9C). Because *NOTCH2* and *NOTCH3* are key effectors of the Notch signaling pathway, we proposed that *Xbp1* may upregulate the Notch signaling pathway through *Jag1* transcription.

GSE144306 and GSE168153 are datasets of the same study, and GSE144306 is an ATAC-seq dataset belonging to the tendon coupling region. The different transcriptional open lengths of *Xbp1* and *Jag1* in the cartilaginous region of tendon fibers are shown in Figure 10(a). This suggests that *Xbp1* is common chromatin developed in different samples, while chromatin opening of *Jag1* is significantly heterogeneous in GSE144306. A global analysis of GSE144306 was also performed, and a heat map was drawn for the analysis of chromatin opening in different samples of the fibrocartilaginous region of the tendons (Figure 10(b)). As a result

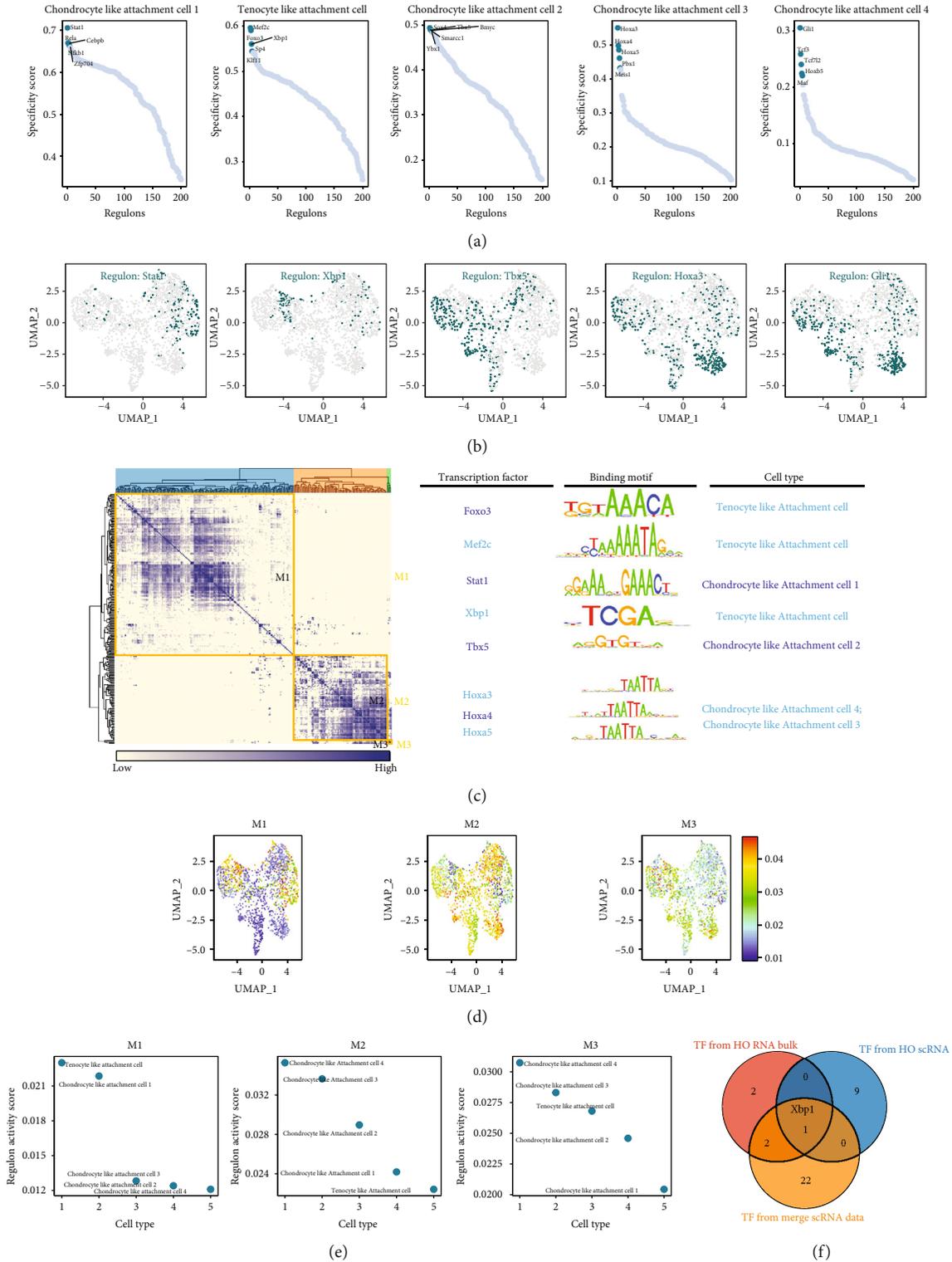


FIGURE 7: XBP1 was found to be a key cotranscriptional regulator in the fibrocartilaginous region of the tendons and ectopic ossified cells. (a) Analysis of transcription factor activities specific to different cell types in the tendon-bone union zone and ectopic ossified cells. The five most transcriptionally active transcription factors were screened for each group of cells. (b) Uniform manifold approximation and projection (UMAP) plots demonstrate the expression characteristics of the core transcription factors in each cell group. (c) Regulatory modules were identified based on the regulon CSI matrix, and the core transcription factors, binding motifs, and corresponding cell types were extracted. (d) The activity of each module in different cells is represented by UMAP plots. (e) Cell types corresponding to the different modules. (f) The Venn diagram reveals that Xbp1 is the common key transcription factor for heterotopic ossification (HO) bulk-RNA-seq, HO scRNA-seq, and the merge scRNA data.

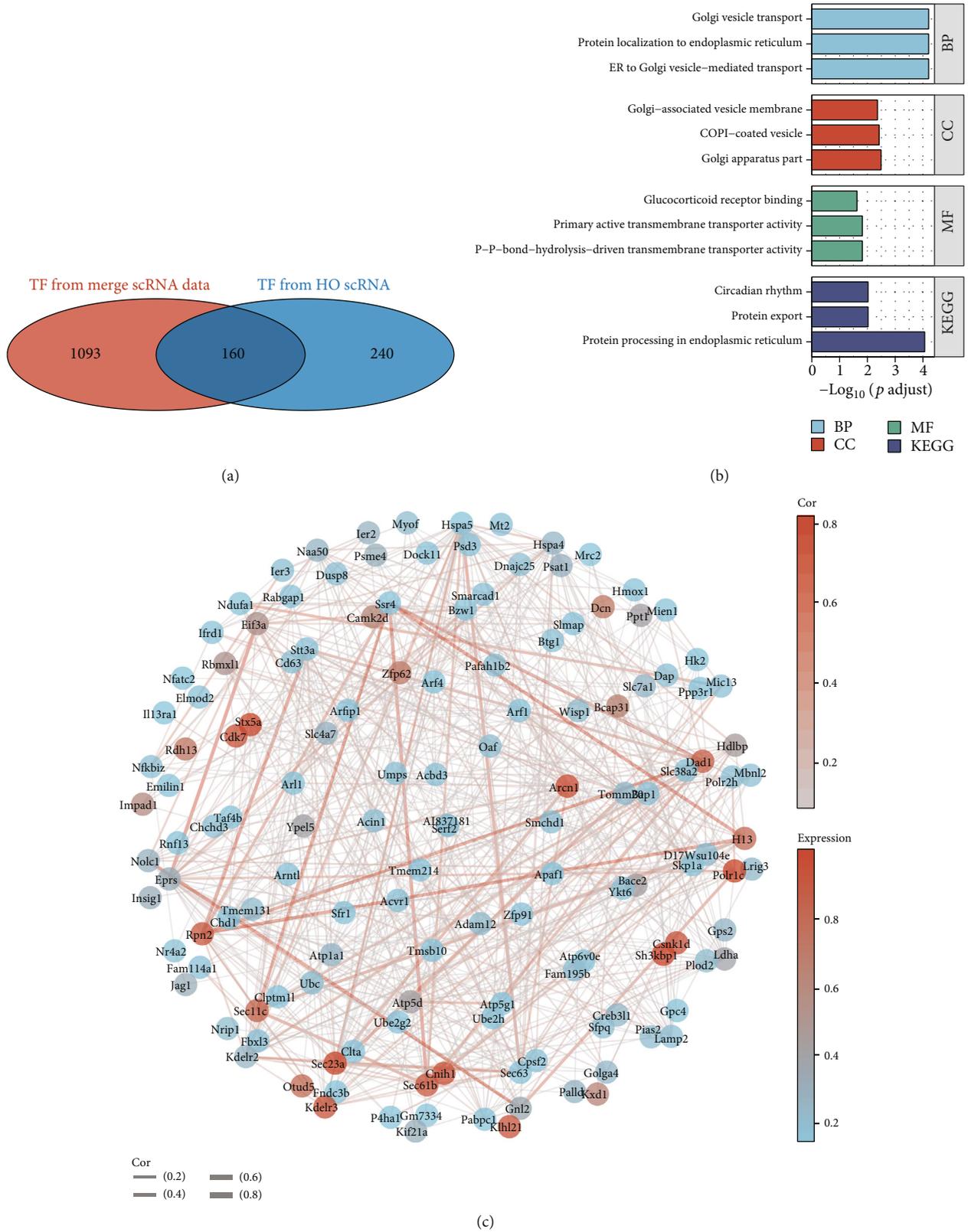


FIGURE 8: Enrichment analysis of downstream genes regulated by XBP1 with protein-protein interaction (PPI) network analysis. (a) Screening of 160 genes that were jointly used as XBP1 regulatory targets in the integrated scRNA data and heterotopic ossification (HO) scRNA data using the Venn diagram. (b) GO and KEGG enrichment analysis for XBP1-regulated genes. (c) PPI network analysis of XBP1-regulated downstream genes.

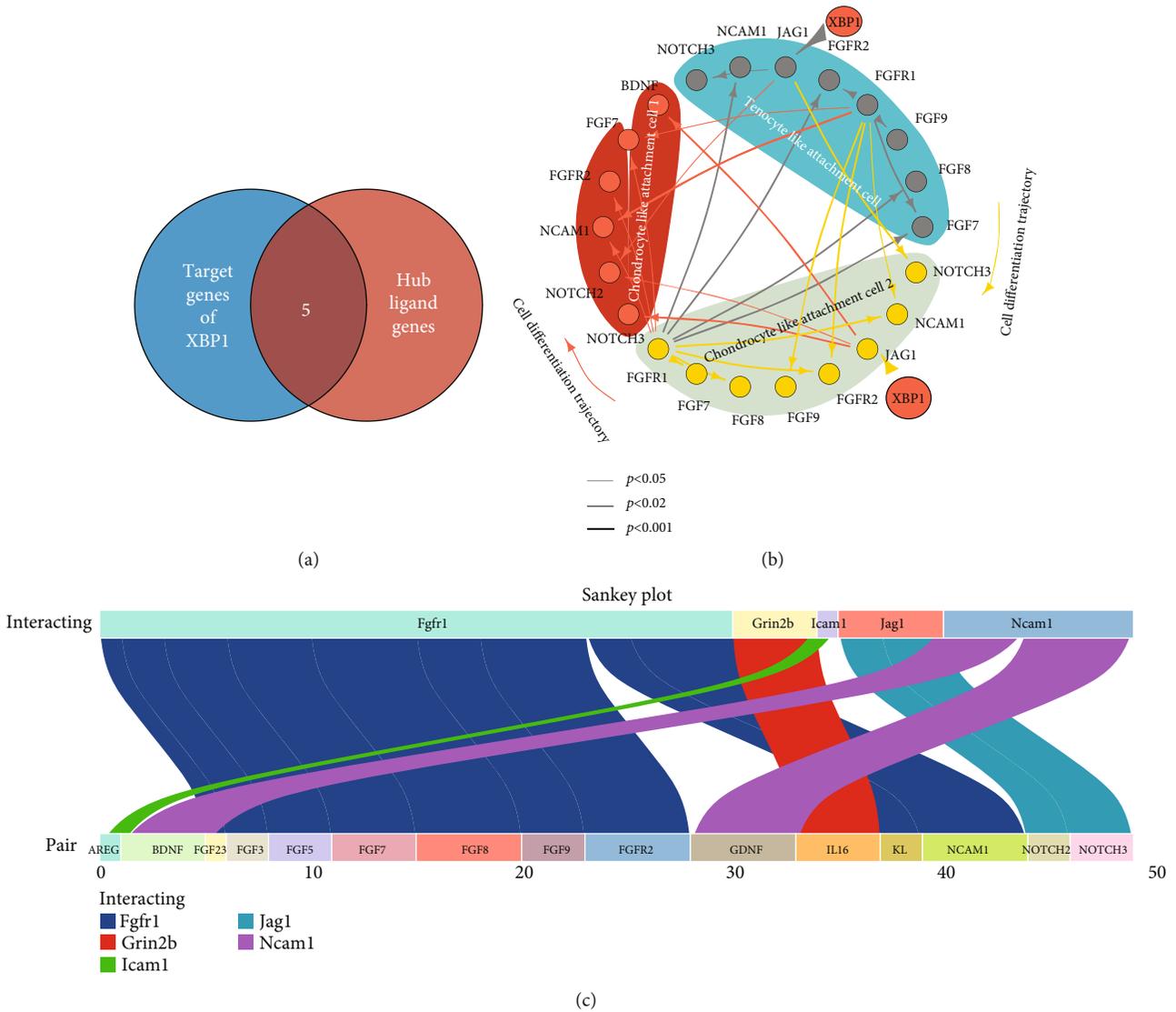
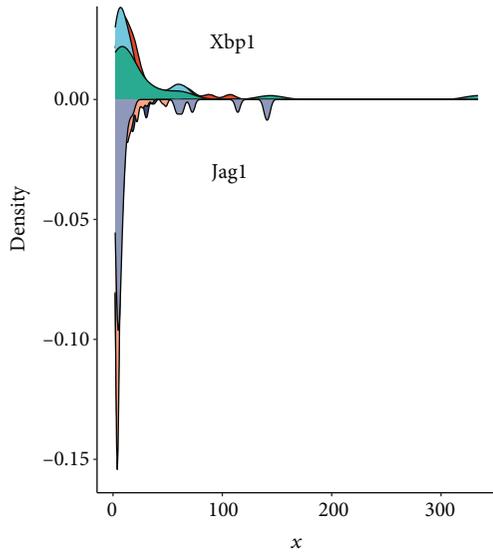


FIGURE 9: Ligand-receptor-based analysis revealed that Xbp1 promotes transcriptional regulation of this ligand gene-mediated receptor regulation of tenocyte-like attachment cell, chondrocyte-like attachment cell 2, and chondrocyte-like attachment cell 1 phenotypes. (a) Intersection analysis suggested that five ligand genes, namely, Fgfr1, Grin2b, Icam1, Jag1, and Ncam1, are potential target genes of Xbp1. (b) A Sankey plot of the potential receptor genes of these five ligand genes: Fgfr1, Grin2b, Icam1, Jag1, and Ncam1. (c) XBP1 mediates the regulation of receptor genes by promoting the transcriptional regulation of five ligand genes (namely, Fgfr1, Grin2b, Icam1, Jag1, and Ncam1) to regulate tenocyte-like attachment cell, chondrocyte-like attachment cell 2, and chondrocyte-like attachment cell 1 phenotypes.

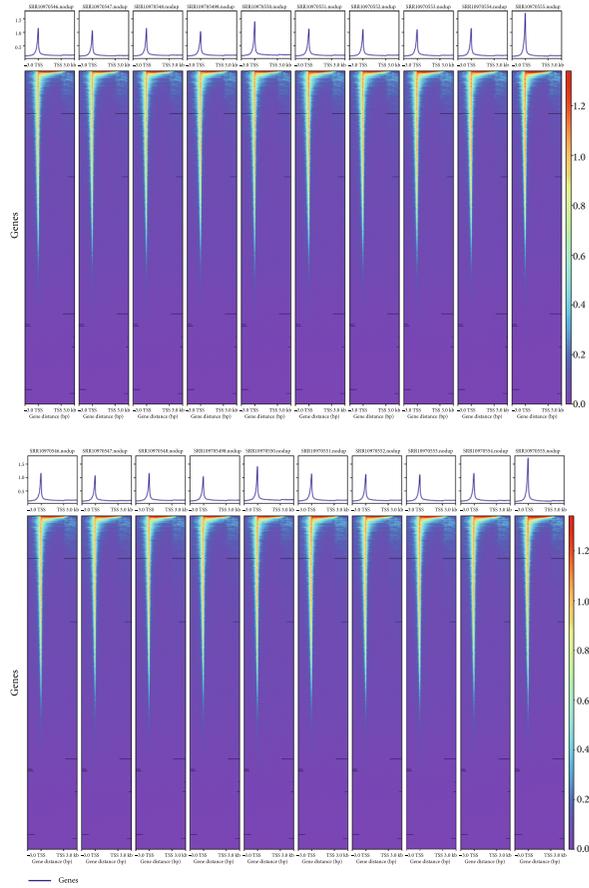
of the ATAC-seq analysis, the MEME Suite database predicted that the promoter region of Jag1 might be regulated by Xbp1 [58, 70]. The promoter of Jag1 was significantly more open in the chondrocyte region (Figure 10(d)). By contrast, Xbp1 did not show this phenomenon (Figure 10(e)). This suggests that the transcriptional activity of Jag1 is significantly upregulated in the chondrocyte region, which may be due to the promoter activation of Jag1. Taken together, we revealed that Xbp1 might upregulate the Notch signaling pathway by promoting Jag1 transcription based on the ligand-receptor analysis and ATAC-seq analysis.

3.9. Crosstalk in the Notch/Hh Signaling Pathway Is Associated with Xbp1 Expression. The GSVA based on GSE94683 revealed that upregulated activities of Hh and Notch signaling

pathway activities were upregulated in HO tissues (Supplementary Figure 9D, 9E). Interestingly, the activity of the Hh signaling pathway was more expressed in HO2 than in HO1, and the activity of the Hh signaling pathway was more expressed in HO1 than in HO2 (Figures 11(a)–11(c)). By contrast, the proposed temporal expression profile of the Hh and Notch signaling pathways suggests that the Hh signaling pathway may be activated before the Notch signaling pathway. The GSEA of XBP1 expression concerning the pathway activity in different datasets was also performed (Figures 11(e)–11(h)). The upregulations of Xbp1 in GSE94683, GSE102929, GSE168153, and GSE138515 were associated with the upregulation of the WNT, Notch, Hh, and TGF beta signaling pathways. This suggests that Xbp1 upregulation may induce the occurrence of ectopic



(a)



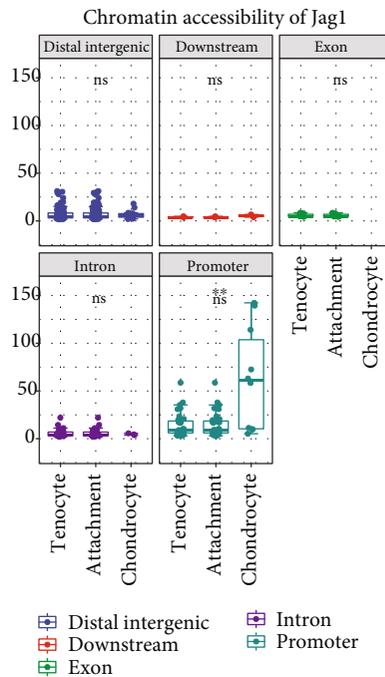
(b)

Database sequences. fa
 Database contains 1 sequence, 200 residues
 Motifs MA0844.1 meme (DNA)

Motifs	Width	Best possible match
MA0844.1	14	AATGCCACGTCATC

Random model letter frequencies (--nrdb--):
 A 0.275 C 0.225 G 0.225 T 0.275

(c)



(d)

FIGURE 10: Continued.

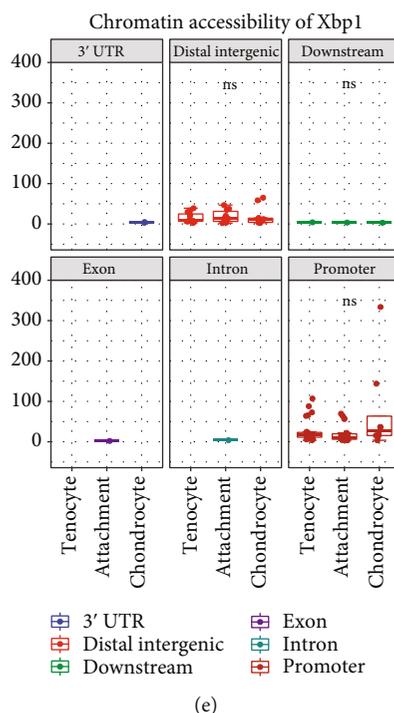


FIGURE 10: ATAC-seq-based analysis reveals that Xbp1 promotes Jag1 transcription. (a) Densitometric analysis of different transcriptional open lengths of Xbp1 and Jag1 in the tendon fibrocartilaginous region. (b) Heat map of chromatin opening analysis of different samples from the fibrocartilaginous region of the tendon. (c) Xbp1 binds to the promoter site of Jag1. (d) Chromatin accessibility of Jag1. (e) Chromatin accessibility of Xbp1.

ossification by upregulating the activities of WNT, Notch, Hh, and TGF beta signaling pathways.

3.10. Screening for Potential MicroRNAs That Induce Upregulation of XBP1 Expression after Acute Ischemic Stroke. Neurogenic heterotopic ossification is a common complication of acute ischemic stroke; however, its exact mechanism remains unknown. A total of 93 microRNAs have been identified as differentially expressed in acute ischemic stroke by both DEseq2 and EdgeR methods that screened the GSE110993 dataset using a Venn diagram (Figure 12(a)). In this study, the target XBP of miRNAs was predicted using the miRWalk database (<http://mirwalk.umm.uni-heidelberg.de/>). Based on the miRwalk dataset, we screened 24 microRNAs that could potentially target XBP1. These microRNAs included hsa-let-7d-3p, hsa-let-7d-5p, hsa-let-7i-5p, hsa-miR-103a-3p, hsa-miR-103b, hsa-miR-130a-3p, hsa-miR-130b-3p, hsa-miR-130b-5p, hsa-miR-15b-3p, hsa-miR-185-5p, hsa-miR-18a-3p, hsa-miR-18a-5p, hsa-miR-193a-5p, hsa-miR-3158-3p, hsa-miR-3184-5p, hsa-miR-320b, hsa-miR-3688-3p, hsa-miR-3688-5p, hsa-miR-378a-3p, hsa-miR-423-3p, hsa-miR-484, hsa-miR-502-3p, hsa-miR-92a-3p, and hsa-miR-942-5p (Figure 12(b)). Schematic depiction of this work was shown in Figure 12(c).

4. Discussion

In this study, we used a single-cell approach to integrate and analyze the developmental lineage of the fibrocartilaginous zone of tendons and the ectopic ossification cell model.

Ectopic ossified cells demonstrated similar transcriptional characteristics to cells in the fibrocartilaginous region of tendons. This study also identified Xbp1 as a common key TF for ectopically ossified cells and cells in the fibrocartilaginous zone of the tendons. To the best of our knowledge, this is the first study to propose that Xbp1 may upregulate the Notch signaling pathway by transcribing Jag1. These results also suggest a potential association of endoplasmic reticulum stress with tendon bone healing and HO.

This study was performed based on the integration analyses of scRNA-seq, bulk RNA-seq, and ATAC-seq data. The CCA in the Seurat package was used for dataset integration, whereby we found that ectopically ossified cells had a similar transcriptional profile to cells in the fibrocartilaginous region of the tendons. Based on the pseudo-temporal trajectory analysis of Monocle 3, this study revealed a potential model of PC differentiation in ectopically ossified tissues. To identify specific TFs associated with ectopic ossification, we used a modified version of the SCENIC method to construct a regulatory network between HO-related targets and TFs. The results showed that Xbp1 was a common key TF for ectopically ossified cells and cells in the fibrocartilaginous region of the tendons. We then performed cellular ligand-receptor analysis using the CellPhoneDB database and software according to the instruction manual, and after further pathway screening, Xbp1 was found to possibly upregulate the Notch signaling pathway by promoting Jag1 transcription. GSE94683 (including nine samples of healthy donors and seven samples of ectopically ossified tissues from patients with HO) was subjected to differential analysis. Five

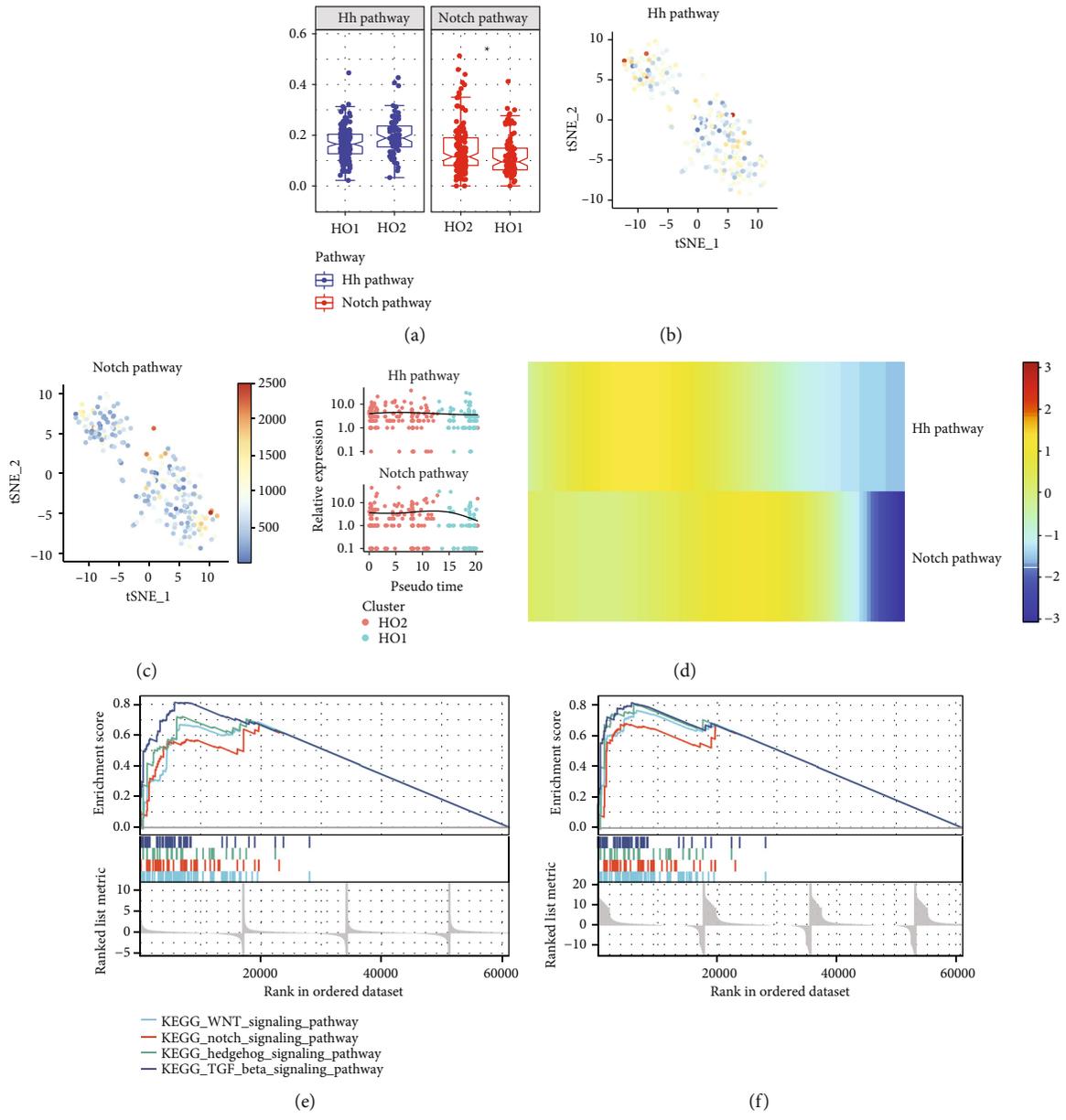


FIGURE 11: Continued.

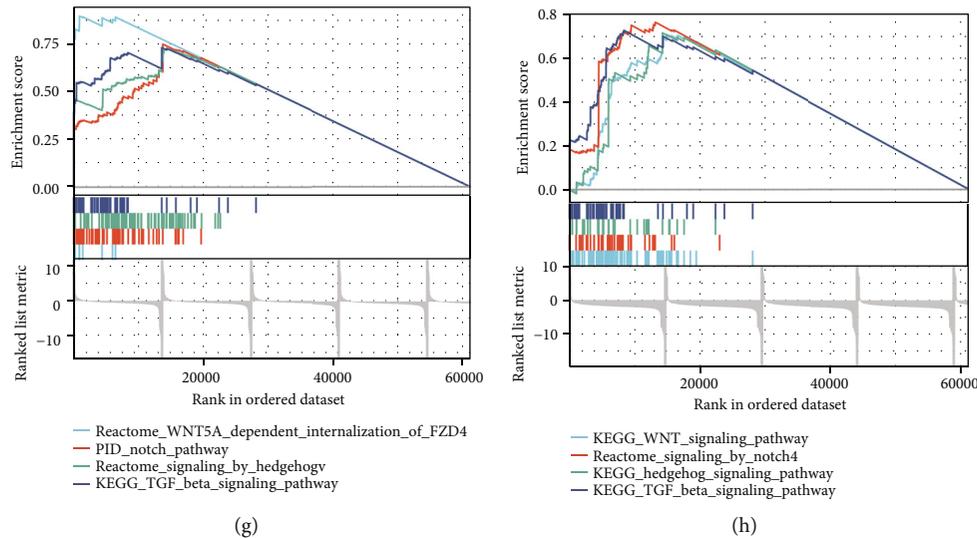


FIGURE 11: Crosstalk in the Notch/Hedgehog (Hh) signaling pathway is associated with Xbp1 expression in ectopic ossified cells. (a) Box line diagrams responding to the difference in the activities of the Hh and Notch signaling pathways in different tissues. (b) Hh signaling pathway activity in mouse tail tendon cells under simulated heterotopic ossification (HO) pathological conditions is indicated by different colors. (c) Notch signaling pathway activity in mouse tail tendon cells under simulated HO pathological conditions is indicated by different colors. (d) Proposed temporal expression characteristics of the Hh and Notch signaling pathways. (e–h) Gene Set Enrichment Analysis of XBP1 differences concerning pathway activity in different datasets: GSE94683 (e), GSE102929 (f), GSE168153 (g), and GSE138515 (h).

TFs (i.e., GLI1, EN1, XBP1, SHOX, and TBX5) and some KEGG pathways (i.e., WNT, Notch, Hh, and TGF beta signaling pathways) were significantly highly expressed in HO tissues.

We first classified pathological cells in the ectopic ossification model into HO1 and HO2 cell subpopulations. The proposed chronological analysis suggests that HO2 is a precursor cell of HO1. Mgp, Bgn, Sox9, Col11a1, Col1a1, Col1a2, Tnmd, Col11a2, Scx, Hivp2, Bcl2, Col2a1, Mxk, Myc, and Dcn were enriched in HO2 cells. Sox9 and Scx were key TFs in the migrating connective tissue between the tendon and bone [34, 71]. Diabetes was found to contribute to the development of tendinopathy through the downregulation of Scx, Mxk, Col1a1, Col1a2, and Bgn [72], and the upregulation of Col1a1, Scx, and Dcn contributed to the control of tendinopathy [73]. Tnmd was expressed at high levels in tendon cells and ligament cells and was found to have great potential in the treatment of tendon repair, which might be related to its coordinated expression of Col2a1 synergistically promoting angiogenesis [74–76]. Mxk controlled the differentiation of tendon and ligament cells, while Bcl2 was associated with the apoptosis and regeneration of tenocytes [77, 78]. Col5a1, Foxc1, Gli3, Gli2, Pecam1, Klf2, Klf4, Zbtb48, Ptch2, Smad7, Sufu, Igfbp5, Tcf, Fbn1, and Jag1 were mainly enriched in HO1 cells. Gli2/3, Tcf, Fbn1, and Col5a1 were molecular markers in cartilage tissues [79–81]. The upregulation of Klf2 and Klf4 was found to promote cartilage stability and resist osteoarticular oxidative stress conditions [82, 83]. Foxc1, Ptch2, and Sufu were found to be essential for the activation of the Indian Hh signaling pathway [84, 85]. Smad7 and Jag1 were found to affect cartilage development through the MAPK or Notch signaling pathway [86, 87]. Igfb5 expression was found to be highly concentrated in developing limb

muscle tissue and was highly correlated with osteogenesis [88]. The expression profiles of HO1 and HO2 cells were more similar to that of chondrocytes and tenocyte-like cells, respectively. Some information also implied the developmental potential of HO2 cells to become HO1 cells. For example, (1) Sox9, Col1a1, and Col2a1 have been shown to play important functions in cartilage development [89, 90]; (2) Mgp was found to be related to the onset of osteoarthritis during cartilage development [91]; (3) Bcl2 was a key protein of the PI3K-AKT signaling pathway, which was closely associated with chondrocyte proliferation [92]; and (4) COL5A1 polymorphism was associated with susceptibility to tendon disease [93]. Notably, *Gli3*, *Cli2*, *Pecam1*, and *Ptch2* were found to be the star genes of the Hh signaling pathway, and *Notch2* and *Jag1* are the star genes of the Notch signaling pathway, suggesting that the activity of the Notch and Hh signaling pathways may be upregulated in HO1 cells [94–97].

In this study, we performed an integrative analysis of single-cell transcriptome data of mouse Achilles tendon, single-cell transcriptome data of mouse tendon junction region, and single-cell transcriptome data of mouse tail tendon cells under simulated HO pathological conditions (including GSE168153, GSE138515, and GSE102929 datasets under simulated HO pathological conditions) was also performed in this study. In Figure 6(b), tenocyte-like attachment cells (*Matn4* and *Col2a1* are cell markers of this cell type) were found to have more HO2 cells, while more HO1 cells were found in chondrocyte-like attachment cell 1 (*Anxa2* and *Col14a1* are cell markers of this cell type). *Matn4* and *Col2a1* are cell markers of tenocyte-like attachment cells. *Col2a1* is a crossover gene located between tenocyte and chondrocyte differentiation [76, 90, 98, 99]. While *Matn4* expression was tightly regulated and served as a

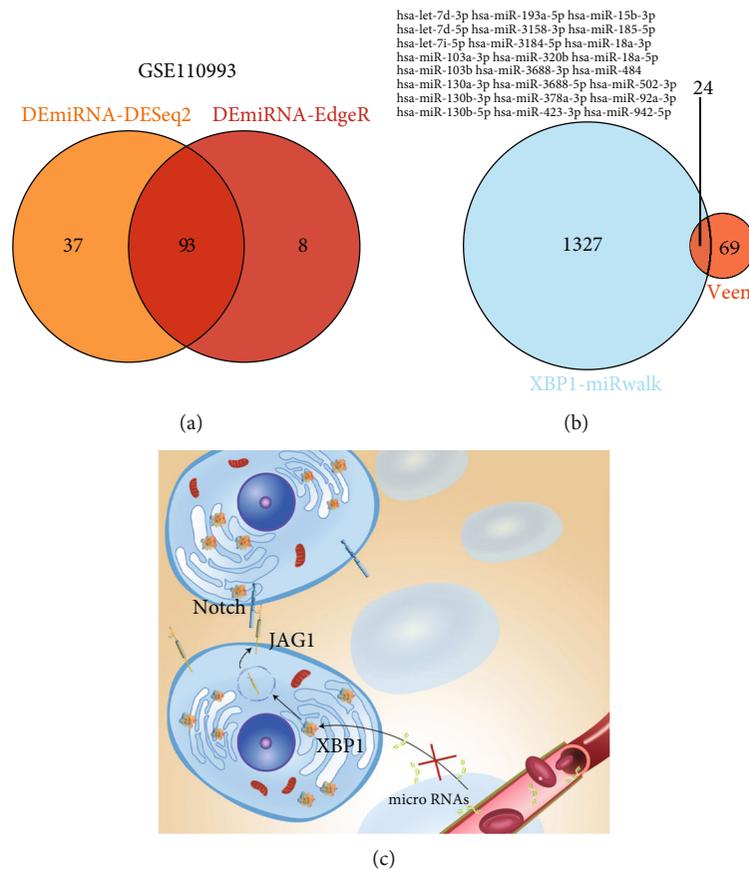


FIGURE 12: Screening for potential microRNAs causing XBP1 upregulation after acute ischemic stroke. (a) A total of 93 microRNAs were identified as differentially expressed in acute ischemic stroke by both DESeq2 and EdgeR methods that screened the GSE110993 dataset using a Venn diagram. (b) A total of 24 microRNAs that potentially target XBP1 were screened based on the miRwalk dataset. (c) Schematic depiction of this work.

marker of cell differentiation, it played a key role in maintaining the stability of articular cartilage [100]. *Anxa2* and *Col14a1* are cell markers of chondrocyte-like attachment cell 1. The upregulation of *Anxa2*, a biomarker of cartilage development, may promote tendon healing [101, 102]. The results showed that *Col14a1* was tendon-associated collagen and was associated with the development of osteoarthritis [103]. These data suggest that ectopically ossified cells have similar transcriptional and developmental characteristics to cells in the fibrocartilaginous zone.

In this study, we first identified the endoplasmic reticulum (ER) stress effector *Xbp1* as a common TF for HO bulk-RNA-seq, HO scRNA-seq, and the integrated scRNA data (including GSE168153, GSE138515, and GSE102929 datasets under simulated HO pathological conditions) using a comprehensive network analysis combined with a modified SCENIC approach. XBP1 was found to be an important regulatory pathway for the unfolding protein response that occurred in response to ER stress and played an important role in both physiological and pathological conditions, and its activity had a significant effect on the progression and prognosis of numerous diseases [104]. In this study, 160 genes were identified as core downstream regulatory targets of XBP1, with significant interactions among them, and they mainly played important roles in the Golgi vesicle transport,

ER protein processing, cellular communication, and translocation. Lei et al. found that mechanical stress can activate the ER stress and MAPK signaling pathways in the posterior longitudinal ligament, which played an important role in the onset of ossification of the posterior longitudinal ligament [105, 106]. The effect of ER stress on cartilage differentiation is regulated by the microenvironment [107]. Recent studies have shown that ER stress regulates the Notch and Hh signaling pathways [108, 109]. As a key effector protein of ER stress, *Xbp1* is positively correlated with the activity of the Notch signaling pathway. The upregulation of *Xbp1* was found to inhibit the Notch pathway effector protein *pcanex*, resulting in ER enlargement [110–112]. In addition, the deletion of XBP1 signaling was found to cause chondrodysplasia and delayed ossification [113]. In the present study, *Xbp1*, a key effector protein of ER stress, was found to be a key TF for the onset of ectopic ossification. These results also suggest a potential association between endoplasmic reticulum stress and tendon bone healing and HO.

Cellular communication is an important process for inducing cell differentiation. We hypothesized that XBP1 may affect intercellular communication by regulating the transcription of ligands. To investigate the similarities and differences in intercellular information exchange for each cell subtype, we used the CellPhoneDB to infer intercellular

communication and constructed a functional model of XBP1 regulation accordingly. We propose that Xbp1 may upregulate the Notch signaling pathway through Jag1 transcription. Based on the results of the chromatin opening analysis, the MEME Suite database predicted that the promoter region of Jag1 might be regulated by Xbp1. ER stress was found to upregulate Jag1 expression, while the relationship between Xbp1 and Jag1 has not been elucidated. To our knowledge, this study is the first to propose that Xbp1 may upregulate the Notch signaling pathway by promoting Jag1 transcription [114]. In GSE94683, GSE102929, GSE168153, and GSE138515, the upregulation of Xbp1 was found to be associated with the upregulation of WNT, Notch, Hh, and TGF beta signaling pathways, which may be due to the crosstalk of these pathways in ectopic ossified tissues. However, the specific regulatory mechanisms remain to be further investigated [7, 115, 116].

The prevalence of HO in stroke is 0.5%-1.2%, which has led to a lack of understanding of the epidemiology and pathogenesis of HO after ischemic stroke [117–119]. Furthermore, the occurrence of HO is dependent on the upregulation of endoplasmic reticulum stress pathways [105, 106, 120–122]. Meanwhile, the activation of endoplasmic reticulum stress-related pathways and the upregulation of XBP1 are key features of the widespread presence of various brain-related diseases, including stroke [123]. Therefore, the couplegulation of endoplasmic reticulum stress-related pathways in HO and ischemic stroke has attracted our attention. The current study found that the expression of microRNAs carried by exosomes in the blood is altered after ischemic stroke, which may affect the function of organs and tissues in the distant compartment [124–126]. We propose the hypothesis that changes in microRNA levels in the peripheral blood exosomes following ischemic stroke may lead to upregulation of stem cell XBP1 expression in muscle tissue and may thus induce neurogenic ectopic ossification. In this study, 24 microRNAs were screened and were found to be potentially associated with the upregulation of XBP1 expression after acute ischemic stroke. However, the effect of circulating blood exosomes on the occurrence of HO remains to be further investigated.

In this study, cell clusters of ectopic ossification phenotypes were isolated in a single-cell dataset of ectopic ossification. Three single-cell transcriptomes (namely, fibrocartilaginous zone single cell, Achilles tendon single-cell transcriptome, and ectopic ossification model single-cell transcriptome) were integrated into one dataset using the CCA method. Ectopically ossified cells were found to colocalize with the fibrocartilaginous zone cells, suggesting that ectopically ossified cells share similar transcriptome characteristics to fibrocartilaginous zone cells. The pseudo-temporal trajectory analysis further supported the above findings. Further TF coexpression network analysis revealed that Xbp1 is a hub TF for HO formation. The ligand-receptor analysis reveals potential pathways through which Xbp1 exerts its transcriptional regulatory role. The single-cell analysis revealed that the Xbp1 and Notch pathway expressions were upregulated in ectopic ossified tissues, and they were positively correlated with each other. Ultimately, we proposed

a potential mechanism by which Xbp1 may upregulate the Notch signaling pathway by promoting Jag1 transcription. In summary, this study integrated six datasets, namely, GSE94683, GSE144306, GSE168153, GSE138515, GSE102929, and GSE110993. The differentiation trajectory and key TFs of ectopic ossification occurrence were systematically analyzed by integrating scRNA-seq, bulk RNA-seq, and ATAC-seq data of the fibrocartilaginous region and HO.

This study revealed similarities between ectopically ossified cells and cells of the fibrocartilaginous zone of tendons and suggested a potential Xbp1/Jag1/Notch signaling pathway. However, a theoretical work with data from databases with no experimental verification may limit the application value of this research. Further functional experiments are still needed to be performed to explore the specific functional mechanisms of Xbp1. In this study, we obtained data from an integrated analysis of multiple databases and planned to conduct further analysis using ectopic ossified tissues and tendon fibrocartilaginous zone cells from the same model. In addition, the regulatory relationships between the Hh and Notch signaling pathways and other key pathways need to be further validated to confirm the activation patterns of the pathways in the single-cell transcriptome. We believe that further collection of clinical samples is also necessary, which will help in clinical diagnosis and translation.

5. Conclusion

A systematic analysis of the differentiation landscape and cellular homogeneity facilitates a molecular understanding of the potential homology between tendon fibrocartilage and ectopic ossified tissue. Furthermore, by identifying Xbp1 as a hub regulator and through ligand-receptor analysis, we propose a potential Xbp1/Jag1/Notch signaling pathway. The findings provide a novel concept for the formation of ectopically ossified tissues, i.e., HO cells may be differentiated from misplaced fibrocartilaginous zone stem cells.

Data Availability

The corresponding author agreed to provide the reader with the original data for a suitable reason.

Conflicts of Interest

The authors declare no conflict of interest.

Authors' Contributions

Yisheng Chen, Yaying Sun, Yuzhen Xu, Wei-Wei Lin, and Zhiwen Luo contributed equally to this work.

Acknowledgments

We are pleased to thank Shanghai qiaoyidao Biotechnology Co., Ltd. for their medical aesthetics service. We also thank Bullet Edits Limited for the linguistic editing and proofreading of the manuscript. This study was supported by a grant

from the National Natural Science Foundation of China (Nos. 81972062 and 81772419).

Supplementary Materials

All supplementary figures were included in the “Supplementary figures” file. (*Supplementary Materials*)

References

- [1] S. Font Tellado, E. R. Balmayor, and M. Van Griensven, “Strategies to engineer tendon/ligament-to-bone interface: biomaterials, cells and growth factors,” *Advanced Drug Delivery Reviews*, vol. 94, pp. 126–140, 2015.
- [2] D. Xu, T. Zhang, J. Qu, J. Hu, and H. Lu, “Enhanced Patella-Patellar tendon healing using combined magnetic fields in a rabbit model,” *The American Journal of Sports Medicine*, vol. 42, no. 10, pp. 2495–2501, 2014.
- [3] H. Lu, C. Chen, J. Qu et al., “Initiation timing of low-intensity pulsed ultrasound stimulation for tendon-bone healing in a rabbit model,” *The American Journal of Sports Medicine*, vol. 44, no. 10, pp. 2706–2715, 2016.
- [4] H. Lu, L. Qin, P. Fok et al., “Low-intensity pulsed ultrasound accelerates bone-tendon junction healing: a partial patellectomy model in rabbits,” *The American Journal of Sports Medicine*, vol. 34, no. 8, pp. 1287–1296, 2006.
- [5] H. H. Lu and S. Thomopoulos, “Functional attachment of soft tissues to bone: development, healing, and tissue engineering,” *Annual Review of Biomedical Engineering*, vol. 15, no. 1, pp. 201–226, 2013.
- [6] K. Atesok, F. H. Fu, M. R. Wolf et al., “Augmentation of Tendon-to-Bone healing,” *The Journal of Bone and Joint Surgery. American Volume*, vol. 96, no. 6, pp. 513–521, 2014.
- [7] G. Yang, B. B. Rothrauff, and R. S. Tuan, “Tendon and ligament regeneration and repair: clinical relevance and developmental paradigm,” *Birth Defects Research. Part C, Embryo Today*, vol. 99, no. 3, pp. 203–222, 2013.
- [8] S. Liu, Y. Sun, F. Wan, Z. Ding, S. Chen, and J. Chen, “Advantages of an attached semitendinosus tendon graft in anterior cruciate ligament reconstruction in a rabbit model,” *The American Journal of Sports Medicine*, vol. 46, no. 13, pp. 3227–3236, 2018.
- [9] B. M. Choi, H. J. Kwak, C. D. Jun et al., “Control of scarring in adult wounds using antisense transforming growth factor-beta 1 oligodeoxynucleotides,” *Immunology and Cell Biology*, vol. 74, no. 2, pp. 144–150, 1996.
- [10] P. L. Hays, S. Kawamura, X. Deng et al., “The role of macrophages in early healing of a tendon graft in a bone tunnel,” *The Journal of Bone and Joint Surgery. American Volume*, vol. 90, no. 3, pp. 565–579, 2008.
- [11] J. Hopkinson-Woolley, D. Hughes, S. Gordon, and P. Martin, “Macrophage recruitment during limb development and wound healing in the embryonic and foetal mouse,” *Journal of Cell Science*, vol. 107, part 5, pp. 1159–1167, 1994.
- [12] D. J. Whitby and M. W. J. Ferguson, “Immunohistochemical localization of growth factors in fetal wound healing,” *Developmental Biology*, vol. 147, no. 1, pp. 207–215, 1991.
- [13] C. Chen, F. Liu, Y. Tang et al., “Book-shaped acellular fibrocartilage scaffold with cell-loading capability and chondrogenic inducibility for tissue-engineered fibrocartilage and bone-tendon healing,” *ACS Applied Materials & Interfaces*, vol. 11, no. 3, pp. 2891–2907, 2019.
- [14] R. Xu, J. Hu, X. Zhou, and Y. Yang, “Heterotopic ossification: mechanistic insights and clinical challenges,” *Bone*, vol. 109, pp. 134–142, 2018.
- [15] K. L. Foley, N. Hebela, M. A. Keenan, and R. J. Pignolo, “Histopathology of periarticular non-hereditary heterotopic ossification,” *Bone*, vol. 109, pp. 65–70, 2018.
- [16] D. Magne and C. Bougault, “What understanding tendon cell differentiation can teach us about pathological tendon ossification,” *Histology and Histopathology*, vol. 30, no. 8, pp. 901–910, 2015.
- [17] I. Miranda, E. Sánchez-Alepuz, J. V. Díaz-Martínez, A. Collado-Sánchez, and I. Peregrín-Nevado, “Calcific tendinopathy of the shoulder with intraosseous extension: experience with arthroscopic treatment and review of the literature,” *Revista Espanola de Cirugia Ortopedica y Traumatologia*, vol. 64, no. 1, pp. 13–21, 2020.
- [18] C. C. Wong, Y. Y. Yeh, T. L. Yang, Y. H. Tsuang, and C. H. Chen, “Augmentation of tendon graft-bone tunnel interface healing by use of bioactive platelet-rich fibrin scaffolds,” *The American Journal of Sports Medicine*, vol. 48, no. 6, pp. 1379–1388, 2020.
- [19] D. Dey, J. Bagarova, S. J. Hatsell et al., “Two tissue-resident progenitor lineages drive distinct phenotypes of heterotopic ossification,” *Science translational medicine*, vol. 8, no. 366, article 366ra163, 2016.
- [20] J. B. Lees-Shepard, S. E. Nicholas, S. J. Stoessel et al., “Palovarotene reduces heterotopic ossification in juvenile FOP mice but exhibits pronounced skeletal toxicity,” *eLife*, vol. 7, article e40814, 2018.
- [21] R. Nishimura, K. Hata, Y. Takahata et al., “Role of signal transduction pathways and transcription factors in cartilage and joint diseases,” *International Journal of Molecular Sciences*, vol. 21, no. 4, p. 1340, 2020.
- [22] K. Matsuo, R. D. Chavez, E. Barruet, and E. C. Hsiao, “Inflammation in fibrodysplasia ossificans progressiva and other forms of heterotopic ossification,” *Current Osteoporosis Reports*, vol. 17, no. 6, pp. 387–394, 2019.
- [23] M. Benjamin and J. R. Ralphs, “Biology of fibrocartilage cells,” *International review of cytology*, vol. 233, pp. 1–46, 2004.
- [24] Y. F. Rui, P. P. Lui, G. Li, S. C. Fu, Y. W. Lee, and K. M. Chan, “Isolation and characterization of multipotent rat tendon-derived stem cells,” *Tissue Engineering Part A*, vol. 16, no. 5, pp. 1549–1558, 2010.
- [25] J. Zhang and J. H. Wang, “Characterization of differential properties of rabbit tendon stem cells and tenocytes,” *BMC Musculoskeletal Disorders*, vol. 11, p. 10, 2010.
- [26] X. Qu, Z. Chen, D. Fan et al., “Notch signaling pathways in human thoracic ossification of the ligamentum flavum,” *Journal of Orthopaedic Research*, vol. 34, no. 8, pp. 1481–1491, 2016.
- [27] H. Liu, J. Xu, and R. Jiang, “Mkx-deficient mice exhibit hedgehog signaling-dependent ectopic ossification in the Achilles tendons,” *Journal of Bone and Mineral Research*, vol. 34, no. 3, pp. 557–569, 2019.
- [28] C. Kan, N. Ding, J. Yang et al., “BMP-dependent, injury-induced stem cell niche as a mechanism of heterotopic ossification,” *Stem Cell Research & Therapy*, vol. 10, no. 1, p. 14, 2019.

- [29] R. Gao, C. Shi, C. Yang, Y. Zhao, X. Chen, and X. Zhou, "Cyclic stretch promotes the ossification of ligamentum flavum by modulating the Indian hedgehog signaling pathway," *Molecular Medicine Reports*, vol. 22, no. 2, pp. 1119–1128, 2020.
- [30] E. Zelzer, E. Blitz, M. L. Killian, and S. Thomopoulos, "Tendon-to-bone attachment: from development to maturity," *Birth Defects Research. Part C, Embryo Today*, vol. 102, no. 1, pp. 101–112, 2014.
- [31] Q. Zhao, K. Zhuang, K. Han et al., "Silencing DVL3 defeats MTX resistance and attenuates stemness via Notch signaling pathway in colorectal cancer," *Pathology, Research and Practice*, vol. 216, no. 8, p. 152964, 2020.
- [32] S. Abe and M. Yamamoto, "Factors involved in morphogenesis in the muscle-tendon-bone complex," *International Journal of Molecular Sciences*, vol. 22, no. 12, p. 6365, 2021.
- [33] H. Asahara, M. Inui, and M. K. Lotz, "Tendons and ligaments: connecting developmental biology to musculoskeletal disease pathogenesis," *Journal of Bone and Mineral Research*, vol. 32, no. 9, pp. 1773–1782, 2017.
- [34] J. Sahlman, R. Inkinen, T. Hirvonen et al., "Premature vertebral endplate ossification and mild disc degeneration in mice after inactivation of one allele belonging to the Col2a1 gene for type II collagen," *Spine (Phila Pa 1976)*, vol. 26, no. 23, pp. 2558–2565, 2001.
- [35] K. N. Evans, B. K. Potter, T. S. Brown, T. A. Davis, E. A. Elster, and J. A. Forsberg, "Osteogenic gene expression correlates with development of heterotopic ossification in war wounds," *Clinical Orthopaedics and Related Research*, vol. 472, no. 2, pp. 396–404, 2014.
- [36] S. Picelli, "Single-cell RNA-sequencing: the future of genome biology is now," *RNA Biology*, vol. 14, no. 5, pp. 637–650, 2017.
- [37] E. A. Rondini and J. G. Granneman, "Single cell approaches to address adipose tissue stromal cell heterogeneity," *The Biochemical Journal*, vol. 477, no. 3, pp. 583–600, 2020.
- [38] A. M. Ranzoni and A. Cvejic, "Single-cell biology: resolving biological complexity, one cell at a time," *Development*, vol. 145, no. 13, article dev163972, 2018.
- [39] X. Kang, Y. Chen, B. Yi et al., "An integrative microenvironment approach for laryngeal carcinoma: the role of immune/methylation/autophagy signatures on disease clinical prognosis and single-cell genotypes," *Journal of Cancer*, vol. 12, no. 14, pp. 4148–4171, 2021.
- [40] W. Chen, Y. Zhao, X. Chen et al., "A multicenter study benchmarking single-cell RNA sequencing technologies using reference samples," *Nature Biotechnology*, vol. 39, 2020.
- [41] L. Wen and F. Tang, "Single-cell sequencing in stem cell biology," *Genome Biology*, vol. 17, no. 1, p. 71, 2016.
- [42] G. Martello and A. Smith, "The nature of embryonic stem cells," *Annual Review of Cell and Developmental Biology*, vol. 30, no. 1, pp. 647–675, 2014.
- [43] L. Laine, T. Kaltenbach, A. Barkun et al., "SCENIC international consensus statement on surveillance and management of dysplasia in inflammatory bowel disease," *Gastroenterology*, vol. 148, no. 3, pp. 639–651.e28, 2015.
- [44] D. Mercatelli, L. Scalambra, L. Triboli, F. Ray, and F. M. Giorgi, "Gene regulatory network inference resources: a practical overview," *Biochimica et Biophysica Acta, Gene Regulatory Mechanisms*, vol. 2020, no. 6, article 194430, 2020.
- [45] M. F. Griffin, H. E. desJardins-Park, S. Mascharak, M. R. Borrelli, and M. T. Longaker, "Understanding the impact of fibroblast heterogeneity on skin fibrosis," *Disease Models & Mechanisms*, vol. 13, no. 6, article dmm044164, 2020.
- [46] S. Toda, N. W. Frankel, and W. A. Lim, "Engineering cell-cell communication networks: programming multicellular behaviors," *Current Opinion in Chemical Biology*, vol. 52, pp. 31–38, 2019.
- [47] E. Canalis, "Notch in skeletal physiology and disease," *Osteoporosis International*, vol. 29, no. 12, pp. 2611–2621, 2018.
- [48] L. M. Francisco, P. T. Sage, and A. H. Sharpe, "The PD-1 pathway in tolerance and autoimmunity," *Immunological Reviews*, vol. 236, no. 1, pp. 219–242, 2010.
- [49] H. Han, J. W. Cho, S. Lee et al., "TRRUST v2: an expanded reference database of human and mouse transcriptional regulatory interactions," *Nucleic Acids Research*, vol. 46, no. D1, pp. D380–D386, 2018.
- [50] A. Butler, P. Hoffman, P. Smibert, E. Papalexi, and R. Satija, "Integrating single-cell transcriptomic data across different conditions, technologies, and species," *Nature Biotechnology*, vol. 36, no. 5, pp. 411–420, 2018.
- [51] X. Zhang, Y. Lan, J. Xu et al., "CellMarker: a manually curated resource of cell markers in human and mouse," *Nucleic Acids Research*, vol. 47, no. D1, pp. D721–D728, 2019.
- [52] D. Aran, A. P. Looney, L. Liu et al., "Reference-based analysis of lung single-cell sequencing reveals a transitional profibrotic macrophage," *Nature Immunology*, vol. 20, no. 2, pp. 163–172, 2019.
- [53] X. Qiu, Q. Mao, Y. Tang et al., "Reversed graph embedding resolves complex single-cell trajectories," *Nature Methods*, vol. 14, no. 10, pp. 979–982, 2017.
- [54] M. Efremova, M. Vento-Tormo, S. A. Teichmann, and R. Vento-Tormo, "CellPhoneDB: inferring cell-cell communication from combined expression of multi-subunit ligand-receptor complexes," *Nature Protocols*, vol. 15, no. 4, pp. 1484–1506, 2020.
- [55] M. W. E. J. Fiers, L. Minnoye, S. Aibar, C. Bravo González-Blas, Z. Kalender Atak, and S. Aerts, "Mapping gene regulatory networks from single-cell omics data," *Briefings in Functional Genomics*, vol. 17, no. 4, pp. 246–254, 2018.
- [56] S. Aibar, C. B. González-Blas, T. Moerman et al., "SCENIC: single-cell regulatory network inference and clustering," *Nature Methods*, vol. 14, no. 11, pp. 1083–1086, 2017.
- [57] S. Suo, Q. Zhu, A. Saadatpour, L. Fei, G. Guo, and G. C. Yuan, "Revealing the critical regulators of cell identity in the mouse cell atlas," *Cell reports*, vol. 25, no. 6, pp. 1436–1445.e3, 2018.
- [58] T. L. Bailey, M. Boden, F. A. Buske et al., "MEME SUITE: tools for motif discovery and searching," *Nucleic acids research*, vol. 37, no. Web Server issue, pp. W202–W208, 2009.
- [59] G. Yu, L. G. Wang, Y. Han, and Q. Y. He, "clusterProfiler: an R package for comparing biological themes among gene clusters," *Omics: a journal of integrative biology*, vol. 16, no. 5, pp. 284–287, 2012.
- [60] A. Subramanian, P. Tamayo, V. K. Mootha et al., "Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles," *Proceedings of the National Academy of Sciences*, vol. 102, no. 43, pp. 15545–15550, 2005.
- [61] S. Hänzelmann, R. Castelo, and J. Guinney, "GSVA: gene set variation analysis for microarray and RNA-seq data," *BMC Bioinformatics*, vol. 14, no. 1, p. 7, 2013.

- [62] W. Zhang, X. Chao, J. Q. Wu et al., "Exploring the potential mechanism of Guchang Zhixie Wan for treating ulcerative colitis by comprehensive network pharmacological approaches and molecular docking validation as well as cell experiments," *Chemistry & Biodiversity*, vol. 18, no. 1, article e2000810, 2021.
- [63] Y. Wang, Z. J. Zhao, X. R. Kang et al., "lnc RNA DLEU2 acts as a miR-181a sponge to regulate SEPP1 and inhibit skeletal muscle differentiation and regeneration," *Aging (Albany NY)*, vol. 12, no. 23, pp. 24033–24056, 2020.
- [64] K. Xu, Z. Zhang, M. Chen et al., "Nesfatin-1 promotes the osteogenic differentiation of tendon-derived stem cells and the pathogenesis of heterotopic ossification in rat tendons via the mTOR pathway," *Frontiers in Cell and Development Biology*, vol. 8, p. 547342, 2020.
- [65] H. Suzuki, Y. Ito, M. Shinohara et al., "Gene targeting of the transcription factor Mohawk in rats causes heterotopic ossification of Achilles tendon via failed tenogenesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 113, no. 28, pp. 7840–7845, 2016.
- [66] S. Kult, T. Olender, M. Osterwalder et al., "Bi-fated tendon-to-bone attachment cells are regulated by shared enhancers and KLF transcription factors," *eLife*, vol. 10, article e55361, 2021.
- [67] H. Feng, W. Xing, Y. Han et al., "Tendon-derived cathepsin K-expressing progenitor cells activate hedgehog signaling to drive heterotopic ossification," *The Journal of Clinical Investigation*, vol. 130, no. 12, pp. 6354–6365, 2020.
- [68] B. Shiroud Heidari, R. Ruan, E. M. De-Juan-Pardo, M. Zheng, and B. Doyle, "Biofabrication and signaling strategies for tendon/ligament interfacial tissue engineering," *ACS Biomaterials Science & Engineering*, vol. 7, no. 2, pp. 383–399, 2021.
- [69] A. J. De Micheli, J. B. Swanson, N. P. Disser et al., "Single-cell transcriptomic analysis identifies extensive heterogeneity in the cellular composition of mouse Achilles tendons," *American Journal of Physiology. Cell Physiology*, vol. 319, no. 5, pp. C885–C894, 2020.
- [70] T. L. Bailey, "STREME: accurate and versatile sequence motif discovery," *Bioinformatics*, vol. 37, article btab203, 2021.
- [71] H. Mutsuzaki and H. Nakajima, "Development of fibrocartilage layers in Achilles tendon entheses in rabbits," *Journal of Rural Medicine*, vol. 16, no. 3, pp. 160–164, 2021.
- [72] Y. F. Wu, H. K. Wang, H. W. Chang, J. Sun, J. S. Sun, and Y. H. Chao, "High glucose alters tendon homeostasis through downregulation of the AMPK/Egr1 pathway," *Scientific Reports*, vol. 7, no. 1, p. 44199, 2017.
- [73] C. C. Lin, P. T. Wu, C. W. Chang et al., "A single-pulsed electromagnetic field enhances collagen synthesis in tendon cells," *Medical Engineering & Physics*, vol. 77, pp. 130–136, 2020.
- [74] V. P. Alexandrov and S. I. Naimov, "A prospectus of tenomodulin," *Folia medica*, vol. 58, no. 1, pp. 19–27, 2016.
- [75] K. Yukata, Y. Matsui, C. Shukunami et al., "Differential expression of tenomodulin and chondromodulin-1 at the insertion site of the tendon reflects a phenotypic transition of the resident cells," *Tissue & Cell*, vol. 42, no. 2, pp. 116–120, 2010.
- [76] C. Shukunami, Y. Yoshimoto, A. Takimoto, H. Yamashita, and Y. Hiraki, "Molecular characterization and function of tenomodulin, a marker of tendons and ligaments that integrate musculoskeletal components," *Japanese Dental Science Review*, vol. 52, no. 4, pp. 84–92, 2016.
- [77] R. Nakamichi, K. Kataoka, and H. Asahara, "Essential role of Mohawk for tenogenic tissue homeostasis including spinal disc and periodontal ligament," *Modern Rheumatology*, vol. 28, no. 6, pp. 933–940, 2018.
- [78] Y. F. Wu, Y. L. Zhou, W. F. Mao, B. Avanesian, P. Y. Liu, and J. B. Tang, "Cellular apoptosis and proliferation in the middle and late intrasynovial tendon healing periods," *The Journal of Hand Surgery*, vol. 37, no. 2, pp. 209–216, 2012.
- [79] M. Wuelling and A. Vortkamp, "Chondrocyte proliferation and differentiation," *Endocrine Development*, vol. 21, pp. 1–11, 2011.
- [80] R. Nishimura, K. Hata, and J. Kida, "Regulation of osteoblasts and chondrocytes by Wnt signaling," *Clinical calcium*, vol. 29, no. 3, pp. 299–307, 2019.
- [81] D. R. Keene, C. D. Jordan, D. P. Reinhardt et al., "Fibrillin-1 in human cartilage: developmental expression and formation of special banded fibers," *The Journal of Histochemistry and Cytochemistry*, vol. 45, no. 8, pp. 1069–1082, 1997.
- [82] X. Gao, S. Jiang, Z. Du, A. Ke, Q. Liang, and X. Li, "KLF2 protects against osteoarthritis by repressing oxidative response through activation of Nrf2/ARE signaling in vitro and in vivo," *Oxidative Medicine and Cellular Longevity*, vol. 2019, 8564618 pages, 2019.
- [83] S. Gurusinghe, N. Bandara, B. Hilbert, G. Trope, L. Wang, and P. Strappe, "Lentiviral vector expression of Klf4 enhances chondrogenesis and reduces hypertrophy in equine chondrocytes," *Gene*, vol. 680, pp. 9–19, 2019.
- [84] M. Yoshida, K. Hata, R. Takashima et al., "The transcription factor Foxc1 is necessary for Ihh-Gli2-regulated endochondral ossification," *Nature Communications*, vol. 6, no. 1, p. 6653, 2015.
- [85] J. A. Caparrós-Martín, M. Valencia, E. Reytor et al., "The ciliary Evc/Evc2 complex interacts with Smo and controls hedgehog pathway activity in chondrocytes by regulating Sufu/Gli3 dissociation and Gli3 trafficking in primary cilia," *Human Molecular Genetics*, vol. 22, no. 1, pp. 124–139, 2013.
- [86] T. Iwai, J. Murai, H. Yoshikawa, and N. Tsumaki, "Smad7 Inhibits Chondrocyte Differentiation at Multiple Steps during Endochondral Bone Formation and Down-regulates p38 MAPK Pathways," *The Journal of Biological Chemistry*, vol. 283, no. 40, pp. 27154–27164, 2008.
- [87] H. J. Cheng, W. T. Hsu, C. N. Chen, and C. Li, "Activation of NOTCH1 by shear force elicits immediate cytokine expression in human chondrocytes," *International Journal of Molecular Sciences*, vol. 21, no. 14, p. 4958, 2020.
- [88] K. McQueeney and C. N. Dealy, "Roles of insulin-like growth factor-I (IGF-I) and IGF-I binding protein-2 (IGFBP2) and -5 (IGFBP5) in developing chick limbs," *Growth Hormone & IGF Research*, vol. 11, no. 6, pp. 346–363, 2001.
- [89] V. Lefebvre and M. Dvir-Ginzberg, "SOX9 and the many facets of its regulation in the chondrocyte lineage," *Connective Tissue Research*, vol. 58, no. 1, pp. 2–14, 2017.
- [90] S. Chen, Z. Xu, J. Shao, P. Fu, and H. Wu, "MicroRNA-218 promotes early chondrogenesis of mesenchymal stem cells and inhibits later chondrocyte maturation," *BMC Biotechnology*, vol. 19, no. 1, p. 6, 2019.
- [91] M. Neidlin, A. Korcari, G. Macheras, and L. G. Alexopoulos, "Cue-signal-response analysis in 3D chondrocyte scaffolds with anabolic stimuli," *Annals of Biomedical Engineering*, vol. 46, no. 2, pp. 345–353, 2018.

- [92] J. Li, G. Yang, S. Liu, L. Wang, Z. Liang, and H. Zhang, "Suv39h1 promotes facet joint chondrocyte proliferation by targeting miR-15a/Bcl2 in idiopathic scoliosis patients," *Clinical Epigenetics*, vol. 11, no. 1, p. 107, 2019.
- [93] J. Brazier, M. Antrobus, G. K. Stebbings et al., "Tendon and ligament injuries in elite rugby: the potential genetic influence," *Sports*, vol. 7, no. 6, p. 138, 2019.
- [94] S. Ohba, "Hedgehog signaling in skeletal development: roles of Indian hedgehog and the mode of its action," *International Journal of Molecular Sciences*, vol. 21, no. 18, p. 6665, 2020.
- [95] A. M. Skoda, D. Simovic, V. Karin, V. Kardum, S. Vranic, and L. Serman, "The role of the hedgehog signaling pathway in cancer: a comprehensive review," *Bosnian Journal of Basic Medical Sciences*, vol. 18, no. 1, pp. 8–20, 2018.
- [96] A. L. Penton, L. D. Leonard, and N. B. Spinner, "Notch signaling in human development and disease," *Seminars in cell & developmental biology*, vol. 23, no. 4, pp. 450–457, 2012.
- [97] S. Zanotti and E. Canalis, "Notch signaling in skeletal health and disease," *European Journal of Endocrinology*, vol. 168, no. 6, pp. R95–103, 2013.
- [98] A. M. McDermott, E. A. Eastburn, D. J. Kelly, and J. D. Boerckel, "Effects of chondrogenic priming duration on mechanoregulation of engineered cartilage," *Journal of Biomechanics*, vol. 125, p. 110580, 2021.
- [99] H. Zhao, W. Zhu, W. Mao, and C. Shen, "Platelet-rich plasma inhibits adriamycin-induced inflammation via blocking the NF- κ B pathway in articular chondrocytes," *Molecular Medicine*, vol. 27, no. 1, p. 66, 2021.
- [100] P. Li, L. Fleischhauer, C. Nicolae et al., "Mice lacking the matrilin family of extracellular matrix proteins develop mild skeletal abnormalities and are susceptible to age-associated osteoarthritis," *International Journal of Molecular Sciences*, vol. 21, no. 2, p. 666, 2020.
- [101] S. Nuerai, J. Ainuer, J. Jiasharete et al., "Kazakh therapy on differential protein expression of Achilles tendon healing in a 7-day postoperative rabbit model," *Journal of Traditional Chinese Medicine*, vol. 31, no. 4, pp. 367–375, 2011.
- [102] T. Kirsch, B. Swoboda, and H. Nah, "Activation of annexin II and V expression, terminal differentiation, mineralization and apoptosis in human osteoarthritic cartilage," *Osteoarthritis and Cartilage*, vol. 8, no. 4, pp. 294–302, 2000.
- [103] C. Karlsson, T. Dehne, A. Lindahl et al., "Genome-wide expression profiling reveals new candidate genes associated with osteoarthritis," *Osteoarthritis and Cartilage*, vol. 18, no. 4, pp. 581–592, 2010.
- [104] W. Xu, C. Wang, and J. Hua, "X-box binding protein 1 (XBP1) function in diseases," *Cell Biology International*, vol. 45, no. 4, pp. 731–739, 2021.
- [105] L. Shi, J. Miao, D. Chen, J. Shi, and Y. Chen, "Endoplasmic reticulum stress regulates mechanical stress-induced ossification of posterior longitudinal ligament," *European Spine Journal*, vol. 28, no. 10, pp. 2249–2256, 2019.
- [106] Y. Chen, H. Yang, J. Miao, X. Liu, X. Wang, and D. Chen, "Roles of the endoplasmic reticulum stress transducer OASIS in ossification of the posterior longitudinal ligament," *Clinical spine surgery*, vol. 30, no. 1, pp. E19–E24, 2017.
- [107] X. Yuan, H. Liu, L. Li et al., "The roles of endoplasmic reticulum stress in the pathophysiological development of cartilage and chondrocytes," *Current Pharmaceutical Design*, vol. 23, no. 11, pp. 1693–1704, 2017.
- [108] L. Zhao, H. Cai, Z. Tang, Q. Cui, Z. Liu, and S. Lu, "Matrine suppresses advanced glycation end products-induced human coronary smooth muscle cells phenotype conversion by regulating endoplasmic reticulum stress-dependent Notch signaling," *European Journal of Pharmacology*, vol. 882, article 173257, 2020.
- [109] V. Sahinturk, S. Kacar, E. Sahin, and N. E. B. Aykanat, "Investigation of endoplasmic reticulum stress and sonic hedgehog pathway in diabetic liver injury in mice," *Life Sciences*, vol. 246, article 117416, 2020.
- [110] T. Yamakawa, K. Yamada, T. Sasamura et al., "Deficient Notch signaling associated with neurogenic pecanex is compensated for by the unfolded protein response in *Drosophila*," *Development*, vol. 139, no. 3, pp. 558–567, 2012.
- [111] M. Qiu, W. Bao, J. Wang et al., "FOXA1 promotes tumor cell proliferation through AR involving the Notch pathway in endometrial cancer," *BMC Cancer*, vol. 14, no. 1, p. 78, 2014.
- [112] S. Rangasamy, F. Monickaraj, C. Legendre et al., "Transcriptomics analysis of pericytes from retinas of diabetic animals reveals novel genes and molecular pathways relevant to blood-retinal barrier alterations in diabetic retinopathy," *Experimental Eye Research*, vol. 195, article 108043, 2020.
- [113] T. L. Cameron, I. L. Gresshoff, K. M. Bell et al., "Cartilage-specific ablation of XBP1 signaling in mouse results in a chondrodysplasia characterized by reduced chondrocyte proliferation and delayed cartilage maturation and mineralization," *Osteoarthritis Cartilage*, vol. 23, no. 4, pp. 661–670, 2015.
- [114] J. Izrailit, A. Jaiswal, W. Zheng, M. F. Moran, and M. Reedijk, "Cellular stress induces TRB3/USP9x-dependent Notch activation in cancer," *Oncogene*, vol. 36, no. 8, pp. 1048–1057, 2017.
- [115] S. Hildebrandt, B. Kampfrath, K. Fischer et al., "ActivinA induced SMAD1/5 signaling in an iPSC derived EC model of fibrodysplasia ossificans progressiva (FOP) can be rescued by the drug candidate Saracatinib," *Stem Cell Reviews and Reports*, vol. 17, no. 3, pp. 1039–1052, 2021.
- [116] Q. He, C. Scott Swindle, C. Wan et al., "Enhanced hematopoietic stem cell self-renewal-promoting ability of clonal primary mesenchymal stromal/stem cells versus their osteogenic progeny," *Stem Cells*, vol. 35, no. 2, pp. 473–484, 2017.
- [117] E. Gurcay, E. A. Ozturk, T. Erdem, A. G. Gurcay, and A. Cakci, "Heterotopic ossification as rare complication of hemiplegia following stroke: two cases," *Brain Injury*, vol. 27, no. 13–14, pp. 1727–1731, 2013.
- [118] F. Genêt, K. Minoee, C. Jourdan, A. Ruet, P. Denormandie, and A. Schnitzler, "Troublesome heterotopic ossification and stroke: Features and risk factors. A case control study," *Brain Injury*, vol. 29, no. 7–8, pp. 866–871, 2015.
- [119] G. Varghese, "Heterotopic ossification," *Physical Medicine and Rehabilitation Clinics of North America*, vol. 3, no. 2, pp. 407–415, 1992.
- [120] M. Furmanik and C. M. Shanahan, "ER stress regulates alkaline phosphatase gene expression in vascular smooth muscle cells via an ATF4-dependent mechanism," *BMC research notes*, vol. 11, no. 1, p. 483, 2018.
- [121] P. R. Dube, L. Birnbaumer, and G. Vazquez, "Evidence for constitutive bone morphogenetic protein-2 secretion by M1 macrophages: Constitutive auto/paracrine osteogenic signaling by BMP-2 in M1 macrophages," *Biochemical and biophysical research communications*, vol. 491, no. 1, pp. 154–158, 2017.

- [122] C. Paganini, L. Monti, R. Costantini et al., “Calcium activated nucleotidase 1 (CANT1) is critical for glycosaminoglycan biosynthesis in cartilage and endochondral ossification,” *Matrix Biology*, vol. 81, pp. 70–90, 2019.
- [123] M. Jiang, S. Yu, Z. Yu et al., “XBP1 (X-box-binding protein-1)-dependent O-GlcNAcylation is neuroprotective in ischemic stroke in young mice and its impairment in aged mice is rescued by Thiamet-G,” *Stroke*, vol. 48, no. 6, pp. 1646–1654, 2017.
- [124] W. Wang, Z. Li, and J. Feng, “The potential role of exosomes in the diagnosis and therapy of ischemic diseases,” *Cytotherapy*, vol. 20, no. 10, pp. 1204–1219, 2018.
- [125] A. M. Zagrean, D. M. Hermann, I. Opris, L. Zagrean, and A. Popa-Wagner, “Multicellular crosstalk between exosomes and the neurovascular unit after cerebral Ischemia. Therapeutic Implications,” *Frontiers in neuroscience*, vol. 12, p. 811, 2018.
- [126] T. Forró, Z. Bajkó, A. Bălaşa, and R. Bălaşa, “Dysfunction of the neurovascular unit in ischemic stroke: highlights on microRNAs and exosomes as potential biomarkers and therapy,” *International Journal of Molecular Sciences*, vol. 22, no. 11, p. 5621, 2021.