

Review Article

Sepsis-Induced Brain Dysfunction: Pathogenesis, Diagnosis, and Treatment

Shangwen Pan , Zheng Lv, Rui Wang, Huaqing Shu, Shiyong Yuan, Yuan Yu ,
and You Shang 

Department of Critical Care Medicine, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

Correspondence should be addressed to Yuan Yu; yuyuanwhuh@hust.edu.cn and You Shang; you_shanghust@163.com

Received 9 November 2021; Revised 30 April 2022; Accepted 28 June 2022; Published 24 August 2022

Academic Editor: Sachchida Nand Rai

Copyright © 2022 Shangwen Pan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dysregulated host response to infection, which cause life-threatening organ dysfunction, was defined as sepsis. Sepsis can cause acute and long-term brain dysfunction, namely, sepsis-associated encephalopathy (SAE) and cognitive impairment. SAE refers to changes in consciousness without direct evidence of central nervous system infection. It is highly prevalent and may cause poor outcomes in sepsis patients. Cognitive impairment seriously affects the life quality of sepsis patients and increases the medical burden. The pathogenesis of sepsis-induced brain dysfunction is mainly characterized by the interaction of systemic inflammation, blood-brain barrier (BBB) dysfunction, neuroinflammation, microcirculation dysfunction, and brain dysfunction. Currently, the diagnosis of sepsis-induced brain dysfunction is based on clinical manifestation of altered consciousness along with neuropathological examination, and the treatment is mainly involves controlling sepsis. Although treatments for sepsis-induced brain dysfunction have been tested in animals, clinical treat sepsis-induced brain dysfunction is still difficult. Therefore, we review the underlying mechanisms of sepsis-induced brain injury, which mainly focus on the influence of systemic inflammation on BBB, neuroinflammation, brain microcirculation, and the brain function, which want to bring new mechanism-based directions for future basic and clinical research aimed at preventing or ameliorating brain dysfunction.

1. Introduction

Sepsis is a life-threatening organ dysfunction syndrome caused by the host's maladjusted responses to infection [1]. Globally, there are >30 million sepsis patients each year [2]. Organ dysfunction is a major complication of sepsis, and sepsis-induced brain dysfunction is highly prevalent and has an early onset [3]. Brain dysfunction is mainly caused by various factors released during sepsis, and clinical examinations have not uncovered any evidence of direct central nervous system (CNS) infection. At acute phase, sepsis-induced brain dysfunction is manifested as sepsis-associated encephalopathy (SAE), delirium, sickness behavior, and cerebral ischemia and hemorrhage, which all associated with cognitive impairment [4]. At long-term phase, cognitive impairment is the main feature of sepsis-induced brain dysfunction [5].

SAE is a main manifestation of sepsis, which is characterized by changes in consciousness that range from confusion to delirium or even coma [6] and affects up to 70% of patients with sepsis [7]. The occurrence of SAE often increases the stay in intensive care unit (ICU) and mortality among sepsis patients [8]. Advancements in medical technology have significantly improved the survival rate of sepsis patients but increased the incidence of long-term sequelae and cognitive impairment, which is as high as 21% [9]. Although such huge challenges stand before us, effective diagnostic strategies and intervention measures in sepsis-induced brain dysfunction, especially SAE and cognitive impairment, are lacking [10]. Currently, only daily neurological examination combined with relevant laboratory examinations can be used to exclusively diagnose SAE, and cognitive instruments were used to diagnose cognitive impairment. Thus, effective techniques for early diagnosis

and treatment and even reversal of SAE and cognitive impairment are urgently needed. Here, we review current knowledge on SAE and cognitive impairment and highlight potential diagnostic and treatment strategies.

2. Pathogenesis

Sepsis can cause brain damage through a variety of mechanisms. Among them systemic inflammation, BBB dysfunction, neuroinflammation, microcirculation dysfunction, and brain dysfunction were studied well. Sepsis can amplify adverse reactions through the interaction of these mechanisms, which at last resulting in white matter damage and cerebral dysfunction (Figure 1).

2.1. System Inflammation. Dysregulation of system inflammatory response is the most important feature of sepsis, which stands throughout the whole process [11]. At acute phase, damage-associated molecular patterns (DAMPs) or pathogen-associated molecular patterns (PAMPs) could be detected by pattern-recognition receptors (PRRs) in immune cells, which at last induce cytokine storm and activate immune system [11]. Currently, the most studied PRRs includes Toll-like receptors (TLRs), nucleotide-binding oligomerization domain- (NOD-) like receptors, retinoic acid-inducible gene- (RIG-) like receptors, mannose-binding lectin, and scavenger receptors. Cytokines including interleukin- (IL-) 1, IL-6, tumor necrosis factor- α (TNF- α), interferon (IFN) regulatory factor 7 (IRF7), and adaptor protein 1 (AP-1) are produced after activating of PRRs during sepsis [12]. Furthermore, sepsis also induces the activation of inflammasome, which promotes the release of cytokines IL-1 β and IL-18 [13]. Interestingly, a new form of programmed cell death, named “pyroptosis,” was suggested to involve in sepsis, which can not only lead to direct cell destruction but also to an inflammatory cascade in sepsis [14, 15]. The mechanisms of pyroptosis were mainly include the canonical caspase-1-dependent pathway and the noncanonical caspase-4/5/11-dependent pathway [14]. In canonical pathway, intracellular PRRs recognize stimuli and cleave procaspase-1 into caspase-1, which then promotes the formation of GSDMD channel and the release of IL-1 β and IL-18 [14]. In noncanonical pathway, HMGB-1 derived from liver cells could promote lipopolysaccharide (LPS) transport into cytoplasm through receptor for advanced glycation end products (RAGE) on vascular endothelial cells and macrophages, which cause the activation of caspase-11 and the formation of GSDMD channel [16]. At the same time, the activated caspase-11 could promote the secretion of IL-1 β and IL-18 through pannexin-1/P2X7/NLRP3 pathway [17]. Recently, caspase-3 and caspase-8 were suggested to mediate pyroptosis. After activated, caspase-3 induces cell pyroptosis and releases proinflammatory cell mediators through cleaved and activated gasdermin E (GSDME) to form GSDME channel [18]. The activated caspase-8 by blocked transforming growth factor β -activated kinase 1 (TAK1) could mediate pyroptosis and inflammatory response in a manner of GSDMD channel [19]. In a word, pyroptosis maybe a reason and a potential therapeutic target for multiple organ dysfunction in sepsis [15]. All immune cells

of the innate immune system are mobilized to participate in the process of sepsis [20]. Neutrophils migrate to the inflammation site, where they perform anti-inflammation and eliminate pathogens [20]. The activated mononuclear/macrophage cells phagocytose, kill pathogens, and present antigens [21]. Effector T cells could cause damage by promoting macrophage activation [22]. Abnormal monocyte metabolism is associated with immunosuppression [23].

With the progression of sepsis, multiple organ failure appears, among which the central nervous system is one of the most dangerous organs [24]. Systemic inflammation caused by sepsis not only affects the acute stage of sepsis-induced brain injury, namely, SAE, but also is closely associated with long-term cognitive impairment in the long-term stage of sepsis [25], suggesting that systemic inflammatory response is not only an important influence factor of sepsis-induced brain injury, but also an important intervention target.

2.2. Changes in the Blood-Brain Barrier (BBB). The increased permeability of BBB during sepsis is increasingly accepted, because vasogenic edema and white matter hyperintensities were presented on MRI in SAE patients, indicating BBB disruption [26]. Although sepsis cause BBB damage is not entirely elucidated, several mechanisms have been postulated. The BBB is primarily comprised of microvascular endothelial cells (ECs), tight junction (TJ) proteins, astrocyte endfeet, pericytes, and capillary basement membrane, which are adversely affected by sepsis [27–30]. In normal physiologic conditions, the BBB serves as a physical barrier because TJ between adjacent ECs restrict molecules from diffusing through ECs, ensuring that most molecular trafficking takes a controlled transcellular route across the BBB. In sepsis patients, the expression of TJ was reduced in the brain tissue, indicating BBB damage [27]. Sepsis could activate Toll-like receptor 4/nuclear factor-k κ -gene binding (TLR4/NF- κ B) pathway to change the structure and function of TJ [31]. Histones released during sepsis could induced TJ disruption [32]. Another way is imbalance the oxidative and antioxidant responses of ECs by inducing reactive oxygen species (ROS), which ultimately damage TJ [33]. Inhibiting endothelial nitric oxide synthase (eNOS) and Guanosine triphosphate cyclohydrolase 1 (GTPCH1) and increasing the activation of caspase-3/7 at last promote EC apoptosis during sepsis [29]. Furthermore, DAMPs that are released during sepsis such as ATP could induce apoptosis by purinergic receptor (P2X7R) in brain ECs [28]. Microglia, astrocytes, pericytes, and neutrophils participate in damaging the BBB in sepsis through inflammatory cascade amplification. Besides, astrocytes could expression of vascular endothelial growth factor A (VEGF-A), followed by activating eNOS, inhibiting the expression of claudin-5, and occluding [34]. Collectively, all these factors are ultimately disrupting the barrier function of the BBB. The integrity of basement membrane is damaged in sepsis. The loss of BBB permeability and integrity is a major cause of sepsis-induced brain dysfunction and resultant systemic damage.

ECs activation plays an important role in BBB integrity and is the earliest event in CNS inflammation upon sepsis

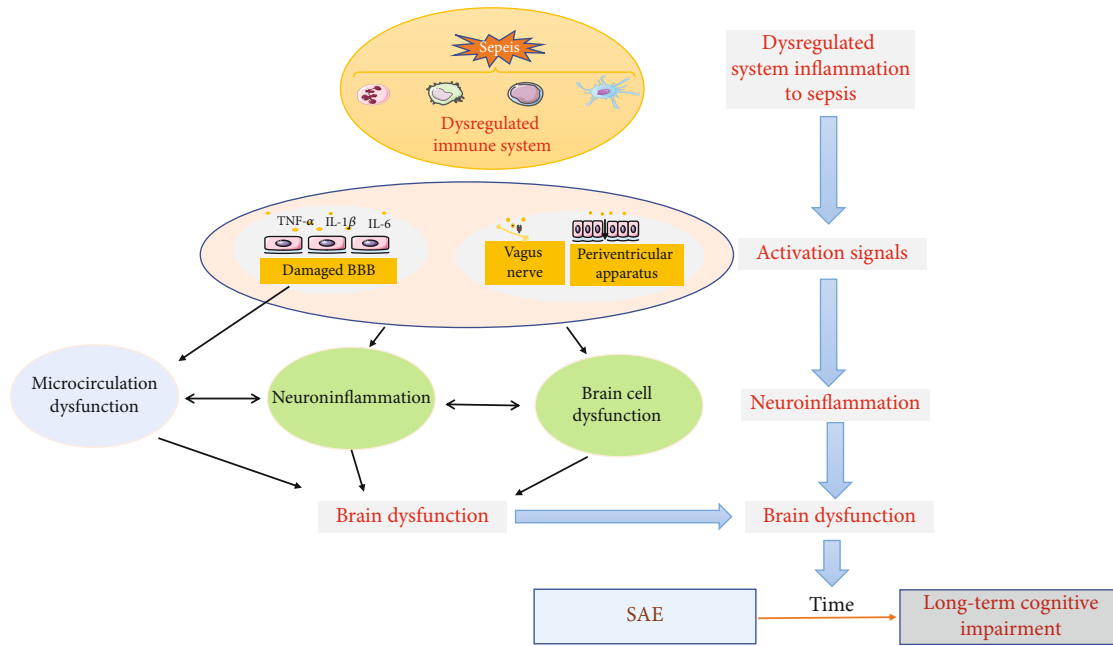


FIGURE 1: Pathophysiology of sepsis-induced brain dysfunction. Sepsis causes inflammation response, which induces neuroinflammation, microcirculation turbulence, and brain dysfunction.

onset [35]. Activated ECs express various adhesion molecules, including tumor necrosis factor receptor superfamily member 5 (CD40), e-selectin, vascular cell adhesion molecule (VCAM), intercellular cell adhesion molecule (ICAM), and inflammatory receptors, including IL-1, TNF- α , and TLR4, which facilitate the entrance of leukocytes and inflammatory mediator into the brain parenchyma [30, 36]. Activation of the EC $\text{I}\kappa\text{B-}\alpha/\text{NF-}\kappa\text{B}$ signaling pathway could produce IL-1 β , TNF- α , IL-6, and other inflammatory cytokines. All these cytokines could bind to their corresponding receptors in microglia [37] and astrocytes [38] and enhance inflammatory responses in the brain parenchyma [35, 39]. Additionally, activated ECs upregulate nitric oxide synthase and cyclooxygenase-2 synthase to aggravate dysfunction of ECs [40–42]. The activated ECs accelerate formation of microthrombus, thereby worsening BBB permeability and exacerbating brain dysfunction [43–45]. Exploratory therapies targeting ECs could alleviate sepsis by suppressing endothelial inflammatory responses, microthrombus formation, and organ dysfunction [46, 47].

BBB destruction disrupts the relative isolation of the CNS, and various neurotoxic substances can directly damage the CNS and exacerbation of gliosis that manifest as the increased cell number and the corresponding marker proteins, Iba1 and GFAP, are stained and enhanced [48]. All of which lead to a neuroinflammatory cascade [35], which may promote sepsis-driven brain dysfunction.

2.3. The Complement System. As an important component of innate immune system, the complement system is involved in the homeostasis of brain [49, 50]. Many organs in the body, such as liver, kidney, and brain, could produce complement proteins [51]. In the brain, microvascular ECs, microglia, astrocyte, and even neuron are the source of com-

plement proteins under certain conditions [51, 52]. Complement activation is a very important factor during sepsis-induced brain dysfunction and may be a potential therapeutic target [53, 54]. Complement C5a levels in cerebrovascular ECs, microglia, and deep neurons increase during sepsis [55]. C5a as a regulatory site influences *Cronobacter sakazakii* related contextual-associated learning through NF- κB and ASK1 pathways [54]. And the C5a neutralizing antibodies or inhibition of its receptor reduces organ dysfunction and BBB damage [56–58]. C3, another important member of the complement system, is also involved in BBB breakdown due to sepsis [59]. In the hippocampus of LPS-stimulated mice, the C3 secreted by astrocyte and C3a receptor (C3aR) expressed by microglia was upregulated [60]. The interaction of C3 and C3aR induced the activation of microglia [60]. Although not explored further, the authors suggested that the activated microglia may induce the loss of inhibitory synapse-related protein through disrupting the transmission of GABAergic synapses, which at last exacerbating cognitive impairment [60]. The complement system also disrupts the BBB via synthesis of inflammatory cytokines and chemokines, which causes edema and neutrophil infiltration [59, 61].

2.4. Neuroinflammation

2.4.1. Activation Signals of Neuroinflammation. During sepsis, homeostasis is maintained via coordination of the nervous, immune, and endocrine systems [62–64]. It is known that sepsis information is transmitted from the periphery to the CNS through the following 3 pathways to regulate the neuroendocrine system, the vegetative nervous system, and behavioral response: (1) the afferent nervous system was represented by the vagus nerve [65]. Inflammatory

cytokines such as IL-1 β bind to IL-1 β or prostaglandin E2 receptor in vagal fibers and increase vagal activity, which at last affects the nucleus tractus solitarius (NTS) by glutamate. The catecholaminergic neurons of the NTS project to different nuclei and cause sickness behaviors [66–68]. Neurotransmitters and neuromodulators derived from bacteria, gamma-aminobutyric acid, noradrenalin, serotonin, dopamine, and acetylcholine could change the state of brain by activating the vagus nerve [69]. Recently, the vagus nerve, as a bridge connecting the gut-brain axis, has been studied extensively. The gut-derived serotonin was released by gut mucosal enterochromaffin cells in response to stimuli including LPS [70, 71]. The increased serotonin by oral selective serotonin reuptake inhibitors upregulating vagal fiber activity, which at last improving depression [72]. Vagus nerve can sense bacterial metabolites (butyrate, propionate, acetate, and valerate) and serotonin through free fatty acid receptors (FFARs) and 5-HT₃/5-HT₄ receptors, which facilitate to transmit signals from gut to the brain [73]. And other different kinds of hormones induced by enteroendocrine cells also transmit peripheral inflammatory signals through the vagus nerve to the autonomic nervous system nucleus, neuroendocrine centers, and behavioral centers, driving corresponding changes [66, 74–76]. The vagus nerve is composed of afferent and efferent nerves, and the activated vagus nerve also acts as an anti-inflammatory mechanism: activating the hypothalamic-pituitary-adrenal axis, thereby producing endogenous steroids and suppressing inflammatory responses [77]. The activated vagus nerve also releases acetylcholine, which binds to nicotinic receptors on the surface of macrophages to suppress inflammatory responses via negative feedback [78]. Vagus activation has also been shown to suppress neuroinflammatory responses by regulating the activation of microglia [74] and the number of astrocytes in the hippocampus and dentate gyrus [79], alleviate the loss of neurons [80], and at last improve sepsis-induced brain dysfunction. Future studies should focus on drug exploration or clinical transformation [74]. (2) Peripheral inflammatory mediators and LPS may directly reach the CNS via the periventricular apparatus [65], which is located between the 3rd and 4th ventricles adjacent to the neuroendocrine nuclei and plant nervous system nuclei. Due to the lacking of BBB and expression of receptors associated with innate and acquired immunity, CNS can directly detect peripheral inflammatory mediators, such as TNF- α , IL-1 β , and IL-6, which activates corresponding nerve nuclei, leading to behavioral changes, fever, and severe nerve damage [75, 81]. (3) Circulating inflammatory mediators enter the CNS via the damaged BBB [30]. Sepsis causes inflammatory mediators to directly access the CNS and to damage the brain parenchyma [27]. Upon entering the CNS, they activate corresponding deep nuclei by influencing neural hormone and cholinergic neurons, gamma-aminobutyric acid, beta-endorphin, and adrenocorticotrophic hormone releasing function. These result in neuroendocrine, behavioral, and cognitive impairment and even affect immune regulation [82–85]. Many mediators, including cytokines, prostaglandins, and nitric oxide (NO), are involved in the activation of proinflammatory responses in the CNS by regulating neu-

rotransmitters and neurosecretion [86]. Moreover, cholinergic and other anti-inflammatory response systems are also activated, suggesting the existence of a proinflammatory/anti-inflammatory homeostasis in the activation signal [87].

2.4.2. Brain Cell Dysfunction. Sepsis produces proinflammatory cytokines that enter the brain parenchyma and cause changes in oxidative stress levels, leading to brain cell dysfunction [88]. The inflammatory cytokines that enter the brain parenchyma bind to receptors on the surface of brain cells and amplify inflammatory responses [30]. Sepsis affects various brain regions differently, with the cortex and hippocampus being highly susceptible [89]. Affected neurons may undergo apoptosis and pyroptosis cellular injury and death in the neural tissues [84, 90]. This may be one of the mechanisms underlying cognitive impairment [91]. Sepsis also disrupts mitochondrial function, which may result in the production of ROS and reactive nitrogen species (RNS) [92]. Additionally, damaged mitochondria can release DAMPs [93]. These effects can cause structural damage to the cell membrane and induce inflammation, causing neuronal apoptosis and cognitive impairment [94].

Microglia, a subgroup of macrophages in the brain, is an important component of glia and has important roles in injury and CNS disease [95]. Recently, myeloid cells in meninges are mainly derived from bone marrow of skull and vertebral, which are transported through vascular channels between the skull and the dura mater [96]. Under some circumstances, such as brain injury or neuroinflammation, these myeloid cells could migrate into parenchyma and differentiated into macrophages to modulate immune response [96]. Therefore, the microglia are constantly self-renewing through myeloid cells in skull and vertebral. In normal physiological conditions, microglia are inhibited by various multiple inhibitory factors. The cytokines TGF- β could induce the quiescent phenotype of microglia by Smad signaling [97]. The complete neuron-microglia connection circuits, CD200 on neuron interacts microglial CD200R, will keep the microglia in its inactivated, resting state [98]. The well-functioning DAP12-trem2 signaling pathway is also a matter of way for inhibiting microglia activation. In this state, microglia are involved in immune surveillance, synaptic modification, and neurogenesis of the surrounding environment through a large number of branches emanating from the cell [99–101]. Microglia also support neuronal survival and growth by secreting neurotrophic factors like insulin growth factor 1 (IGF1), brain-derived neurotrophic factor (BDNF), transforming growth factor β (TGF β), and nerve growth factor (NGF) [102, 103]. Sepsis relieves the inhibitory mechanisms and releases various other factors like TNF, iNOS, or glucose and confers proinflammatory function to microglia, which produce a range of proinflammatory and neurotoxic factors, thereby expanding inflammatory responses and neuronal damage in the CNS. The activated microglia are divided into two phenotypes based on antigen markers and function: M1 and M2. M1 is the classic activation of microglia, mainly expressing surface antigens CD16, CD32, and CD86 and secreting IL-1, IL-6, and TNF- α , which mediate inflammatory response and produce cytotoxic effects.

These effects contribute to brain injury and cognitive dysfunction [104]. Activated microglia may also exacerbate brain dysfunction by altering the permeability of the BBB [105]. M2 phenotype is an alternative activation of microglia, mainly expressing antigens chitinase 3 like Protein3 (Chi3l3), arginase-1 (ARG-1), and CD206. Secretion of insulin like growth factor-1 (IGF-1) and transforming TGF- β and other anti-inflammatory factors can inhibit the excessive inflammatory response. M2 phenotype microglia can also secrete neurotrophic factors and play a protective role on neurons. Increasing the proportion of M2 microglia can improve the brain dysfunction in sepsis [106]. In conclusion, as the first line of defense against pathogens or injury, microglia are crucial in the maintenance of CNS homeostasis.

Astrocytes maintain CNS homeostasis through a wide range of functions, including ion homeostasis and neurotransmitter metabolism, fluid balance, regulation of local blood flow, neurogenesis, maintenance of synaptic connectivity, and plasticity [107]. Astrocytes express a wide range of receptors for DAMPs and PAMPs, including TLRs, NLRs, double-stranded RNA-dependent protein kinases, scavenger receptors, mannose receptor, complement components, and mediators like CXC chemokine ligand-10 (CXCL10), chemokine (C-C motif) ligand 2 (CCL2), IL-6, and B-cell-activating factor of the TNF family (BAFF) [108]. After activation, astrocytes secrete proinflammatory factors that induce and/or regulate neuroinflammation. Astrocyte-derived factors with proinflammatory activity are represented by the following: (i) chemokines (including monocyte chemoattractant protein-1 (MCP-1/CCL2), CCL5 (RANTES), CCL7, CCL8, CCL12, CXCL1, CXCL8 (IL-8), CXCL9, IFN- γ -inducible protein-10 (IP-10/CXCL10), CXCL12, and CXCL16), (ii) cytokines and growth factors (including IL-1- β , IL-6, IL-11, IL-15, IL-17, TNF- α , BAFF, and VEGF), (iii) intracellular signaling factors (including NF- κ B, SOCS3, and Act1), and (iv) small intercellular effector molecules (including PGE and NO) [109]. These proinflammatory cytokines exacerbate neuronal damage, leading to brain dysfunction in sepsis. Astrocytes also regulate the neuroinflammation during sepsis by controlling microglial activation. Microglia activation in inflamed brain triggers a pronounced neurotoxic phenotype characterized by the release of multiple cytokines and ROS/RNS, which contribute to cell death in specific vulnerable brain areas [110]. During sepsis, PAMPs could bind to the corresponding receptors on the surface of astrocytes, such as TLR4, activate the NLRP3 inflammasome to induce pyroptosis, and release histones to damage neurons [111]. By inhibiting glutamate reuse by astrocytes, sepsis affects neuronal function. Astrocytes are an important part of the BBB and control its permeability. Upon activation, astrocytes produce VEGF-A and thymidine phosphorylase (TYMP/endothelial cell growth factor 1, ECGF1), which suppress the expression of TJ proteins in cerebral ECs, thereby enhancing breakdown of the BBB [34]. In response to endotoxemia, astrocytes also secrete CCL11, which impairs learning and memory in the adult brain and triggers microglia migration and ROS production, thereby causing hippocampal neuronal damage, behavioral changes, and memory impairment [112].

Neurons can be damaged through a variety of mechanisms during sepsis. Activated microglia can cause neuronal damage by releasing inflammatory cytokines and ROS [113, 114]. Microglia can also induce the transformation of A1 astrocytes to injury neurons [115]. In addition to secreting inflammatory cytokines [116], astrocytes also affect the release of neurotransmitters, which at last leads to neuronal injury [117]. Sepsis can dysfunction the autophagy and pyroptosis homeostasis [118] and activate ferroptosis [119] and endoplasmic reticulum stress of neurons, all of which could lead to neuronal damage [120]. Abnormal activation of neuronal membrane receptors can transmit stimulus signals and induce PANoptosis [121]. Recently, the interactions of organs are getting a lot of attention. The gut can damage neurons through the accumulation of cytokines [122]. In conclusion, in sepsis, various direct or indirect factors could lead to neuronal damage and aggravate the development of cognitive impairment.

2.5. Microcirculation Dysfunction. Normal microcirculation is essential for maintaining CNS function. Sepsis triggers coagulation disorder by activating ECs, which enhances coagulation cascade activity and promotes microthrombus formation [43]. The activated brain endothelium activates thrombin, which regulates coagulation via prothrombin cleavage by Factor X. Thrombin then converts soluble fibrinogen to fibrin and activates platelets, resulting in microocclusions [44]. Continuous microthrombus formation exacerbates focal ischemia by occluding the vasculature beyond the initial occlusion sites. The lack of oxygen and nutrients supply further aggravates EC activation and induces coagulation dysfunction, leading to ischemia or hemorrhagic injury [123]. Moreover, basic and clinical studies show that sepsis damages the regulatory function of cerebral vasomotor and blood pressure autoregulation [124], affects cerebral perfusion, and aggravates brain injury [125]. The degree of damage to the cerebral microcirculation negatively correlated with the prognosis of sepsis-induced brain dysfunction. Microcirculatory dysfunction is characterized by rapid onset and clearly precedes changes in neurovascular coupling and systemic circulation [126]. However, there are few simple and effective methods of clinically detecting cerebral microcirculation. Although systemic circulation parameters may change with sepsis, their applicability to microcirculation is unclear. At present, evaluation and optimization of cerebral perfusion are still inconclusive, but necropsy reports have confirmed that sepsis it causes multiple microinfarcts, especially in areas with relatively low cerebral blood flow. And an MRI study revealed that patients with sepsis are at a higher risk of ischemic stroke.

In conclusion, impaired microcirculation may contribute to the pathogenesis of sepsis-induced brain injury, especially, sepsis-induced cognitive impairment. Currently, there are 2 hypotheses on sepsis-induced cognitive impairment: (a) the hypothesis that neurodegeneration involves microglial activation and (b) the hypothesis that impaired microcirculation involves blood vessels. These hypotheses are interwoven and warrant further investigation.

2.6. Brain Dysfunction. Although neuroinflammation usually occurs in a diffuse form, patients with sepsis usually suffer from multiple factors and some brain areas are particularly sensitive to neuroinflammation or lack BBB protection, making them more vulnerable to direct attack by peripheral inflammatory cytokines. The hippocampus is particularly vulnerable to damage during sepsis because inflammation, ischemia, hypoxia, and blood sugar disorders can all injure the hippocampus. Moreover, these changes may cooccur during sepsis. Inhibition of oxidative stress in the hippocampus may reduce injuries and cognitive impairment in sepsis [127]. Additionally, other areas of the brain, including the cortex, cerebellum, and brain stem, are also damaged by sepsis [128–130]. Thus, it is believed that sepsis-driven brain injury presents in a diffuse form and is closely associated with cognitive impairment.

Sepsis-induced brain stem dysfunction results in changes in consciousness as well as cardiovascular and immune system dysfunction and contributes to poor patient prognoses [30]. The brain stem controls immune responses through the sympathetic and parasympathetic nervous systems [131]. During sepsis, changes in cholinergic neurotransmitters are also an important driver of brain stem dysfunction. Brainstem cholinergic pathways can diminish cardiovascular and neuroinflammatory actions during endotoxemia [132]. The brain stem nucleus is susceptible to sepsis and treatment for this can alleviate sepsis-induced brain dysfunction, including neuroinflammation and cognitive dysfunction [132].

Additionally, sepsis also has driven damage of the neurotransmitter system [133], which at last promotes the incidence of brain injury [134, 135]. These neurotransmitter systems include acetylcholine, GABA, dopamine, norepinephrine, serotonin, and glutamate [136–139]. During sepsis, neurotransmitter synthesis is also altered by neurotoxic amino acids like NO, tryptophan, and phenylalanine. Metabolic dysfunction due to liver and kidney failure caused by sepsis and various drugs can affect neurotransmitter synthesis and release.

3. Clinical Symptoms of Sepsis-Induced Brain Dysfunction

Clinically, sepsis-induced brain dysfunction is characterized by focal neurological deficits, cognitive impairments, depression, attention decline, mood disorders, and movement-coordination problems, as well as reduced rationality, awareness, comprehension, intelligence, mental processing, and social interaction. Psychomotor agitation, anxiety syndrome, reduced visual representation (episodic and semantic memory), loss of visual acuity, executive and intellectual changes, and disturbed circadian rhythm have also been observed in cases of sepsis-induced brain dysfunction [4]. Moreover, simultaneous changes in cerebral and cardiovascular function have been reported, whereby cardiopulmonary resuscitation and artificial ventilation were used as remedies [65]. Sepsis-induced brain dysfunction also affects peripheral circulation and its link with brain parenchymal signal intensity [45]. Impaired cerebral microvasculature and decreased microcirculation, as well as overall reduction in total and

perfused blood vessels and functional red blood cell capillary densities, are key features of sepsis-induced brain dysfunction. Another key feature is brain microcirculatory abnormalities during the onset and progression of sepsis induced brain dysfunction [140]. Thus, because sepsis-induced brain dysfunction lacks specific neurological indicators, clinicians should use exclusion diagnosis based on patient history to determine if brain dysfunction is due to sepsis.

4. Diagnosis

Diagnosis of sepsis-induced brain dysfunction is exclusionary and requires that other potential causes of neurological dysfunction, including drug effects, metabolic disorders, primary central diseases like meningitis, encephalitis, cerebrovascular diseases, and epilepsy, and noninfectious systemic inflammatory reactions like burns, severe pancreatitis, and trauma, are first excluded [141]. Neurological examination is the basic means of identifying patients with sepsis-induced brain dysfunction but is not suitable for mechanically ventilated patients in deep sedation. Electroencephalogram can be effective but should be combined with other examinations for comprehensive assessment [142, 143].

Currently, some biomarkers have been used to assess sepsis-induced brain dysfunction. But the evidence is insufficient, which has not been promoted clinically (Table 1).

5. Treatments

Currently, there are no specific treatments for sepsis-induced brain dysfunction and treatment mainly focusses on symptoms and may include control of sepsis and minimizing the injury to the CNS. Symptomatic treatment of sepsis-induced brain dysfunction does not differ significantly from sepsis treatment. Early resuscitation is considered a key therapeutic strategy against sepsis. Fast fluid restoration is proposed as a primary measure so as to restore hemodynamic stability and systemic oxygen delivery, which reduces neuroinflammation, stroke volume, and need for vasopressor agents. However, this approach carries some risks, including hyperchloremic metabolic acidosis, hyperkalemia, and pathologic immune activation, as well as cellular damage, bleeding disorders, renal failure, or life-threatening allergic responses. After fluid therapy during the early resuscitation process, vasopressor therapy that links with a normal arterial pressure can reduce the severity of sepsis.

In recent years, specific treatments for sepsis-induced brain dysfunction have been sought. Vagus nerve stimulation attenuates sepsis-induced peripheral inflammation and activates afferent nerve fibers at axonal projections. This stimulation attenuates the expression of proinflammatory cytokines in the brain. Vagus nerve stimulation also reduces sepsis-induced hypotension, disseminated intravascular coagulation, fibrinolytic activity, and systemic organ dysfunction. Free radical generation and oxidative stress contribute to the progression of sepsis-induced brain damage. Thus, antioxidant therapies have been proposed for managing sepsis-associated brain dysfunction. In response to stress, the hypothalamus secretes corticotropin releasing hormone,

TABLE 1: Suggested biomarkers to monitor sepsis-induced brain dysfunction.

Biomarkers	Significance	Location
C-reactive protein (CRP) and Procalcitonin (PCT)	Higher CRP levels indicated prolonged acute brain dysfunction [149]	Plasma
C-type natriuretic peptide (NT-proCNP)	High-peak concentration of NT-proCNP in the early phase of sepsis could predict SAE [150]	Plasma
IL-6, IL-8, IL-10, TNF- α and S-100 β	Negatively associated with delirium free days [151]	Plasma
Neurofilament (Nf)	Nf could predict poorer cognitive outcome in SAE patients [151]	Cerebrospinal fluid (CSF) and plasma
Adiponectin, Tau, and neopterin	Significantly higher in patients with delirium [152]	Plasma

TABLE 2: Suggested treatments to sepsis-induced brain dysfunction.

Therapies	Mechanism
Vagus nerve stimulation	Regulation inflammatory response to protect organs [74]
Antioxidant therapies	Reduces oxidative stress to improve brain function [74]
Glucocorticoids	Regulate the secretion of hormones and anti-inflammatory [144]
CNI-1493	Regulation neuroinflammation by inhibiting P38/MAPK signaling [145]
Hypothermia	Change antibiotic pharmacokinetics [146]
IDO	Influences neuroinflammation [147]
Rg1	Anti-inflammation, suppress apoptosis, and autophagic degradation [148]
CC16	Anti-inflammatory, antioxidant, regulation autophagy [153]

resulting in cortisol secretion by adrenal glands. Hence, glucocorticoids have been considered for treating sepsis-induced brain dysfunction [144]. Surrogate markers and modulators of neuroimmune axis have also been considered for treating sepsis-induced brain dysfunction. CNI-1493 is a guanlylhydrazone that reduces neuroinflammation by inhibiting P38/MAPK signaling. Moreover, α - or β -adrenergic receptor modulation may also stimulate recovery from dysregulated immune responses [145]. Hypothermia has been considered a standard strategy [146]. The enzyme indoleamine 2,3-dioxygenase (IDO), which influences inflammatory processes, has been considered as a therapeutic target for treating CNS disorders. IDO inhibition not only restored adaptive immunity and energy metabolism but also improved cognitive function in sepsis [147]. The ginsenoside, Rg1, an important component of ginseng, has been reported to suppress apoptosis and autophagic degradation in the hippocampus during sepsis-induced brain dysfunction. Rg1 is reported to attenuate brain atrophy and to reduce histopathological alterations in the hippocampus. It reduces the levels of the inflammatory mediators, TNF- α , IL-1 β , and IL-6, the expression of activated microglial marker, Iba1, microglia morphological changes like rounded cell bodies and shrunken neurites, macrophage infiltration, neuroinflammation, and caspase-3 activation in neurons. These Rg1-induced neuroprotective effects decreased behavioral defects in the sepsis model [148]. Treatment with recombinant club cell protein (CC16), which has anti-inflammatory and antioxidant effects, reduced sepsis-associated pathological changes in brain tissue by suppressing p38/MAPK signaling. However, neuroprotection from recombinant CCL16 involved augmented LC3II and suppressed QSTM1/p62 expression, along with the division of large autophagosomes into smaller autophagic vacuoles. This led to an

increased neuronal survival due to reduced apoptosis. Depending on the protective agent, increased autophagy often protects from sepsis-induced apoptosis. A rat model of sepsis-induced brain dysfunction revealed that low-dose dexamethasone enhances autophagy, as revealed by inhibited mTOR signaling, increased LC3-II/LC3-I ratio and decreased p62/QSTM1 in cortical neurons. Although adjunctive therapy in patients with severe sepsis-induced brain dysfunction may improve BBB dysfunction, further studies are needed to confirm this (Table 2). Despite these treatments' advances in animal, clinical applications are still to be explored.

6. Conclusion and Outlook

Brain dysfunction due to sepsis is often overlooked despite its high incidence and contribution to increased mortality in ICU patients. The pathophysiological mechanisms underlying sepsis-induced brain dysfunction are very complex and are mainly driven by inflammation. These processes affect brain cell metabolism by inducing oxidative stress and altering neurotransmission. The inflammatory processes mainly involve endothelial activation, impaired microcirculation, BBB defects, inflammatory mediators, and microglial cell activation. A comprehensive neurological examination is essential for diagnosing sepsis-induced brain dysfunction and should be performed daily. Active control of sepsis is the cornerstone of treating sepsis-associated brain dysfunction and should be done in a holistic framework that includes high-dose antibiotic therapy and fluid support therapy. On this basis, further research is needed to develop effective treatments for brain dysfunction. Potential future treatment strategies include vagus nerve stimulation and

the regulation of neuroinflammation regulation, neuroendocrine function, and neuroimmunity.

Future research on sepsis or brain dysfunction needs to address several issues. First, quick and accurate strategies for diagnosing sepsis-induced brain dysfunction are needed. The rapidly advancing imaging techniques are getting us closer solving this problem. New neuroimaging methods that target neuroinflammation, including PET, SPECT, and new MRI protocols that have been applied in multiple sclerosis and neurodegenerative diseases may in future become applicable to sepsis. Secondly, a better understanding of pathophysiological mechanisms underlying sepsis-triggered brain dysfunction is needed. There is no doubt that the relationship between sepsis and brain dysfunction will become a major research area in the future. Currently, several treatments have been investigated using animal models, but they are all in the early stages of research and have not reached clinical trial stages. In view of its high incidence, more effective treatments are urgently needed for sepsis-induced brain dysfunction.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

This work was supported by the Scientific Research Project of Health Commission of Hubei Province (WJ2021M248).

References

- [1] M. Singer, C. S. Deutschman, C. W. Seymour et al., "The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3)," *JAMA*, vol. 315, no. 8, pp. 801–810, 2016.
- [2] C. Fleischmann, A. Scherag, N. K. Adhikari et al., "Assessment of global incidence and mortality of hospital-treated sepsis. current estimates and limitations," *American Journal Of Respiratory And Critical Care Medicine*, vol. 193, no. 3, pp. 259–272, 2016.
- [3] T. E. Gofton and G. B. Young, "Sepsis-associated encephalopathy," *Nature Reviews Neurology*, vol. 8, no. 10, pp. 557–566, 2012.
- [4] P. F. Czempik, M. P. Pluta, and L. J. Krzych, "Sepsis-associated brain dysfunction: a review of current literature," *International Journal of Environmental Research and Public Health*, vol. 17, no. 16, 2020.
- [5] T. J. Iwashyna, E. W. Ely, D. M. Smith, and K. M. Langa, "Long-term cognitive impairment and functional disability among survivors of severe sepsis," *JAMA*, vol. 304, no. 16, pp. 1787–1794, 2010.
- [6] M. Ebersoldt, T. Sharshar, and D. Annane, "Sepsis-associated delirium," *Intensive Care Medicine*, vol. 33, no. 6, pp. 941–950, 2007.
- [7] C. N. Widmann and M. T. Heneka, "Long-term cerebral consequences of sepsis," *The Lancet Neurology*, vol. 13, no. 6, pp. 630–636, 2014.
- [8] R. Sonnevile, E. de Montmollin, J. Poujade et al., "Potentially modifiable factors contributing to sepsis-associated encephalopathy," *Intensive Care Medicine*, vol. 43, no. 8, pp. 1075–1084, 2017.
- [9] A. Calsavara, V. Nobre, T. Barichello, and A. L. Teixeira, "Post-sepsis cognitive impairment and associated risk factors: A systematic review," *Australian Critical Care*, vol. 31, no. 4, pp. 242–253, 2018.
- [10] H. E. Wang, M. M. Kabeto, M. Gray et al., "Trajectory of cognitive decline after sepsis," *Critical Care Medicine*, vol. 49, no. 7, pp. 1083–1094, 2021.
- [11] T. Manabe and M. T. Heneka, "Cerebral dysfunctions caused by sepsis during ageing," *Nature Reviews Immunology*, vol. 22, pp. 1–15, 2021.
- [12] C. Lv and L. Huang, "Xenobiotic receptors in mediating the effect of sepsis on drug metabolism," *Acta Pharmaceutica Sinica B*, vol. 10, no. 1, pp. 33–41, 2020.
- [13] L. G. Danielski, A. D. Giustina, S. Bonfante, T. Barichello, and F. Petronilho, "The NLRP3 inflammasome and its role in sepsis development," *Inflammation*, vol. 43, no. 1, pp. 24–31, 2020.
- [14] X. Zheng, W. Chen, F. Gong, Y. Chen, and E. Chen, "The role and mechanism of pyroptosis and potential therapeutic targets in sepsis: a review," *Frontier in Immunology*, vol. 12, article 711939, 2021.
- [15] Y. L. Gao, J. H. Zhai, and Y. F. Chai, "Recent advances in the molecular mechanisms underlying pyroptosis in sepsis," *Mediators of Inflammation*, vol. 2018, Article ID 5823823, 2018.
- [16] D. Tang, H. Wang, T. R. Billiar, G. Kroemer, and R. Kang, "Emerging mechanisms of immunocoagulation in sepsis and septic shock," *Trends in Immunology*, vol. 42, no. 6, pp. 508–522, 2021.
- [17] D. Yang, Y. He, R. Munoz-Planillo, Q. Liu, and G. Nunez, "Caspase-11 requires the pannexin-1 channel and the purinergic P2X7 pore to mediate pyroptosis and endotoxin shock," *Immunity*, vol. 43, no. 5, pp. 923–932, 2015.
- [18] C. Rogers, T. Fernandes-Alnemri, L. Mayes, D. Alnemri, G. Cingolani, and E. S. Alnemri, "Cleavage of DFNA5 by caspase-3 during apoptosis mediates progression to secondary necrotic/pyroptotic cell death," *Nature Communications*, vol. 8, p. 14128, 2017.
- [19] P. Orning, D. Weng, K. Starheim et al., "Pathogen blockade of TAK1 triggers caspase-8-dependent cleavage of gasdermin D and cell death," *Science*, vol. 362, no. 6418, pp. 1064–1069, 2018.
- [20] X. Wen, B. Xie, S. Yuan, and J. Zhang, "The "Self-Sacrifice" of immunocytes in sepsis," *Frontiers in Immunology*, vol. 13, article 833479, 2022.
- [21] A. Sica, M. Erreni, P. Allavena, and C. Porta, "Macrophage polarization in pathology," *Cellular and Molecular Life Sciences*, vol. 72, no. 21, pp. 4111–4126, 2015.
- [22] R. D. Stout and J. Suttles, "T cell signaling of macrophage function in inflammatory disease," *Frontiers in Bioscience-Landmark*, vol. 2, pp. d197–d206, 1997.
- [23] F. Venet and G. Monneret, "Advances in the understanding and treatment of sepsis-induced immunosuppression," *Nature Reviews Nephrology*, vol. 14, no. 2, pp. 121–137, 2018.
- [24] C. Caraballo and F. Jaimes, "Organ dysfunction in sepsis: an ominous trajectory from infection to death," *The Yale Journal Of Biology And Medicine*, vol. 92, no. 4, pp. 629–640, 2019.
- [25] V. T. A. Peters, L. E. Meijer-van, M. I. Bergkamp et al., "Risk of dementia and structural brain changes following

- nonneurological infections during 9-year follow-up,” *Critical Care Medicine*, vol. 50, no. 4, pp. 554–564, 2022.
- [26] D. J. Stubbs, A. K. Yamamoto, and D. K. Menon, “Imaging in sepsis-associated encephalopathy—insights and opportunities,” *Nature Reviews Neurology*, vol. 9, no. 10, pp. 551–561, 2013.
- [27] K. Erikson, H. Tuominen, M. Vakkala et al., “Brain tight junction protein expression in sepsis in an autopsy series,” *Critical Care*, vol. 24, no. 1, p. 385, 2020.
- [28] K. Wang, M. Sun, Z. Juan et al., “The improvement of sepsis-associated encephalopathy by P2X7R inhibitor through inhibiting the Omi/HtrA2 apoptotic signaling pathway,” *Behavioural Neurology*, vol. 2022, Article ID 3777351, 2022.
- [29] L. Liu, H. Zhang, Y. Shi, and L. Pan, “Prostaglandin E1 improves cerebral microcirculation through activation of endothelial NOS and GRPCH1,” *Journal of Molecular Neuroscience*, vol. 70, no. 12, pp. 2041–2048, 2020.
- [30] M. Gu, X. L. Mei, and Y. N. Zhao, “Sepsis and cerebral dysfunction: BBB damage, neuroinflammation, oxidative stress, apoptosis and autophagy as key mediators and the potential therapeutic approaches,” *Neurotoxicity Research*, vol. 39, no. 2, pp. 489–503, 2021.
- [31] H. C. Zhou, C. A. Guo, W. W. Yu et al., “Zizyphus jujuba cv. Muzao polysaccharides enhance intestinal barrier function and improve the survival of septic mice,” *Journal of Food Biochemistry*, vol. 45, no. 5, article e13722, 2021.
- [32] N. Villalba, S. Baby, B. J. Cha, and S. Y. Yuan, “Site-specific opening of the blood-brain barrier by extracellular histones,” *Journal of Neuroinflammation*, vol. 17, no. 1, p. 281, 2020.
- [33] Z. Zhao, J. Hu, X. Gao, H. Liang, and Z. Liu, “Activation of AMPK attenuates lipopolysaccharide-impaired integrity and function of blood-brain barrier in human brain microvascular endothelial cells,” *Experimental and Molecular Pathology*, vol. 97, no. 3, pp. 386–392, 2014.
- [34] C. Chapouly, A. A. Tadesse, S. Horng et al., “Astrocytic TYMP and VEGFA drive blood-brain barrier opening in inflammatory central nervous system lesions,” *Brain*, vol. 138, no. 6, pp. 1548–1567, 2015.
- [35] M. C. Kodali, H. Chen, and F. F. Liao, “Temporal unsnarling of brain’s acute neuroinflammatory transcriptional profiles reveals panendothelitis as the earliest event preceding microgliosis,” *Molecular psychiatry*, vol. 26, no. 8, pp. 3905–3919, 2020.
- [36] H. Zhou, G. Andonegui, C. H. Wong, and P. Kuberski, “Role of endothelial TLR4 for neutrophil recruitment into central nervous system microvessels in systemic inflammation,” *The Journal of Immunology*, vol. 183, no. 8, pp. 5244–5250, 2009.
- [37] N. P. Hailer, C. Vogt, H. W. Korf, and F. Dehghani, “Interleukin-1beta exacerbates and interleukin-1 receptor antagonist attenuates neuronal injury and microglial activation after excitotoxic damage in organotypic hippocampal slice cultures,” *European Journal of Neuroscience*, vol. 21, no. 9, pp. 2347–2360, 2005.
- [38] A. Fernandes, A. Barateiro, A. S. Falcao et al., “Astrocyte reactivity to unconjugated bilirubin requires TNF-alpha and IL-1beta receptor signaling pathways,” *GLIA*, vol. 59, no. 1, pp. 14–25, 2011.
- [39] M. L. Wong, P. B. Bongiorno, A. Al-Shekhlee, A. Esposito, P. Khatri, and J. Licinio, “IL-1 beta, IL-1 receptor type I and iNOS gene expression in rat brain vasculature and perivascular areas,” *Neuroreport*, vol. 7, no. 15-17, pp. 2445–2448, 1996.
- [40] D. Freyer, R. Manz, A. Ziegenhorn et al., “Cerebral endothelial cells release TNF-alpha after stimulation with cell walls of *Streptococcus pneumoniae* and regulate inducible nitric oxide synthase and ICAM-1 expression via autocrine loops,” *The Journal of Immunology*, vol. 163, no. 8, pp. 4308–4314, 1999.
- [41] O. Handa, J. Stephen, and G. Cepinskas, “Role of endothelial nitric oxide synthase-derived nitric oxide in activation and dysfunction of cerebrovascular endothelial cells during early onsets of sepsis,” *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 295, no. 4, pp. H1712–H1719, 2008.
- [42] K. Matsumura, C. Cao, M. Ozaki, H. Morii, K. Nakadate, and Y. Watanabe, “Brain endothelial cells express cyclooxygenase-2 during lipopolysaccharide-induced fever: light and electron microscopic immunocytochemical studies,” *Journal of Neuroscience*, vol. 18, no. 16, pp. 6279–6289, 1998.
- [43] H. Wang, L. J. Hong, J. Y. Huang et al., “P2RX7 sensitizes Mac-1/ICAM-1-dependent leukocyte-endothelial adhesion and promotes neurovascular injury during septic encephalopathy,” *Cell Research*, vol. 25, no. 6, pp. 674–690, 2015.
- [44] G. E. Jarvis, B. T. Atkinson, J. Frampton, and S. P. Watson, “Thrombin-induced conversion of fibrinogen to fibrin results in rapid platelet trapping which is not dependent on platelet activation or GPIb,” *British journal of pharmacology*, vol. 138, no. 4, pp. 574–583, 2003.
- [45] D. C. Nwafor, A. L. Brichacek, A. S. Mohammad et al., “Targeting the blood-brain barrier to prevent sepsis-associated cognitive impairment,” *Journal of Central Nervous System Disease*, vol. 11, p. 593300084, 2019.
- [46] T. Li, H. Chen, X. Shi et al., “HSF1 alleviates microthrombosis and multiple organ dysfunction in mice with sepsis by upregulating the transcription of tissue-type plasminogen activator,” *Thrombosis and Haemostasis*, vol. 121, no. 8, pp. 1066–1078, 2021.
- [47] D. Maier-Begandt, H. S. Comstra, S. A. Molina et al., “A venous-specific purinergic signaling cascade initiated by Pan-nexin 1 regulates TNFalpha-induced increases in endothelial permeability,” *Science Signaling*, vol. 14, no. 672, 2021.
- [48] L. Zhang, X. Peng, Y. Ai et al., “Amitriptyline reduces sepsis-induced brain damage through TrkA signaling pathway,” *Journal of Molecular Neuroscience*, vol. 70, no. 12, pp. 2049–2057, 2020.
- [49] D. Ricklin, E. S. Reis, and J. D. Lambris, “Complement in disease: a defence system turning offensive,” *Nature Reviews Nephrology*, vol. 12, no. 7, pp. 383–401, 2016.
- [50] D. C. Mastellos, “Complement emerges as a masterful regulator of CNS homeostasis, neural synaptic plasticity and cognitive function,” *Experimental Neurology*, vol. 261, pp. 469–474, 2014.
- [51] B. P. Morgan, “Complement in the pathogenesis of Alzheimer’s disease,” *Seminars in Immunopathology*, vol. 40, no. 1, pp. 113–124, 2018.
- [52] J. J. Alexander, “Blood-brain barrier (BBB) and the complement landscape,” *Molecular Immunology*, vol. 102, pp. 26–31, 2018.
- [53] X. Liang, T. Wu, Q. Chen et al., “Serum proteomics reveals disorder of lipoprotein metabolism in sepsis,” *Life Sci Alliance*, vol. 4, no. 10, 2021.

- [54] P. Vinay, C. Karen, K. Balamurugan, and K. E. Rajan, "Cronobacter sakazakii infection in early postnatal rats impaired contextual-associated learning: a putative role of C5a-mediated NF-kappaB and ASK1 pathways," *Journal of Molecular Neuroscience*, vol. 71, no. 1, pp. 28–41, 2021.
- [55] M. A. Flierl, P. F. Stahel, B. M. Touban et al., "Bench-to-bedside review: burn-induced cerebral inflammation—a neglected entity?," *Critical Care*, vol. 13, no. 3, p. 215, 2009.
- [56] F. S. Zetoune and P. A. Ward, "Role of complement and histones in sepsis," *Frontiers in Medicine*, vol. 7, p. 616957, 2020.
- [57] M. A. Flierl, P. F. Stahel, D. Rittirsch et al., "Inhibition of complement C5a prevents breakdown of the blood-brain barrier and pituitary dysfunction in experimental sepsis," *Critical Care*, vol. 13, no. 1, p. R12, 2009.
- [58] A. Jacob, B. Hack, E. Chiang, J. G. Garcia, R. J. Quigg, and J. J. Alexander, "C5a alters blood-brain barrier integrity in experimental lupus," *The FASEB Journal*, vol. 24, no. 6, pp. 1682–1688, 2010.
- [59] A. Jacob, J. R. Brorson, and J. J. Alexander, "Septic encephalopathy: inflammation in man and mouse," *Neurochemistry International*, vol. 58, no. 4, pp. 472–476, 2011.
- [60] S. M. Li, B. Li, L. Zhang et al., "A complement-microglial axis driving inhibitory synapse related protein loss might contribute to systemic inflammation-induced cognitive impairment," *International Immunopharmacology*, vol. 87, p. 106814, 2020.
- [61] J. J. Alexander, A. Jacob, P. Cunningham, L. Hensley, and R. J. Quigg, "TNF is a key mediator of septic encephalopathy acting through its receptor, TNF receptor-1," *Neurochemistry International*, vol. 52, no. 3, pp. 447–456, 2008.
- [62] U. Andersson and K. J. Tracey, "Reflex principles of immunological homeostasis," *Annual Review Of Immunology*, vol. 30, pp. 313–335, 2012.
- [63] K. A. Muscatell, K. Dedovic, G. M. Slavich et al., "Greater amygdala activity and dorsomedial prefrontal-amygdala coupling are associated with enhanced inflammatory responses to stress," *Brain, behavior, and Immunity*, vol. 43, pp. 46–53, 2015.
- [64] C. M. Hueston and T. Deak, "The inflamed axis: the interaction between stress, hormones, and the expression of inflammatory-related genes within key structures comprising the hypothalamic-pituitary-adrenal axis," *Physiology & Behavior*, vol. 124, pp. 77–91, 2014.
- [65] R. Sonnevile, F. Verdonk, C. Rauturier et al., "Understanding brain dysfunction in sepsis," *Annals of Intensive Care*, vol. 3, no. 1, p. 15, 2013.
- [66] M. Ek, M. Kurosawa, T. Lundeberg, and A. Ericsson, "Activation of vagal afferents after intravenous injection of interleukin-1beta: role of endogenous prostaglandins," *Journal of Neuroscience*, vol. 18, no. 22, pp. 9471–9479, 1998.
- [67] R. Dantzer, "Cytokine-induced sickness behavior: where do we stand?," *Brain, Behavior, and Immunity*, vol. 15, no. 1, pp. 7–24, 2001.
- [68] M. M. Moughnyeh, K. M. Brawner, B. A. Kennedy et al., "Stress and the gut-brain axis: implications for cancer," *Inflammation and Sepsis Journal of Surgical Research*, vol. 266, pp. 336–344, 2021.
- [69] J. F. Cryan and T. G. Dinan, "Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour," *Nature Reviews Neuroscience*, vol. 13, no. 10, pp. 701–712, 2012.
- [70] M. Kidd, B. I. Gustafsson, I. Drozdov, and I. M. Modlin, "IL1beta- and LPS-induced serotonin secretion is increased in EC cells derived from Crohn's disease," *Neurogastroenterology & Motility*, vol. 21, no. 4, pp. 439–450, 2009.
- [71] J. M. Yano, K. Yu, G. P. Donaldson et al., "Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis," *Cell*, vol. 161, no. 2, pp. 264–276, 2015.
- [72] N. K. McVey, J. Bienenstock, A. Bharwani et al., "Oral selective serotonin reuptake inhibitors activate vagus nerve dependent gut-brain signalling," *Scientific Reports*, vol. 9, no. 1, p. 14290, 2019.
- [73] K. G. Margolis, J. F. Cryan, and E. A. Mayer, "The microbiota-gut-brain axis: from motility to mood," *Gastroenterology*, vol. 160, no. 5, pp. 1486–1501, 2021.
- [74] W. J. Huffman, S. Subramaniam, R. M. Rodriguiz, W. C. Wetsel, W. M. Grill, and N. Terrando, "Modulation of neuroinflammation and memory dysfunction using percutaneous vagus nerve stimulation in mice," *Brain Stimulation*, vol. 12, no. 1, pp. 19–29, 2019.
- [75] A. V. Catarina, G. Branchini, L. Bettoni, J. R. De Oliveira, and F. B. Nunes, "Sepsis-associated encephalopathy: from pathophysiology to progress in experimental studies," *Molecular Neurobiology*, vol. 58, no. 6, pp. 2770–2779, 2021.
- [76] S. Cussotto, K. V. Sandhu, T. G. Dinan, and J. F. Cryan, "The neuroendocrinology of the microbiota-gut-brain axis: a behavioural perspective," *Frontiers in Neuroendocrinology*, vol. 51, pp. 80–101, 2018.
- [77] M. Rosas-Ballina and K. J. Tracey, "Cholinergic control of inflammation," *Journal of Internal Medicine*, vol. 265, no. 6, pp. 663–679, 2009.
- [78] K. J. Tracey, "Reflex control of immunity," *Nature Reviews Immunology*, vol. 9, no. 6, pp. 418–428, 2009.
- [79] L. Venkatasamy, D. Nizamutdinov, J. Jenkins, and L. A. Shapiro, "Vagus nerve stimulation ameliorates cognitive impairment and increased hippocampal astrocytes in a mouse model of gulf war illness," *Neurosci Insights*, vol. 16, p. 2006320936, 2021.
- [80] Y. Ano, R. Ohya, T. Yamazaki et al., "Hop bitter acids containing a beta-carbonyl moiety prevent inflammation-induced cognitive decline via the vagus nerve and noradrenergic system," *Scientific Report*, vol. 10, no. 1, p. 20028, 2020.
- [81] J. F. Thayer and E. M. Sternberg, "Neural aspects of immunomodulation: focus on the vagus nerve," *Brain Behaviour Immunity*, vol. 24, no. 8, pp. 1223–1228, 2010.
- [82] B. Peeters, P. Meersseman, P. S. Vander et al., "ACTH and cortisol responses to CRH in acute, subacute, and prolonged critical illness: a randomized, double-blind, placebo-controlled, crossover cohort study," *Intensive Care Medicine*, vol. 44, no. 12, pp. 2048–2058, 2018.
- [83] N. N. Santos-Junior, L. Costa, C. Catalao, A. Kanashiro, T. Sharshar, and M. Rocha, "Impairment of osmotic challenge-induced neurohypophysial hormones secretion in sepsis survivor rats," *Pituitary*, vol. 20, no. 5, pp. 515–521, 2017.
- [84] S. Pan, Y. Wu, L. Pei et al., "BML-111 reduces neuroinflammation and cognitive impairment in mice with sepsis via the SIRT1/NF-kappaB signaling pathway," *Frontier In Cellular Neuroscience*, vol. 12, p. 267, 2018.
- [85] D. B. Hoover, "Cholinergic modulation of the immune system presents new approaches for treating inflammation," *Pharmacology & Therapeutics*, vol. 179, pp. 1–16, 2017.

- [86] G. R. Johnston and N. R. Webster, "Cytokines and the immunomodulatory function of the vagus nerve," *British Journal of Anaesthesia*, vol. 102, no. 4, pp. 453–462, 2009.
- [87] Q. Zhai, D. Lai, P. Cui et al., "Selective activation of basal forebrain cholinergic neurons attenuates polymicrobial sepsis-induced inflammation via the cholinergic anti-inflammatory pathway," *Crit Care Medicine*, vol. 45, no. 10, pp. e1075–e1082, 2017.
- [88] S. Z. Zhu, W. P. Huang, L. Q. Huang et al., "Huperzine A protects sepsis associated encephalopathy by promoting the deficient cholinergic nervous function," *Neuroscience Letters*, vol. 631, pp. 70–78, 2016.
- [89] T. Barichello, J. J. Fortunato, A. M. Vitali et al., "Oxidative variables in the rat brain after sepsis induced by cecal ligation and perforation," *Critical Care Medicine*, vol. 34, no. 3, pp. 886–889, 2006.
- [90] X. Sun, R. Zhou, Y. Lei, J. Hu, and X. Li, "The ligand-gated ion channel P2X7 receptor mediates NLRP3/caspase-1-mediated pyroptosis in cerebral cortical neurons of juvenile rats with sepsis," *Brain Research*, vol. 1748, article 147109, 2020.
- [91] X. E. Xu, L. Liu, Y. C. Wang et al., "Caspase-1 inhibitor exerts brain-protective effects against sepsis-associated encephalopathy and cognitive impairments in a mouse model of sepsis," *Brain Behaviour Immunity*, vol. 80, pp. 859–870, 2019.
- [92] L. G. Danielski, A. D. Giustina, M. P. Goldim et al., "Vitamin B6 reduces neurochemical and long-term cognitive alterations after polymicrobial sepsis: involvement of the kynurenine pathway modulation," *Molecular Neurobiology*, vol. 55, no. 6, pp. 5255–5268, 2018.
- [93] M. Harland, S. Torres, J. Liu, and X. Wang, "Neuronal mitochondria modulation of LPS-induced neuroinflammation," *Journal Neuroscience*, vol. 40, no. 8, pp. 1756–1765, 2020.
- [94] L. Zhang, Y. Jiang, S. Deng et al., "S100B/RAGE/Ceramide signaling pathway is involved in sepsis-associated encephalopathy," *Life Science*, vol. 277, p. 119490, 2021.
- [95] L. Castro, C. F. Goncalves-de-Albuquerque, and A. R. Silva, "Polarization of microglia and its therapeutic potential in sepsis," *International Journal of Molecular Sciences*, vol. 23, no. 9, 2022.
- [96] A. Cugurra, T. Mamuladze, J. Rustenhoven et al., "Skull and vertebral bone marrow are myeloid cell reservoirs for the meninges and CNS parenchyma," *Science*, vol. 373, no. 6553, 2021.
- [97] L. D. Estrada, L. Oliveira-Cruz, and D. Cabrera, "Transforming growth factor beta type I role in neurodegeneration: implications for Alzheimer's disease," *Current Protein and Peptide Science*, vol. 19, no. 12, pp. 1180–1188, 2018.
- [98] K. Biber, H. Neumann, K. Inoue, and H. W. Boddeke, "Neuronal 'On' and 'Off' signals control microglia," *Trends Neuroscience*, vol. 30, no. 11, pp. 596–602, 2007.
- [99] A. Nimmerjahn, F. Kirchhoff, and F. Helmchen, "Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo," *Science*, vol. 308, no. 5726, pp. 1314–1318, 2005.
- [100] R. C. Paolicelli, G. Bolasco, F. Pagani et al., "Synaptic pruning by microglia is necessary for normal brain development," *Science*, vol. 333, no. 6048, pp. 1456–1458, 2011.
- [101] A. Sierra, J. M. Encinas, J. J. Deudero et al., "Microglia shape adult hippocampal neurogenesis through apoptosis-coupled phagocytosis," *Cell Stem Cell*, vol. 7, no. 4, pp. 483–495, 2010.
- [102] A. Bessis, C. Bechade, D. Bernard, and A. Roumier, "Microglial control of neuronal death and synaptic properties," *GLIA*, vol. 55, no. 3, pp. 233–238, 2007.
- [103] E. Polazzi and B. Monti, "Microglia and neuroprotection: from in vitro studies to therapeutic applications," *Progress in Neurobiology*, vol. 92, no. 3, pp. 293–315, 2010.
- [104] M. Li, J. Hu, Y. Peng, J. Li, and R. Ren, "CircPTK2-miR-181c-5p-HMGB1: a new regulatory pathway for microglia activation and hippocampal neuronal apoptosis induced by sepsis," *Molecular Medicine*, vol. 27, no. 1, p. 45, 2021.
- [105] K. Haruwaka, A. Ikegami, Y. Tachibana et al., "Dual microglia effects on blood brain barrier permeability induced by systemic inflammation," *Nature Communications*, vol. 10, no. 1, p. 5816, 2019.
- [106] S. C. Tauber, M. Djukic, J. Gossner, H. Eiffert, W. Bruck, and R. Nau, "Sepsis-associated encephalopathy and septic encephalitis: an update," *Expert Review of Anti-Infective Therapy*, vol. 19, no. 2, pp. 215–231, 2021.
- [107] A. Verkhratsky and M. Nedergaard, "Physiology of astroglia," *Physiological Reviews*, vol. 98, no. 1, pp. 239–389, 2018.
- [108] C. Farina, F. Aloisi, and E. Meinl, "Astrocytes are active players in cerebral innate immunity," *Trends in Immunology*, vol. 28, no. 3, pp. 138–145, 2007.
- [109] Y. Dong and E. N. Benveniste, "Immune function of astrocytes," *GLIA*, vol. 36, no. 2, pp. 180–190, 2001.
- [110] N. Chaudhry and A. K. Duggal, "Sepsis associated encephalopathy," *Advances in Medicine*, vol. 2014, Article ID 762320, 2014.
- [111] Y. B. Sun, H. Zhao, D. L. Mu et al., "Dexmedetomidine inhibits astrocyte pyroptosis and subsequently protects the brain in in vitro and in vivo models of sepsis," *Cell Death & Disease*, vol. 10, no. 3, p. 167, 2019.
- [112] S. Hasegawa-Ishii, M. Inaba, H. Umegaki, K. Unno, K. Wakabayashi, and A. Shimada, "Endotoxemia-induced cytokine-mediated responses of hippocampal astrocytes transmitted by cells of the brain-immune interface," *Scientific Report*, vol. 6, p. 25457, 2016.
- [113] H. Chen, B. Dong, Y. Shi, Y. Yu, and K. Xie, "Hydrogen alleviates neuronal injury and neuroinflammation induced by microglial activation via the nuclear factor erythroid 2-related factor 2 pathway in sepsis-associated encephalopathy," *Neuroscience*, vol. 466, pp. 87–100, 2021.
- [114] Q. Han, Q. Lin, P. Huang et al., "Microglia-derived IL-1beta contributes to axon development disorders and synaptic deficit through p38-MAPK signal pathway in septic neonatal rats," *Journal Neuroinflammation*, vol. 14, no. 1, p. 52, 2017.
- [115] T. Xiao, H. Ji, X. Shangguan, S. Qu, Y. Cui, and J. Xu, "NLRP3 inflammasome of microglia promotes A1 astrocyte transformation, neo-neuron decline and cognition impairment in endotoxemia," *Biochemical and Biophysical Research Communications*, vol. 602, pp. 1–7, 2022.
- [116] J. Shi, H. Xu, M. J. Cavagnaro, X. Li, and J. Fang, "Blocking HMGB1/RAGE signaling by berberine alleviates A1 astrocyte and attenuates sepsis-associated encephalopathy," *Frontiers in Pharmacology*, vol. 12, p. 760186, 2021.
- [117] S. Satarker, S. L. Bojja, P. C. Gurram, J. Mudgal, D. Arora, and M. Nampoothiri, "Astrocytic glutamatergic transmission and its implications in neurodegenerative disorders," *Cells-Basel*, vol. 11, no. 7, 2022.
- [118] Y. Lei, R. Zhou, X. Sun et al., "The pannexin-1 channel regulates pyroptosis through autophagy in a mouse model of

- sepsis-associated encephalopathy," *Annals Translational Medicine*, vol. 9, no. 24, p. 1802, 2021.
- [119] Z. Xie, M. Xu, J. Xie et al., "Inhibition of ferroptosis attenuates glutamate excitotoxicity and nuclear autophagy in a CLP septic mouse model," *Shock*, vol. 57, no. 5, pp. 694–702, 2022.
- [120] M. Li, Y. Zhang, and J. Wang, "Endoplasmic reticulum stress regulates cell injury in lipopolysaccharide-induced nerve cells," *Journal International Medical Research*, vol. 48, no. 9, p. 1220749314, 2020.
- [121] R. Zhou, J. Ying, X. Qiu et al., "A new cell death program regulated by toll-like receptor 9 through p38 mitogen-activated protein kinase signaling pathway in a neonatal rat model with sepsis associated encephalopathy," *Chinese Medical Journal*, 2022.
- [122] S. Xi, Y. Wang, C. Wu, W. Peng, Y. Zhu, and W. Hu, "Intestinal epithelial cell exosome launches IL-1beta-mediated neuron injury in sepsis-associated encephalopathy," *Frontier Cell Infection Microbiology*, vol. 11, p. 783049, 2021.
- [123] T. Sharshar, D. Annane, G. L. de la Grandmaison, J. P. Brouland, N. S. Hopkinson, and G. Francoise, "The neuropathology of septic shock," *Brain Pathology*, vol. 14, no. 1, pp. 21–33, 2004.
- [124] C. S. Burkhart, M. Siegemund, and L. A. Steiner, "Cerebral perfusion in sepsis. CRIT," *Care*, vol. 14, no. 2, p. 215, 2010.
- [125] T. Csipo, B. R. Cassidy, P. Balasubramanian, D. A. Drevets, Z. I. Ungvari, and A. Yabluchanskiy, "Endothelial dysfunction and impaired neurovascular coupling responses precede cognitive impairment in a mouse model of geriatric sepsis," *Frontier Aging Neuroscience*, vol. 13, article 644733, 2021.
- [126] B. Rosengarten, S. Wolff, S. Klatt, and R. T. Schermuly, "Effects of inducible nitric oxide synthase inhibition or norepinephrine on the neurovascular coupling in an endotoxic rat shock model," *Critical Care*, vol. 13, no. 4, p. R139, 2009.
- [127] Y. H. Cui, S. F. Zhou, Y. Liu et al., "Injection of anti-proBDNF attenuates hippocampal-dependent learning and memory dysfunction in mice with sepsis-associated encephalopathy," *Frontier Neuroscience*, vol. 15, p. 665757, 2021.
- [128] M. G. Granja, L. P. Alves, M. Leardini-Tristao et al., "Inflammatory, synaptic, motor, and behavioral alterations induced by gestational sepsis on the offspring at different stages of life," *Journal Neuroinflammation*, vol. 18, no. 1, p. 60, 2021.
- [129] M. Rocha, A. Vieira, M. Michels et al., "Effects of S100B neutralization on the long-term cognitive impairment and neuroinflammatory response in an animal model of sepsis," *Neurochemistry International*, vol. 142, article 104906, 2021.
- [130] S. Hasegawa-Ishii, M. Inaba, and A. Shimada, "Widespread time-dependent changes in tissue cytokine concentrations in brain regions during the acute phase of endotoxemia in mice," *Neurotoxicology*, vol. 76, pp. 67–74, 2020.
- [131] A. M. Kressel, T. Tsaava, Y. A. Levine et al., "Identification of a brainstem locus that inhibits tumor necrosis factor," *Proceedings of the National Academy of Sciences*, vol. 117, no. 47, pp. 29803–29810, 2020.
- [132] M. Y. Sallam, S. M. El-Gowilly, M. A. Fouda, M. M. Abd-Alhaseeb, and M. M. El-Mas, "Brainstem cholinergic pathways diminish cardiovascular and neuroinflammatory actions of endotoxemia in rats: Role of NFkappaB/alpha7/alpha4beta2AChRs signaling," *Neuropharmacology*, vol. 157, p. 107683, 2019.
- [133] Y. Kadoi, S. Saito, F. Kunimoto, T. Imai, and T. Fujita, "Impairment of the brain beta-adrenergic system during experimental endotoxemia," *Journal Surgical Research*, vol. 61, no. 2, p. 496–502, 1996.
- [134] M. C. Barbosa-Silva, M. N. Lima, D. Battaglini et al., "Infectious disease-associated encephalopathies. CRIT," *CARE*, vol. 25, no. 1, p. 236, 2021.
- [135] J. L. Stollings, K. Kottis, G. Chanques, B. T. Pun, P. P. Pandharipande, and E. W. Ely, "Delirium in critical illness: clinical manifestations, outcomes, and management," *Intensive Care Medicine*, vol. 47, no. 10, pp. 1089–1103, 2021.
- [136] B. Mei, J. Li, and Z. Zuo, "Dexmedetomidine attenuates sepsis-associated inflammation and encephalopathy via central alpha2A adrenoceptor," *Brain Behaviour Immunity*, vol. 91, pp. 296–314, 2021.
- [137] D. B. Hoover, M. D. Poston, S. Brown et al., "Cholinergic leukocytes in sepsis and at the neuroimmune junction in the spleen," *International Immunopharmacology*, vol. 81, p. 106359, 2020.
- [138] M. H. Ji, L. Zhang, M. J. Mao, H. Zhang, J. J. Yang, and L. L. Qiu, "Overinhibition mediated by parvalbumin interneurons might contribute to depression-like behavior and working memory impairment induced by lipopolysaccharide challenge," *Behavioural Brain Research*, vol. 383, p. 112509, 2020.
- [139] F. Li, B. Zhang, S. Duan et al., "Small dose of L-dopa/Benserazide hydrochloride improved sepsis-induced neuroinflammation and long-term cognitive dysfunction in sepsis mice," *Brain Research*, vol. 1737, article 146780, 2020.
- [140] F. S. Taccone, F. Su, C. Pierrakos et al., "Cerebral microcirculation is impaired during sepsis: an experimental study," *Critical Care*, vol. 14, no. 4, p. R140, 2010.
- [141] E. Iacobone, J. Bailly-Salin, A. Polito, D. Friedman, R. D. Stevens, and T. Sharshar, "Sepsis-associated encephalopathy and its differential diagnosis," *Critical Care Medicine*, vol. 37, 10 Supplement, pp. S331–S336, 2009.
- [142] O. Urdanibia-Centelles, R. M. Nielsen, E. Rostrup et al., "Automatic continuous EEG signal analysis for diagnosis of delirium in patients with sepsis," *Clinical Neurophysiology*, vol. 132, no. 9, pp. 2075–2082, 2021.
- [143] N. D. Pantzaris, C. Platanaki, K. Tsiotsios, I. Koniari, and D. Velissaris, "The use of electroencephalography in patients with sepsis: a review of the literature," *Journal of Translational Internal Medicine*, vol. 9, no. 1, pp. 12–16, 2021.
- [144] Y. Ma, T. Matsuwaki, K. Yamanouchi, and M. Nishihara, "Glucocorticoids suppress the protective effect of cyclooxygenase-2-related signaling on hippocampal neurogenesis under acute immune stress," *Molecular Neurobiology*, vol. 54, no. 3, pp. 1953–1966, 2017.
- [145] D. Agac, L. D. Estrada, R. Maples, L. V. Hooper, and J. D. Farrar, "The beta2-adrenergic receptor controls inflammation by driving rapid IL-10 secretion," *Brain Behaviour Immunity*, vol. 74, pp. 176–185, 2018.
- [146] D. W. Choi, J. H. Park, S. Y. Lee, and S. H. An, "Effect of hypothermia treatment on gentamicin pharmacokinetics in neonates with hypoxic-ischaemic encephalopathy: a systematic review and meta-analysis," *Journal of Clinical Pharmacy and Therapeutics*, vol. 43, no. 4, pp. 484–492, 2018.
- [147] C. M. Comim, V. Freiberger, L. Ventura et al., "Inhibition of indoleamine 2,3-dioxygenase 1/2 prevented cognitive impairment and energetic metabolism changes in the hippocampus of adult rats subjected to polymicrobial sepsis," *Journal Neuroimmunology*, vol. 305, pp. 167–171, 2017.

- [148] Y. Li, F. Wang, and Y. Luo, "Ginsenoside Rg1 protects against sepsis-associated encephalopathy through beclin 1-independent autophagy in mice," *Journal Surgical Research*, vol. 207, pp. 181–189, 2017.
- [149] S. McGrane, T. D. Girard, J. L. Thompson et al., "Procalcitonin and C-reactive protein levels at admission as predictors of duration of acute brain dysfunction in critically ill patients," *Critical Care*, vol. 15, no. 2, p. R78, 2011.
- [150] J. Ehler, T. Saller, M. Wittstock et al., "Diagnostic value of NT-proCNP compared to NSE and S100B in cerebrospinal fluid and plasma of patients with sepsis-associated encephalopathy," *Neuroscience Letters*, vol. 692, pp. 167–173, 2019.
- [151] B. A. Khan, A. J. Perkins, N. K. Prasad et al., "Biomarkers of delirium duration and delirium severity in the ICU," *Critical Care Medicine*, vol. 48, no. 3, pp. 353–361, 2020.
- [152] K. S. Simons, M. van den Boogaard, E. Hendriksen et al., "Temporal biomarker profiles and their association with ICU acquired delirium: a cohort study," *Critical Care*, vol. 22, no. 1, p. 137, 2018.
- [153] R. Zhou, Y. Qu, Q. Huang, X. Sun, D. Mu, and X. Li, "Recombinant CC16 regulates inflammation, oxidative stress, apoptosis and autophagy via the inhibition of the p38MAPK signaling pathway in the brain of neonatal rats with sepsis," *Brain Research*, vol. 1725, article 146473, 2019.