

Review Article

Psoriasis between Autoimmunity and Oxidative Stress: Changes Induced by Different Therapeutic Approaches

Marija V. Medovic ^{1,2} **Vladimir Lj. Jakovljevic** ^{3,4} **Vladimir I. Zivkovic** ³
Nevena S. Jeremic ⁵ **Jovana N. Jeremic** ⁵ **Sergey B. Bolevich** ⁴
Ana B. Ravic Nikolic ^{1,2} **Vesna M. Milicic** ^{1,2} and **Ivan M. Srejavic** ³

¹Department of Dermatovenerology, University of Kragujevac, Faculty of Medical Sciences, Svetozara Markovica 69, 34000 Kragujevac, Serbia

²University Clinical Center Kragujevac, Zmaj Jovina 30, 34000 Kragujevac, Serbia

³Department of Physiology, University of Kragujevac, Faculty of Medical Sciences, Svetozara Markovica 69, 34000 Kragujevac, Serbia

⁴I.M. Sechenov First Moscow State Medical University, Department of Human Pathophysiology, Moscow, Russian Federation, Trubetskaya Str. 2, 119992 Moscow, Russia

⁵Department of Pharmacy, University of Kragujevac, Faculty of Medical Sciences, Svetozara Markovica 69, 34000 Kragujevac, Serbia

Correspondence should be addressed to Vladimir Lj. Jakovljevic; drvladakbg@yahoo.com

Received 22 October 2021; Accepted 15 February 2022; Published 11 March 2022

Academic Editor: Dragan Hrnčić

Copyright © 2022 Marija V. Medovic et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Psoriasis is defined as chronic, immune-mediated disease. Regardless of the development of new therapeutic approaches, the precise etiology of psoriasis remains unknown and speculative. The aim of this review was to systematize the results of previous research on the role of oxidative stress and aberrant immune response in the pathogenesis of psoriasis, as well as the impact of certain therapeutic modalities on the oxidative status in patients with psoriasis. Complex immune pathways of both the innate and adaptive immune systems appear to be major pathomechanisms in the development of psoriasis. Oxidative stress represents another important contributor to the pathophysiology of disease, and the redox imbalance in psoriasis has been reported in skin cells and, systemically, in plasma and blood cells, and more recently, also in saliva. Current immune model of psoriasis begins with activation of immune system in susceptible person by some environmental factor and loss of immune tolerance to psoriasis autoantigens. Increased production of IL-17 appears to be the most prominent role in psoriasis pathogenesis, while IL-23 is recognized as master regulator in psoriasis having a specific role in cross bridging the production of IL-17 by innate and acquired immunity. Other proinflammatory cytokines, including IFN- γ , TNF- α , IL-1 β , IL-6, IL-22, IL-26, IL-29, or IL-36, have also been reported to play important roles in the development of psoriasis. Oxidative stress can promote inflammation through several signaling pathways. The most noticeable and most powerful antioxidative effects exert various biologics compared to more convenient therapeutic modalities, such as methotrexate or phototherapy. The complex interaction of redox, immune, and inflammatory signaling pathways should be focused on further researches tackling the pathophysiology of psoriasis, while antioxidative supplementation could be the solution in some refractory cases of the disease.

1. Introduction

Psoriasis was firstly described in detail by Robert Willan, founder of dermatology as a medical specialty [1]. The prevalence of psoriasis varies from 0.51% to 11.43% in adults,

making psoriasis one of the most important global health problems [2]. According to the World Health Organization, psoriasis is classified as one of the most serious noninfectious diseases, due to complications that develop during the course of the disease and affection of multiple organ

systems [3, 4]. Psoriasis is defined as chronic, inflammatory, recurrent, incurable, and noncontagious disease, characterized by sharply demarcated erythematous skin lesions with overlying silver hyperkeratotic plaques, accompanied by systemic manifestations [3].

1.1. Clinical Presentation of Psoriasis. The most distinctive characteristic of psoriasis are well-defined, symmetric, raised skin lesions most commonly located on the knees, elbows, scalp, and trunk [5]. Such clinical presentation is a characteristic for plaque psoriasis or psoriasis vulgaris, the most common form of psoriasis, but psoriasis also may appear as guttate, pustular (von Zumbusch psoriasis) or erythrodermic psoriasis (Figure 1). Skin changes are usually accompanied by pruritus, itching, pain, cracking, bleeding, and flaking of the skin. Furthermore, psoriasis is recognized as a risk factor for many pathological conditions, including cardiovascular diseases, gastrointestinal disorders, malignant tumors, infections, and mood disorders [6]. The most common comorbidity of psoriasis is psoriatic arthritis, usually defined as heterogeneous inflammatory arthritis which affects joints and enthesal tissues [7]. Such interconnection of psoriasis and the variety of comorbidities probably arises from, on the one hand, complex etiological and pathophysiological basis of the disease, and on the other hand, the fact that psoriasis remains unrecognized and untreated for a long period [8, 9].

Psoriasis vulgaris is generally equally present among the sexes, but it develops somewhat earlier in women than in men [3]. The severity of psoriasis is usually classified using Psoriasis Area and Severity Index (PASI) and Body Surface Area (BSA) [10]. Due to the common association of psoriasis and psoriatic arthritis, several screening questionnaires are performed for early recognition of psoriatic arthritis, such as Psoriasis and Arthritis Screening Questionnaire (PASQ), Psoriasis Epidemiology Screening Tool (PEST), and Toronto Psoriatic Arthritis Screen (ToPAS) [11].

1.2. Main Pathophysiological Features of Psoriasis. Psoriasis is defined as chronic, immune-mediated disease. Regardless of the development of new therapeutic approaches, the precise etiology of psoriasis remains unknown and speculative. Complex immune pathways of both the innate and adaptive immune systems appear to be pathophysiological basis in the development of psoriasis [12, 13]. Furthermore, epidemiological investigations indicated the importance of genetic component in the development of psoriasis [14]. Firstly, it was noticed that psoriasis concordance is higher in monozygotic twins [15]. Recent genetic studies identified almost 60 psoriasis susceptibility loci which could interfere with the development of psoriasis [14]. Various environmental factors, such as stress, mechanical trauma, and streptococcal infections are considered to trigger and not cause the disease. In predisposed persons, various factors may provoke the onset of disease or exacerbation of existing symptoms.

Due to the complex pathophysiological mechanisms that affect various tissues and organs, psoriasis may be defined as a systemic disorder with the predominant skin representation. Cutaneous psoriatic manifestations occur as a result

of disruption of skin homeostasis due to immune-mediated aberrant differentiation of keratinocytes [12]. Interleukin-(IL-) 23 and IL-17 are recognized as key immune mediators that mediate not only the development of skin lesions but also the occurrence of psoriasis-associated comorbidities [3]. Skin lesions, followed by extracutaneous comorbidities and chronic course of the disease, significantly affect the quality of life of the psoriasis-suffering patients resulting in the development of anxiety and depression [16].

Psoriasis is a chronic, incurable disease, so treatment of psoriasis can reduce skin lesions, but not provide a complete cure. The psoriatic therapy varies depending on the surface of affected skin—PASI [17]. The most effective therapeutics in psoriasis treatment are biologic therapies, recently developed drugs that target some immune pathway. Drugs from this group act in different ways and target diverse cytokines: tumor necrosis factor α (TNF- α) (adalimumab, certolizumab pegol, etanercept, and infliximab), IL-12/IL-23p40 (ustekinumab), IL-17A (ixekizumab, secukinumab), IL-17 receptor (brodalumab), and IL-23p19 (guselkumab, risankizumab, and tildrakizumab) [18] (Table 1). Biologics are usually used in severe cases of psoriasis. Mild disease is treated with topical preparations combined with phototherapy, and moderate psoriasis is treated with immunomodulatory therapy.

Reactive oxygen species (ROS) represent important regulators of immune response [19]. The imbalance in production and elimination of ROS results in oxidative stress and, consequently, oxidative damage of various cellular structures. Proinflammatory processes involved in the development of autoimmune disorders are combined with increased production of ROS and oxidative stress. Redox imbalance in psoriasis exists both in skin cells and, systemically, in plasma and blood cells [20]. Thus, ROS and oxidative stress appear to be an important step in the psoriasis pathophysiological cascade. The aim of this review is to show the relationship between redox balance and applied therapy in patients suffering from psoriasis.

1.3. Autoimmune Nature of Psoriasis. The main feature of autoimmune disorders is destruction of healthy tissues by the host immune system upon misidentification and recognition of its own tissue structures as foreign. The results of the immunological studies in the recent decades showed involvement of both innate and acquired immunity and importance of T cells in pathogenesis of psoriasis [21–23]. T helper (Th) cells are classified as Th1 cells, which predominantly secrete TNF- α , interferon- (IFN-) γ , and IL-2, and Th2 cells, which secrete IL-4, IL-5, IL-10, and IL-13 [24]. Determination of naïve T cells toward Th1 or Th2 subpopulation depends on the stimulation by IL-12, which mediates Th1 differentiation, or IL-2, which mediates Th2 differentiation [25]. Due to increased levels of TNF- α and IL-12 in psoriatic lesions, psoriasis is defined as Th1-mediated disease [26]. IL-23 is another important cytokine in the development of psoriasis because it shifts the differentiation of naïve T cells toward proinflammatory Th17 cells [27]. These findings are the backbone of the development of new therapeutic approaches such as biologic therapies. Apart from T cells, as



FIGURE 1: Continued.

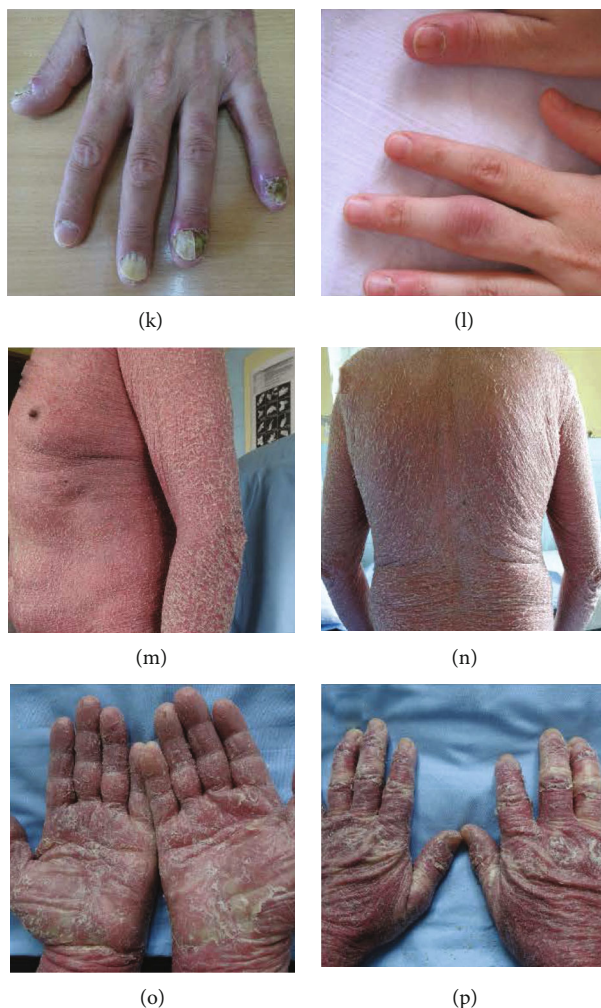


FIGURE 1: Clinical manifestations of psoriasis. Psoriasis vulgaris manifested through typical erythematous plaques with silvery scales of various sizes from guttate lesions through nummular to giant lumbar plaque (a). Psoriasis vulgaris and guttate psoriasis (b, d). Generalized psoriasis (c, h, i). Inverse psoriasis (g). Psoriatic lesions on the feet and hands (e, f, j, k), including psoriatic nail dystrophy (f, k) and psoriatic arthritis (l). Erythrodermic psoriasis, erythema, and squamous cover 90% of the skin (m, n, o, p). Pictures presented are part of the private collection of Vesna M. Milicic and Ana B. Ravic Nikolic.

TABLE 1: Biologics in psoriasis treatment.

Name of the drug	Biological target
Ustekinumab	IL-12 and IL-23—p40 subunit
Guselkumab	
Tildrakizumab	IL-23—p19 subunit
Risankizumab	
Secukinumab	
Ixekizumab	IL-17
Bimekizumab	
Brodalumab	IL-17 receptor
Etanercept	
Infliximab	
Adalimumab	TNF- α
Certolizumab pegol	
Golimumab	

representatives of acquired immunity, neutrophils, dendritic antigen presenting cells (APCs), and Natural Killer T (NKT) cells in pathogenesis of psoriasis play an important role [28]. One of the first described histopathological hallmarks of psoriasis was Munro's microabscesses containing neutrophils [29]. Increased neutrophil activity in psoriatic plaques results in increased ROS production [30] (Figure 2, Table 2). ROS act as stimulators of dendritic APCs to present antigens to T cells, which further stimulate proliferation of keratinocytes [29]. ROS also have a role of second messengers in inflammatory signaling cascades involving activation of mitogen-activated protein kinase (MAPK), nuclear factor-kappa B (NF- κ B). NKT cells encompass heterologous cells which share some features of the natural killer (NK) and T cells. In psoriatic patients, NKT cells interact with other components of immune system, thereby supporting the creation of proinflammatory environment. Subset of NKT cells, NKT17 cells, can produce IL-17, and they are found in many tissues, among others in the skin, but their

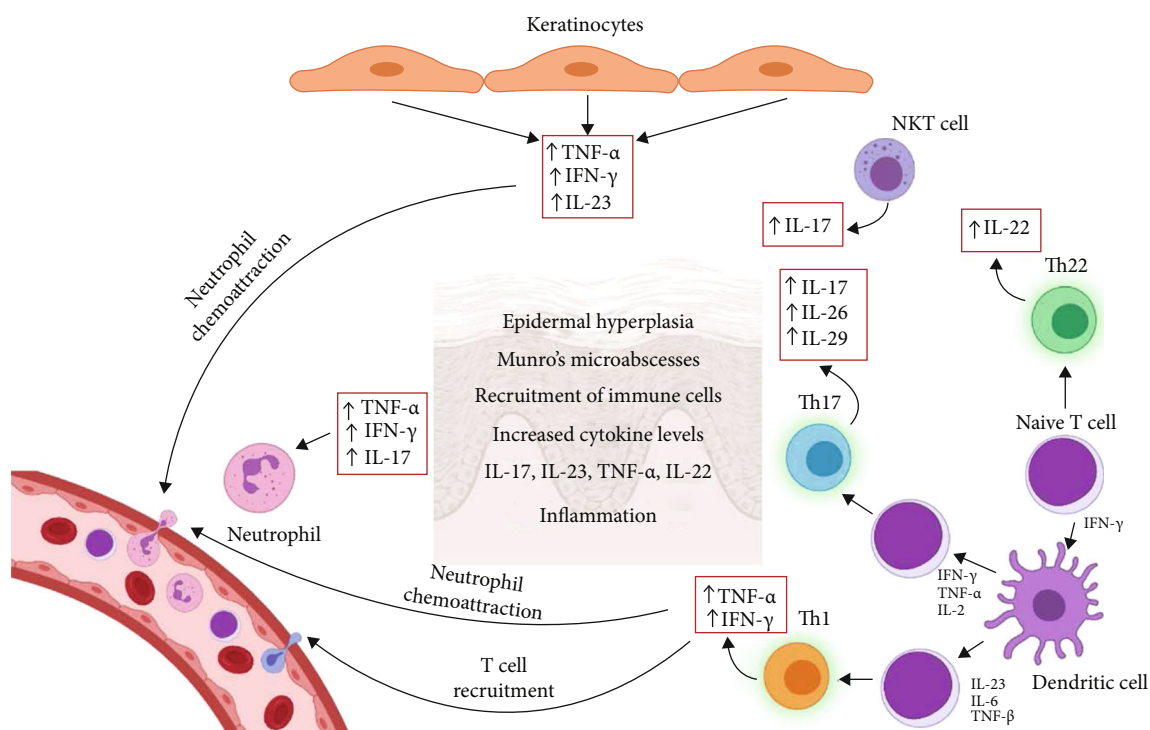


FIGURE 2: Immune response in psoriasis. Complex interaction of various parts of innate (dendritic cells, NKT cells, and neutrophils) and acquired immunity (T cells) in pathophysiology of psoriasis. IFN: interferon; IL: interleukin; NKT: natural killer T; Th: T-helper cell; TGF: transforming growth factor; TNF: tumour necrosis factor.

TABLE 2: Main articles included in the summary of signaling cascades shown in Figure 2.

Article	Main findings
Marble et al. <i>J Dermatol Sci.</i> 2007; 48(2):87-101. [26]	CD11c+ dendritic cells, CD68+ macrophages and TNF- α + cells are increased in psoriatic lesions
Ten Bergen et al. <i>Scand J Immunol.</i> 2020; 92(4):e12946. [27]	TNF- α /IL-23/IL-17 axis appears to have central role in the pathophysiology of psoriasis
Bos et al. <i>Br J Dermatol.</i> 2005; 152(6):1098-107. [28]	Activation of innate immunity, represented by the activity of NKT cells, dendritic cells, neutrophils, and keratinocytes, is crucial in pathogenesis of psoriasis.
Chiang et al. <i>Front Immunol.</i> 2019; 10:2376. [29]	The abundant presence of neutrophils in the psoriatic skin lesions and formation of Munro's microabscesses serves as a typical histopathologic hallmark of psoriasis.
Kim and Krueger. <i>Dermatol Clin.</i> 2015; 33(1):13-23. [13]	Keratinocytes appear to be important regulators of immune responses, involved in increased production of TNF- α or IFN- γ in psoriasis.
Hawkes et al. <i>J Immunol.</i> 2018; 201(6):1605-1613. [34]	Psoriasis is characterized by the presence of multiple T lymphocyte subsets (Th1, Th17, and Th22).
Stephen-Victor et al. <i>PLoS Pathog.</i> 2016; 12(6):e1005624 [37]	Th17 cells, besides IL-17, produce TNF- α , IL-26, and IL-29, which further stimulate release of proinflammatory mediators.
Wang et al. <i>J Cell Mol Med.</i> 2019; 23(12):7926-7932. [38]	
Levin and Gottlieb. <i>J Am Acad Dermatol.</i> 2014; 70(3):555-61. [46]	IL-23 is recognized as a key regulator in psoriasis due to specific role in cross bridging the production of IL-17 by innate and acquired immunity.
Langrish et al. <i>Immunol Rev.</i> 2004; 202:96-105. [47]	

role in the pathogenesis of psoriasis is not fully understood [31, 32].

Keratinocytes play an important role in physiological orchestration of innate and acquired immune responses in the skin due to various pathological stimuli. Injured keratinocytes may produce TNF- α and IFN- γ [13]. TNF- α induces increased neutrophil accumulation in injured skin, and IFN- γ increases recruitment of Th1 cells. Psoriatic plaques contain increased number of T lymphocytes which produce IFN- γ , IL-17, and IL-22, labeled as Th1, Th17, and Th22 cells, respectively [13].

Dendritic APCs may have a crucial role in pathogenesis of psoriasis due to the activation of T cells and production of various proinflammatory cytokines. Population of skin dendritic cells include epidermal dendritic cells (Langerhans cells) and dermal dendritic cells (myeloid and plasmacytoid dendritic cells). CD11c is recognized as the correct marker of myeloid dendritic cells, while blood dendritic cell antigen (BDCA) is used for identification of different human subsets of dendritic cells [33]. Myeloid CD11c⁺BDCA-1⁻ dermal dendritic cells are recognized as proinflammatory dendritic cells which induce activation and clonal expansion of CD4⁺ and CD8⁺ T cells and their stimulation in the production of IFN- γ , IL-17, and IL-22 [34]. Furthermore, it was shown that content of CD11c⁺BDCA-1⁻ dermal dendritic cells is increased in psoriatic lesion, but their number is significantly reduced upon effective therapeutic approach [35].

The current immune model of psoriasis begins with the activation of immune system in susceptible persons by some environmental factor and a loss of immune tolerance to psoriasis autoantigens [34]. TNF- α and IFN- γ , secreted by dendritic cells, induce polarization and expansion of IL-17 and IL-22 secreting T cells (Th17 and Th22 cells), resulting in significant increase of the IL-17 and IL-22 production (Figure 2, Table 2). IL-17 is recognized as the key cytokine in the development of psoriasis. There are six isoforms of IL-17 (IL-17A–IL-17F), whereby IL-17A has the most prominent role in psoriasis pathogenesis [36]. Th17 cells, besides IL-17, produce TNF- α , IL-26, and IL-29. IL-17A, alone or synergistically with TNF- α , induces the release of proinflammatory molecules from keratinocytes and enhances aberrant proliferation of keratinocytes leading to epidermal hyperplasia. Increased production of IL-26 and IL-29 by Th17 stimulate further release of proinflammatory mediators which recruit Th1 cells into psoriatic skin lesions [37, 38]. Increased production of IL-17 results in increased secretion of IL-19, IL-22, and IL-36 which also contribute to the development of epidermal hyperplasia [39, 40]. IL-22 in psoriatic lesions is secreted not only by CD4⁺ and CD8⁺ T cells known as Th22 and Tc22 cells but also by Th17, mast cells, and others [41]. IL-22 enhances migration of keratinocytes, increases epidermal thickness, decreases keratinocyte differentiation, and stimulates secretion of various molecules which act as chemokines, neutrophil chemoattractants [41]. IL-22 proinflammatory action is weaker than IL-17, but IL-22 mainly acts synergistically with IL-17 and TNF- α . Dendritic cells also secrete IL-23 which acts via the IL-23 receptor located on naïve T cells and promotes their differentiation into Th17 [42]. IL-23 belongs to the IL-12 cyto-

kine family. It is a composite cytokine containing two subunits IL-23p19 and IL-12p40 [43]. Acting together with TNF- α , IL-1 β , and IL-6, IL-23 stimulates the differentiation of Th17 and Tc17 cells and the conversion of regulatory T cells (T_{reg}) into Th17 cells [44]. After the differentiation of naïve T cells to Th17 cells due to various stimuli such as transforming growth factor (TGF)- β and IL-6, the presence of IL-23 is necessary to maintain the Th17 phenotype [25, 45]. Binding of IL-23 to its receptor on Th17 cells initiates signaling pathway, which results in the facilitation of IL-17 expression and increased levels of IL-17A in plasma [46]. Besides the action of IL-23 on T cells and induction of them to produce IL-17, thus provoking the inflammatory autoimmune response (acquired immunity), it can also stimulate the production of IL-17 by NK cells and neutrophils (innate immunity) [47, 48]. Thus, IL-23 is recognized as a master regulator in psoriasis having a specific role in cross bridging the production of IL-17 by innate and acquired immunity. The administration of IL-23 in mice induced epidermal hyperplasia and increased expression of both IL-17A and IL-22 [49]. Psoriasis is defined as Th17-mediated disease, but results of a growing number of studies indicate the central role of IL-23 in the development of psoriasis due to its effects on sustention of cytotoxic Th17 cells and production of IL-17 and IL-22 (Figure 2, Table 2). Altogether, various parts of acquired and innate immune system create complex signaling pathways in the development of psoriasis.

1.4. Oxidative Stress in Pathogenesis of Psoriasis. Oxidative stress is usually defined as an imbalance between the production of ROS or reactive nitrogen species (RNS) and antioxidative capacity. Decreased antioxidative ability may be the consequence of decreased activity of antioxidative enzymes (such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx)) or/and decreased concentration of scavenging antioxidants, both endogenous (reduced glutathione (GSH)) and exogenous (vitamin C, vitamin E, carotenoids, and others). At low, physiological concentrations, ROS/RNS have important roles as signaling molecules in regulatory cascades of different biological processes, but excess ROS/RNS and consequent oxidative stress induce oxidation of various cellular structures (DNA, lipids, and proteins) leading to cell death [50, 51].

The skin and thus keratinocytes are continuously exposed to various external stressful stimuli, including ultraviolet (UV) radiation of the sun, and oxygen from the air. It is assumed that more than 50% of skin damage induced by UV radiation is mediated by ROS/RNS [52]. Furthermore, different toxic substances, as well as their metabolites, directly or indirectly initiate the production of various prooxidative molecules in keratinocytes [53]. Oxidative stress can promote inflammation through several signaling pathways including NF- κ B, mitogen-activated protein kinases (MAPKs), and STAT3 (Signal Transducer and Activator of Transcription 3) [54]. MAPKs represent a family of serine-threonine protein kinases encompassing several members: extracellular signal-regulated kinases (ERKs), c-Jun N-terminal kinases (JNKs), and the p38 MAPKs [55]. Increased presence of ROS and impaired antioxidative

potential directly induce increased activation of NF- κ B [56]. Immunohistochemical analysis of psoriatic skin lesions showed increased levels of phosphorylated ERK1/2 and p38 MAPK [57, 58]. It was shown that propranolol induced psoriasis-like skin inflammation through increase of oxidative stress as well as NF- κ B and MAPK p38 activation and subsequent secretion of IL-23 [59]. Results of this research confirmed the crucial role of oxidative stress in IL-23/IL-17 axis of Th17-related psoriasis-like skin inflammation (Figure 3, Table 3). ROS produced in the skin also act as chemoattractant for neutrophils and, furthermore, increased number of neutrophils combined with high levels of ROS may result in the activation of neutrophils and further increase of ROS production [60, 61]. Augmented inflammatory response further facilitates production of ROS and decreases already weakened antioxidative capacity which makes psoriasis a chronic inflammatory disease [62].

The nuclear factor erythroid 2-related factor 2 (Nrf2) is redox-sensitive transcription factor involved in the regulation of keratinocyte proliferation and expression of keratin [63]. There are ambiguous data regarding the role of Nrf2 in pathogenesis of psoriasis. Lee et al. pointed out increased oxidative damage in psoriatic skin lesions combined with decreased expression of Nrf2, while dimethyl fumarate, as an Nrf2 activator, upregulated Nrf2 levels in HaCaT keratinocyte cell line and promoted growth inhibition and apoptosis [64]. The authors assumed that the increased production of ROS and oxidative stress interfere with dysregulation of the Nrf2 signaling cascade. Another study also showed a decrease of ROS and increase of nuclear accumulation of Nrf2 after the application of antioxidant, followed by the reduction of vascular endothelial growth factor (VEGF) and the reduction of keratinocyte proliferation in a similar experimental model [65]. On the other hand, Yang et al. presented increased nuclear-localized Nrf2 in psoriatic epidermis compared to normal skin [63]. Increased expression of Nrf2 was linked with higher expression of psoriasis-related keratins K6, K16, and K17. Furthermore, it was shown that IL-17 and IL-22 enhance the proliferation of psoriasis-related keratins via Nrf2 signaling. It appears that redox signaling and Nrf2 poses divergent functions in the development of psoriasis, probably due to the cellular localization of Nrf2; however, many questions are to be answered further.

Several studies indicated an increased risk for the psoriasis occurrence in persons with polymorphisms of specific genes that are related to the regulation of redox balance (Figure 3, Table 3). Asefi et al. indicated an increased risk for the development of psoriasis in persons bearing 55M allele for paraoxonase 1 (PON1) [66]. PON1 is hydrolytic enzyme bound to high-density lipoprotein (HDL), able to break down lipid peroxides. It is assumed that enzymatic activity of PON1 is crucial for protective effect of HDL. The PON1 55M allele in psoriatic patients was found to be associated with higher malondialdehyde (MDA) levels, apolipoprotein B, and lipoprotein (a), suggesting interference of oxidative stress and disturbances in lipid metabolism in the pathogenesis of psoriasis [66]. Another study also revealed lower PON1 activity in psoriasis-suffering patients due to

PON1 polymorphism related to decreased antioxidative activity and different lipid levels [67]. Glutathione S-transferases (GSTs) are a group of enzymes involved in catalytic regulation of the conjugation of GSH to various substrates, thus providing protection against various detrimental factors including oxidative stress and inflammation. Some polymorphisms of GST genes were significantly more common in patients with psoriasis compared to the healthy population [68]. Furthermore, null polymorphisms for GSTs were related to increased sensitivity psoralen-ultraviolet A (PUVA) photochemotherapy [69]. The activity of SOD also appears to be an important factor in the development of psoriasis. Knock out (KO) of extracellular SOD in mice induced more intense IL-23-mediated skin inflammation characterized with elevated accumulation of CD4⁺ T cells, CD11b⁺ macrophages, and CD11c⁺ dendritic cells accompanied by increased expression of proinflammatory cytokines [70]. Naïve CD4⁺ T cells were more differentiated into the Th17 cell in extracellular SOD KO mice compared to the wild type (WT) controls. The previous study showed decreased activity of SOD and CAT in erythrocytes of psoriatic patients [71]. Levels of MDA were higher in these patients, combined with decreased activity of antioxidative enzymes. An interplay between various components of the immune system, ROS/RNS, and antioxidative system creates intertwined signaling pathways.

Analyzing the differences in various pro- and antioxidants in stimulated and unstimulated saliva of psoriatic patients and healthy individuals, it was shown that several prooxidative markers were increased in patients with plaque psoriasis [72]. Contrary to the previous study [71], antioxidative enzymes were significantly higher not only in saliva but also in erythrocytes, of psoriatic patients compared to healthy control [72]. The same authors also showed significantly higher levels of TNF- α , IFN- γ , and IL-2, nitric oxide (NO), and nitrotyrosine in saliva of psoriatic patients compared to healthy individuals [73]. Furthermore, it was shown that various inflammation-related proteins and microbiota were changed in saliva of psoriatic patients [74, 75]. Thus, oxidative stress biomarkers, such as total oxidative status or oxidative stress index, as well as inflammatory markers could be considered as diagnostic tool in psoriasis.

1.5. Therapeutic Approaches in Psoriasis. Various treatments have been used for psoriasis patients, from topical medications that contain corticosteroids, retinoid derivatives, synthetic vitamin D3 analogues, tar, or anthralin to systemic drugs with a different mechanism of action. However, all therapeutic modalities have transient curative effects, and it is also difficult to predict the occurrence of exacerbations and determine which drug delays their occurrence most effectively. Most of the therapeutics used in psoriasis curation are not suitable for a long-term use due to serious side effects and high costs [54]. Until recently, drugs such as methotrexate, acitretin, cyclosporine, dexamethasone, and salicylic acid were most commonly used in psoriasis treatment. Many of these drugs have limited clinical efficacy due to different shortcomings including low absorption capacity, inconsistent drug release, low target tissue

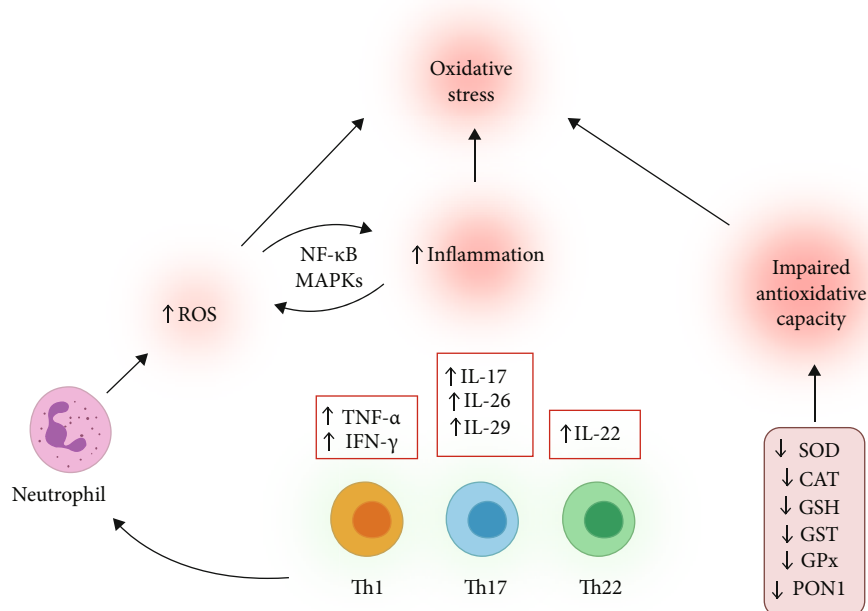


FIGURE 3: Mechanisms involved in increasing oxidative stress in psoriasis and interactions between oxidative stress and inflammation. IFN: interferon; IL: interleukin; MAPKs: mitogen-activated protein kinases, NF- κ B: nuclear factor kappa B; ROS: reactive oxygen species; Th: T-helper cell; TNF: tumour necrosis factor.

TABLE 3: Papers included in the summary of signaling cascades shown in Figure 3.

Article	Main findings
Lai et al. Redox Rep. 2018; 23(1):130-135. [54]	ROS induces proliferation and differentiation of Th17/Th1/Th22 cells.
Johansen et al. Br J Dermatol. 2005; 152(1):37-42. [57]	Activity of the MAPKs and ERK1/2 is increased in psoriatic skin.
Müller et al. Autophagy. 2020; 16(8):1380-1395. [59]	ROS are important mediators in IL23A secretion via NF- κ B and MAPK pathways.
Zhou et al. Free Radic Biol Med. 2009; 47(7):891-905. [62]	Increased inflammation facilitates production of ROS and decreases already weakened antioxidative capacity
Srivastava et al. Indian J Dermatol Venereol Leprol. 2018; 84(1):39-44. [68]	Polymorphisms in the GST genes may result in increased production of ROS that could influence the pathogenesis of psoriasis.
Lee et al. J Invest Dermatol. 2013; 133(3):732-741. [70]	SOD deficiency resulted in more severe IL-23-mediated psoriasis-like skin inflammation.
Drewa et al. Med Sci Monit. 2002; 8(8):BR338-43. [71]	SOD and CAT activities were significantly lower in psoriatic patients.

selectivity, and retention of drug molecules in the target tissue, as well as various adverse reactions [76].

Groundbreaking shifts in psoriasis therapy were achieved with the introduction of drugs that target key immune cascades in the pathogenesis of psoriasis. Biologic therapies or biologics represent progress in the treatment of psoriatic patients in the last decade, due to improved efficacy and tolerability of the drugs and consequently improved quality of life of the patients [77]. Novel biological

drugs, which target TNF- α , the p40 subunit of IL-12 and IL-23, or IL-17 receptors, are effective in treating psoriasis and reducing the PASI score [78] (Table 1). Interestingly, inhibition of IL-1, IL-6, or IFN- γ failed to achieve a significant clinical effect in psoriasis [79]. TNF- α inhibitors include etanercept, infliximab, adalimumab, certolizumab pegol, and golimumab [80]. Inhibitors of IL-17 include secukinumab, ixekizumab, and bimekizumab, while brodalumab is inhibitor of IL-17 receptor [80]. Inhibition of 12p40 subunit,

which is common for IL-12 and IL-23, represents the mechanism of action of ustekinumab and briakinumab, while guselkumab, tildrakizumab, and risankizumab are specific human antibodies targeting the p19 subunit of IL-23, thus blocking the biologic activity of IL-23 [80, 81].

Due to considerable prevalence of psoriasis in general population, as well as comorbidities related to psoriasis, new therapeutic modalities are constantly being investigated. In patients with severe cases of psoriasis in which established therapeutic procedures did not give desired results, the application of cell therapy is being examined, which includes the hematopoietic stem cell transplantation (HSCT) and mesenchymal stromal cell (MSC) [82, 83]. In addition to the fact that mesenchymal cell therapy brings the possibility of a complete recovery of patients with psoriasis, there are also a number of possible adverse effects that reduce the aspiration for their use. The possible adverse effects of cell therapy include neoplastic proliferation, graft versus host reaction, localized skin reactions, and a lack of efficacy [82]. Botulinum toxin has also been shown to be an effective drug in treatment of plaque-type psoriasis [84]. Hydrogen sulfide (H_2S) is recognized as important mediator of various physiological processes and a crucial antioxidative molecule. Due to the disturbed redox balance in psoriasis, therapeutic value of H_2S should be investigated [85]. Antioxidative support to applied antipsoriatic drugs could be an important factor in the reduction of keratinocyte proliferation and remission of the disease [65, 86, 87].

1.6. Effects of Biologic Therapeutics on Oxidative Stress

1.6.1. Effects of IL-12 and IL-23 Inhibitors on Oxidative Stress. Ustekinumab and briakinumab bind to the p40 subunit which is common to IL-12 and IL-23 preventing the interaction of these cytokines with their receptors. Both drugs are fully human monoclonal antibodies. Ustekinumab was approved by the US Food and Drug Administration (FDA) and the European Medicines Agency (EMA) in 2009 for treatment of mild to severe cases of psoriasis, while all clinical trials for briakinumab were discontinued due to the increased risk for myocardial infarction, cerebrovascular accident, and cardiac death [88, 89].

There are very limited data regarding the effects of ustekinumab on redox balance in patients with psoriasis. In a randomized clinical trial, the effects of ustekinumab (IL-12/IL-23 inhibitor), etanercept (TNF- α inhibitor), or cyclosporine were compared regarding the heart function and oxidative stress in patients with psoriasis [90]. After the four-month treatment, the MDA level in patients treated with ustekinumab was significantly decreased compared to TNF- α inhibition, where it was unchanged, or cyclosporine, where it was even increased. Another antioxidative effect of anti-IL-23 antibody was shown in experimental cerebral ischemia [91]. The application of anti-IL-23 antibody decreased the production of ROS and MDA levels in the serum and brain. The antioxidative mechanism achieved by blocking of IL-23 in this experimental model involved targeting the immune specific Janus kinase 2- (JAK2-) STAT3 pathway. Clinical efficacy of IL-12 and IL-23 blockade (ustekinumab) is presented in Figure 4.

1.6.2. Effects of IL-17A Inhibitors on Oxidative Stress. Antioxidative effect of IL-17 inhibition was first reported in a 42-year-old Caucasian woman suffering from plaque psoriasis [92]. One month after secukinumab therapy, plasma levels of lipid measured as thiobarbituric acid reactive substances (TBARS) were significantly decreased, while total antioxidant capacity was significantly improved. It was also noticed that neutrophils, monocytes, and lymphocytes decreased the production of ROS. In three murine experimental models of psoriasis-like skin disease, levels of IL-17A correlated with the severity of the disease and vascular dysfunction [93]. Treatment with anti-IL-17A antibody efficiently eradicated cutaneous lesions and decreased peripheral oxidative stress in two (CD11c-IL-17A^{ind/ind} mice and mice with imiquimod induced psoriasis) assessed the models of psoriasis. Interestingly, in the experimental model of severe psoriasis using K14-IL-17A^{ind/+} mice, neither skin lesions nor peripheral oxidative stress was improved [93]. Another study compared the effects of secukinumab (IL-17A inhibitor), cyclosporine, or methotrexate (MTX) treatment on the left ventricular function and oxidative stress in patients with psoriasis [94]. Secukinumab exerted most beneficial effects on the improvement of the left ventricular function and the decrease of MDA and protein carbonyl as markers of oxidative stress. On the other hand, MTX did not change the measured parameters for oxidative stress, while cyclosporine even caused them to increase [90]. The results of the Eding et al. showed that MutT Homolog 1 (MTH1) levels were increased in skin lesions of imiquimod-induced psoriatic mice [95]. Inhibition of MTH1 resulted in both decrease of ROS and oxidative stress and IL-17-mediated inflammation. Assessing the effect of IL-17A overexpression in CD4-IL-17A^{ind/+} mice, it was shown that IL-17A overexpression was accompanied by increased peripheral ROS/RNS levels, as well as increased ROS/RNS production by spleen CD11b⁺ cells [97]. Increased production of IL-17A, followed by an increase in oxidative stress, resulted in endothelial dysfunction and vascular damage. Clinical efficiency of IL-17 inhibition (secukinumab) is shown in Figure 5.

1.6.3. Effects of TNF α Blockers on Oxidative Stress. TNF- α alone does not elicit a significant response to keratinocytes. However, in combination with IL-17A and other cytokines, TNF- α is a significant element of the cytokine milieu in psoriasis. Powerful synergism between TNF- α and IL-17A enhances the effects of IL-17A. In addition, it leads to increased expression of IL-17R by keratinocytes [97]. TNF- α inhibitors (etanercept, adalimumab, and infliximab) are human fusion proteins used in treatment psoriasis. Aiming to investigate the effects of infliximab, as TNF- α blocker, on redox balance of psoriatic patients, Barygina and coworkers have measured various oxidative stress biomarkers in plasma and white blood cells [98]. After six-month infliximab treatment, plasma levels of MDA and protein carbonyl content were decreased, while the activity of NADPH oxidase was decreased compared to untreated patients. The most significant limitation of this study is the very small number of patients, given that only 29 patients in total were

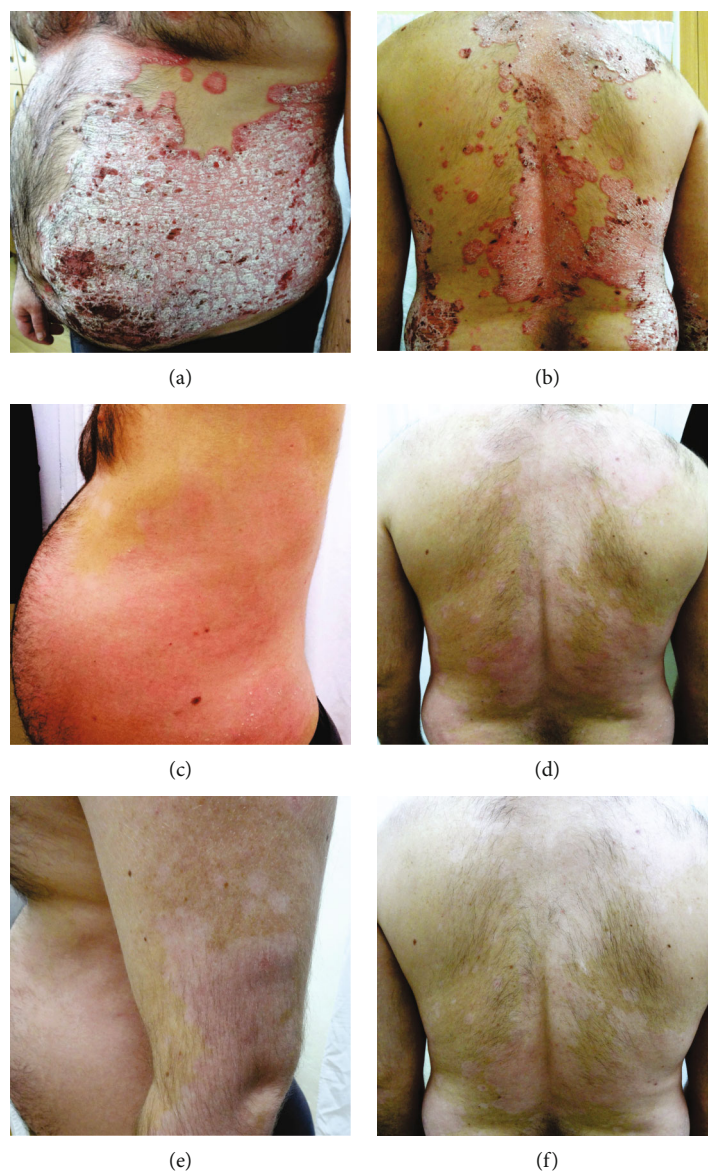


FIGURE 4: Clinical efficacy of IL-12 and IL-23 blockade (ustekinumab). Patient before starting therapy (a, b), after 8 weeks from the introduction of the IL-12/IL-23 blocker (c, d), and after 16 weeks after the introduction of the IL-12/IL-23 blocker (e, f). Pictures presented are part of the private collection of Vesna M. Milicic and Ana B. Ravic Nikolic.

included. In another study dealing with the role of etanercept, another $\text{TNF-}\alpha$ inhibitor, the authors measured plasma, total antioxidative capacity, and PON1 activity [99]. Etanercept application for 24 weeks significantly improved total antioxidative capacity and PON1 activity combined with reduction of inflammatory markers (C-reactive protein). Oxidative stress was also reduced by etanercept in experimental peritonitis [100]. Inhibition of $\text{TNF-}\alpha$ by adalimumab, human monoclonal antibody against $\text{TNF-}\alpha$, showed significant reduction of oxidative stress in experimental model of vascular dementia [100]. Comparing the effects of ustekinumab, etanercept, or cyclosporine on oxidative stress in psoriasis-suffering patients, ustekinumab exerted most powerful antioxidative effect [92].

Overall, it can be concluded that all biologics exert antioxidative capacity but novel study comparing the antioxi-

dative effects of biologics of different mechanism of action should offer new insights into interfering networks of oxidative stress, immune response, and inflammation.

1.7. Effects of Immunomodulatory Therapy on Oxidative Stress. Methotrexate (MTX) is a well-established drug for systemic treatment of psoriasis due to its antiproliferative, immunosuppressive, and anti-inflammatory properties. Its exact underlining mechanisms are not fully elucidated [102]. The antiproliferative effects of MTX are based on inhibitory effect on folate-dependent enzymes, such as dihydrofolate reductase, resulting in decreased synthesis of DNA and reduced cell proliferation [103]. Immunosuppressive and anti-inflammatory ability of MTX are a consequence of increased adenosine release, decreased release of proinflammatory cytokines ($\text{TNF-}\alpha$ and IL-1), favored apoptosis

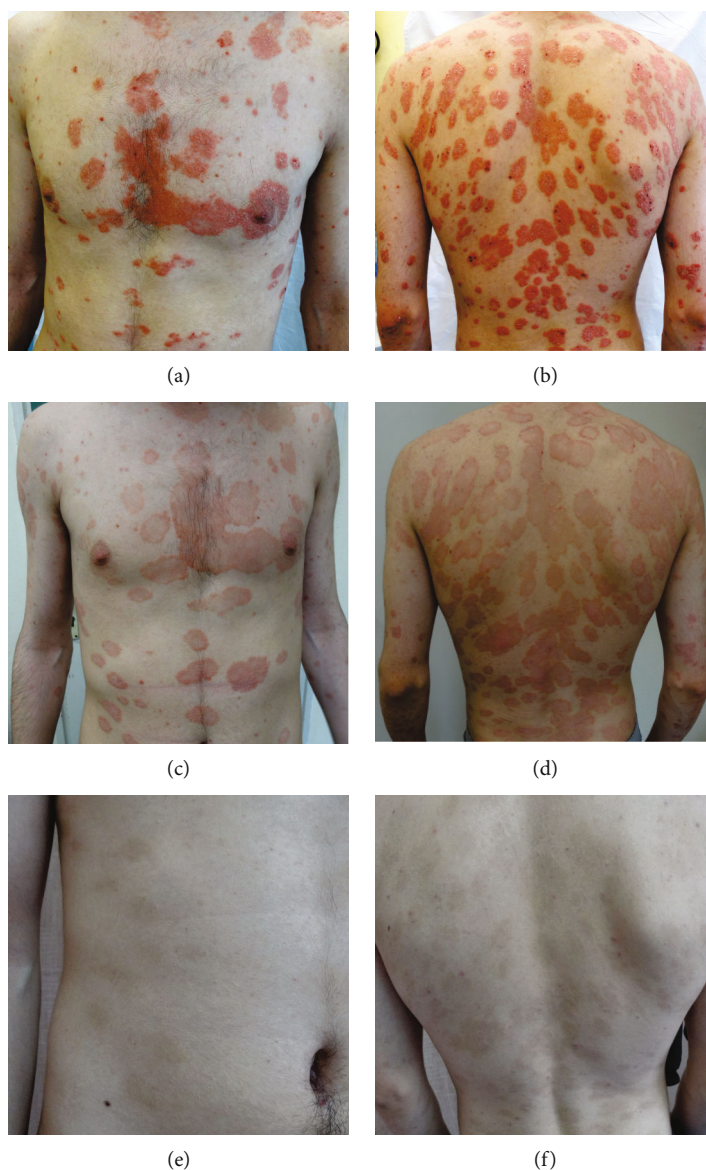


FIGURE 5: Clinical efficacy of IL-17A blockade (secukinumab). Patient before starting therapy (a, b), after 8 weeks from the introduction of the IL-17A blocker (c, d), and after 16 weeks after the introduction of the IL-17A blocker (e, f). Pictures presented are part of the private collection of Vesna M. Milicic and Ana B. Ravic Nikolic.

of activated T cells, and decreased chemotaxis of neutrophils [103].

A twelve-week MTX treatment in psoriatic patients induced augmentation of oxidative stress, reflected through increased levels of MDA and reduction of nitrate/nitrite, SOD, and CAT activity and TAS in the plasma [104]. The authors proposed that increased ROS and decreased NO production, combined with increased caspase-3 expression, represent the mechanism of MTX-mediated induction of apoptosis. Experimental study in an isolated hepatocytes rat confirmed prooxidative effects of MTX [105]. Antioxidants decreased ROS availability and reduced MTX-induced cytotoxicity. MTX also induced mitochondria swelling, decrease of ATP and GSH amount, and release of cytochrome c. The assessment of expression of isoenzymes of GST and cytochrome (CYP) families showed that expression

was increased in patients with psoriasis compared to healthy controls [106]. After 12-week MTX treatment, the expression of these enzymes, important in regulation of redox balance, did not change significantly, although there was marked clinical improvement. Prooxidative effects of MTX were shown in other tissues and organs. In the MTX treated rats, the levels of GSH, SOD, and CAT testicular were significantly decreased, combined with increased DNA and tissue damage [107]. MTX also showed prooxidative features in liver and neural tissues due to decreased activity of SOD, CAT, or GPx [108, 109].

Conversely, the results of several studies implicated antioxidative effects of MTX. Zimmerman et al. showed that MTX inhibits the generation of MAA-protein adducts, formed by MDA and acetaldehyde interaction [110]. MAA-protein adducts have high immunogenic and

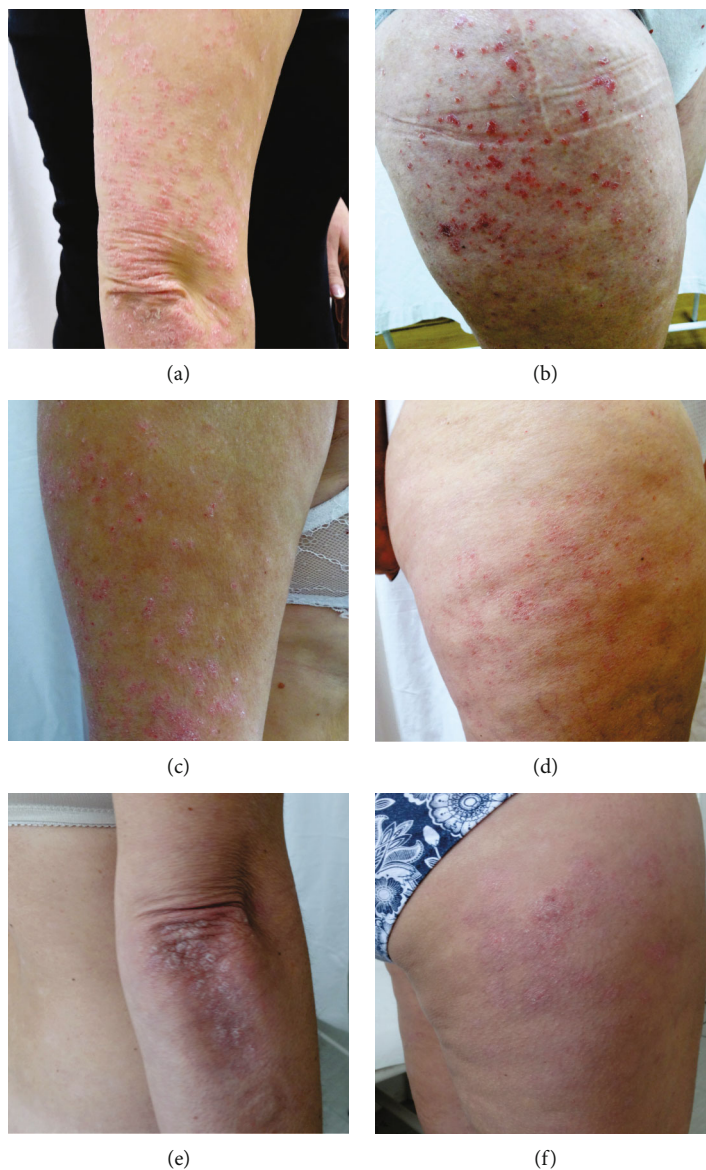


FIGURE 6: Clinical efficacy of methotrexate (MTX). Patient before starting therapy (a, b), after 8 weeks from the introduction of the MTX (15 mg per week) (c, d), and after 16 weeks after the introduction of the MTX (15 mg per week) (e, f). Pictures presented are part of the private collection of Vesna M. Milicic and Ana B. Ravic Nikolic.

prooxidative potential, so they are not only markers of oxidative stress, but also play an active role in the development of immune disorders. MTX significantly reduced the production of MAA-protein adducts as well as the level of free radicals *in vitro*. The reduction of free radicals and antioxidative action of MTX was achieved by the ability of MTX to directly scavenge O_2^- . Another study on the effects of 24 weeks of MTX therapy on oxidative stress showed antioxidative properties of MTX in psoriatic patients [111]. The levels of MDA significantly decreased, while TAS markedly increased after MTX therapy in psoriatic patients. Comparing to healthy controls the oxidative stress was higher in psoriatic patients both at the beginning and at the end of the MTX therapy protocol.

Analyzing results of different studies, it can be concluded that MTX has somewhat contradictory impact on redox bal-

ance. This could be the consequence of its prooxidative and proapoptotic effects, and on the other hand, of decreased production of ROS through inhibition of inflammation-mediated pathways [112]. Clinical efficacy MTX is shown in Figure 6.

1.8. Effects of UVB and PUVA Therapy on Oxidative Stress. Phototherapy is often used in the treatment of skin diseases. It is a type of light therapy that mimics the effects of exposure to sunlight. Established phototherapies in psoriatic patients include PUVA radiation (psoralen in combination with UVA radiation) and narrow-band UVB (NB-UVB) [113]. The phototherapeutic procedures are usually effective in psoriasis treatment, but due to potential side-effects, including carcinogenesis, it can be used only for short-term treatment of the disease [113]. Increased ROS production

during PUVA therapy, due to psoralen sensitization and UVA exposure, increases already increased levels of superoxide anion radical (O_2^-) and forms the basis of photocarcinogenesis [114].

Production of ROS due to PUVA therapy may cause oxidative damage to various macromolecules. The analysis of the PUVA effects on oxidative damage of DNA showed that suberythemal dose induced increased urinary extraction of 8-Oxo-2'-deoxyguanosine (8-oxo-dG) as a marker of DNA oxidation [115]. 8-oxo-dG in urine reached the peak 4 days after PUVA irradiation. *In vitro* study on HaCaT keratinocyte cell line also showed increased 8-oxo-dG production after exposure to therapeutic doses of PUVA and NB-UVB [116].

In the 12-week follow-up study, oxidative stress biomarkers, TBARS, and total antioxidant status (TAS) were significantly improved due to NB-UVB and PUVA therapy [117]. PUVA treatment appears to be more effective in the reduction of oxidative stress comparing to NB-UVB. The reduction of oxidative stress markers was connected with the reduction of inflammation. A study of similar design also showed the reduction of oxidative stress biomarkers, TBARS and TAS, after 12-week long PUVA and NB-UVB therapy [118]. Short-term increase of ROS production upon PUVA treatment, combined with the reduction of inflammation, could improve antioxidative potential and thus prolonged PUVA treatment results in the reduction of oxidative stress.

2. Conclusion

Oxidative stress and the aberrant immune response remain two crucial pathophysiological mechanisms in psoriasis. Their interdependence and interconnectedness represent the key to understanding etiopathogenesis of psoriasis. A large number of factors that can influence the development of psoriasis, from genetic predisposition to lifestyle, make the tangle of causes and consequences still insufficiently understood. Novel therapeutic options, primarily biological therapy, have significantly improved the quality of life of patients with psoriasis. However, the success of the applied therapy seems to depend on the reduction of both the proinflammatory and prooxidative components of the disease. The most noticeable and most powerful antioxidative effects exert various biologics compared to more convenient therapeutic modalities, such as methotrexate or phototherapy. Intersections of redox, immune, and inflammatory signaling pathways should be the focus of further researches dealing with pathophysiology of psoriasis, while antioxidative supplementation could be the solution in some refractory cases of the disease.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors would like to thank Mr. Dusan Tomasevic for technical support. Figures 2 and 3 are created using BioRen-

der. This work was supported by grant from the Faculty of Medical Sciences, University of Kragujevac, Serbia (project no. JP 27/20).

References

- [1] F. S. Glickman, "Lepra, psora, psoriasis," *Journal of the American Academy of Dermatology*, vol. 14, no. 5, pp. 863–866, 1986.
- [2] I. M. Michalek, B. Loring, and S. M. John, "A systematic review of worldwide epidemiology of psoriasis," *Journal of the European Academy of Dermatology and Venereology*, vol. 31, no. 2, pp. 205–212, 2017.
- [3] C. E. M. Griffiths, A. W. Armstrong, J. E. Gudjonsson, and J. N. W. N. Barker, "Psoriasis," *Lancet*, vol. 397, no. 10281, pp. 1301–1315, 2021.
- [4] W. H. Boehncke and M. P. Schön, "Psoriasis," *Lancet*, vol. 386, no. 9997, pp. 983–994, 2015.
- [5] E. Muñoz-Aceituno, L. Martos-Cabrera, M. C. Ovejero-Benito, A. Reolid, F. Abad-Santos, and E. Daudén, "Pharmacogenetics update on biologic therapy in psoriasis," *Medicina (Kaunas, Lithuania)*, vol. 56, no. 12, p. 719, 2020.
- [6] J. Takeshita, S. Grewal, S. M. Langan et al., "Psoriasis and comorbid diseases: epidemiology," *Journal of the American Academy of Dermatology*, vol. 76, no. 3, pp. 377–390, 2017.
- [7] A. Ogdie and P. Weiss, "The epidemiology of psoriatic arthritis," *Rheumatic Diseases Clinics of North America*, vol. 41, no. 4, pp. 545–568, 2015.
- [8] E. J. Horn, K. M. Fox, V. Patel, C. F. Chiou, F. Dann, and M. Lebwohl, "Are patients with psoriasis undertreated? Results of National Psoriasis Foundation survey," *Journal of the American Academy of Dermatology*, vol. 57, no. 6, pp. 957–962, 2007.
- [9] A. W. Armstrong, A. D. Robertson, J. Wu, C. Schupp, and M. G. Lebwohl, "Undertreatment, treatment trends, and treatment dissatisfaction among patients with psoriasis and psoriatic arthritis in the United States: findings from the National Psoriasis Foundation surveys, 2003-2011," *JAMA Dermatology*, vol. 149, no. 10, pp. 1180–1185, 2013.
- [10] U. Mrowietz, K. Kragballe, K. Reich et al., "Definition of treatment goals for moderate to severe psoriasis: a European consensus," *Archives of Dermatological Research*, vol. 303, no. 1, pp. 1–10, 2011.
- [11] P. J. Mease, D. D. Gladman, P. Helliwell et al., "Comparative performance of psoriatic arthritis screening tools in patients with psoriasis in European/North American dermatology clinics," *Journal of the American Academy of Dermatology*, vol. 71, no. 4, pp. 649–655, 2014.
- [12] G. Tonel and C. Conrad, "Interplay between keratinocytes and immune cells—recent insights into psoriasis pathogenesis," *The International Journal of Biochemistry & Cell Biology*, vol. 41, no. 5, pp. 963–968, 2009.
- [13] J. Kim and J. G. Krueger, "The immunopathogenesis of psoriasis," *Dermatologic Clinics*, vol. 33, no. 1, pp. 13–23, 2015.
- [14] F. Capon, "The genetic basis of psoriasis," *International Journal of Molecular Sciences*, vol. 18, no. 12, article 2526, 2017.
- [15] D. L. Duffy, L. S. Spelman, and N. G. Martin, "Psoriasis in Australian twins," *Journal of the American Academy of Dermatology*, vol. 29, no. 3, pp. 428–434, 1993.
- [16] C. Ohata, Y. Kanai, K. Murotani, H. Kitabayashi, and S. Imafuku, "Difference in health-related quality of life

- between anxiety and depressive symptoms in Japanese patients with plaque psoriasis: the ProLOGUE study,” *Journal of the European Academy of Dermatology and Venereology*, vol. 36, pp. e57–e59, 2022.
- [17] K. A. Papp and M. G. Lebwohl, “Onset of action of biologics in patients with moderate-to-severe psoriasis,” *Journal of Drugs in Dermatology*, vol. 17, no. 3, pp. 247–250, 2017.
- [18] S. K. Mahil, M. C. Ezejimofor, L. S. Exton et al., “Comparing the efficacy and tolerability of biologic therapies in psoriasis: an updated network meta-analysis,” *The British Journal of Dermatology*, vol. 183, no. 4, pp. 638–649, 2020.
- [19] Z. Yang, Z. Min, and B. Yu, “Reactive oxygen species and immune regulation,” *International Reviews of Immunology*, vol. 39, no. 6, pp. 292–298, 2020.
- [20] E. Ambrożewicz, P. Wójcik, A. Wroński et al., “Pathophysiological alterations of redox signaling and endocannabinoid system in granulocytes and plasma of psoriatic patients,” *Cell*, vol. 7, no. 10, article 159, 2018.
- [21] J. R. Bjerke, “In situ characterization and counting of mononuclear cells in lesions of different clinical forms of psoriasis,” *Acta Dermato-Venereologica*, vol. 62, no. 2, pp. 93–100, 1982.
- [22] H. Hammar, S. Q. Gu, A. Johannesson, K. G. Sundkvist, and P. Biberfeld, “Subpopulations of mononuclear cells in microscopic lesions of psoriatic patients. Selective accumulation of suppressor/cytotoxic T cells in epidermis during the evolution of the lesion,” *The Journal of Investigative Dermatology*, vol. 83, no. 6, pp. 416–420, 1984.
- [23] S. L. Gottlieb, P. Gilleaudeau, R. Johnson et al., “Response of psoriasis to a lymphocyte-selective toxin (DAB₃₈₉IL-2) suggests a primary immune, but not keratinocyte, pathogenic basis,” *Nature Medicine*, vol. 1, no. 5, pp. 442–447, 1995.
- [24] T. Biedermann, M. Röcken, and J. M. Carballido, “TH1 and TH2 lymphocyte development and regulation of TH cell-mediated immune responses of the skin,” *The Journal of Investigative Dermatology. Symposium Proceedings*, vol. 9, no. 1, pp. 5–14, 2004.
- [25] M. J. Gooderham, K. A. Papp, and C. W. Lynde, “Shifting the focus – the primary role of IL-23 in psoriasis and other inflammatory disorders,” *Journal of the European Academy of Dermatology and Venereology*, vol. 32, no. 7, pp. 1111–1119, 2018.
- [26] D. J. Marble, K. B. Gordon, and B. J. Nickoloff, “Targeting TNF α rapidly reduces density of dendritic cells and macrophages in psoriatic plaques with restoration of epidermal keratinocyte differentiation,” *Journal of Dermatological Science*, vol. 48, no. 2, pp. 87–101, 2007.
- [27] L. L. Ten Bergen, A. Petrovic, A. Krogh Aarebrot, and S. Appel, “The TNF/IL-23/IL-17 axis—head-to-head trials comparing different biologics in psoriasis treatment,” *Scandinavian Journal of Immunology*, vol. 92, no. 4, article e12946, 2020.
- [28] J. D. Bos, M. A. de Rie, M. B. Teunissen, and G. Piskin, “Psoriasis: dysregulation of innate immunity,” *The British Journal of Dermatology*, vol. 152, no. 6, pp. 1098–1107, 2005.
- [29] C. C. Chiang, W. J. Cheng, M. Korinek, C. Y. Lin, and T. L. Hwang, “Neutrophils in psoriasis,” *Frontiers in Immunology*, vol. 10, article 2376, 2019.
- [30] S. A. Belambri, L. Rolas, H. Raad, M. Hurtado-Nedelec, P. M. Dang, and J. El-Benna, “NADPH oxidase activation in neutrophils: role of the phosphorylation of its subunits,” *European Journal of Clinical Investigation*, vol. 48, article e12951, Suppl 2, 2018.
- [31] N. Milosavljevic, M. Gazdic, B. Simovic Markovic et al., “Mesenchymal stem cells attenuate acute liver injury by altering ratio between interleukin 17 producing and regulatory natural killer T cells,” *Liver Transplantation*, vol. 23, no. 8, pp. 1040–1050, 2017.
- [32] S. Bagchi, Y. He, H. Zhang et al., “CD1b-autoreactive T cells contribute to hyperlipidemia-induced skin inflammation in mice,” *The Journal of Clinical Investigation*, vol. 127, no. 6, pp. 2339–2352, 2017.
- [33] L. C. Zaba, J. Fuentes-Duculan, N. J. Eungdamrong et al., “Psoriasis is characterized by accumulation of immunostimulatory and Th1/Th17 cell-polarizing myeloid dendritic cells,” *The Journal of Investigative Dermatology*, vol. 129, no. 1, pp. 79–88, 2009.
- [34] J. E. Hawkes, B. Y. Yan, T. C. Chan, and J. G. Krueger, “Discovery of the IL-23/IL-17 signaling pathway and the treatment of psoriasis,” *Journal of Immunology*, vol. 201, no. 6, pp. 1605–1613, 2018.
- [35] L. M. Johnson-Huang, M. A. Lowes, and J. G. Krueger, “Putting together the psoriasis puzzle: an update on developing targeted therapies,” *Disease Models & Mechanisms*, vol. 5, no. 4, pp. 423–433, 2012.
- [36] Y. Iwakura, H. Ishigame, S. Saijo, and S. Nakae, “Functional specialization of interleukin-17 family members,” *Immunity*, vol. 34, no. 2, pp. 149–162, 2011.
- [37] E. Stephen-Victor, H. Fickenscher, and Bayry, J., “IL-26: an emerging proinflammatory member of the IL-10 cytokine family with multifaceted actions in antiviral, antimicrobial, and autoimmune responses,” *PLoS Pathogens*, vol. 12, no. 6, article e1005624, 2016.
- [38] J. M. Wang, A. F. Huang, W. D. Xu, and L. C. Su, “Insights into IL-29: emerging role in inflammatory autoimmune diseases,” *Journal of Cellular and Molecular Medicine*, vol. 23, no. 12, pp. 7926–7932, 2019.
- [39] E. Witte, G. Kokolakis, K. Witte et al., “IL-19 is a component of the pathogenetic IL-23/IL-17 cascade in psoriasis,” *The Journal of Investigative Dermatology*, vol. 134, no. 11, pp. 2757–2767, 2014.
- [40] Q. Jiang, G. Yang, F. Xiao et al., “Role of Th22 cells in the pathogenesis of autoimmune diseases,” *Frontiers in Immunology*, vol. 12, article 688066, 2021.
- [41] A. Chiricozzi, P. Romanelli, E. Volpe, G. Borsellino, and M. Romanelli, “Scanning the immunopathogenesis of psoriasis,” *International Journal of Molecular Sciences*, vol. 19, no. 1, article 179, 2018.
- [42] L. Zhou, I. I. Ivanov, R. Spolski et al., “IL-6 programs T_H-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways,” *Nature Immunology*, vol. 8, no. 9, pp. 967–974, 2007.
- [43] D. M. Floss, J. M. Moll, and J. Scheller, “IL-12 and IL-23-close relatives with structural homologies but distinct immunological functions,” *Cell*, vol. 9, no. 10, article 2184, 2020.
- [44] H. J. Bovenschen, P. C. van de Kerkhof, P. E. van Erp, R. Woestenenk, I. Joosten, and H. J. Koenen, “Foxp3+ regulatory T cells of psoriasis patients easily differentiate into IL-17A-producing cells and are found in lesional skin,” *The Journal of Investigative Dermatology*, vol. 131, no. 9, pp. 1853–1860, 2011.

- [45] N. J. Wilson, K. Boniface, J. R. Chan et al., “Development, cytokine profile and function of human interleukin 17-producing helper T cells,” *Nature Immunology*, vol. 8, no. 9, pp. 950–957, 2007.
- [46] A. A. Levin and A. B. Gottlieb, “Specific targeting of interleukin-23p19 as effective treatment for psoriasis,” *Journal of the American Academy of Dermatology*, vol. 70, no. 3, pp. 555–561, 2014.
- [47] C. L. Langrish, B. S. McKenzie, N. J. Wilson, R. de Waal Malefyt, R. A. Kastelein, and D. J. Cua, “IL-12 and IL-23: master regulators of innate and adaptive immunity,” *Immunological Reviews*, vol. 202, no. 1, pp. 96–105, 2004.
- [48] Y. A. Rodriguez-Rosales, J. D. Langereis, M. A. J. Gorris et al., “Immunomodulatory aged neutrophils are augmented in blood and skin of psoriasis patients,” *The Journal of Allergy and Clinical Immunology*, vol. 148, no. 4, pp. 1030–1040, 2021.
- [49] H. L. Rizzo, S. Kagami, K. G. Phillips, S. E. Kurtz, S. L. Jacques, and A. Blauvelt, “IL-23-mediated psoriasis-like epidermal hyperplasia is dependent on IL-17A,” *Journal of Immunology*, vol. 186, no. 3, pp. 1495–1502, 2011.
- [50] B. Halliwell, “Biochemistry of oxidative stress,” *Biochemical Society Transactions*, vol. 35, no. 5, pp. 1147–1150, 2007.
- [51] B. Halliwell, “Free radicals and antioxidants: updating a personal view,” *Nutrition Reviews*, vol. 70, no. 5, pp. 257–265, 2012.
- [52] E. F. Bernstein, D. B. Brown, M. D. Schwartz, K. Kaidbey, and S. M. Ksenzenko, “The polyhydroxy acid gluconolactone protects against ultraviolet radiation in an in vitro model of cutaneous photoaging,” *Dermatologic Surgery*, vol. 30, no. 2, pp. 189–196, 2004.
- [53] T. Bitto and C. Nishigori, “Impact of reactive oxygen species on keratinocyte signaling pathways,” *Journal of Dermatological Science*, vol. 68, no. 1, pp. 3–8, 2012.
- [54] R. Lai, D. Xian, X. Xiong, L. Yang, J. Song, and J. Zhong, “Proanthocyanidins: novel treatment for psoriasis that reduces oxidative stress and modulates Th17 and Treg cells,” *Redox Report*, vol. 23, no. 1, pp. 130–135, 2018.
- [55] J. M. Kyriakis and J. Avruch, “Mammalian MAPK signal transduction pathways activated by stress and inflammation: a 10-year update,” *Physiological Reviews*, vol. 92, no. 2, pp. 689–737, 2012.
- [56] K. N. Schmidt, P. Amstad, P. Cerutti, and P. A. Baeuerle, “The roles of hydrogen peroxide and superoxide as messengers in the activation of transcription factor NF- κ B,” *Chemistry & Biology*, vol. 2, no. 1, pp. 13–22, 1995.
- [57] C. Johansen, K. Kragballe, M. Westergaard, J. Henningsen, K. Kristiansen, and L. Iversen, “The mitogen-activated protein kinases p38 and ERK1/2 are increased in lesional psoriatic skin,” *The British Journal of Dermatology*, vol. 152, no. 1, pp. 37–42, 2005.
- [58] X. J. Yu, C. Y. Li, H. Y. Dai et al., “Expression and localization of the activated mitogen-activated protein kinase in lesional psoriatic skin,” *Experimental and Molecular Pathology*, vol. 83, no. 3, pp. 413–418, 2007.
- [59] G. Müller, C. Lübow, and G. Weindl, “Lysosomotropic beta blockers induce oxidative stress and IL23A production in Langerhans cells,” *Autophagy*, vol. 16, no. 8, pp. 1380–1395, 2020.
- [60] J. Pleńkowska, M. Gabig-Cimińska, and P. Mozolewski, “Oxidative stress as an important contributor to the pathogenesis of psoriasis,” *International Journal of Molecular Sciences*, vol. 21, no. 17, article 6206, 2020.
- [61] N. Vorobjeva, A. Prikhodko, I. Galkin et al., “Mitochondrial reactive oxygen species are involved in chemoattractant-induced oxidative burst and degranulation of human neutrophils in vitro,” *European Journal of Cell Biology*, vol. 96, no. 3, pp. 254–265, 2017.
- [62] Q. Zhou, U. Mrowietz, and M. Rostami-Yazdi, “Oxidative stress in the pathogenesis of psoriasis,” *Free Radical Biology & Medicine*, vol. 47, no. 7, pp. 891–905, 2009.
- [63] L. Yang, X. Fan, T. Cui, E. Dang, and G. Wang, “Nrf2 promotes keratinocyte proliferation in psoriasis through up-regulation of keratin 6, keratin 16, and keratin 17,” *The Journal of Investigative Dermatology*, vol. 137, no. 10, pp. 2168–2176, 2017.
- [64] Y. J. Lee, J. H. Bae, S. G. Kang et al., “Pro-oxidant status and Nrf2 levels in psoriasis vulgaris skin tissues and dimethyl fumarate-treated HaCaT cells,” *Archives of Pharmacological Research*, vol. 40, no. 9, pp. 1105–1116, 2017.
- [65] W. Wang and H. Wang, “Astilbin reduces ROS accumulation and VEGF expression through Nrf2 in psoriasis-like skin disease,” *Biological Research*, vol. 52, no. 1, p. 49, 2019.
- [66] M. Asefi, A. Vaisi-Raygani, F. Bahrehmand et al., “Paraoxonase 1 (PON1) 55 polymorphism, lipid profiles and psoriasis,” *The British Journal of Dermatology*, vol. 167, no. 6, pp. 1279–1286, 2012.
- [67] A. A. Hernández-Collazo, O. Pérez-Méndez, V. López-Olmos et al., “Association between rs662 (A > G) and rs854560 (A > T) polymorphisms in PON1 gene and the susceptibility for psoriasis in mestizo population of Western Mexico,” *Molecular Biology Reports*, vol. 48, no. 1, pp. 183–194, 2021.
- [68] D. S. L. Srivastava, V. K. Jain, P. Verma, and J. P. Yadav, “Polymorphism of glutathione S-transferase M1 and T1 genes and susceptibility to psoriasis disease: a study from North India,” *Indian Journal of Dermatology, Venereology and Leprology*, vol. 84, no. 1, pp. 39–44, 2018.
- [69] S. H. Ibbotson, R. S. Dawe, A. T. Dinkova-Kostova et al., “Glutathione S-transferase genotype is associated with sensitivity to psoralen-ultraviolet A photochemotherapy,” *The British Journal of Dermatology*, vol. 166, no. 2, pp. 380–388, 2012.
- [70] Y. S. Lee, I. S. Cheon, B. H. Kim, M. J. Kwon, H. W. Lee, and T. Y. Kim, “Loss of extracellular superoxide dismutase induces severe IL-23-mediated skin inflammation in mice,” *The Journal of Investigative Dermatology*, vol. 133, no. 3, pp. 732–741, 2013.
- [71] G. Drewa, E. Krzyżyńska-Malinowska, A. Woźniak et al., “Activity of superoxide dismutase and catalase and the level of lipid peroxidation products reactive with TBA in patients with psoriasis,” *Medical Science Monitor*, vol. 8, no. 8, article BR338-343, 2002.
- [72] A. Skutnik-Radziszewska, M. Maciejczyk, K. Fejfer et al., “Salivary antioxidants and oxidative stress in psoriatic patients: can salivary total oxidant status and oxidative status index be a plaque psoriasis biomarker?,” *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 9086024, 12 pages, 2020.
- [73] A. Skutnik-Radziszewska, M. Maciejczyk, I. Flisiak et al., “Enhanced inflammation and nitrosative stress in the saliva and plasma of patients with plaque psoriasis,” *Journal of Clinical Medicine*, vol. 9, no. 3, article 745, 2020.

- [74] G. Ganzetti, A. Campanati, A. Santarelli et al., "Salivary interleukin-1 β : oral inflammatory biomarker in patients with psoriasis," *The Journal of International Medical Research*, vol. 44, 1_suppl, pp. 10–14, 2016.
- [75] D. Belstrøm, J. M. Eiberg, C. Enevold et al., "Salivary microbiota and inflammation-related proteins in patients with psoriasis," *Oral Diseases*, vol. 26, no. 3, pp. 677–687, 2020.
- [76] P. Gisondi, F. Bellinato, M. Bruni, G. De Angelis, and G. Girolomoni, "Methotrexate vs secukinumab safety in psoriasis patients with metabolic syndrome," *Dermatologic Therapy*, vol. 33, no. 6, article e14281, 2020.
- [77] P. Biswasroy, D. Pradhan, B. Kar, G. Ghosh, and G. Rath, "Recent advancement in topical nanocarriers for the treatment of psoriasis," *AAPS PharmSciTech*, vol. 22, no. 5, p. 164, 2021.
- [78] J. W. Marson, M. L. Snyder, and M. G. Lebwohl, "Newer therapies in psoriasis," *The Medical Clinics of North America*, vol. 105, no. 4, pp. 627–641, 2021.
- [79] M. A. Richard, J. P. Lacour, M. P. Konstantinou et al., "Secukinumab efficacy in reducing the severity and the psychosocial impact of moderate-to-severe psoriasis as assessed by the simplified psoriasis index: results from the IPSI-PSO study," *Journal of the European Academy of Dermatology and Venereology*, vol. 35, no. 3, pp. 677–684, 2021.
- [80] J. Baliwag, D. H. Barnes, and A. Johnston, "Cytokines in psoriasis," *Cytokine*, vol. 73, no. 2, pp. 342–350, 2015.
- [81] F. M. Ghazawi, F. Mahmood, L. Kircik et al., "A review of the efficacy and safety for biologic agents targeting IL-23 in treating psoriasis with the focus on tildrakizumab," *Frontiers in Medicine*, vol. 8, article 702776, 2021.
- [82] A. Ruggiero, G. Fabbrocini, E. Cinelli, S. S. Ocampo Garza, E. Camela, and M. Megna, "Anti-interleukin-23 for psoriasis in elderly patients: guselkumab, risankizumab and tildrakizumab in real-world practice," *Clinical and Experimental Dermatology*, vol. 47, no. 3, 2021.
- [83] A. A. B. Eita, A. M. Zaki, and S. A. Mahmoud, "Serum 8-isoprostane levels in patients with resistant oral lichen planus before and after treatment with lycopene: a randomized clinical trial," *BMC Oral Health*, vol. 21, no. 1, article 343, 2021.
- [84] Y. Chen, Q. Yu, Y. Hu, and Y. Shi, "Current research and use of mesenchymal stem cells in the therapy of autoimmune diseases," *Current Stem Cell Research & Therapy*, vol. 14, no. 7, pp. 579–582, 2019.
- [85] J. Abou-Rahal, L. Abdullah, M. Kurban, and O. Abbas, "Letter to the editor regarding the article"Rossi A, Magri F, Michelini S, et al. New onset of alopecia areata in a patient with SARS-COV-2 infection: Possible pathogenetic correlations?," *Journal of Cosmetic Dermatology*, vol. 20, no. 7, article 2004, 2021.
- [86] Q. Xiao, L. Xiong, J. Tang, L. Li, and L. Li, "Hydrogen sulfide in skin diseases: a novel mediator and therapeutic target," *Oxidative Medicine and Cellular Longevity*, vol. 2021, Article ID 6652086, 11 pages, 2021.
- [87] F. Guarneri, L. Bertino, G. Pioggia, M. Casciaro, and S. Gangemi, "Therapies with antioxidant potential in psoriasis, vitiligo, and lichen planus," *Antioxidants (Basel)*, vol. 10, no. 7, article 1087, 2021.
- [88] A. Winiarska-Mieczan, T. Mieczan, and G. Wójcik, "Importance of redox equilibrium in the pathogenesis of psoriasis-impact of antioxidant-rich diet," *Nutrients*, vol. 12, no. 6, article 1841, 2020.
- [89] L. Puig, "The role of IL 23 in the treatment of psoriasis," *Expert Review of Clinical Immunology*, vol. 13, no. 6, pp. 525–534, 2017.
- [90] P. Traczewski and L. Rudnicka, "Briakinumab for the treatment of plaque psoriasis," *BioDrugs*, vol. 26, no. 1, pp. 9–20, 2012.
- [91] I. Ikonomidis, E. Papadavid, G. Makavos et al., "Lowering interleukin-12 activity improves myocardial and vascular function compared with tumor necrosis factor- α antagonism or cyclosporine in psoriasis," *Circulation. Cardiovascular Imaging*, vol. 10, no. 9, article e006283, 2017.
- [92] L. Fan and L. Zhou, "Anti-IL-23 exerted protective effects on cerebral ischemia-reperfusion injury through JAK2/STAT3 signaling pathway," *Molecular Biology Reports*, vol. 48, no. 4, pp. 3475–3484, 2021.
- [93] M. Becatti, M. L. Urban, G. Taurisano et al., "Secukinumab reduces plasma oxidative stress in psoriasis: a case-based experience," *Dermatologic Therapy*, vol. 31, no. 5, article e12675, 2018.
- [94] R. Schüler, A. Brand, S. Klebow et al., "Antagonization of IL-17A attenuates skin inflammation and vascular dysfunction in mouse models of psoriasis," *The Journal of Investigative Dermatology*, vol. 139, no. 3, pp. 638–647, 2019.
- [95] G. Makavos, I. Ikonomidis, I. Andreadou et al., "Effects of interleukin 17A inhibition on myocardial deformation and vascular function in psoriasis," *The Canadian Journal of Cardiology*, vol. 36, no. 1, pp. 100–111, 2020.
- [96] C. Bivik Eding, I. Köhler, D. Verma et al., "MTH1 inhibitors for the treatment of psoriasis," *The Journal of Investigative Dermatology*, vol. 141, no. 8, pp. 2037–2048.e4, 2021.
- [97] R. Schüler, P. Efentakis, J. Wild et al., "T cell-derived IL-17A induces vascular dysfunction via perivascular fibrosis formation and dysregulation of \cdot NO/cGMP signaling," *Oxidative Medicine and Cellular Longevity*, vol. 2019, Article ID 6721531, 15 pages, 2019.
- [98] M. Noack, A. Beringer, and P. Miossec, "Additive or synergistic interactions between IL-17A or IL-17F and TNF or IL-1 β depend on the cell type," *Frontiers in Immunology*, vol. 10, p. 1726, 2019.
- [99] V. V. Barygina, M. Becatti, G. Soldi et al., "Altered redox status in the blood of psoriatic patients: involvement of NADPH oxidase and role of anti-TNF- α therapy," *Redox Report*, vol. 18, no. 3, pp. 100–106, 2013.
- [100] T. Bacchetti, A. Campanati, G. Ferretti, O. Simonetti, G. Liberati, and A. M. Offidani, "Oxidative stress and psoriasis: the effect of antitumour necrosis factor- α inhibitor treatment," *The British Journal of Dermatology*, vol. 168, no. 5, pp. 984–989, 2013.
- [101] Y. Yildirim, E. G. Cellad, A. V. Kara et al., "Effect of intraperitoneal etanercept on oxidative stress in rats with peritonitis," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 9418468, 6 pages, 2016.
- [102] J. J. Xu, S. Guo, R. Xue et al., "Adalimumab ameliorates memory impairments and neuroinflammation in chronic cerebral hypoperfusion rats," *Aging (Albany NY)*, vol. 13, no. 10, pp. 14001–14014, 2021.
- [103] M. Czarnecka-Operacz and A. Sadowska-Przytocka, "The possibilities and principles of methotrexate treatment of psoriasis - the updated knowledge," *Postepy Dermatologii Alergologii*, vol. 31, no. 6, pp. 392–400, 2014.
- [104] G. Piskin, V. M. Heydendael, M. A. de Rie, J. D. Bos, and M. B. Teunissen, "Cyclosporin A and methotrexate are

- equally effective in reducing T cell numbers in psoriatic skin lesions but have no consistent effect on IFN- γ and IL-4 expression in psoriatic skin in situ,” *Archives of Dermatological Research*, vol. 294, no. 12, pp. 559–562, 2003.
- [105] T. Elango, H. Dayalan, P. Gnanaraj, H. Malligarjunan, and S. Subramanian, “Impact of methotrexate on oxidative stress and apoptosis markers in psoriatic patients,” *Clinical and Experimental Medicine*, vol. 14, no. 4, pp. 431–437, 2014.
- [106] A. Al Maruf, P. J. O'Brien, P. Naserzadeh, R. Fathian, A. Salimi, and J. Pourahmad, “Methotrexate induced mitochondrial injury and cytochrome c release in rat liver hepatocytes,” *Drug and Chemical Toxicology*, vol. 41, no. 1, pp. 51–61, 2018.
- [107] O. Akbulak, A. S. Karadag, N. Akdeniz et al., “Evaluation of oxidative stress via protein expression of glutathione S-transferase and cytochrome p450 (CYP450) isoenzymes in psoriasis vulgaris patients treated with methotrexate,” *Cutaneous and Ocular Toxicology*, vol. 37, no. 2, pp. 180–185, 2018.
- [108] S. G. Felemban, M. A. Aldubayan, A. H. Alhowail, and I. S. Almami, “Vitamin B17 ameliorates methotrexate-induced reproductive toxicity, oxidative stress, and testicular injury in male rats,” *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 4372719, 11 pages, 2020.
- [109] M. Roghani, H. Kalantari, M. J. Khodayar et al., “Alleviation of liver dysfunction, oxidative stress and inflammation underlies the protective effect of ferulic acid in methotrexate-induced hepatotoxicity,” *Drug Design, Development and Therapy*, vol. Volume 14, pp. 1933–1941, 2020.
- [110] J. U. Welbat, S. Naewla, W. Pannangrong, A. Sirichoat, A. Aranarochana, and P. Wigmore, “Neuroprotective effects of hesperidin against methotrexate-induced changes in neurogenesis and oxidative stress in the adult rat,” *Biochemical Pharmacology*, vol. 178, article 114083, 2020.
- [111] M. C. Zimmerman, D. L. Clemens, M. J. Duryee et al., “Direct antioxidant properties of methotrexate: inhibition of malondialdehyde-acetaldehyde-protein adduct formation and superoxide scavenging,” *Redox Biology*, vol. 13, pp. 588–593, 2017.
- [112] D. Boda, C. Negrei, F. Nicolescu, and C. Balalau, “Assessment of some oxidative stress parameters in methotrexate treated psoriasis patients,” *Farmácia*, vol. 62, pp. 704–710, 2014.
- [113] A. García-Sánchez, A. G. Miranda-Díaz, and E. G. Cardona-Muñoz, “The role of oxidative stress in physiopathology and pharmacological treatment with pro- and antioxidant properties in chronic diseases,” *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 2082145, 16 pages, 2020.
- [114] A. Mei-Yen Yong, L. Juay Kang Hng, E. Tan Yu Yuan, S. Woo Jia Qian, and C. Tan Lixian, “Targeted and localized phototherapy for atopic dermatitis and psoriasis: an eight-year experience in a tertiary dermatological centre in Singapore,” *Photodermatology, Photoimmunology & Photomedicine*, vol. 37, no. 6, pp. 571–574, 2021.
- [115] P. Filipe, I. Emerit, A. Alaoui Youssefi et al., “Oxyradical-mediated clastogenic plasma factors in psoriasis: increase in clastogenic activity after PUVA,” *Photochemistry and Photobiology*, vol. 66, no. 4, pp. 497–501, 1997.
- [116] M. S. Cooke, M. D. Evans, K. Patel et al., “Induction and Excretion of Ultraviolet-Induced 8-Oxo-2'-deoxyguanosine and Thymine Dimers In Vivo : Implications for PUVA,” *The Journal of Investigative Dermatology*, vol. 116, no. 2, pp. 281–285, 2001.
- [117] H. Orimo, Y. Tokura, R. Hino, and H. Kasai, “Formation of 8-hydroxy-2'-deoxyguanosine in the DNA of cultured human keratinocytes by clinically used doses of narrowband and broadband ultraviolet B and psoralen plus ultraviolet A,” *Cancer Science*, vol. 97, no. 2, pp. 99–105, 2006.
- [118] S. Coimbra, H. Oliveira, F. Reis et al., “Psoriasis therapy and cardiovascular risk factors,” *American Journal of Clinical Dermatology*, vol. 11, no. 6, pp. 423–432, 2010.
- [119] S. Coimbra, H. Oliveira, F. Reis et al., “Erythroid disturbances before and after treatment of Portuguese psoriasis vulgaris patients: a cross-sectional and longitudinal study,” *American Journal of Clinical Dermatology*, vol. 13, no. 1, pp. 37–47, 2012.