Review Article

Noncoding RNA as Diagnostic and Prognostic Biomarkers in Cerebrovascular Disease

Ruiyuan Weng,1,2,3,4 Zhiwen Jiang,1,2,3,4 and Yuxiang Gu1,2,3,4

1Department of Neurosurgery, Huashan Hospital, Shanghai Medical College, Fudan University, China
2Neurosurgical Institute of Fudan University, China
3Shanghai Clinical Medical Center of Neurosurgery, China
4Shanghai Key Laboratory of Brain Function and Restoration and Neural Regeneration, China

Correspondence should be addressed to Yuxiang Gu; guyuxiang1972@126.com

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Noncoding RNAs (ncRNAs), such as microRNAs, long noncoding RNAs, and circular RNAs, play an important role in the pathophysiology of cerebrovascular diseases (CVDs). They are effectively detectable in body fluids, potentially suggesting new biomarkers for the early detection and prognosis of CVDs. In this review, the physiological functions of circulating ncRNAs and their potential role as diagnostic and prognostic markers in patients with cerebrovascular diseases are discussed, especially in acute ischemic stroke, subarachnoid hemorrhage, and moyamoya disease.

1. Introduction

Cerebrovascular disease (CVD) is one of the leading causes of disability and mortality worldwide. Recent epidemiological studies have shown that CVD was one of the top ten leading causes of total years of life lost worldwide, especially in China that is of great severity [1, 2]. They mainly include ischemic and hemorrhagic events, but also rare diseases such as moyamoya disease.

The human genome is characterized by a number of noncoding RNAs (ncRNAs) of unknown function. The ncRNAs are involved in many cellular processes and include microRNAs (miRNAs), long noncoding RNAs (lncRNAs), circular RNAs, and transfer RNA-derived small RNAs (tsRNAs). In addition to their intracellular activity, ncRNAs are released within extracellular vesicles in the blood, which makes them potential biomarkers in different pathological conditions [3]. Several studies have reported that circulating ncRNAs could be measured both in tissues and in biological fluids, supporting their potential use as diagnostic and prognostic markers [4–8].

In this review, we discuss the physiological functions of circulating ncRNAs and their potential role as diagnostic markers, prognostic markers, and therapeutic targets in patients with cerebrovascular diseases, including acute ischemic stroke (AIS), subarachnoid hemorrhage (SAH), and moyamoya disease (MMD). Current challenges and future perspectives are also discussed.

2. Biogenesis and Transport of ncRNAs

A large number of studies have demonstrated that CVDs may affect the expression level of ncRNAs in body fluids. A multitude of ncRNAs have been described in body fluids, such as serum, plasma, urine, and breast milk [9]. Different techniques have been used, including RNA sequencing, microarray screening, and real-time quantitative polymerase chain reaction (RT-qPCR) [10].

Apparently, the ncRNAs existing in body fluids mostly come from specific cells, tissues, or organs, which are quite relevant to the disease conditions, whereas there are only a few studies that provided potential mechanisms involving ncRNAs in some disease conditions. So far, five transport mechanisms of ncRNA have been described (Figure 1): (1) exosomes, (2) microparticles, (3) apoptotic bodies, (4) ribonucleoproteins, including argonaute-2 (AGO2), nucleophosmin-
1 (NPM1), and high-density lipoproteins (HDLs), and (5) direct cellular connections, such as gap junctions, synapses, and intercellular bridges [10–16]. After secretion to the extracellular space, ncRNAs target specific cells and organs and exercise certain functions. Extracellular vesicles, like exosomes, microparticles, and apoptotic bodies, can transfer cargos from parental cells to recipient cells, achieving cell-to-cell communication, whereas the exact process how these extracellular vesicles are able to recognize the target cells remains unknown [10]. Ribonucleoproteins are other important carriers of ncRNAs. More than 90% of circulating miRNAs transfer through this pathway [10, 17]. The ncRNAs are quite stable and protected from RNases in body fluids, indicating that they are promising biomarkers in assessing the pathophysiological changes in the body. However, further studies are required to clarify the relationship between circulating ncRNAs and their carriers, how they interact with target cells, and what functions they exert [18].

3. ncRNA in Acute Ischemic Stroke

Effective management of patients with acute cerebrovascular disease relies on precise diagnosis and timely treatment, especially those with acute ischemic stroke (AIS). AIS accounts for around 80% of strokes and only has a narrow therapeutic window [19]. However, the accurate diagnosis of AIS can be disturbed by stroke mimics and other types of strokes. A computed tomography (CT) scan is usually able to detect a stroke from a blood clot or bleeding within the brain, but around half of patients are false negatives [20, 21]. MR diffusion-weighted imaging (DWI) is highly sensitive in detecting and localizing acute ischemic brain lesions but is limited by the length of the procedure, the lack of availability in remote areas, expensive costs, and prehospital settings [22]. Blood-based biomarkers with high sensitivity and specificity are therefore attractive, but those available to date have poor diagnostic accuracy [23].

Several questions in AIS diagnosis and treatment that remain to be solved can be briefly concluded as follows: the first is how to diagnose the AIS with high effectiveness and precision, which means that physicians should not only identify AIS immediately but also exclude other differential diagnoses like ICH, SAH, and stroke mimic confidently and further ascertain the stroke etiology (TOAST type) to instruct therapeutic strategies. In this setting, highly sensitive and specific biomarkers are needed in clinical practice. The second one is how to evaluate the short-term and long-term outcomes after stroke, which can be beneficial for planning subsequent therapeutic strategies and decide whether an aggressive treatment is helpful to promote post-stroke recovery. To answer these questions, various investigations that focus on the ncRNA-based biomarkers are summarized as follows.

3.1. miRNAs as Diagnostic Biomarkers in AIS. Circulating miRNAs have been reported as potential diagnostic and prognostic biomarkers in several studies (Table 1). Using RNA sequencing data, miR-125a-5p, miR-125b-5p, and
### Table 1: The studies about ncRNA as diagnostic and prognostic biomarkers in AIS patients.

<table>
<thead>
<tr>
<th>Reference</th>
<th>Sample size</th>
<th>ncRNA</th>
<th>Platform</th>
<th>Sample</th>
<th>Findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Long et al. [27]</td>
<td>2013</td>
<td>miR-30a, miR-126, let-7b</td>
<td>RT-qPCR</td>
<td>Plasma</td>
<td>miR-30a and miR-126 were significantly downregulated in AIS patients, while let-7b was downregulated in large-artery atherosclerosis but upregulated in other kinds of AIS patients</td>
</tr>
<tr>
<td>Leung et al. [28]</td>
<td>2014</td>
<td>miR-124-3p, miR-16</td>
<td>RT-qPCR</td>
<td>Plasma</td>
<td>miR-124-3p was significantly upregulated in ICH patients, while miR-16 was significantly upregulated in AIS patients</td>
</tr>
<tr>
<td>Wu et al. [25]</td>
<td>2015</td>
<td>miR-15, miR-16, miR-17-5p</td>
<td>RT-qPCR</td>
<td>Serum</td>
<td>These 3 miRNAs were significantly upregulated in AIS patients</td>
</tr>
<tr>
<td>Wang et al. [31]</td>
<td>2015</td>
<td>miR-29b</td>
<td>RT-qPCR</td>
<td>Peripheral leukocyte</td>
<td>miR-29b was downregulated in AIS patients and was negatively associated with the NIHSS score and infarct volume</td>
</tr>
<tr>
<td>Rainer et al. [35]</td>
<td>2016</td>
<td>miR-124-3p, miR-16</td>
<td>RT-qPCR</td>
<td>Plasma</td>
<td>Higher miR-124-3p was associated with mRS &gt; 2 and mortality within 90 days; miR-16 was the opposite</td>
</tr>
<tr>
<td>Huang et al. [36]</td>
<td>2016</td>
<td>miR-132</td>
<td>RT-qPCR</td>
<td>Serum</td>
<td>IncRNA expression profiling changes after stroke and can change over time</td>
</tr>
<tr>
<td>Dykstra-Aiello et al. [39]</td>
<td>2016</td>
<td>miR-9-5p, miR-9-3p, miR-124-3p, miR-128-3p</td>
<td>RNA-seq and RT-qPCR</td>
<td>CSF</td>
<td>miR-9-5p, miR-9-3p, miR-124-3p, and miR-128-3p were higher in bigger infarct size patients</td>
</tr>
<tr>
<td>Sørensen et al. [37]</td>
<td>2017</td>
<td>miR-150-5p</td>
<td>RT-qPCR</td>
<td>Plasma</td>
<td>Lower miR-150-5p was significantly associated with mortality within 90 days</td>
</tr>
<tr>
<td>Scherrer et al. [32]</td>
<td>2017</td>
<td>miR-223</td>
<td>RT-qPCR</td>
<td>Exosome</td>
<td>Upregulated miRNA was associated with the high NIHSS score and mRS &gt; 2 within 90 days</td>
</tr>
<tr>
<td>Jin and Xing [33]</td>
<td>2017</td>
<td>miR-126, miR-378, miR-101, miR-222, miR-218, miR-206</td>
<td>RT-qPCR</td>
<td>Plasma</td>
<td>These 3 miRNAs’ upregulation can identify the AIS from HC and TIA patients</td>
</tr>
<tr>
<td>Chen et al. [34]</td>
<td>2017</td>
<td>miR-125a-5p, miR-125b-5p, miR-143-5p</td>
<td>RNA-seq in the discovery cohort; RT-qPCR in the validation and republication cohort</td>
<td>Plasma</td>
<td>These 2 miRNAs were significantly downregulated in AIS patients</td>
</tr>
<tr>
<td>Tiedt et al. [24]</td>
<td>2017</td>
<td>miR-221-3p, miR-382-5p</td>
<td>RT-qPCR</td>
<td>Serum</td>
<td>Uregulated MIAT was associated with NIHSS scores, mRS, high-sensitivity C-reactive protein, and infarct volume</td>
</tr>
<tr>
<td>Wang et al. [26]</td>
<td>2017</td>
<td>miR-125b, miR125a, let-7b, let-7e, miR-7-2-3p, miR-1908</td>
<td>Microarray in the discovery cohort; RT-qPCR in the validation cohort</td>
<td>Serum</td>
<td>miRNAs were associated with the TOAST subtype</td>
</tr>
<tr>
<td>Zhu et al. [40]</td>
<td>2018</td>
<td>miR-150-5p</td>
<td>RT-qPCR</td>
<td>Peripheral leukocyte</td>
<td>miR-150-5p was significantly associated with mortality within 90 days</td>
</tr>
<tr>
<td>Gui et al. [30]</td>
<td>2019</td>
<td>miR-223</td>
<td>RT-qPCR</td>
<td>Exosome</td>
<td>Upregulated miRNA was associated with the high NIHSS score and mRS &gt; 2 within 90 days</td>
</tr>
</tbody>
</table>
miR-143-3p were constructed to discriminate between AIS and transient ischemic attack (TIA) patients and healthy controls. Longitudinal analysis showed a significant increase in miR-125a-5p, miR-125b-5p, and miR-143-3p during the first 24 hours after AIS, with miR-125b-5p and miR-143-3p returning to normal levels within 48 hours after AIS, which reveals a good discrimination in the acute phase. A random forest classification model presented good performance in differentiating AIS and healthy controls [24]. Another study showed that serum miR-15a, miR-16, and miR-17-5p significantly increased in patients with AIS compared to healthy controls [25]. Similarly, Wang et al. reported that serum miRNA-221-3p and miRNA-382-5p were downregulated in patients with AIS [26]. The downregulation of plasma miR-30a, miR-126, and let-7b was also described in patients with AIS [27].

The miRNAs are also thought to identify stroke subtypes (Table 1). As reported by Leung et al., miR-124-3p and miR-16 are potential diagnostic biomarkers between intracerebral hemorrhage (ICH) and AIS. Plasma concentration of miR-124-3p was significantly higher, whereas the concentration of miR-16 was significantly lower in patients with ICH than in AIS patients within 24 hours after the stroke onset [28]. These findings indicated that plasma miRNAs have the potential to distinguish ICH from AIS. Kalani et al. comprehensively profiled miRNAs across acute stroke subtypes through next-generation sequencing. The sequencing data were put into LASSO regression analysis to classify AIS, ICH, and subarachnoid hemorrhage (SAH) [29]. In discriminating different ischemic stroke subtypes, miRNA can also act as an effective biomarker. A total of 87 patients with AIS and 13 healthy subjects were recruited. The ROC analysis demonstrated that miR-125b, miR-125a, let-7b, and let-7e discriminate between acute stroke due to cardiac embolism and other subtypes of stroke. Besides, miR-7-2-3p and miR-1908 showed significant AUC in both large-artery atherosclerosis and lacunar infarct patients [30].

3.2. miRNAs as Prognostic Biomarkers in AIS. The predictive value of miRNAs in patients with AIS is shown in Table 1. Wang et al. reported that miR-29b was significantly downregulated in patients with AIS and negatively associated with National Institute of Health Stroke Scale (NIHSS) scores and stroke volume. Of note, the overexpression of miR-29b reduced the infarct volume and brain edema and infarct volume of the brain in mice [31]. In a prospective cohort study, miRNA-150-5p was negatively associated with mortality in patients with AIS within 90 days after the stroke onset [32]. Some miRNAs related to angiogenesis, including miR-126, miR-378, and miR-101, were negatively correlated with NIHSS scores, while miR-222, miR-218, and miR-206 were positively associated with NIHSS scores [33]. The exosomal miR-223 was positively associated with AIS occurrence, stroke severity, and short-term outcomes [34]. In another study conducted by Rainer et al., plasma miR-124-3p was elevated in patients with AIS who died within 3 months after hospital admission, while miR-16 was better associated with survival [35]. A high level of serum miR-132 correlated with post-stroke cognitive impairment [36], while several brain-enriched miRNAs (miR-9-5p, miR-9-3p, miR-124-3p, and miR-128-3p) were elevated in patients with infarcts larger than 2 cubic meters [37]. Overall, several miRNAs are potential biomarkers associated with the prognosis of AIS.

3.3. Other Biomarkers in AIS. Other ncRNAs also play an important role in diagnostic and prognostic assessment of AIS (Table 1). Recently, Zuo et al. suggested that increased plasma levels of circFUND1C1, circPDSS5B, and circCDC14A may be useful to diagnose AIS. In addition, an opposite trend was observed in patients with good modified Rankin Scale (mRS) scores within 7 days, suggesting their prognostic value [38]. Like other ncRNAs, IncRNAs may change in patients with AIS. In a case-control study, a large array of differentially expressed IncRNAs were detected in blood. Specifically, IncRNA NR-002332 and IncRNA A131606 were upregulated, while IncRNA C10 and IncRNA E57-2 were downregulated [39]. At the same time, IncRNAs in the blood were associated with clinical outcomes: the IncRNA MIAT was significantly upregulated and correlated with the NIHSS score, mRS score, serum C-reactive protein, and infarct volume [40].

3.4. ncRNA as a Therapeutic Target in AIS. The secondary brain injuries are common problems after AIS, such as brain...
edema, ischemic reperfusion injury, and hemorrhagic transformation. Recent studies have demonstrated that ncRNAs make great contribution to exacerbate or attenuate these injury processes through affecting neuroinflammation, neural apoptosis, oxidative stress, microglia activation, and excitotoxicity. Besides, neuroprotective ncRNA or their mimics can serve as promising drugs for their ability to penetrate the blood-brain barrier (BBB). Therefore, ncRNA may play a crucial role in improving AIS outcomes when serving as the therapeutic target. For example, ncRNA can influence the AIS process through regulating the oxidative stress process [41]. The upregulation of miR-424 pre- and poststroke in middle cerebral artery occlusion (MCAO) mice can decrease the cerebral infarction volume and brain edema by inhibiting oxidative stress, cellular apoptosis, and microglia activation [42], while miR-106b-5p antagonist could also decrease the infarction volume and neurological deficit in rats by regulating oxidative stress after AIS [43]. Other regulation of ncRNAs can also attenuate the oxidative stress following ischemic stroke. For example, the downregulation of miR-93 and miR-182 or the upregulation of miR-23a-3p, and miR-99a protected the AIS brain [44–47]. miR-93 antagomir alleviates ischemic injury through the Nrf2/HO-1 antioxidant pathway [47]. miR-182 promoted nitric oxide (NO) and 3-nitrotyrosine (3-NT) production and caspase-3 expression, while reducing superoxide dismutase (SOD) and manganese SOD (MnSOD) activities [46]. On the contrary, miR-23a-3p attenuated oxidative stress injury by reducing the production of NO and 3-NT and increasing the production of SOD and MnSOD [44]. All these researches indicated that miRNA might be a promising therapeutic target by attenuating the oxidative stress process.

Other miRNAs, such as miR-223 [48], miR-29b [49], miR-29c [50], miR-17-92 [51], miR-124 [52], miR-210 [53], miR-139-5p [54], miR-let-7c-5p [55], miR-107 [56], miR-207 [57], miR-335 [58], miR-22 [59], miR-9 [60], miR-378 [61], miR-122 [62], miR-210 [63], miR-455 [64], and miR-363 [65], can also reduce infarction volume and improve outcomes in animal models through various mechanisms. For example, miR-223 lowered the levels of glutamate receptors to reduce excitotoxicity and has a therapeutic role after stroke [48]. miR-139-5p agomir reduced neural apoptosis by inhibiting human growth transformation-dependent protein (HGTDP-P), providing a new therapeutic insight [54].

On the contrary, the overexpression of miR-145 [66], miR-320a [67], miR-497 [68], miR-let-7f [69], miR-181a [70], miR-181b [71], miR-103-1 [72], miR-30a [73], miR-124 [74], miR-134 [75], miR-200c [76], miR-155 [77], miR-24 [78], miR-182 [46], miR-493 [79], miR-383 [80], miR-106b-5p [43], miR-15a/16-1 [81], miR-30d-5p [82], miR-337 [83], and miR-337-3p [84] exacerbates the infarction volumes, edema, and neuroinflammation. For instance, inhibition of miR-377 decreased cerebral infarct volume and suppressed cerebral inflammation but promoted angiogenesis in MCAO rats [83]. Reducing poststroke miR-200c was also a potential target to mitigate infarction volume and neurological deficit by inducing reelin expression in mice [76].

4. ncRNAs in Aneurysmal Subarachnoid Hemorrhage

Subarachnoid hemorrhage (SAH) is a medical emergency, accounting for 2–7% of all stroke cases, mostly due to aneurysm rupture in over 80% of cases [85]. The diagnosis of SAH mainly relies on CT scan. If the CT scan is not definitive, the next recommended diagnostic tool is the lumbar puncture [86]. Besides, SAH is also a life-threatening disease with a case fatality of 25–35%, most of which results from the subsequent cerebral vasospasm (CVS) and delayed cerebral infarction (DCI) who survive at the first bleeding event [85–89]. Therefore, the development of a predictive biomarker would be helpful to prevent the development of CVS and understand the precise mechanism behind CVS. The emerging ncRNAs suggest future developments in the diagnostic and prognostic assessment of SAH.

4.1. ncRNAs as Diagnostic Biomarkers in SAH. Several studies have demonstrated that the ncRNA signature in SAH was quite different from that without SAH (Table 2). A microarray analysis and RT-qPCR were utilized to confirm the association among health controls, SAH with DCI, and SAH without DCI. This single study demonstrated that serum miR-132 and miR-324 were upregulated in SAH, compared with healthy controls, while the differences between DCI and non-DCI were not statistically significant. A possible reason was that the sample size was insufficient [90]. Using a similar methodology, Lai et al. reported that miR-502-5p, miR-1297, and miR-4320 were overexpressed in patients with SAH. Seven days after diagnosis, serum miR-502-5p and miR-1297 were significantly higher in patients with SAH. Additionally, at the 7th day after SAH, serum miR-502-5p and miR-1297 levels were significantly higher in patients with increased higher World Federation of Neurological Surgeons (WFNS) and mRS at the ninth month after stroke, which can represent the worse progression of SAH [91]. Another research also determined that serum miR-1297 was directly associated with a higher WFNS grade, Hunt-Hess grade, higher Fisher score, and poor one-year outcome [92]. In cerebrospinal fluid, miR-92a and let-7b decreased, whereas miR-491 increased over time in patients with SAH [93].

4.2. ncRNAs as Prognostic Biomarkers in SAH. With the help of the differential expression profile of ncRNA in SAH patients’ body fluids, ncRNAs are simultaneously competent to distinguish or predict some severe complications after SAH (Table 2). Styli et al. reported that miR-27a-3p, miR-516a-5p, miR-566, and miR-1197 were expressed in cerebrospinal fluid (CSF) differently between patients with cerebral vasospasm (CVS) and those without [94]. In another study, Lu et al. described that 4 circulating miRNAs (miR-4532, miR-4463, miR-1290, and miR-4793) differentiated patients with SAH with delayed cerebral infarction (DCI) from those without DCI by using a machine learning method [95]. A further prospective case-control study demonstrated that an array of miRNA profiles were overexpressed in the CSF of patients with SAH compared with healthy controls. Of interest, the angiographic CVS after SAH was associated...
with an increase in miR-132-3p, -19b-3p, -210-3p, -221-3p, and -484 [96], miR-15a and Kruppel-like factor 5 (KLF5), a potent modulator of miR-15a expression, may also be involved in CVS [97]. Several lncRNAs were also investigated in patients with SAH. According to Pan et al., lncRNAs ZFAS1 and MALAT1 were significantly upregulated, whereas lncRNAs LINC00261 and LINC01619 were downregulated in patients with SAH and CVS compared with those without CVS. Moreover, two lncRNAs (MALAT1 and LINC01619) accurately predicted CVS in around 90% of cases [98].

### 4.3. ncRNA as a Therapeutic Target in SAH

Several investigations have demonstrated that regulation of ncRNA influences many pathophysiological processes after SAH, including apoptosis, autophagy, neuroinflammation, and brain edema. Therefore, ncRNAs serve as promising therapeutic targets in SAH by regulating these underlying processes. For example, circulating exosomal miR-193b-3p treatment suppressed the expression and activity of HDAC3, reducing inflammation reaction in mice after SAH [99]. Intracerebroventricular injection of miR-103-3p antagonist before SAH reduced BBB permeability and improved neurological function [100]. Additionally, by downregulating iNOS and inhibiting the NF-κB signaling pathway, miR-195-5p attenuated white matter injury and SAH-induced vasospasm [101]. Extracellular vesicle derived from the mesenchymal stem cell could transfer miR-21 to neurons, promoting neuronal survival and improving cognitive function after SAH [102]. Liang et al. showed that, in a rat SAH model, IncRNA MEG3 overexpression increased cell apoptosis [103]. Besides, other studies demonstrated that the regulation of miR-26b [104], miR-706 [105], miR-206 [106], miR-675, and let-7a [107] could also affect the brain injury and outcomes after SAH. These results might provide a deeper understanding of the pathophysiological processes in brain injury after SAH, as well as potential therapeutic targets for the translational researches.

### 5. ncRNA Biomarkers in Moyamoya Disease

Moyamoya disease (MMD) is a rare, chronic, and progressive disorder of blood vessels in the brain. MMD is characterized by progressive occlusion of the internal carotid artery or its terminal branches, associated with the formation of collateral vessels at the base of the brain [108]. It is associated with vascular cognitive impairment [109, 110] and an increased risk of stroke [111, 112]. Although the diagnosis and treatment of MMD are of high standard, the efficiency still needs improvement. According to the current guidelines from Japan [113], the diagnosis requires an angiogram. The diagnosis is mainly characterized by the morphological abnormalities of cerebral arteries but not etiological or pathogenetic abnormalities [114]. Thus, diagnostic biomarkers that can reflect the disease process are eagerly awaited.

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**Table 2: The studies about ncRNA as diagnostic and prognostic biomarkers in SAH patients.**

<table>
<thead>
<tr>
<th>Reference</th>
<th>Sample size</th>
<th>ncRNA</th>
<th>Platform</th>
<th>Sample</th>
<th>Findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Su et al. [90] 2015</td>
<td>20 HC and 40 SAH</td>
<td>miR-132, miR-324</td>
<td>Microarray and RT-qPCR</td>
<td>Serum</td>
<td>miR-132 and miR-324 were higher in SAH patients</td>
</tr>
<tr>
<td>Powers et al. [93]2016</td>
<td>8 SAH</td>
<td>—</td>
<td>NanoString array and RT-qPCR</td>
<td>CSF</td>
<td>miRNA expression pattern changed over time after SAH</td>
</tr>
<tr>
<td>Lai et al. [91] 2017</td>
<td>10 HC and 60 SAH</td>
<td>miR-502-5p, miR-1297, miR-4320</td>
<td>Microarray and RT-qPCR</td>
<td>Serum</td>
<td>miR-502 and miR-1297 were associated with WFNS and mRS at 9 months</td>
</tr>
<tr>
<td>Styli et al. [94] 2017</td>
<td>4 HC and 20 SAH</td>
<td>miR-27a-3p, miR-516a-5p, miR-566, and miR-1197</td>
<td>NanoString array</td>
<td>CSF</td>
<td>These miRNAs were differentially expressed between SAH patients with and without CVS</td>
</tr>
<tr>
<td>Lu et al. [95] 2017</td>
<td>20 HC and 40 HC</td>
<td>miR-4532, miR-4463, miR-1290, and miR-4793</td>
<td>RT-qPCR</td>
<td>Plasma</td>
<td>4-miRNA characterized SAH patients with DCI</td>
</tr>
<tr>
<td>Bache et al. [96] 2017</td>
<td>10 HC and 27 SAH</td>
<td>miR-21 and miR-221</td>
<td>High-throughput RT-qPCR</td>
<td>CSF</td>
<td>The dysregulation of miR-15a and KLF4 after SAH may result in CVS</td>
</tr>
<tr>
<td>Kikkawa et al. [97] 2017</td>
<td>3 HC and 8 SAH</td>
<td>miR-15a</td>
<td>Microarray and RT-qPCR</td>
<td>Plasma and CSF</td>
<td>miR-1297 was associated with WFNS, Hun-Hess grade, and Fisher score and 1-year mRS</td>
</tr>
<tr>
<td>Sheng et al. [92] 2018</td>
<td>40 HC and 128 SAH</td>
<td>miR-1297</td>
<td>RT-qPCR</td>
<td>Serum</td>
<td>These two IncRNA signatures can predict the occurrence of CVS</td>
</tr>
<tr>
<td>Pan et al. [98] 2020</td>
<td>Discovery: 10 HC and 20 SAH, Validation: 65 SAH with and without CVS</td>
<td>IncRNA MALAT1, IncRNA LINC01619</td>
<td>RT-qPCR</td>
<td>CSF</td>
<td>These two IncRNA signatures can predict the occurrence of CVS</td>
</tr>
</tbody>
</table>

Abbreviation: CSF: cerebrospinal fluid; CVS: cerebral vasospasm; DCI: delayed cerebral infraction; HC: health control; mRS: modified Rankin Scale; RT-qPCR: real-time quantitative polymerase chain reaction; SAH: subarachnoid hemorrhage; WFNS: World Federation of Neurological Surgeons.
Some studies focused on ncRNAs in MMD (Table 3). In 2014, Dai et al. reported that serum miR-106b, miR-130a, miR-126, and miR-125a-3p were differentially expressed in patients with MMD compared with healthy controls [115]. Wang et al. suggested that transfer RNA-derived fragments (tRFs) are associated with the disease [116]. Ma et al. profiled the circRNA transcriptome of circulating neutrophils in asymptomatic patients with MMD, showing a differential expression of hsa-circRNA-100146, hsa-circRNA-102534, hsa-circRNA-036592, hsa-circRNA-405463, and hsa-circRNA-405324 [117]. Similar methods have been implemented also in the whole blood of patients with MMD [118]. Another study investigated exosome miRNAs from CSF samples of 31 patients and 31 health controls, showing that miR-3679-5p, miR-6165, miR-6760-5p, and miR-574-5p had significant diagnostic power for discriminating between patients and healthy controls [119].

### 6. Current Challenges and Future Perspectives

Although several ncRNA biomarkers are currently rapidly developing, the use of ncRNA as effective biomarkers in the clinical settings still has to face some unavoidable challenges. Several ncRNAs have been tested as biomarkers in cerebrovascular diseases. They have achieved some success in distinguishing sick people from healthy controls, but sample size in these studies was relatively small and the ability of ncRNAs to discriminate between different disease subtypes or other similar diseases needs further study. At present, the sensitivity, specificity, and reproducibility of ncRNAs are not at their best. Furthermore, the standardization of sampling and testing specimens collected from different body fluids requires more investigation, since different blood centrifugation conditions, sample storage conditions, sequencing platforms, and ncRNA isolation kits can affect the outcomes [120–122]. Moreover, patients from real-life conditions have several comorbidities that may influence the expression of circulating ncRNAs, such as hypertension, diabetes mellitus, and cancer [123, 124]. Even different lifestyles, like smoking or dietary structures, affect the expression of circulating ncRNAs, such as miR-126, miR-130a, and miR-125a, which influence the diagnosis of coronary artery disease when using miR-126 as a ncRNA-based biomarker [125, 126]. Additionally, the exact regulatory mechanisms of ncRNA and their physical functions are not clear. The analysis of confounding factors that affect ncRNA expression would be difficult.

Despite the shortcomings mentioned before, ncRNA biomarkers are still promising biomarkers in cerebrovascular diseases. The first ncRNA-based biomarker, IncRNA prostate cancer antigen 3, approved by FDA in 2012, has been routinely utilized in prostate cancer diagnosis, which stimulated the further development of ncRNA biomarkers in other diseases [127, 128]. ncRNAs are abundant and easily detectable in body fluids and are especially appealing as biomarkers because they are not prone to RNase degradation and remain stable in stored samples [129–131].

With the rapid development of artificial intelligence and machine learning (ML), we have never been closer to help physicians making data-driven medical decisions. The concentration of circulating ncRNAs may become essential to diagnose disease and predict outcomes. Several attempts have been made to determine their potential as biomarkers,
using random forest algorithms, support vector machines, and LASSO [4–8, 24]. However, the significance of ncRNAs in cerebrovascular diseases remains poor. Further studies using ML are ongoing and will shed new light on the topic.

This review discusses the important researches about the present situation and the advance in diagnostic and prognostic ncRNA biomarkers of several cerebrovascular diseases. Although the researches of ncRNA in cerebrovascular disease remain at the preclinical stage, such studies gave us clues of understanding cerebrovascular pathophysiology and finally would drive us to a more accurate diagnosis and prognosis for cerebrovascular diseases. Further studies using ML are ongoing and will shed new light on the topic.

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this article.

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