Research Article

Hydroxychloroquine Potentiates Apoptosis Induced by PPARα Antagonist in 786-O Clear Cell Renal Cell Carcinoma Cells Associated with Inhibiting Autophagy

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Received 22 November 2020; Revised 14 March 2021; Accepted 5 April 2021; Published 19 April 2021

1. Introduction

Renal cell carcinoma is one of the malignant tumors derived from normal renal tubular epithelial cells, and clear cell renal cell carcinoma (ccRCC) is its major pathological pattern [1]. According to epidemiological investigations, the morbidity of renal cell carcinoma is ranked second among the various urinary system neoplasms [2]. Metastasis occurs in about 1/3rd of the initially diagnosed renal cell carcinoma patients [3]. Owing to the presence of genetic mutations, renal cell carcinoma can resist traditional chemotherapeutic drugs and radiation [4]. Antiangiogenetic and tyrosine kinase inhibitors, such as sunitinib and sorafenib, have been developed and applied in clinical studies [5, 6]. Unfortunately, the resistance of renal cell carcinoma to these drugs has been reported in recent years [7, 8]. Hence, it is essential to develop novel targeted drugs for renal cell carcinoma.

Peroxisome proliferator-activated receptor (PPAR) is a member of the steroid hormone receptor superfamily that consists of three major subtypes, namely, PPARα, PPARβ/δ, and PPARγ [9]. PPARα heterodimer with retinoid X receptor (RXR) can bind to peroxisome proliferator responsive element (PPRE) and regulate the expression of several genes associated with lipid, glucose, and amino acid metabolism, as well as inflammatory response [10]. Over the years, researchers have discovered that PPARα may be a potential target in managing leukemia, melanoma, breast cancer, colon cancer, and glioblastomas [11–15]. With regard to ccRCC, Aboud et al. reported that the PPARα antagonist, GW6471, can induce apoptosis and cell cycle arrest [16]. PPARα may
be one of the potential targets for treating renal cell carcinoma.

Autophagy is a mechanism in which organelles or macromolecular proteins are "wrapped" into autophagosomes. Small molecules are generated in the lysosomes from the decomposed organelles or macromolecules for recycling by the eukaryotic cells [17]. Autophagy-related genes (ATGs) code most of the autophagy-associated proteins for the initialization of autophagy and the formation of autophagosomes. Autophagic flux is an integral process of autophagy [18]. The basal level of autophagy can regulate proliferation, growth, and differentiation to achieve cellular homeostasis [19, 20]. When cells are in stress, such as during cell cycle arrest, energy deficiency, and growth inhibition, the level of autophagy is enhanced so that they can adapt themselves to the modified conditions and avoid death or other adverse outcomes [21, 22]. The role of autophagy in tumor cell growth and death is unclear, but some researchers believe that it has a protective role by enhancing apoptosis or inducing autophagic cell death, which exerts a "double sword effect" on the fate of the tumor cells [23].

It is unclear whether GW6471 can induce autophagy; besides, its influence on the fate of the ccRCC cells also remains unknown. Therefore, in this study, we attempted to find whether autophagy can be induced in the 786-O ccRCC cell line by GW6471. Furthermore, we tried to ascertain the role of GW6471 in the 786-O cells. We discovered that the expression of PPARα was low in highly differentiated ccRCC tissues and 786-O cells but high in poorly differentiated ccRCC tissues. Subsequently, we came up with evidence that GW6471 can decrease the viability of the 786-O cells and induce integral autophagy. When the 786-O cells were cotreated with GW6471 and hydroxychloroquine, which is an inhibitor of autophagy, the viability was much lower than when treated with GW6471 or hydroxychloroquine alone, and apoptosis was promoted. Moreover, when human kidney 2 (HK-2) cells were cotreated with GW6471 and hydroxychloroquine, there was no significant effect on cell viability. Thus, the combination of hydroxychloroquine and GW6471 may serve as a novel and potentially beneficial strategy for managing ccRCC. Moreover, this approach is likely to be safe as it has only minimal effects on the normal renal tissues.

2. Materials and Methods

2.1. Cell Lines and Culture. The 786-O ccRCC cell line and the HK-2 human normal kidney tubular epithelial cell line were obtained from the Cell Bank of the Chinese Academy of Science. The two cell lines were cultured in Roswell Park Memorial Institute-1640 medium (Gibco, USA) supplemented with 10% fetal bovine serum (Gibco, USA) and 1% penicillin-streptomycin (Gibco, USA) at 37°C in an incubator (Thermo, USA) with 5% CO2 and saturated humidity.

2.2. Drugs. The PPARα antagonist GW6471 and the agonist WY14643 were purchased from MCE (USA), and hydroxychloroquine was sourced from Dulai Bio (China).

2.3. Immunohistochemical (IHC) Staining. The ccRCC tissues and the adjacent normal tissues of 40 patients were collected from the pathology department of the Third Affiliated Hospital of Soochow University, and all tissues were promptly embedded in paraffin by the staffs. Subsequently, all samples were deparaffinized in xylene and hydrated in a gradient of ethanol (100%, 95%, and 100%). The tissues were then heated in ethylene diamine tetraacetic acid (EDTA) for antigen retrieval and washed thrice with phosphate-buffered saline (PBS), 3 minutes for each wash. Afterwards, Immunohistochemical Staining Kit (Maxim, China) was applied for IHC staining with rabbit anti-human PPARα primary polyclonal antibody (Protech, USA, diluted in 1:100) according to the manufacturer’s instructions. The tissues were dehydrated in another gradient of ethanol (85%, 95%, and 100%). The nuclei were dyed by hematoxylin. All tissues were covered with microscope slides and observed using a fluorescent optical microscope in the optical mode (Olympus IX77, Japan). The semivalue of the optical density (OD) in IHC was measured using the software ImageJ (USA). The differentiation grade of the ccRCC tissues was analyzed independently by two experienced staffs of the pathology department. The patients’ characteristics of age and sex were listed in Table 1. The collection and use of the tissues were approved by the Ethics Committee of The Third Affiliated Hospital of Soochow University.

2.4. RNA Extraction and Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR). Total RNA of the 786-O and HK-2 cells was extracted using the Total RNA-pure Kit (FORE GENE, China) and dissolved in RNase-free ddH2O (FORE GENE, China) as per the manufacturer’s instructions. Next, the RNA samples were utilized for generating cDNA using the RT Easy™ I Kit (FORE GENE, China). Finally, the cDNA samples were employed for qRT-PCR with the Real-Time PCR Easy™ Kit (FORE GENE, China). The amplification was performed using ABI7500 (Applied Biosystems, USA). The primers of PPARα and GAPDH were synthesized by Takara Biotechnology (Takara, China), and the sequences are listed in Table 2. GAPDH was applied as an internal control, and the relative expression level of PPARα was calculated by the 2−ΔΔCT method [24]. The detailed procedure for qRT-PCR is given in Table 3.

2.5. Cell Viability Assay. The 786-O and HK-2 cells were seeded (2 × 103) in 96-well plates and incubated for adherence. They were treated on the next day with different groups of drugs for 6, 12, 24, or 48 hours. Then, 10 μL of Cell Counting Kit-8(CCK-8) reagent (Phygene, China) was added to each well. After incubation for 2 hours, the absorbance (A) of each well at 450 nm was measured by a microplate reader (Thermo, USA). The percentage of viable cells was calculated using the formula given below:

\[
\text{Cell Viability (\%)} = \left( \frac{A(\text{experimental group}) - A(\text{reagent blank})}{A(\text{untreated group}) - A(\text{reagent blank})} \right) \times 100%.
\]
2.6. Acridine Orange Staining. The 786-O cells were seeded \((1 \times 10^4)\) in 24-well plates for adherence and treated with different groups of drugs for 24 hours. Before staining, each well was washed with PBS for 15 minutes, three times, and the cells were fixed using 4% poly-formalin for 10 minutes. The dye acridine orange (AO) (Solarbio, China, diluted to \(10 \mu g/mL\) before staining) was added to each well. After 15 minutes, each well was again washed with PBS for 15 minutes, three times. All the wells were observed using a fluorescent optical microscope in the mode of fluorescence. The nuclei were dyed by AO in green, and the fluorescence of the lysosomes was in orange [25].

2.6.1. Apoptosis Assay by Flow Cytometry. The 786-O cells were collected after treatment and washed twice with PBS. Apoptosis was measured using the Annexin V-FITC/Propidium Iodide (PI) Double Fluorescent Dye Kit (Phygene, China) as per the manufacturer’s instructions. The percentage of apoptotic cells was calculated using FACS CantoII Flow Cytometry (BD, USA). Four “gates” were divided by a cross in the images of the flow cytometry: surviving cells were located in the lower left [Annexin V (-) and PI (-)]; early apoptotic cells were located in the lower right [Annexin V (+) and PI (-)]; late apoptotic cells were located in the upper right [Annexin V (+) and PI (+)]; necrotic cells were located in the upper left [Annexin V (-) and PI (+)].

2.7. Western Blotting. The collected 786-O cells were treated with drugs for 24 hours and washed twice with PBS. Cell lysis was determined using the Lysis Buffer Kit (KeyGEN, China) on ice according to the manufacturer’s instructions. All samples were centrifuged at 4°C and 12000 g/min for 5 minutes. The protein concentration was estimated using the Protein Quantitation Assay Kit (KeyGEN, China) as per the instructions. 5 × loading buffer was added to all the protein samples and heated to 100°C for 5 minutes. The proteins were separated in 8%, 10%, or 15% sodium dodecyl sulfate-polyacrylamide gel electrophoresis. Later, the protein in the gels was transferred to polyvinylidene difluoride membranes (Millipore, USA), and the membranes were blocked with 5% nonfat milk dissolved in Tris-buffered saline (TBST) for 1 hour. When blocking was accomplished, the membranes were incubated with diluted rabbit anti-human primary antibodies by TBST overnight (ABclonal, China, dilution of every antibody: PPARα: 1 : 2000, LC3 : 1: 1000, p62/Sequestosome-1 : 1 : 1000, PARP : 1 : 1000, Caspase-3 : 1 : 1000 GAPDH : 1 : 4000). On the next day, the membranes were washed by TBST for 15 minutes, three times, and incubated with goat anti-rabbit horseradish peroxidase (HRP) conjugated secondary antibody (ABclonal, China, diluted 1 : 8000 using TBST) for 1 hour. After incubating with the secondary antibody, the membranes were washed with TBST for 10 minutes, three times. Finally, the protein signal was detected using the Enhanced Chemiluminescent Plus reagent (Millipore, USA) and visualized by a protein imaging system (Tanon, China). The semivalue of the optical density in Western blotting was measured using the software ImageJ.

2.8. Statistical Analysis and Image Capture. Each experiment was performed at least three times independently. All value data were presented as mean ± SEM. The value data were analyzed, and the diagram was drawn using the software Prism 6 (USA). All comparisons of the value data were analyzed using unpaired Student’s \(t\)-test, one-way ANOVA analysis with Fisher Least Significant Difference (LSD) test, and chi-square test in this research of variance. All IHC and fluorescent images were captured by the software.
3. Results

3.1. PPARα Was Poorly Expressed in the Highly Differentiated ccRCC Tissues and 786-O Cell Line but Highly Expressed in the Poorly Differentiated ccRCC Tissues. In the first part of this research, the expression of PPARα was investigated in the ccRCC tissues and 786-O cell line. As depicted in Figures 1(a) and 1(b), IHC was performed for detecting the expression of PPARα in the ccRCC tissues. The results suggested that the expression of PPARα was lower in the highly differentiated ccRCC tissues than in the adjacent normal tissues. However, PPARα was highly expressed in the poorly differentiated ccRCC tissues, which was similar to that of the adjacent normal tissues. In the 786-O cell line, as illustrated in Figures 1(c)–1(e), we observed that the expressions of the PPARα mRNA and protein were lower when compared with the HK-2 cells, as inferred from qRT-PCR and Western blotting. On the other hand, we investigated the correlation of PPARα expression in ccRCC tissues among some clinical factors, including sex, age, and grade of differentiation. In Table 4, the expression of PPARα in ccRCC tissues was related to the grades of differentiation, but not to sex or age of patients.

3.2. GW6471 Decreased the Viability of the 786-O Cells in a Dose-Dependent but Not Time-Dependent Manner, and GW6471 Have No Effect of Cell Viability on HK-2 Cells with Dose-Dependent or Time-Dependent. Next, we performed a CCK-8 assay for detecting the variations in the viability of the 786-O cells and HK-2 cells treated with GW6471. As shown in Figure 2(a), the viability of the 786-O cells treated with GW6471 decreased in a dose-dependent (0-75 μM) manner. Furthermore, it was noted that WY14643 had no significant effect on the viability of the 786-O cells at any of the tested concentrations (0-75 μM). However, Figure 2(b) demonstrates that when the 786-O cells were treated with 25 μM of GW6471 for 12 hours, there was no significant difference in cell viability when compared with the cells in the 6h group. On the other hand, when the 786-O cells were treated with 25 μM of GW6471 for 24 hours, the cell viability was lowered when compared with the 6h and 12h groups. However, the viability of the 786-O cells treated with GW6471 for 48 hours was not significantly different when compared with those treated for 24 hours. Finally, as shown in Figure 2(c), there was no significant influence on viability in HK-2 cells treated with GW6471 or WY14643 at any of the tested concentrations (0-75 μM); Figure 2(d) demonstrates that when HK-2 cells treated with 25 μM GW6471 or WY14643 for 6, 12, 24, and 48 hours, and there was also no significant effect on cell viability. These results indicated that GW6471 can decrease the viability of the 786-O cells in a dose-dependent but not time-dependent manner, and GW6471 has no effect on cell viability in HK-2 cells with dose-dependent or time-dependent.

3.3. GW6471 Induced Integral Autophagy in 786-O Cells. GW6471 can decrease cell viability and induce cell cycle arrest and apoptosis in the 786-O cells; hence, we investigated whether GW6471 can induce autophagy in the 786-O cells [16]. Based on existing references about other human tumors treated with GW6471 and our own preliminary experiments, we chose 25 μM of GW6471 and WY14643 and 50 μM of hydroxychloroquine to treat the cells for a duration of 24 hours for the rest of the experiments in this research [26]. Hydroxychloroquine (or chloroquine, which was applied in some studies on autophagy) is an autophagy blocker that can inhibit the fusing of autophagosomes with lysosomes [27]. We first utilized AO for dyeing the lysosomes, as shown in Figures 3(a) and 3(b), and discovered that the fluorescent signal of the lysosomes normalized to per cell was brighter in the group treated with GW6471 than in the untreated and WY14643 treated groups. In the group treated with GW6471 and hydroxychloroquine, the fluorescent signal was much brighter than that in the group treated with GW6471 or hydroxychloroquine alone. As shown in Figures 3(c) and 3(d), the ratio of LC3-II/LC3-I, a protein which can be used to monitor autophagy induction and analysis, was found to decrease in the GW6471 alone group but increase in the GW6471 with hydroxychloroquine group, as shown in Figures 3(e) and 3(f) [27]. The results of AO dyeing and Western blotting indicated that GW6471 induced integral autophagy in the 786-O cells.

3.4. On Cotreatment of GW6471 and Hydroxychloroquine, the Cell Viability of 786-O Cells Was Further Decreased. Autophagy can affect the fate of the tumor cells; hence, we explored whether inhibiting autophagy can alter the viability

### Table 2: Sequence of primers of GAPDH and PPARα.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Sequence of upstream primer</th>
<th>Sequence of downstream primer</th>
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<tbody>
<tr>
<td>GAPDH</td>
<td>5'-GACACCGTCAAGGCTGAGAAC-3'</td>
<td>5'-TGGTTAAGACGCCAGTGGGA-3'</td>
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<tr>
<td>PPARα</td>
<td>5'-CCATCGGGAGGAGTCTG-3'</td>
<td>5'-CTACATTGCTGTCCAATGCTCCAC-3'</td>
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### Table 3: The procedure of qRT-PCR.

<table>
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<th>Temperature</th>
<th>Time</th>
<th>Cycle</th>
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<tr>
<td>1</td>
<td>94°C</td>
<td>3 min</td>
<td>1</td>
</tr>
<tr>
<td>2</td>
<td>94°C</td>
<td>5-10 s</td>
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</tr>
<tr>
<td></td>
<td>60°C</td>
<td>25 s</td>
<td>40</td>
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Image-Pro-Insight (USA). \( P < 0.05 \) was considered as a statistically significant difference.
of the 786-O cells treated with GW6471 subsequently. As presented in Figure 4, cotreatment with GW6471 and hydroxychloroquine for 24 hours decreased the viability of the 786-O cells more profoundly than the treatment with GW6471, hydroxychloroquine alone, and WY14643 with hydroxychloroquine.

3.5. On Cotreatment of Hydroxychloroquine and GW6471, the Ratio of Apoptotic Cells Was Promoted in 786-O Cells. The viability of the 786-O cells treated with a combination of GW6471 and hydroxychloroquine was lower than that of the cells treated with GW6471 or hydroxychloroquine alone. We investigated the mechanism of death in the 786-O cells

Figure 1: PPARα was poorly expressed in high differentiation of the ccRCC tissues and 786-O cell line, but highly expressed in poorly differentiation of ccRCC tissues. (a) IHC staining for PPARα among different grades of differentiation in ccRCC tissues when compared with the adjacent normal tissues. (b) Mean OD of IHC staining (normal: adjacent normal tissues; HD: highly differentiation; PD: poorly differentiation). (c) The expression of PPARα mRNA in 786-O and HK-2 cells by qRT-PCR normalized by GAPDH. (d) Western blotting for determining the expression of PPARα protein in 786-O and HK-2 cells. (e) Semiquantification of PPARα in 786-O and HK-2 cells normalized by GAPDH (n = 3, unpaired Student’s t-test, *P < 0.05).
treated with the combination of GW6471 and hydroxychloroquine. Flow cytometry was performed, and as shown in Figures 5(a) and 5(b), the ratio of apoptotic cells in the GW6471 group was higher than that in the untreated and WY14643 with or without hydroxychloroquine groups. When the 786-O cells were cotreated with GW6471 and hydroxychloroquine, the ratio of the apoptotic cells was higher than that in the cells treated with GW6471 alone. Subsequently, we performed western blotting for detecting cleaved-PARP and cleaved caspase-3, which are the two markers of cellular apoptosis. As inferred from Figures 5(c)–5(e), the expressions of cleaved PARP and cleaved caspase-3 in the GW6471, hydroxychloroquine alone, and WY14643 with hydroxychloroquine groups were higher than those in the untreated and WY14643 alone groups. When the 786-O cells were co-treated with GW6471 and hydroxychloroquine, the expressions of cleaved PARP and cleaved caspase-3 were the highest among all the experimental groups. The results displayed in Figure 5 indicate that hydroxychloroquine potentiated apoptosis in the 786-O cells treated with GW6471.

3.6. On Cotreatment of GW6471 and Hydroxychloroquine, HK-2 Cells Showed No Significant Effect on Cell Viability. We finally explored whether GW6471 combined with hydroxychloroquine can influence the viability of the HK-2 cells. As indicated in Figure 6, when the HK-2 cells were treated with the drugs for 24 hours, there was no significant effect on cell viability in any of the experimental groups, including the group of GW6471 with hydroxychloroquine. This result illustrates that cotreatment with GW6471 and hydroxychloroquine might not have a significant cytotoxic effect on the HK-2 cells.

4. Discussion

PPARα can regulate nutrient metabolism as a transcription factor. In hepatocytes, PPARα is highly expressed and can regulate carnitine palmitoyltransferase 1A and medium chain acyl-coenzyme A dehydrogenase, thereby enhancing fatty acid β-oxidation [28]. In the metabolism of glucose, PPARα heterodimer with RXR can bind to the genes encoding phosphofructokinase-1 (PFK-1) and pyruvate kinase (PK) with the sequence promoter, thus, regulating the expression of these two enzymes [29]. Additionally, in PPARα−/− mice, arginine is accumulated in the plasma and the level of nitric oxide (NO) is lowered [30], which implies that PPARα may influence amino acid metabolism. The expression of PPARα in the ccRCC cells has been researched in the past. In the present investigation, our results demonstrated that the expression of PPARα was low in the highly differentiated ccRCC tissues and the 786-O cell line but high in the poorly differentiated ccRCC tissues. The predominant morphological feature of the ccRCC cells is the presence of abundant lipid droplets in the cytoplasm [31]. Accumulation of lipid droplets in the ccRCC cells is associated with several genes, including the underexpression of carnitine palmitoyltransferase 1A (CPT1A) and the overexpression of the fatty acid synthase (FASN), perilipin-3 [32–34]. Moreover, Du et al. reported that the accumulation of lipid droplets in the 786-O cells is associated with the low expression of CPT1A, and glucose is the major ingredient for the formation of the lipid droplets in the 786-O cells [35]. However, Wettersten et al. reported that CPT1A may catalyze the synthesis of acyl-carnitines from free fatty acids for anti-inflammatory action in high-grade ccRCC [36]. The CPT1A gene can be regulated by PPARα [37]. Based on our research of PPARα expression in different grades of ccRCC tissues and previous studies by other investigators, it could be stated that PPARα might exert complex and variable effects on proliferation and growth in different grades of ccRCC. Finally, as the results of Table 4, the expression of PPARα in ccRCC tissues was correlated to the grade of differentiation, but not to the sex or age of patients. PPARα may be a potential marker for differentiating grades of ccRCC tissues. In the future, more samples of ccRCC tissue will be collected for investigating the correlation between the expression of PPARα in ccRCC and clinically relevant profiles.

The role of PPARα in the treatment of different tumors is also complex and variable. PPARα is poorly expressed in melanoma, and fenofibrate (an agonist of PPARα) can inhibit the metastasis of melanoma via the downregulation of AKT and ERK1/2 phosphorylation [38]. PPARα is highly expressed in parangangioma. When the parangangioma cell lines were treated with GW6471, cellular apoptosis was promoted and migration was inhibited via the repression of the PI3K/GSK3/β-catenin pathway [26]. On the other hand, PPARα is overexpressed in pancreatic cancer tissues when compared with their adjacent normal tissues. However, clofibrate (another PPARα agonist) can induce apoptosis and sensitize the cells to radiation through the downregulation of the Wnt/β-catenin pathway. In the ccRCC cells, glycolysis is the major pathway for energy generation. GW6471 can block glycolysis and induce cell cycle arrest via the downregulation of the oncogene c-Myc [39]. In this research, we discovered that although PPARα was poorly expressed in the 786-O cells when compared with the HK-2 cells, the viability of the former cells can be decreased by GW6471 in a dose-dependent but not time-dependent manner. Our results are similar to those obtained by Aboud et al. [16]. However, we discovered that WY14643 has no significant effect on the viability of the 786-O cells. Based on the

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<tr>
<th>Table 4: The relationship between PPARα expression and some clinical factors of ccRCC patients (n = 40, chi-square test).</th>
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<tbody>
<tr>
<td>Clinical factors</td>
</tr>
<tr>
<td>Sex</td>
</tr>
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<td>Male</td>
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<td>High</td>
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metabolism of ccRCC, we speculated that the low expression of PPARα may be essential for maintaining the basal level of glycolysis in the 786-O cells. The reason for WY14643 not having a significant effect on the viability of the 786-O cells remains to be explored in future studies.

In renal cell carcinoma, the basal level of autophagy plays an important role in cellular growth and proliferation. Ma et al. applied IHC and discovered that the expression of ATG9 is higher in the ccRCC tissues than in the adjacent normal tissues and that this elevated expression is an independent risk factor of prognosis in ccRCC [40]. Lu et al. reported that UNC-51-like kinase 1 (ULK1, ATG1) is another autophagy-associated protein and that the high expression of ULK1 is an independent risk factor of prognosis in ccRCC [41]. When cells are under stress, autophagy can be induced via the suppression of the PI3K/AKT/mTOR and MAPK/ERK/mTOR pathways or the activation of the AMPK/mTOR pathway [42–44]. Based on the findings of Aboud et al. and ours, we further explored whether autophagy can be induced by GW6471 in the 786-O cells. AO is a fluorescent dye that can stain the acidic vesicular organelles in red and the nuclei in green [25]. Hydroxychloroquine is a weakly alkaline drug that can slightly increase the pH of the lysosomes, inhibit autophagosome-lysosome fusion, and induce an autophagy-independent severe disorganization of the Golgi and endo-lysosomal systems, which may cause fusion impairment [45]. When autophagy is induced in the cells and the autophagic flux is fluent, the AO signal can be

Figure 2: GW6471 decreased the viability of the 786-O cells in a dose-dependent but not time-dependent manner, and GW6471 has no effect of cell viability on HK-2 cells with dose-dependent or time-dependent. (a) Cell viability of 786-O cells treated with different levels of GW6471 and WY14643 (0–75 μM) for 24 h by CCK-8 assay. (b) Cell viability of 786-O cells treated with 25 μM GW6471 or WY14643 for 0, 12, 24, and 48 h by CCK-8 assay. (c) Cell viability of HK-2 cells treated with different levels of GW6471 and WY14643 (0–75 μM) for 24 h by CCK-8 assay. (d) Cell viability of HK-2 cells treated with 25 μM GW6471 and WY14643 for 6, 12, 24, and 48 h by CCK-8 assay. (n = 3, unpaired Student’s t-test *P < 0.05).
detected significantly in the cells. Since the autophagic flux is blocked by hydroxychloroquine, lysosomes and autophagosomes can be accumulated and observed in the cells [46].

We discovered that when the 786-O cells were treated with GW6471 alone, the fluorescent signal of the lysosomes dyed with AO was brighter than that of the untreated and...
and induce autophagy via the downregulation of the PI3K/AKT/mTOR pathway. When the OS-RC-2 renal cell carcinoma cells were cotreated with sunitinib and chloroquine, the decrease in cell viability was more than that of the cells treated with sunitinib alone, which could be attributed to the promotion of apoptosis [54]. Mammalian target of rapamycin (mTOR) is a sensor of nutrition and cellular growth, and it consists of two subtypes, namely, mTOR1 and mTOR2. mTOR can regulate autophagy as a key protein by “perceiving” the variations in the upstream signals of PI3K/AKT, MAPK/ERK, and AMPK [55]. In renal cell carcinoma, the expression of mTOR is higher than that in the normal renal tissues and is therefore acclaimed as a target in the treatment of RCC [56]. Zheng et al. reported that the mTOR dual inhibitor AZD-2014 can inhibit cell growth and induce autophagy in the 786-O and OS-RC-2 cells. In the two cell lines treated with a combination of AZD-2014 and chloroquine, cell viability was decreased further by the promotion of apoptosis [57]. This finding implies that AZD-2014 and chloroquine may have a synergistic effect in killing renal cell carcinoma cells. Autophagy induced by drugs in renal cell carcinoma may enhance cellular death. Ubenimex, an inhibitor of aminopeptidase, can enhance immunity and is considered as a potential drug for treating renal cell carcinoma [58]. Ubenimex can induce autophagy in the 786-O and OS-RC-2 cells, and when the two cell lines were cotreated with ubenimex and rapamycin (an inhibitor of mTOR), cell growth was inhibited. However, when 3-methyladenine (3-MA), another autophagy blocker, was combined with ubenimex, there was no significant difference in cell viability when compared with ubenimex alone [59]. This result alludes that the autophagic flux enhanced by ubenimex is correlated with cell death in renal cell carcinoma. Sorafenib is another inhibitor of angiogenesis and tyrosine kinase and is also applied for treating renal cell carcinoma [60]. The drug can inhibit cellular growth and induce autophagy via the downregulation of the PI3K/AKT/mTOR pathway in the 786-O and ACHN cells of renal cell carcinoma. When the cells were cotreated with 3-MA and sorafenib (or knockdown of the ATG5 gene), the toxic effect of sorafenib was reversed and cell viability was higher when compared to treatment with sorafenib alone [61]. This observation implies that autophagy induced by sorafenib can facilitate cell death in renal cell carcinoma. Interestingly, in our research, flow cytometry was performed for detecting the proportion of apoptotic cells, and the results indicated that the proportion of apoptotic cells in the GW6471 with hydroxychloroquine group was higher than that in the GW6471, hydroxychloroquine alone, and WY14643 with chloroquine groups. With the use of western blotting, we showed that when the 786-O cells were cotreated with GW6471 and hydroxychloroquine, the expressions of cleaved PARP and cleaved caspase-3 were higher than those in the cells treated with GW6471 or hydroxychloroquine alone. Thus, the combined use of GW6471 and hydroxychloroquine decreased the viability of the 786-O cells and promoted apoptosis. This result suggests that inhibiting autophagy can potentiate apoptosis in the 786-O cells treated with GW6471. These results discussed in this paragraph were summarized as a schematic diagram in Figure 7.
Though the role of autophagy in cell death is still unclear, some researchers have explained the relationship between the two with regard to the degradation of the organelles by the autophagosome and the generation of energy by autophagy. Some organelles such as mitochondria and endoplasmic reticulum and lipid droplets can be degraded by autophagosomes [62–64]. Wang et al. reported that gambogic acid can induce apoptosis and autophagy in pancreatic cancer. The level of reactive oxygen species (ROS) was found to increase due to the promotion of damaged mitochondria. It was observed that cotreatment of the pancreatic cells with gambogic acid and chloroquine lowered the cell viability to a greater extent and that the level of ROS was accentuated since the scavenging of the damaged mitochondria was inhibited [65]. In lung cells, perfluoroalkyl acid can decrease cell viability and induce autophagy by engulfing the damaged endoplasmic reticulum and avoiding excessive endoplasmic

![Figure 5: On cotreatment of hydroxychloroquine and GW6471, the ratio of apoptotic cells was promoted in 786-O cells. (a) Flow cytometry was performed for detecting apoptotic cells. (b) Ratio of apoptotic cells was calculated by flow cytometry. (c) The expression of cleaved-PARP and cleaved caspase-3 was analyzed by western blotting. (d) Semiquantification of cleaved-PARP normalized by GAPDH. (e) Semiquantification of cleaved caspase-3 normalized by GAPDH. (*P < 0.05, compared with untreated group, ##compared with group of GW6471).]

![Figure 6: On cotreatment of GW6471 and hydroxychloroquine, the HK-2 cells showed no significant effect on cell viability. (n = 3, CCK-8 assay).]
cotreatment with GW6471 and hydroxychloroquine was potentiated in 786-O cells cotreated with GW6471 and hydroxychloroquine in 786-O cells. GW6471 can induce apoptosis in ccRCC cells, with minimal side effects on normal renal tissues. According to the results of our research, we observed that PPARα was poorly expressed in highly differentiated ccRCC tissues and 786-O cell line, which was applied as a cellular model in this research. Hence, our findings may be valid for treating highly differentiated ccRCC only. However, using only one ccRCC cell line for analyzing is the limitation in our research, so we intend to apply more renal cell carcinoma cell lines such as Caki-2 or UM-RC-2 for validating the roles of PPARα in different pathological patterns of RCC.

5. Conclusion

Our research has shown that PPARα is poorly expressed in highly differentiated ccRCC tissues and 786-O cells but highly expressed in poorly differentiated ccRCC tissues. Autophagy can be induced by GW6471 in the 786-O cells and blocked by hydroxychloroquine. The combined use of hydroxychloroquine and GW6471 was found to be more effective in killing the 786-O cells than the use of GW6471 or hydroxychloroquine alone, which could be attributed to the promotion of apoptosis. Furthermore, this approach had little effect on the viability of the HK-2 cells, thereby asserting its safety on normal renal tissues. According to the results of this study, the combined use of GW6471 and hydroxychloroquine may serve as a novel and potentially beneficial clinical therapy for highly differentiated ccRCC. However, further in vivo studies and clinical trials are required. Moreover, the reason for PPARα agonists not having a significant effect on the proliferation of the 786-O cells and the signaling pathway of autophagy induced by GW6471 in the 786-O cells needs to be explored in the future.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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