

**Supplementary Table 1: Comparison of SVF cell counts obtained by manual counting vs. automated counting methods.**

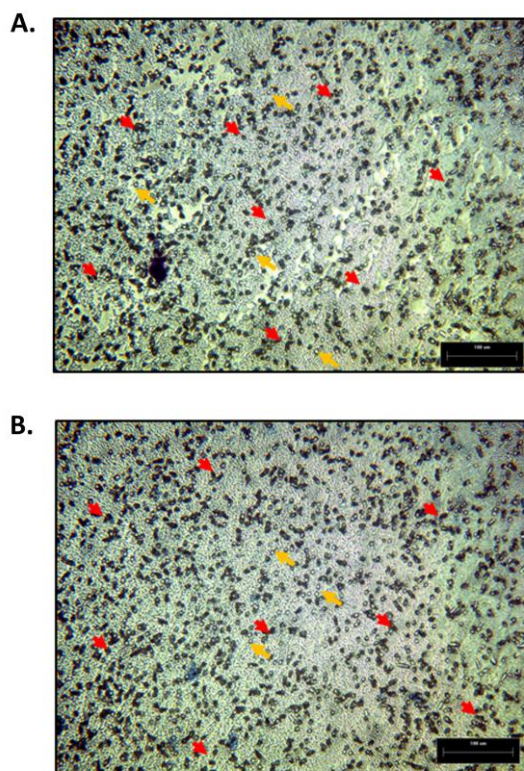
<b>Sample ID</b>	<b>Manual count (x 10<sup>6</sup>)</b>	<b>Automated count (x 10<sup>6</sup>)</b>
Sample 1	2.56	2.09
Sample 2	0.52	0.69
Sample 3	1.81	1.93
Sample 4	1.89	1.71
Sample 5	2.77	1.64
Sample 6	8.05	3.43
Sample 7	5.55	6.14
Sample 8	6.45	5.17
Mean±SD	3.7±2.6	2.8±1.9

**Supplementary Table 2: Yield and viability of SVF from lipoaspirates processed before and after passing through peristaltic pump.**

Sample ID	Control		Per. Pump.	
	Yield (x10 <sup>5</sup> /gram)	Viability %	Yield (x10 <sup>5</sup> /gram)	Viability %
Sample 1	2.4	95%	1.7	94%
Sample 2	2.6	98%	3.0	98%
Sample 3	0.56	98%	0.87	98%
Mean±SD	<b>1.85±1.1</b>	<b>96.7±2.3%</b>	<b>1.86±1.1</b>	<b>97.0±1.7%</b>

Difference between the control and per. pump. groups is not significant (p=0.99, paired t-test).

### Supplementary figure 1



**Supplementary figure 1. Retention of SVF cells on 5 µm polycarbonate track etch filters (PCTE).** The aqueous phase of digested lipoaspirate tissue was sequentially filtered through nylon filters of 100 µm and 35 µm pore size and SVF cells were ultimately recovered on a 5 µm PCTE filter. The 5 µm PCTE filter was then fixed in 1% paraformaldehyde for 20 min, stained with 0.1% toluidine blue (in 1% formaldehyde solution) for 1 h and then rinsed with water. Cells retained on the filter were visualized by light microscopy. Micrographs show a representative set of images from two different samples (A) & (B). Arrowheads indicate stained SVF cells (red) and filter pores (yellow). Scale bars=100 µm.