

# Dyskinesia in Parkinson's Disease Therapy

Guest Editors: Anna Rosa Carta, Andrea Giuffrida, and Gilberto Fisone





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Parkinson's Disease

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## Editorial

# Dyskinesia in Parkinson's Disease Therapy

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L-DOPA is still regarded as the standard pharmacotherapy for the treatment of the motor symptoms of Parkinson's disease (PD). However, the efficacy of this drug is limited by the emergence of dystonic and choreic involuntary movements, generally referred to as dyskinesia. Current interventions to treat dyskinesia are mainly based on continuous delivery of L-DOPA, administration of glutamatergic drugs (i.e., amantadine), replacement or combined administration of L-DOPA with less dyskinetic, albeit less effective, dopaminergic agonists, and deep-brain stimulation of discrete regions of the basal ganglia.

Preclinical research, on the other hand, is searching for novel approaches to the treatment of L-DOPA-induced dyskinesia, targeting alternative neurotransmitter systems, such as the serotonin, adenosine, and opioid systems, or identifying abnormalities in intracellular signal transduction and synaptic plasticity associated to this condition. This Special Issue discusses recent breakthroughs in this direction and, at the same time, provides an update of the clinical features and management of L-DOPA-induced dyskinesia.

The article by J. Guridi et al. describes the mechanisms underlying the various forms of dyskinesia produced by prolonged administration of L-DOPA. Basic pathophysiological mechanisms and current treatments are presented in detail and critically discussed.

I. Aviles-Olmos et al. focus on the comparison between L-DOPA-induced dyskinesia and graft-induced dyskinesia, an analogous condition observed in response to transplantation of fetal dopaminergic cells in PD patients. This article also provides a timely discussion of the use of gene therapy in PD and of its effects on dyskinesia.

The article by N. Tambasco et al. presents the clinical and epidemiological characteristics of dyskinesia and describes the management of this condition, based on the use of various types of dopaminergic agonists. On the same line, J. C. P. Piedad and A. E. Cavanna discuss more in detail the use of pramipexole, an agonist at dopamine D<sub>2</sub>-type receptors, in the treatment of dyskinesia.

A number of signaling components potentially implicated in dyskinesia have been discovered during the last decade. In this regard, V. Ghiglieri et al. provide a comprehensive overview of the mechanisms and the possible pathological consequences of abnormalities affecting various forms of corticostriatal plasticity associated to L-DOPA-induced dyskinesia.

Recently, studies have pointed to the serotonergic system as a key player in the aberrant effects produced by L-DOPA and linked to the emergence of dyskinesia. The article by S. Navailles and P. De Deurwaerdère describes the evidence at the basis of this hypothesis and discusses the use of therapeutic approaches designed to prevent this phenomenon. On the same subject, E. Shin et al. discuss the experimental and clinical evidence implicating the serotonin system in L-DOPA- and graft-induced dyskinesia.

Drugs acting as antagonists at adenosine A<sub>2A</sub> receptors have attracted considerable interest as antiparkinsonian and antidyskinetic agents. The article by M. Morelli et al. provides a critical appraisal of the potential therapeutic properties of these compounds, as determined in experimental models of PD and L-DOPA-induced dyskinesia.

Finally, G. Frazzitta et al. present an interesting study showing the beneficial effects produced on dyskinesia by

intensive physiotherapeutic rehabilitation. These results are discussed with regard to similar studies performed in animal models of PD.

### **Acknowledgments**

We hope that the reader will find these articles interesting and informative both at the preclinical and clinical level. We are very grateful to the contributors and to all the colleagues involved in the reviewing process.

*Anna Rosa Carta  
Andrea Giuffrida  
Gilberto Fisone*

## Review Article

# Clinical Features, Pathophysiology, and Treatment of Levodopa-Induced Dyskinesias in Parkinson's Disease

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Dyskinetic disorders are characterized by excess of motor activity that may interfere with normal movement control. In patients with Parkinson's disease, the chronic levodopa treatment induces dyskinetic movements known as levodopa-induced dyskinesias (LID). This paper analyzed the pathophysiology, clinical manifestations, pharmacological treatments, and surgical procedures to treat hyperkinetic disorders. Surgery is currently the only treatment available for Parkinson's disease that may improve both parkinsonian motor syndrome and LID. However, this paper shows the different mechanisms involved are not well understood.

## 1. Introduction

Hyperkinetic or dyskinetic disorders are characterized by excessive muscular activity that may interfere with normal movement control. Dyskinesias include different types of movement disorders such as chorea-ballism, dystonia, myoclonus, tics, and tremor. In patients with Parkinson's disease (PD), chronic levodopa treatment may induce various dyskinetic movements (levodopa induced dyskinesias (LID)) which are classified according to the phenomenology and also their temporal presentation in relation with the effect of levodopa.

The association between levodopa and the induction of dyskinesias was recognized soon after the introduction of levodopa [1, 2]. In the past, levodopa therapy was associated with the development of motor complications in about 80% of patients within 5 years of treatment [3, 4]. In patients with young onset PD, the incidence of LID was higher and ensued more rapidly [2, 5]. Currently, with the introduction and widespread use of dopaminergic agonists, the overall treatment exposure to levodopa is decreasing, especially in the first years of treatment; nevertheless, progression of the nigrostriatal deficit will facilitate the onset of LID at a later point in time. Thus, LID continues to be a common and important cause of disability in PD and one of the main reasons for recommending surgical treatment.

In this paper we describe the major clinical features, main pathophysiological and pharmacological abnormalities associated with LIDs, and the drug and surgical treatments currently available.

## 2. Clinical Presentation

LID may be divided into various presentation forms (Figure 1) [6].

- (1) "Peak dose" or "on" period dyskinesia related to high plasma levels of levodopa, in parallel with the maximal antiparkinsonian benefit. These are typically choreic in nature and predominantly involve the neck, trunk, and upper limbs, but dystonic movements may also occur.
- (2) Diphasic dyskinesia appears at the onset and offset of the levodopa effect, coinciding with arising and decaying plasma levodopa levels. This is characterized by repetitive and stereotyped repetitive, slow (<4 Hz) movements of the lower limbs often coinciding with 4 Hz tremor in the upper limbs [4], indicating the patient is not fully "on". In severe cases, the movements of the legs may lose the repetitive and stereotypic nature and resemble ballism. In a small proportion of patients, diphasic dyskinesias are very

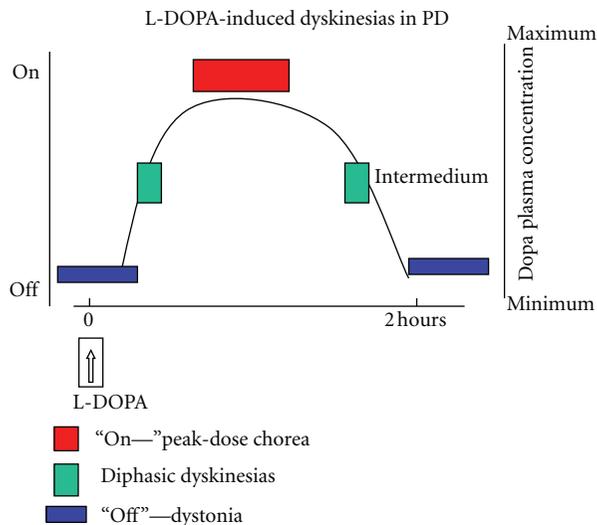


FIGURE 1: Relationship between LID and DOPA plasma level. “Peak of dose” or “on” period dyskinesia is correlated to high level of levodopa and in parallel with the maximal clinical benefit. Diphasic dyskinesia appears at the onset and offset of the levodopa effect in relationship with increment or decrement of plasma level. “Off” period dystonia is characterized by painful postures in lower extremities and is correlated with the lowest levodopa level. Generally a full spectrum of the three types is present in patients with motor fluctuations.

prominent while walking, drastically interfering with gait, and giving rise to a picturesque pattern [7]. Dystonic posture may also occur, although much less frequently.

- (3) “Off” period dystonia, characterized by fixed and painful postures more frequently affecting the feet, but which can be segmental or generalized in distribution.

A combination of any of these 3 types or indeed, all of them, may be observed in some patients throughout the levodopa (“off-on-off”) cycle. Until now LID, by definition, were associated with levodopa intake and, to a much lesser extent, with dopamine agonists used in monotherapy. Two more recent situations whereby dyskinesias can be induced in patients with PD despite not being treated with dopaminergic drugs are (1) patients treated with fetal cell transplants [8]; (2) patients treated with deep brain stimulation (DBS) of the subthalamic nucleus (STN) [9]. The former has no practical implications as experimental trials with fetal cell transplants are not a therapeutic option, but STN-DBS is fairly frequently applied. In the latter instance, adjusting the current parameters usually results in control of dyskinesias.

### 3. Pathophysiology of LID

Levodopa is converted into dopamine (DA) in many brain regions and “a priori” there are several sites where its dyskinesogenic effect could occur. The striatal origin of LIDs was suspected as soon as the problem was recognized in the early 1970’s but there were no experimental or clinical proofs.

Direct proofs arose unexpectedly when fetal mesencephalic cells transplanted into the striatum in experimental trials for PD were associated with dyskinesias with a similar clinical pattern than LID [10, 11].

Two main factors are involved in the origin of LID. (1) Degree of dopaminergic nigro-striatal depletion, which is related to disease duration and severity. (2) The pharmacokinetics and mechanism of action of levodopa, which delivers a discontinuous or pulsatile stimulation of dopaminergic receptors [3, 12]. Together, degree of nigro-striatal lesion and the action of levodopa interact to induce changes in corticostriatal transmission and plastic synaptic abnormalities in striatal spiny neurons, which ultimately may alter the physiological activity of striatopallidal circuits, leading to abnormal pattern of neuronal activity underlying LID [13, 14]. A direct demonstration of the link between short acting dopaminergic stimulation and changes in basal ganglia output was provided several years ago. It was shown that once or twice a day levodopa or apomorphine administration in parkinsonian monkeys induced dyskinesias which were associated with a reduction in the main firing frequency of globus pallidus internus (GPi) neurons [15, 16]. Similar results have been described in parkinsonian patients who were administered apomorphine during pallidal surgery. Here, the turning from the “off” parkinsonian condition to the “on” mobile state plus LID was associated with a significant reduction in the mean neuronal firing rate of the GPi and STN [17–19]. In addition, STN and GPi activity was decreased when assessed by regional brain uptake of 2-deoxyglucose, which measures afferent synaptic activity, in MPTP monkeys with dyskinesias induced by dopaminergic drugs [20]. Thus, reduced GPi inhibitory output activity to the thalamus leads to disinhibition of the thalamocortical projection, facilitating the abnormal recruitment of cortical motor areas which ultimately give rise to dyskinetic movements. In simple terms therefore, dyskinesias in general and LID in particular may be understood as the reverse of the parkinsonian state, whereby the latter is mainly characterized by overactivity of the STN and GPi output, leading to overinhibition of the thalamus and decreased thalamocortical activity (Figure 2) [20–24].

The metabolic activity reduction and firing reduction and firing frequency changes in firing pattern of GPi activity to the thalamus are thought to produce an increase in thalamocortical drive leading to dyskinesia.

Which striatopallidal circuits, if any, may be preferentially mediate LID has been a matter of discussion over the years.  $D_2$  mediated activation of the striato-pallidal projections in the “indirect” basal ganglia circuit was favored for a long time. Thus, pharmacological manipulation of the “indirect” circuit induces dyskinesias in monkeys which are similar to LID. For example, this is achieved by injecting bicuculline, a  $\gamma$ -amino butyric acid (GABA) antagonist into the globus pallidus externus (GPe), which results in increased GPe efferent activity and overinhibition of the STN [25] or by blocking STN glutamatergic projection, which provokes GPi neuronal hypoactivity and involuntary movements in the monkey [26, 27]. Moreover, it is well known that STN lesion

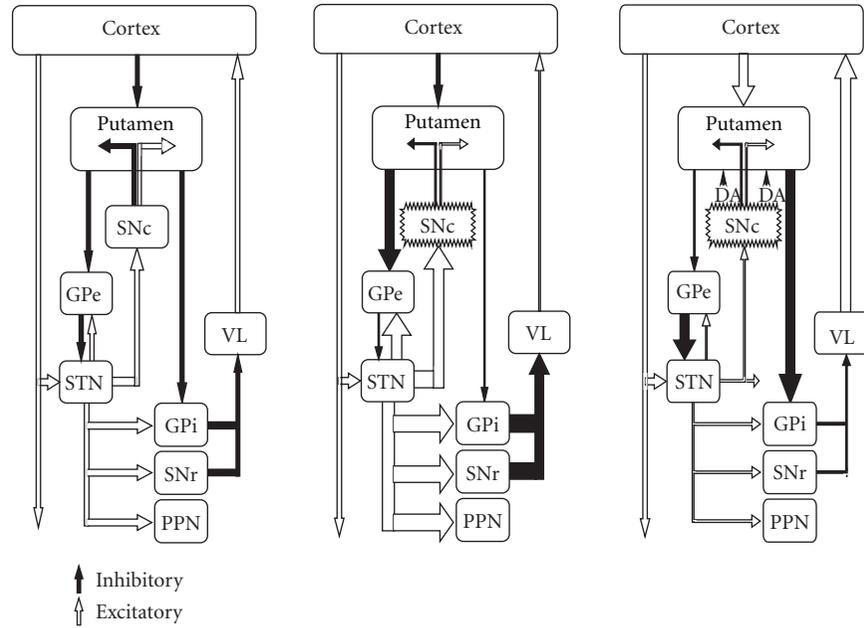


FIGURE 2: Classic model of basal ganglia in normal condition, parkinsonian, and dyskinetic conditions. During LID the different population of striatal cells from direct and indirect circuit are opposite to parkinsonian state. LID would result from a decrease in the inhibitory pathway by striatal neurons in the indirect pathway to the GPe, leading to an inhibitory increment over the STN and consequently reducing STN and GPi/SNr activity. This is facilitated by the increase in the inhibitory striatal activity of GPi by the direct pathway from striatum.

induces hemichorea-ballism, and both deep brain stimulation (DBS) and subthalamotomy in PD patients may induce dyskinesias that are identical to those triggered by levodopa. On the other hand, more recently molecular changes in the striatum and the effects of some dopaminergic drugs have suggested, that LID are mediated by  $D_1$  receptor activation in the “direct” circuit [28, 29]. Thus, increased activity in the signaling by activation of  $D_1$  receptors has been encountered both in animal models and PD patients with LID [30–33].  $D_1$  receptor is abnormally abundant at the plasma membrane of striatal neurons and it seems to be dysregulated in LID by alterations in intraneuronal trafficking [34]. In addition, some interesting findings have suggested a relevant role for  $D_3$  receptor in the pathophysiology of LID [35, 36].

It is also important to consider the changes related to glutamatergic striatal input. The striatum receives massive cortical and thalamic glutamatergic inputs, which are increased in the parkinsonian state [37]. This has been suggested as the mechanism mediating loss of spines in medium spiny neurons [38], which in turn could render the striatum vulnerable to large changes in dopamine availability following levodopa treatment in PD. Recent evidence suggests that the expression, proportion and location of striatal NMDA glutamate receptors may play a paramount role in the molecular mechanisms mediating LID. In the 6-hydroxydopamine (OHDA) rat model it has been shown that the ratio of  $NR_{2b}/NR_{2a}$  is increased and there is a shift to the extra-synaptic space of the  $NR_{2b}$  receptor subunit in dyskinetic rats [39].

Recently, optogenetics was applied to selectively block the protein DARPP-32 in medium spiny neurons of the

“direct” striatonigral projection, resulting in marked LID reduction in the rat model, whereas blockade of striatopallidal neurons giving rise to the “indirect circuit” produced a robust increase in locomotor activity and reduced cataleptic response to haloperidol [40].

Finally, dopaminergic drugs act not only in the striatum but also on other basal ganglia nuclei, the thalamus and cortex, all of which are dopamine depleted in variable extent in PD. The possible action of levodopa and other dopaminergic drugs modulating firing activity of the GPe, GPi and STN should not be underestimated and is still pending definitive studies.

Altogether, there is increasing evidence that overlapping mechanisms underlie the appearance of LID. They seem to converge in alterations of the striatal synaptic function in response to the loss of dopaminergic input and to subsequent replacement of dopamine by pharmacological means [29]. This concept, defined as striatal plasticity, occurs through functional processes such as long term potentiation, long term depression, or a maladaptive form of plasticity invoked as depotentiation [33, 41]. In the presence of exogenous levodopa, distinct patterns of synaptic aberrant plasticity developed in both the direct and indirect pathways, and so a new perspective is open whereby LID in PD could be considered as a network disorder [42]. Indeed, two recent studies comparing LID versus non-LID groups of patients found an increase in the structural signal of the gray-matter focused on the inferior frontal gyrus (IFG) particularly in the right hemisphere, whereas a functional MRI study pointed to an increased task-related activity in the supplementary motor area and reduced activity in the right IFG. These data

suggest that changes in the right IFG reflect neuroplasticity following from years of increased use of executive control to override involuntary movements in LID [43].

In conclusion, the dopaminergic system controls the excitability of the striatum and other basal ganglia nuclei leading to modulation of neuronal firing rates and patterns. LID may originate in striatal spiny neurons, mainly in the putamen leading to reduced mean discharge rate, abnormal firing pattern, and pathological oscillatory activity that are transmitted throughout striatopallidal projection to the thalamocortical projection.

#### 4. LID Pharmacological Treatments

Three main therapeutic strategies have been used to treat LID in PD.

- (1) Prevention of LID development by early use of dopamine agonist drugs and reduced levodopa dose intake at the beginning of treatment.
- (2) Symptomatic treatment, once LID developed, with putative antidyskinetic interventions.
- (3) Reverting dyskinesias by continuous dopaminergic stimulation to achieve a wider therapeutic window, reducing "off" hours while improving dyskinesias.

*4.1. Prevention of LID.* The use of neuroprotective drugs to slow disease progression has been extensively explored. L-deprenyl (selegiline), in an extension of the DATATOP study, failed to produce a significant reduction in the incidence of dyskinesias [44].

The only group that has demonstrated to some extent a reduction in the risk of developing dyskinesias is the dopamine agonists. Several placebo-controlled studies compared the evolution of patients initiated with a dopamine agonist (ropinirol, pramipexol, and cabergoline) and standard levodopa. Rascol et al. in a comprehensive, double-blind parallel study, compared the efficacy of ropinirol and levodopa over a period of 5 years in 268 patients with early PD [5]. The analysis of the time to onset of dyskinesia showed a significant difference in favor of ropinirol. The cumulative incidence of dyskinesia at fifth year, regardless of levodopa supplementation, was 20% in the ropinirol group and 45% in the levodopa group. The mean daily dose of ropinirol was 15 mg but the majority of the patients enrolled in that group required supplementary treatment with levodopa [5]. When patients receiving ropinirole monotherapy required the addition of levodopa, the risk of developing dyskinesias increased, and eventually, during followup, did not differ significantly from that associated with levodopa alone [45]. The use of ropinirole as monotherapy with only later addition of levodopa over 10-year follow-up delayed the onset of dyskinesias by up to 3 years [46]. Moreover, the prolonged-release form of ropinirole recently demonstrated a delay in the onset of dyskinesias compared with increasing doses of levodopa [47]. These clinical observations under control conditions confirmed experimental data in the MPTP monkey showing that ropinirol alone or in combination with

low-dose levodopa delayed dyskinesia onset while improving motor performance [48].

The CALM-PD was a randomized controlled trial that evaluated the risk of developing dyskinesias in patients with early PD treated initially with either pramipexole or levodopa, followed by a maintenance phase during which open-label levodopa-carbidopa was permitted as needed [49]. After 24 months, pramipexole-treated patients were receiving a mean daily dose of 2.78 mg pramipexole plus 264 mg levodopa, compared with 509 mg levodopa for those receiving only this agent. There were fewer pramipexole-treated patients that reached the primary endpoint of time to first occurrence of wearing off, dyskinesias, or on-off motor fluctuations (27.8% versus 50.7%). Patients in the pramipexole group also had a significantly lower incidence of dyskinesias (9.9% versus 30.7%) [49]. After a mean 6-year follow-up, over 90% of patients ended up receiving levodopa therapy regardless of their initial treatment assignment. Compared to those taking pramipexole, patients initially treated with levodopa had significantly more dyskinesias (20.4% versus 36.8%), but there was no difference in the incidence of disabling or painful dyskinesias [50].

The ergot derivative cabergoline holds a long half-life ( $\approx 72$  hours) and therefore may be administered once daily. In a double-blind multicenter trial on 419 patients naive to treatment, comparing cabergoline and levodopa as initial therapy for PD, motor complications were significantly delayed and occurred less frequently in cabergoline-treated patients compared to levodopa-treated patients [51].

An evidence-based review compared the results of studies published on early treatment of PD with dopamine agonists with similar studies using levodopa [52]. Cabergoline, pramipexole, and ropinirol were similarly effective in reducing the risk of LID, although reduction was slightly greater for pramipexole and ropinirol than for cabergoline. The latter is no longer used widely because of the associated risk of cardiac valvulopathy [53].

A concern encountered in the three studies was that, whereas treatment with a dopamine agonist reduced the risk of dyskinesia, this was associated with less antiparkinsonian benefit. Currently, three dopamine agonists provide longer stimulation of DA receptors, by delay-release per oral route (for pramipexol and ropinirole) and transdermal application (rotigotine). The efficacy of these new dopamine agonists formulations on LID has not been specifically assessed yet. It remains also open to future analysis to determine whether the initial benefit on LID of treatment with a dopamine agonist is carried forward over the long-term evolution once levodopa is added to the regimen. In addition, several issues related to the design of the studies have been raised by critical voices. Our own view, which is generally shared by most movement disorder neurologists, is that the severity of LID observed in clinical practice has been considerably reduced over the last decade, coinciding with the earlier use of dopamine agonists and the associated possibility of reducing levodopa daily dose. Thus, while more definitive data are being compiled, we favor the prevailing concept of starting therapy with a dopamine agonist, particularly in patients who are 65 years old or younger at the time of diagnosis.

This approach has been tempered by the more recent realization of a variety of impulse control disorders (ICD) associated with the use of dopamine agonists. Whether or not pathological impulsivity in PD patients will be also reduced by the use of long-acting dopamine agonists, it is too early to tell. We hope this will be the case by the analogy and shared pathophysiological mechanisms of LID and ICD [54].

*4.2. LID as a Clinical Management Problem: Symptomatic Treatments.* This is the commonest clinical scenario. Patients have already developed LID and the clinician has to attempt to control the abnormal movements by adjusting antiparkinsonian drugs or adding agents capable of reducing LID without increasing motor disability. The difficulty in achieving therapeutic efficacy is directly related to the severity and complexity of PD in each individual subject. Thus, LID are relatively easy to control when they are mild and occur in patients with a wide therapeutic window, but may be difficult or impossible to treat pharmacologically in advanced patients who exhibit all forms of LID and fall into severe "off" episodes when they are not dyskinetic. We shall review here the different individual pharmacological approaches available to treat LID but commonly, in many instances of clinical practice one needs to combine several options aiming to control both fluctuations and dyskinesias.

*4.2.1. Dopamine Agonists.* Any one of the above mentioned dopamine agonists may be added with the intention of reducing levodopa dose and avoiding peak of dose on-dyskinesias associated with high levodopa plasma levels while controlling the severity of "off" motor state. Belanger et al. first examined the possibility of reducing LID by using a small dose of cabergoline [55]. During treatment, they found LID in the levodopa group but not in the levodopa + cabergoline group, which suggests that a small dose of a long-acting D<sub>2</sub> agonist combined with low doses of levodopa could reduce the incidence of LID in patients with PD. This study supports a commonly applied clinical strategy. The practical problem in many instances arises when the reduction in levodopa doses precludes achieving a sufficiently good anti-parkinsonian response, a situation poorly tolerated by most patients.

A partial D<sub>2</sub> receptor agonist may represent an interesting alternative for the treatment of PD and dyskinesias. These drugs, characterized by having lower intrinsic activity at the receptor level than full agonists, act as either functional agonist or antagonist, depending on the levels of endogenous dopamine. Preclamol has a selective dopamine mixed agonist-antagonist profile for both pre and postsynaptic receptors. Its action in patients with disabling "on-off" fluctuations was compared against placebo and subcutaneous apomorphine [56], showing a mild but significant anti-kinetic effect which was of lesser magnitude than that achieved with subcutaneous apomorphine but caused less dyskinesia. Aripiprazole is an antipsychotic drug showing partial agonist activity for D<sub>2</sub> and 5HT<sub>2A</sub>, and antagonist for 5HT<sub>2A</sub> receptors. Lieberman postulated that this drug may be able to reduce dyskinesias without enhancing parkinsonism [57], and a small pilot study was positive, [58]. However

further studies are required to investigate its antidyskinetic capacity.

*4.2.2. Dopamine Antagonists.* The use of drugs that block the dopaminergic system has been a classical approach for the treatment of dyskinesias in general. D<sub>2</sub> antagonists, like haloperidol, olanzapine, tiapride, and sulpiride, and presynaptic dopamine-depleting drugs, like reserpine and tetra-benazine, have all proven useful in the management of hemichorea-ballism, tardive dyskinesias, and tics. These same drugs are also effective in reducing or suppressing LID in PD, but this is invariably associated with marked motor worsening after a variable period (ranging from hours to weeks). In clinical practice, therefore, they are neither useful nor recommended.

Recent observations increasingly suggest that atypical neuroleptic drugs, which are able to block D<sub>3</sub> receptors preferentially, can be beneficial for patients with movement disorders. Oh et al. evaluated the effects of an atypical antipsychotic drug which is antagonistic of 5HT<sub>2A/C</sub> and D<sub>2/3</sub> receptors, quetiapine, on motor behavior in the OHDA lesioned rat, and in MPTP treated monkeys [59]. In unilaterally lesioned rats, quetiapine reversed the shortening of the motor response to levodopa challenge produced by treatment during 3 weeks with levodopa twice daily. Quetiapine also normalized the short-duration response to acute injection of agonists either for D<sub>1</sub> receptor (SKF38392) or D<sub>2</sub> (quinpirole) in rats that had received levodopa in chronic administration. Quetiapine had no effect on parkinsonian manifestations when given alone to OHDA lesioned rats or MPTP monkeys, but did substantially reduce LID when administered together with levodopa. Katzenschlager et al. assessed the effect of quetiapine on dyskinesias in a double-blind cross-over study in 9 patients with PD, receiving different doses of quetiapine or placebo at night [60]. On 50 mg/day quetiapine, a slight reduction in LID severity was observed on a visual analog scale but this improvement was not reflected in the patients' overall impression of treatment effect. Durif et al. investigated the efficacy of clozapine in the treatment of LID in 50 patients during a 10-week, double-blind, placebo-controlled, multicenter trial. During a levodopa challenge the maximal LID score was significantly decreased in the clozapine group (mean dose ≈40 mg/day), which led to the conclusion that clozapine is effective in the treatment of LID in severe PD [61].

*4.2.3. Glutamatergic Antagonists.* The N-methyl-D aspartate (NMDA) receptor is thought to mediate excitotoxicity in the basal ganglia, but the use of NMDA antagonists in humans has generally been limited because of adverse effects associated with a non-selective blockade. Metman et al., in a double-blind cross-over study, showed that 3 weeks' treatment with dextrometorphan was able to reduce dyskinesias by 30–40% while maintaining the response to levodopa. In recent years amantadine, which is believed to increase dopamine release from presynaptic uptake sites, has become popular as an antidyskinetic drug based on its putative anti-NMDA action [62]. Del Dotto et al. evaluated the effect of a 2-hour intravenous amantadine or placebo infusion

against LID in 9 PD patients with motor fluctuations and severely disabling peak-dose dyskinesias [63]. Intravenous amantadine acutely improved LID by 50%, without losing the antiparkinsonian benefit of levodopa along the 5-week, double-blind cross-over trial. In another study, Luginger et al. assessed LID severity by self-scoring diaries after oral levodopa challenges and found them to be reduced by approximately 50% after amantadine treatment compared with baseline or placebo control [64]. Further studies also found a positive effect for amantadine on LID [65, 66]. Moreover, in a recent trial in advanced PD patients receiving amantadine continuously over 1 year, a withdrawal of amantadine led to a significant increase of dyskinesias in those patients when double-blind switched to placebo, while no change occurred in those maintained on amantadine. This supports the notion of a sustained antidyskinetic effect of amantadine beyond one year of therapy. Our own view is that, on an individual basis, amantadine may result in a drastic amelioration of LID and is therefore worth trying in the absence of contraindications. The antidyskinetic effect is probably exerted at the level of the STN as amantadine failed to control dyskinesias evoked by subthalamotomy in patients who had previously responded markedly well [67].

Merello et al. evaluated the efficacy of memantine on the pharmacological response to levodopa and the induction of LID [68]. In 12 patients, in opposition to recent findings with amantadine, no effect on LID was observed. Nevertheless, several reports described a benefit of memantine in PD patients with cognitive impairment and LID with regard to dyskinesia control [69, 70]. No effect was found for riluzole on LID [71, 72]. In general, the high expectations that were raised with the potential therapeutic impact of antihypertensive drugs for PD have so far been disappointed.

**4.2.4. Drugs Acting on the Serotonergic System.** The serotonergic system projects quite profusely to the striatum and also to other key basal ganglia nuclei (i.e., STN, GPe, GPi), exerting an inhibitory effect on dopamine striatal transmission. Durif et al. found a 47% improvement in LID severity induced by apomorphine in 7 patients with PD treated with fluoxetine [73], out of any reduction in antiparkinsonian benefits. Bupirone has a complex mechanism of action, which aside from its 5HT<sub>1A</sub> properties includes partial dopamine agonism and mild opiate and noradrenergic antagonism [74]. Bonifati et al. in a double-blind, placebo-controlled, cross-over study, found that bupirone significantly lessened the severity of LID in 5 out of 7 patients [75]. Meco et al., in an open-label study including 20 parkinsonian patients, found that mirtazapine, an  $\alpha_2$  antagonist, 5HT<sub>1A</sub> agonist, and 5HT<sub>2</sub> antagonist, may be effective in reducing LID [76].

**4.2.5. Drugs Acting on the Opioid System.** The opioid striatal neurons may play a role in the induction of dyskinesias. In MPTP monkeys Samadi et al. investigated the effect of different doses of naloxone and naltrexone (opioid receptor antagonists) on the dyskinetic response to the D<sub>1</sub> agonist SKF-82958, the D<sub>2</sub> agonist quinpirole and levodopa [77]. They found that joint administration of naloxone or naltrexone

together with dopaminergic agents led to a significant reduction in the severity of dyskinesias without reducing antiparkinsonian efficacy. Recently, the selective  $\mu$  opioid antagonist ADL5510 provided almost complete alleviation of LID without compromising reversal of parkinsonian disability in the MPTP lesioned macaque model of PD [78]. In PD patients, Carroll et al. conducted a placebo-controlled, double-blind, cross-over trial to examine the potential effect of cannabis on LID in PD [79]. Seventeen patients completed the trial and cannabis was well tolerated with no pro- or anti-parkinsonian action, but there was no evidence of a treatment effect on LID. Thus, despite many experimental suggestions, there is no drug currently employed clinically to manipulate the opioid system for the treatment of LID.

**4.2.6. Noradrenergic Drugs.** The close relationship between the dopaminergic, adrenergic and noradrenergic systems has led to the assessment of a possible antidyskinetic effect of a few drugs acting on those systems. Carpentier et al. found a significant 40% improvement in dyskinesia scores in PD patients treated with a low dose of propranolol [80]. Other studies have shown how the  $\alpha_2$  adrenoceptor antagonist idazoxan can significantly reduce LID in monkey and rat models as well as in advanced PD patients [81, 82]. Rascol et al. reported improvement of LID without reappearance of parkinsonian symptoms in 18 patients treated with idazoxan [83]. Another  $\alpha_2$  antagonist, fipamezole, reduced the severity of LID by 23% and 31% at 60 mg, and 90 mg respectively, without affecting antiparkinsonian response. Currently, further trials are being carried out [84].

**4.2.7. Adenosine A<sub>2A</sub> Antagonists.** Adenosine A<sub>2A</sub> receptors are found in the striatum and thalamus and colocalize with dopamine D<sub>2</sub> receptors. Adenosine A<sub>2A</sub> antagonists regulate dopamine and glutamate release in the brain, and they may improve motor symptoms as novel compensatory mode for loss of dopamine signaling with associated NMDA antagonism [85]. The trials target symptoms associated with dopamine replacement and therapy of dyskinesia, such as istradefylline [86, 87]. However, recent trial outcomes showed that istradefylline did not improve motor behavior or "off" times in PD patients compared with earlier results [88–92]. Preladenant showed, in a phase II placebo-controlled dose-ranging trial of 253 PD patients receiving stable dopaminergic therapy, an increase in awake time spent in the on-state of 1.4 h/day compared to 0.2 h/day in the placebo group, without overall worsening of dyskinesias [93]. The long-term antidyskinetic effect of preladenant needs ascertainment.

**4.2.8. Other Drug Treatments.** Levetiracetam, an antiepileptic drug, has been evaluated against LID with mixed results in several open-label studies [94–98]. The most promising data come from a study of 9 patients experiencing LID for at least 25% of waking hours [98]. After 60 days treatment with a mean of 625 mg of levetiracetam, patients experienced a 42% increase in the "on" time without LID or with nontroublesome dyskinesia in absence of significant change in the "off" time. Pardoprunox is a mixed dopamine agonist/antagonist D<sub>2</sub> and D<sub>3</sub>, and a full agonist

at 5HT<sub>1a</sub> receptors. It also binds with lower affinity to D<sub>4</sub>,  $\alpha_1$  adrenergic, and 5HT<sub>7</sub> receptors [99, 100]. Due to its unique pharmacologic profile, pramipexole might have a lower tendency than other dopaminergic therapies to cause dyskinesias or neuropsychiatric side effects [93, 99–101]. Safinamide is an antiparkinsonian agent that is also in advance state of development to reach clinical practice. It has a dual mechanism of action, as it is a MAO<sub>B</sub> inhibitor and also reduces overactivity of glutamatergic signaling by inhibiting glutamate release [102, 103]. On this prospect, AFQ056 recently achieved a significant and relevant antidyskinetic clinical effect without reducing the antiparkinsonian benefits of dopaminergic therapy [104]. Recently, low-frequency transcranial magnetic stimulation has also been applied to the treatment of LID, showing transient experimental improvements in preliminary study [105].

**4.2.9. Practical Considerations.** There appear to be many drugs that are capable of reducing LID severity. In occasional patients the therapeutic impact of any one of the treatments summarized above may be strikingly positive, but in the majority of patients it is limited to mild and short-lasting improvement. Nevertheless, these treatments are generally well tolerated and worth trying, when available, in patients in whom other therapeutic measurements cannot be afforded. In our experience, the degree of symptomatic control of LID mainly depends upon the complexity of dyskinesias and severity of “off” periods. This may be schematically summarized as follows: (1) in patients with mild but bothersome peak-dose dyskinesias, readjust the levodopa schedule, and consider adding a dopamine agonist. If this approach fails, any one of the drugs discussed above may be tried out; (2) for patients with intense peak-dose dyskinesias, consider switching treatment to provide continuous dopaminergic stimulation; (3) patients with severe peak-dose dyskinesias and diphasic dyskinesias probably require surgical treatment (Table 1).

**4.3. Continuous Dopaminergic Stimulation.** Since the introduction of the concept of continuous dopaminergic stimulation in the 1980s [3, 106–108], it has been realized that constant delivery of dopaminergic drugs is associated with a reduction in LID severity. Over the past decade, further evidence has accumulated to support the notion that continuous stimulation of dopamine receptors may even reverse some of the changes induced by chronic pulsatile levodopa administration. The antidyskinetic response to this approach is not immediate and it may take several weeks of continuous infusion before becoming apparent. The initial pivotal study using continuous delivery was published by Mouradian et al., who used levodopa intravenously for 7–12 days to a small group ( $n = 12$ ) of patients with advanced PD [109]. They found a progressive attenuation of LID and improvement of the “on–off” fluctuations. Levodopa is too acid to be delivered intravenously or subcutaneously in practice, a problem by and large resolved with the development of duodenal levodopa infusion. This has been used with clear benefit to improve motor complications and quality of life despite the obvious practical limitations [110–113]. Very

recently, the first double-blind, placebo controlled study assessing the effect of duodenal levodopa carried out in North America has been disclosed. However, the technique is complex, expensive, and potential long-term adverse effects are under debate, such as axonal polyneuropathy and vitamin B complex deficiency [114, 115]. The infusion of the duodenal levodopa gel, which also contains the dopa-decarboxylase inhibitor carbidopa, is currently available only in certain countries.

Further alternative strategies of oral intake were also tested, such as controlled release levodopa/carbidopa formulations, but they did not delay the onset of motor complications [116]. The STRIDE-PD study, initiating levodopa with entacapone, failed to reduce the frequency or delay the onset of LID [117]; an inadequate dosing schedule perturbing the putative continuous stimulation expected to be achieved with this treatment and a bias in the treatment group toward more severe disease have been suggested as potential confounders [118]. IPX066 might be soon available and it may be used to attain and maintain therapeutic levodopa plasma concentrations with a potential antidyskinetic efficacy [119].

In line with continuous delivery procedures, dopamine agonists that operate via the subcutaneous route, such as lisuride and apomorphine, are associated with a reduction in LID. The majority of trials used infusions during the daytime but stopped at night to reduce the risk of severe psychiatric complications. Stocchi et al. compared the long-term incidence of dyskinesias in patients treated with subcutaneous infusion of lisuride (plus supplementary oral levodopa as needed) versus patients treated with standard levodopa orally, and showed that patients receiving lisuride infusions experienced a reduction in the incidence of dyskinesia and motor fluctuations, compared with patients receiving standard therapies [120]. The benefit lasted over the 4 years of follow-up and this study also endorsed earlier results indicating that continuous lisuride infusion can be fairly well tolerated and beneficial for patients' motor complications, provided they have not previously developed severe psychiatric complications [121, 122].

Similarly, Manson et al. reviewed their experience in 64 patients treated with subcutaneous apomorphine infusions [123]. Forty-five patients were successfully converted to monotherapy and discontinued all other dopaminergic drugs during the daytime infusion. LID were reduced by 64% in the monotherapy group compared to 30% in those on polytherapy. Another retrospective evaluation over a 5-year period of 82 patients receiving apomorphine obtained a similar outcome [124], with average follow-up of  $\approx 20$  months, 5 mg/h dose, and 14 hours/day duration. Patients improved in severity of dyskinesia by 31% as assessed by the UPDRS dyskinesia evaluation, injection-site adverse events being the main reason for discontinuation of treatment. These results confirmed that monotherapy with infusions of apomorphine may reset peak-dose dyskinesia threshold in patients treated with levodopa, while further reducing off-period disability. Katzenschlager et al. prospectively assessed the antidyskinetic effect of continuous subcutaneous apomorphine using subjective and objective measures and response to a levodopa

TABLE 1

Practical suggestions for pharmacological management of LID
(1) The optimal therapeutic approach for LID is to try avoiding their development
(2) Start PD treatment with an agonist if possible, particularly in young onset patients
(3) Save levodopa as long as you can hold the patient's requirements for daily life activities
(4) Adjust the drug schedule: reduce total daily doses and/or shorten the intake intervals
(5) Add amantadine 200–400 mg/day
(6) Low doses of quetiapine or clozapine may be helpful
(7) Propose continuous drug delivery devices: duodenal levodopa/carbidopa gel or subcutaneous apomorphine
(8) For refractory cases, when indication is set by an expert and the risks are assumable by the patient, surgery is the treatment of choice

challenge [125]. By the sixth month the mean levodopa dose had been reduced by 55% and the daily “off” time in patients' diaries was reduced by 38%. Levodopa challenge showed a reduction of 40–44% in the dyskinesia scores and patients' self-assessment scores reflected these significant changes positively. Overall, these results reinforce the concept that replacement of oral short-acting antiparkinsonian drugs with medication capable of providing more continuous dopamine receptor stimulation may at least partially avoid or reverse the sensitization process believed to mediate the development of LID. In theory, therefore, therapy with infusions capable of providing continuous dopaminergic stimulation might be the pharmacological treatment of choice for advanced PD patients. Nevertheless, the degree of control of LID achieved with infusions is not complete in many patients. Pharmacological tolerance appears in a large proportion after some time on treatment. It occurs more readily the more severe the underlying disease is, leading to “off” episodes or exacerbation of diphasic dyskinesias. The latter may cause a very troublesome dyskinetic status [122]. At this point, surgical treatment may still be the only and best therapeutic option for a proportion of patients with severe LID.

## 5. Surgical Treatments

The three main surgical targets for PD are the thalamus, GPi, and STN. In this section we review the antidyskinetic effect of stereotactic surgery directed towards these 3 different targets using either ablative surgery or DBS.

### 5.1. The Thalamus and LID

**5.1.1. Vim-Thalamotomy.** During the 1960s the ventral lateral nucleus (VL) of the thalamus was determined as the best target to remove tremor in PD. This target was later defined from physiology as the ventralis intermedialis (Vim) and it became established as the target of choice for tremor of any origin [126–128]. Despite the thousands of thalamotomies performed over the years, no formal and prospective evaluation of the response of thalamic surgery against LID has been reported in the literature. Some reports described how the development of LID was prevented in patients with a previous thalamotomy [129–132], others described

how the lesion improved tremor and also LID [133–135], or the concept that LID improvement should be correlated with daily levodopa reduction after surgery following tremor suppression [136]. However, there are also reports where no benefit was obtained with Vim-thalamotomy in patients with LID [137].

The best study, which is somewhat an exception to the above, is the observational paper published by Narabayashi et al. [138]. These authors report an interesting and detailed study of the effect of thalamotomy against LID, dividing the patients according to the thalamic target selected for surgery. The patients subjected to lesions in the ventralis oralis anterior nucleus (Voa) or posterior (Vop) prior to the introduction of levodopa did not develop LID, but patients subjected to Vim-thalamotomy for tremor did develop dyskinesias when levodopa was introduced as treatment [138]. The conclusion reached by Narabayashi et al. was that the GPi-Voa/Vop pathway mediated LID and lesions restricted to the Vim to treat tremor were not effective against LID. Interestingly, similar results were reported by Page et al. in parkinsonian monkeys with LID induced by dopamine agonists. Thalamotomy performed in the pallidal territory removed LID, but lesions in the nigral or cerebellar terminal territory of the thalamus had no antidyskinetic effect [139].

**5.1.2. Vim-DBS.** The introduction of high frequency stimulation coupled with stereotactic surgery supposed a marked advance for patients with movement disorders. Vim-DBS was initially performed as an additional contralateral treatment to patients who had had a previous thalamotomy [140]. In a group of parkinsonian patients, Benabid et al. described significant tremor improvement after Vim-DBS, which was accompanied by inconsistent responses or no alleviation of LID [141]. Similar results were obtained in other studies [142–144]. In contrast, successful alleviation or suppression of LID was described in association with a different positioning of the electrode which supposedly impinges upon the Vim and the Centromedian-parafascicular nucleus (CM-Pf) [145–147]. However, a more recent study in MPTP treated monkeys revealed that lesion of the CM-Pf had no effect against parkinsonian features or LID [148]. In conclusion, the available data indicate that Vim lies outside the pathways underlying LID and, accordingly, Vim's surgery conveys no effect against LID.

## 5.2. Surgery of the GPi and LID

**5.2.1. Pallidotomy.** Posteroventral pallidotomy was reintroduced as a treatment for PD, applying Leksell's concepts, by Laitinen et al. in 1992 [149]. The clinical response to pallidal lesion included a significant benefit of the cardinal features on the contralateral side and, unexpectedly according to the basal ganglia model, a large impact against LID. Thus, pallidotomy has been shown to portray a very significant and long lasting effect against peak dose dyskinesia, diphasic dyskinesia, and also "off" period dystonia on the side contralateral to the lesion. This antidyskinetic effect is enduring and long-lasting, for at least 10 years [150, 151], with a benefit that occurred without a significant reduction in daily levodopa dose.

**5.2.2. GPi-DBS and LID.** In the first multicentre DBS Cooperative Multicentre Study after GPi-DBS, patients showed a 76% reduction in LID severity ( $P < 0.0001$ ) with no change in levodopa doses at 1 and 4 years follow up [152, 153]. Longer follow-up (5-6 years) continued to show that GPi-DBS maintained a significant improvement of LID with a significant increase in "on" time without LID [154]. Levodopa was not significantly reduced compared with baseline [155].

## 5.3. STN Surgery and LID

**5.3.1. Subthalamotomy.** The STN plays a capital role in the pathophysiology of parkinsonian and dyskinetic states. This anatomical target is typically considered a prodyskinetic structure and classically avoided in patients with severe LID. Subthalamotomy is performed on occasional patients, more frequently in countries where DBS is not affordable, with fairly good general results [156].

Assessing the evolution of LID after subthalamotomy is limited by the relatively reduced number of patients reported, and by the variables in controlling some important factors, such as levodopa dose pre- and postsurgery, surgical procedure, lesion placement and volume. A recent analysis described how in a group of 68 patients "peak dose dyskinesias" increased on the side contralateral to the lesion during the first postoperative year but decreased after two to three years, showing no significant change versus baseline at the last assessment. In the ipsilateral side to the lesion, LID increased significantly with the progressive increment of levodopa suggesting that the operated side has had an antidyskinetic effect. Diphasic dyskinesias and "off" period dystonia also improved significantly ( $P < 0.01$ ) contralateral to the lesion at 12th and 24th months after surgery [156].

**5.3.2. STN-DBS.** Bilateral STN-DBS is currently the surgical procedure most often selected for PD patients given the large impact against "off" medication severity and the associated reduction in the daily levodopa dose [153, 157–162]. STN-DBS has generally been associated with significant reduction in LID, closely correlated with levodopa dosage reduction. Subthalamic stimulation appears to improve the whole spectrum of LID, such as peak dose dyskinesia (30%),

biphasic dyskinesia (50%), and "off" dystonia (90%) with a 47% reduction in levodopa dosage as reported by Krack et al. [157]. DBS-STN also increases "on" time without LID and reduces "off" time periods [161–163]. After 5-6 years of follow-up, LID scores were significantly improved by 83.3% in total, with 75% reduction in dyskinesia duration and 100% drop of disability compared with baseline [153, 155, 161]. Levodopa reduction was also significantly reduced in the long term compared with baseline preoperative data (30%) [155]. In a survey of 38 studies involving 737 patients treated in 34 neurosurgical centers, STN-DBS improved LID assessed by UPDRS-IV scores 94% at 12 months in the on-stimulation/"on" medication state in comparison with "on" preoperative medication scores [163].

How STN-DBS may improve LID is not well understood [164]. For most authors, LID improvement by STN-DBS may be directly correlated with levodopa reduction [165–169]. However, it is difficult to interpret these studies, because there are very few patients who maintained similar levodopa equivalent doses after surgery. Thus, fluctuations and LID disappeared in patients with levodopa withdrawal postimplantation as Vingenhoerst et al. described, whereas they persisted in those patients on medication 2 years after surgery [167]. Similarly, another group reported that 1 year after implantation, patients receiving levodopa displayed a 47% LID reduction, whereas the reduction was 90% of LID in patients who did not receive levodopa ( $P < 0.003$ ) [168]. On the other hand, the antidyskinetic response after STN-DBS could be related with the effect of continuous high frequency stimulation, providing antidyskinetic efficacy on its own [170–172]. This may be supported by some instances where improvement of LID occurred despite maintaining the same daily dose of levodopa [170]. Thus, STN surgery could induce a stable and continuous functional state with reduced fluctuations in basal ganglia network, somehow mimicking the effect of continuous dopaminergic stimulation.

Finally, it has also been suggested that the antidyskinetic effect of STN-DBS (as well as subthalamotomy) may be due to an effect on the dorsal border of the nucleus, reaching the lenticularis fasciculus and zona incerta. In this context some studies have suggested that the real subthalamic target may be the region above the dorsal border of the nucleus [173–175].

In conclusion, STN-DBS probably interferes with abnormal discharge pattern in basal ganglia output nuclei associated with the parkinsonian condition, improving PD, and permitting a reduction of chronic levodopa therapy. The latter is likely responsible for the anti-LID effect. On the other hand, it is also possible that high frequency stimulation of the STN could modify the patterns of neuronal firing and the rhythms associated with LID having "per se" an antidyskinetic effect [176].

## 6. Conclusions

Most PD patients develop motor fluctuations and LID during chronic evolution and on levodopa treatment. Motor complications are directly related with disease progression and the effects of chronic levodopa therapy. Once

established, LID remains unabated throughout evolution. Pharmacological management is not simple but in recent years, the proportion of patients suffering severe LID has declined considerably, mainly in relation with the use smaller dose of levodopa. Surgical treatment has a potent anti-dyskinetic effect whose value has to be judged for every particular patient against the risk. LID is no longer the major cause of disability in PD patients nor a problem lacking several treatment options.

## References

- [1] A. Barbeau, "Long term assessment of levodopa therapy in Parkinson's disease," *Canadian Medical Association Journal*, vol. 112, no. 12, pp. 1379–1380, 1975.
- [2] C. D. Marsden and J. D. Parkes, "Success and problems of long term levodopa therapy in Parkinson's disease," *The Lancet*, vol. 1, no. 8007, pp. 345–349, 1977.
- [3] J. A. Obeso, F. Grandas, J. Vaamonde et al., "Motor complications associated with chronic levodopa therapy in Parkinson's disease," *Neurology*, vol. 39, supplement 2, no. 11, pp. 11–19, 1989.
- [4] M. R. Luquin, O. Scipioni, J. Vaamonde, O. Gershanik, and J. A. Obeso, "Levodopa-induced dyskinesias in Parkinson's disease: clinical and pharmacological classification," *Movement Disorders*, vol. 7, no. 2, pp. 117–124, 1992.
- [5] O. Rascol, D. J. Brooks, A. D. Korczyn, P. P. De Deyn, C. E. Clarke, and A. E. Lang, "A five-year study of the incidence of dyskinesia in patients with early Parkinson's disease who were treated with ropinirole or levodopa," *The New England Journal of Medicine*, vol. 342, no. 20, pp. 1484–1491, 2000.
- [6] J. A. Obeso, M. C. Rodriguez-Oroz, M. Rodriguez, M. R. DeLong, and C. W. Olanow, "Pathophysiology of levodopa-induced dyskinesias in Parkinson's disease: problems with the current model," *Annals of Neurology*, vol. 47, no. 4, pp. S22–S34, 2000.
- [7] E. Růžička, K. Zárubová, J. G. Nutt, and B. R. Bloem, "'Silly Walks' in Parkinson's disease: unusual presentation of dopaminergic-induced dyskinesias," *Movement Disorders*, vol. 26, no. 9, pp. 1783–1784, 2011.
- [8] C. R. Freed, P. E. Greene, R. E. Breeze et al., "Transplantation of embryonic dopamine neurons for severe Parkinson's disease," *The New England Journal of Medicine*, vol. 344, no. 10, pp. 710–719, 2001.
- [9] P. Limousin, P. Krack, P. Pollak et al., "Electrical stimulation of the subthalamic nucleus in advanced Parkinson's disease," *The New England Journal of Medicine*, vol. 339, no. 16, pp. 1105–1111, 1998.
- [10] C. W. Olanow, J. H. Kordower, A. E. Lang, and J. A. Obeso, "Dopaminergic transplantation for Parkinson's disease: current status and future prospects," *Annals of Neurology*, vol. 66, no. 5, pp. 591–596, 2009.
- [11] C. W. Olanow, J. M. Gracies, C. G. Goetz et al., "Clinical pattern and risk factors for dyskinesias following fetal nigral transplantation in parkinson's disease: a double blind video-based analysis," *Movement Disorders*, vol. 24, no. 3, pp. 336–343, 2009.
- [12] C. W. Olanow, A. H. V. Schapira, and O. Rascol, "Continuous dopamine-receptor stimulation in early Parkinson's disease," *Trends in Neurosciences*, vol. 23, supplement 10, pp. S117–S126, 2000.
- [13] M. A. Cenci, C. S. Lee, and A. Björklund, "L-DOPA-induced dyskinesia in the rat is associated with striatal overexpression of prodynorphin- and glutamic acid decarboxylase mRNA," *European Journal of Neuroscience*, vol. 10, no. 8, pp. 2694–2706, 1998.
- [14] P. Calabresi, M. D. Filippo, V. Ghiglieri, N. Tambasco, and B. Picconi, "Levodopa-induced dyskinesias in patients with Parkinson's disease: filling the bench-to-bedside gap," *The Lancet Neurology*, vol. 9, no. 11, pp. 1106–1117, 2010.
- [15] M. Filion, L. Tremblay, and P. J. Bedard, "Effects of dopamine agonists on the spontaneous activity of globus pallidus neurons in monkeys with MPTP-induced parkinsonism," *Brain Research*, vol. 547, no. 1, pp. 152–161, 1991.
- [16] S. M. Papa, R. Desimone, M. Fiorani, and E. H. Oldfield, "Internal globus pallidus discharge is nearly suppressed during levodopa-induced dyskinesias," *Annals of Neurology*, vol. 46, no. 5, pp. 732–738, 1999.
- [17] R. Levy, J. O. Dostrovsky, A. E. Lang, E. Sime, W. D. Hutchison, and A. M. Lozano, "Effects of apomorphine on subthalamic nucleus and globus pallidus internus neurons in patients with Parkinson's disease," *Journal of Neurophysiology*, vol. 86, no. 1, pp. 249–260, 2001.
- [18] A. M. Lozano, A. E. Lang, R. Levy, W. Hutchison, and J. Dostrovsky, "Neuronal recordings in Parkinson's disease patients with dyskinesias induced by apomorphine," *Annals of Neurology*, vol. 47, supplement 1, no. 4, pp. S141–S146, 2000.
- [19] M. Merello, J. Balej, M. Delfino, A. Cammarota, O. Betti, and R. Leiguarda, "Apomorphine induces changes in GPi spontaneous outflow in patients with Parkinson's disease," *Movement Disorders*, vol. 14, no. 1, pp. 45–49, 1999.
- [20] I. J. Mitchell, S. Boyce, M. A. Sambrook, and A. R. Crossman, "A 2-deoxyglucose study of the effects of dopamine agonists on the Parkinsonian primate brain. Implications for the neural mechanisms that mediate dopamine agonist-induced dyskinesia," *Brain*, vol. 115, pp. 809–824, 1992.
- [21] R. L. Albin, "The pathophysiology of chorea/ballism and Parkinsonism," *Parkinsonism and Related Disorders*, vol. 1, no. 1, pp. 3–11, 1995.
- [22] R. L. Albin, A. B. Young, and J. B. Penney, "The functional anatomy of basal ganglia disorders," *Trends in Neurosciences*, vol. 12, no. 10, pp. 366–375, 1989.
- [23] A. R. Crossman, "Primate models of dyskinesia: the experimental approach to the study of basal ganglia-related involuntary movement disorders," *Neuroscience*, vol. 21, no. 1, pp. 1–40, 1987.
- [24] M. R. DeLong, "Primate models of movement disorders of basal ganglia origin," *Trends in Neurosciences*, vol. 13, no. 7, pp. 281–285, 1990.
- [25] A. R. Crossman, I. J. Mitchell, M. A. Sambrook, and A. Jackson, "Chorea and myoclonus in the monkey induced by gamma-aminobutyric acid antagonism in the lentiform complex. The site of drug action and a hypothesis for the neural mechanisms of chorea," *Brain*, vol. 111, pp. 1211–1233, 1988.
- [26] I. Hamada and M. R. DeLong, "Excitotoxic acid lesions of the primate subthalamic nucleus result in transient dyskinesias of the contralateral limbs," *Journal of Neurophysiology*, vol. 68, no. 5, pp. 1850–1858, 1992.
- [27] I. J. Mitchell, A. Jackson, M. A. Sambrook, and A. R. Crossman, "The role of the subthalamic nucleus in experimental chorea. Evidence from 2-deoxyglucose metabolic mapping and horseradish peroxidase tracing studies," *Brain*, vol. 112, pp. 1533–1548, 1989.
- [28] A. Berthet, E. Bezard, G. Porras et al., "L-DOPA impairs proteasome activity in parkinsonism through D<sub>1</sub> dopamine

- receptor," *Journal of Neuroscience*, vol. 32, no. 2, pp. 681–691, 2012.
- [29] M. M. Iravani, A. C. McCreary, and P. Jenner, "Striatal plasticity in Parkinson's disease and L-DOPA induced dyskinesia," *Parkinsonism and Related Disorders*, vol. 181, supplement 1, pp. S123–S125, 2012.
- [30] I. Aubert, C. Guigoni, K. Håkansson et al., "Increased D<sub>1</sub> dopamine receptor signaling in levodopa-induced dyskinesia," *Annals of Neurology*, vol. 57, no. 1, pp. 17–26, 2005.
- [31] R. Bordet, S. Ridray, J. C. Schwartz, and P. Sokoloff, "Involvement of the direct striatonigral pathway in levodopa-induced sensitization in 6-hydroxydopamine-lesioned rats," *European Journal of Neuroscience*, vol. 12, no. 6, pp. 2117–2123, 2000.
- [32] C. Guigoni, S. Dovero, I. Aubert et al., "Levodopa-induced dyskinesia in MPTP-treated macaques is not dependent on the extent and pattern of nigrostriatal lesioning," *European Journal of Neuroscience*, vol. 22, no. 1, pp. 283–287, 2005.
- [33] B. Picconi, D. Centonze, K. Håkansson et al., "Loss of bidirectional striatal synaptic plasticity in L-DOPA-induced dyskinesia," *Nature Neuroscience*, vol. 6, no. 5, pp. 501–506, 2003.
- [34] A. Berthet, G. Porras, E. Doudnikoff et al., "Pharmacological analysis demonstrates dramatic alteration of D<sub>1</sub> dopamine receptor neuronal distribution in the rat analog of L-DOPA-induced dyskinesia," *Journal of Neuroscience*, vol. 29, no. 15, pp. 4829–4835, 2009.
- [35] E. Bézard, S. Ferry, U. Mach et al., "Attenuation of levodopa-induced dyskinesia by normalizing dopamine D<sub>3</sub> receptor function," *Nature Medicine*, vol. 9, no. 6, pp. 762–767, 2003.
- [36] C. Fiorentini, C. Busi, E. Gorruso, C. Gotti, P. Spano, and C. Missale, "Reciprocal regulation of dopamine D<sub>1</sub> and D<sub>3</sub> receptor function and trafficking by heterodimerization," *Molecular Pharmacology*, vol. 74, no. 1, pp. 59–69, 2008.
- [37] Y. Smith, D. Raju, B. Nanda, J. F. Pare, A. Galvan, and T. Wichmann, "The thalamostriatal systems: anatomical and functional organization in normal and parkinsonian states," *Brain Research Bulletin*, vol. 78, no. 2-3, pp. 60–68, 2009.
- [38] R. M. Villalba and Y. Smith, "Neuroglial plasticity at striatal glutamatergic synapses in Parkinson's disease," *Frontiers in Systems Neuroscience*, vol. 5, article 68, 2011.
- [39] F. Gardoni, B. Picconi, V. Ghiglieri et al., "A critical interaction between NR2B and MAGUK in L-DOPA induced dyskinesia," *Journal of Neuroscience*, vol. 26, no. 11, pp. 2914–2922, 2006.
- [40] H. S. Bateup, E. Santini, W. Shen et al., "Distinct subclasses of medium spiny neurons differentially regulate striatal motor behaviors," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 33, pp. 14845–14850, 2010.
- [41] V. Ghiglieri, B. Picconi, and P. Calabresi, "Direct and indirect pathways in levodopa-induced dyskinesia: a more complex matter than a network imbalance," *Movement Disorders*, vol. 25, no. 11, pp. 1527–1529, 2010.
- [42] P. Belujon, D. J. Lodge, and A. A. Grace, "Aberrant striatal plasticity is specifically associated with dyskinesia following levodopa treatment," *Movement Disorders*, vol. 25, no. 11, pp. 1568–1576, 2010.
- [43] A. R. Aron and J. Obeso, "Is executive control used to compensate for involuntary movements in levodopa-induced dyskinesia?" *Movement Disorders*, vol. 27, no. 3, pp. 339–340, 2012.
- [44] I. Shoulson, "DATATOP: a decade of neuroprotective inquiry," *Annals of Neurology*, vol. 44, supplement 1, no. 3, pp. S160–S166, 1998.
- [45] O. Rascol, D. J. Brooks, A. D. Korczyn et al., "Development of dyskinesias in a 5-year trial and ropinirole and L-dopa," *Movement Disorders*, vol. 21, no. 11, pp. 1844–1850, 2006.
- [46] R. A. Hauser, O. Rascol, A. D. Korczyn et al., "Ten-year follow-up of Parkinson's disease patients randomized to initial therapy with ropinirole or levodopa," *Movement Disorders*, vol. 22, no. 16, pp. 2409–2417, 2007.
- [47] R. L. Watts, K. E. Lyons, R. Pahwa et al., "Onset of dyskinesia with adjunct ropinirole prolonged-release or additional levodopa in early Parkinson's disease," *Movement Disorders*, vol. 25, no. 7, pp. 858–866, 2010.
- [48] E. C. Maratos, M. J. Jackson, R. K. B. Pearce, and P. Jenner, "Antiparkinsonian activity and dyskinesia risk of ropinirole and L-DOPA combination therapy in drug naive MPTP-lesioned common marmosets (*Callithrix jacchus*)," *Movement Disorders*, vol. 16, no. 4, pp. 631–641, 2001.
- [49] R. Holloway, "A randomized controlled trial comparing pramipexole with levodopa in early Parkinson's disease: design and methods of the CALM-PD study," *Clinical Neuropharmacology*, vol. 23, no. 1, pp. 34–44, 2000.
- [50] R. Holloway, K. Marek, K. Biglan et al., "Long-term effect of initiating pramipexole vs levodopa in early Parkinson disease," *Archives of Neurology*, vol. 66, no. 5, pp. 563–570, 2009.
- [51] F. Bracco, A. Battaglia, C. Chouza et al., "The long-acting dopamine receptor agonist cabergoline in early Parkinson's disease: final results of a 5-year, double-blind, levodopa-controlled study," *CNS Drugs*, vol. 18, no. 11, pp. 733–746, 2004.
- [52] R. Inzelberg, E. Schechtman, and P. Nisipeanu, "Cabergoline, pramipexole and ropinirole used as monotherapy in early Parkinson's disease: an evidence-based comparison," *Drugs and Aging*, vol. 20, no. 11, pp. 847–855, 2003.
- [53] A. Pinero, P. Marcos-Alberca, and J. Fortes, "Cabergoline-related severe restrictive mitral regurgitation," *The New England Journal of Medicine*, vol. 353, no. 18, pp. 1976–1977, 2005.
- [54] V. Voon, P. O. Fernagut, J. Wickens et al., "Chronic dopaminergic stimulation in Parkinson's disease: from dyskinesias to impulse control disorders," *The Lancet Neurology*, vol. 8, no. 12, pp. 1140–1149, 2009.
- [55] N. Bélanger, L. Grégoire, A. H. Tahar, and P. J. Bédard, "Chronic treatment with small doses of cabergoline prevents dopa-induced dyskinesias in Parkinsonian monkeys," *Movement Disorders*, vol. 18, no. 12, pp. 1436–1441, 2003.
- [56] Z. Pirtosek, M. Merello, A. Carlsson, and G. Stern, "Prelam and parkinsonian fluctuations," *Clinical Neuropharmacology*, vol. 16, no. 6, pp. 550–554, 1993.
- [57] J. A. Lieberman, "Dopamine partial agonists: a new class of antipsychotic," *CNS Drugs*, vol. 18, no. 4, pp. 251–267, 2004.
- [58] G. Meco, P. Stirpe, F. Editto et al., "Aripiprazole in l-dopa-induced dyskinesias: a one-year open-label pilot study," *Journal of Neural Transmission*, vol. 116, no. 7, pp. 881–884, 2009.
- [59] J. D. Oh, F. Bibbiani, and T. N. Chase, "Quetiapine attenuates levodopa-induced motor complications in rodent and primate parkinsonian models," *Experimental Neurology*, vol. 177, no. 2, pp. 557–564, 2002.
- [60] R. Katzschlager, A. J. Manson, A. Evans, H. Watt, and A. J. Lees, "Low dose quetiapine for drug induced dyskinesias in Parkinson's disease: a double blind cross over study," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 75, no. 2, pp. 295–297, 2004.

- [61] F. Durif, B. Debilly, M. Galitzky et al., "Clozapine improves dyskinesias in Parkinson disease A double-blind, placebo-controlled study," *Neurology*, vol. 62, no. 3, pp. 381–388, 2004.
- [62] L. V. Metman, P. Del Dotto, P. J. Blanchet, P. Van Den Munckhof, and T. N. Chase, "Blockade of glutamatergic transmission as treatment for dyskinesias and motor fluctuations in Parkinson's disease," *Amino Acids*, vol. 14, no. 1–3, pp. 75–82, 1998.
- [63] P. Del Dotto, N. Pavese, G. Gambaccini et al., "Intravenous amantadine improves levodopa-induced dyskinesias: an acute double-blind placebo-controlled study," *Movement Disorders*, vol. 16, no. 3, pp. 515–520, 2001.
- [64] E. Luginger, G. K. Wenning, S. Bösch, and W. Poewe, "Beneficial effects of amantadine on L-dopa-induced dyskinesias in Parkinson's disease," *Movement Disorders*, vol. 15, no. 5, pp. 873–878, 2000.
- [65] B. J. Snow, L. Macdonald, D. McAuley, and W. Wallis, "The effect of amantadine on levodopa-induced dyskinesias in Parkinson's disease: a double-blind, placebo-controlled study," *Clinical Neuropharmacology*, vol. 23, no. 2, pp. 82–85, 2000.
- [66] A. Thomas, D. Iacono, A. L. Luciano, K. Armellino, A. Di Iorio, and M. Onofrij, "Duration of amantadine benefit on dyskinesia of severe Parkinson's disease," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 75, no. 1, pp. 141–143, 2004.
- [67] M. Merello, S. Perez-Lloret, J. Antico, and J. A. Obeso, "Dyskinesias induced by subthalamotomy in Parkinson's disease are unresponsive to amantadine," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 77, no. 2, pp. 172–174, 2006.
- [68] M. Merello, M. I. Nouzeilles, A. Cammarota, and R. Leiguarda, "Effect of memantine (NMDA antagonist) on Parkinson's disease: a double-blind crossover randomized study," *Clinical Neuropharmacology*, vol. 22, no. 5, pp. 273–276, 1999.
- [69] J. Lökk, "Memantine can relieve certain symptoms in Parkinson's disease," *Lakartidningen*, vol. 101, no. 23, pp. 2003–2006, 2004.
- [70] S. Varanese, J. Howard, and A. Di Rocco, "NMDA antagonist memantine improves levodopa-induced dyskinesias and "on-off" phenomena in Parkinson's disease," *Movement Disorders*, vol. 25, no. 4, pp. 508–510, 2010.
- [71] C. A. Braz, V. Borges, and H. B. Ferraz, "Effect of riluzole on dyskinesia and duration of the on state in Parkinson disease patients: a double-blind, placebo-controlled pilot study," *Clinical Neuropharmacology*, vol. 27, no. 1, pp. 25–29, 2004.
- [72] W. Bara-Jimenez, T. D. Dimitrova, A. Sherzai, M. Aksu, and T. N. Chase, "Glutamate release inhibition ineffective in levodopa-induced motor complications," *Movement Disorders*, vol. 21, no. 9, pp. 1380–1383, 2006.
- [73] F. Durif, M. Vidailhet, A. M. Bonnet, J. Blin, and Y. Agid, "Levodopa-induced dyskinesias are improved by fluoxetine," *Neurology*, vol. 45, no. 10, pp. 1855–1858, 1995.
- [74] B. Kleedorfer, A. J. Lees, and G. M. Stern, "Buspirone in the treatment of levodopa induced dyskinesias," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 54, no. 4, pp. 376–377, 1991.
- [75] V. Bonifati, E. Fabrizio, R. Cipriani, N. Vanacore, and G. Meco, "Buspirone in levodopa-induced dyskinesias," *Clinical Neuropharmacology*, vol. 17, no. 1, pp. 73–82, 1994.
- [76] G. Meco, E. Fabrizio, S. Di Rezze, A. Alessandri, and L. Pratesi, "Mirtazapine in L-dopa-induced dyskinesias," *Clinical Neuropharmacology*, vol. 26, no. 4, pp. 179–181, 2003.
- [77] P. Samadi, L. Grégoire, and P. J. Bédard, "The opioid agonist morphine decreases the dyskinetic response to dopaminergic agents in parkinsonian monkeys," *Neurobiology of Disease*, vol. 16, no. 1, pp. 246–253, 2004.
- [78] J. B. Koprach, S. H. Fox, T. H. Johnston et al., "The selective mu-opioid receptor antagonist adl5510 reduces levodopa-induced dyskinesia without affecting antiparkinsonian action in mptp-lesioned macaque model of Parkinson's disease," *Movement Disorders*, vol. 26, no. 7, pp. 1225–1233, 2011.
- [79] C. B. Carroll, P. O. Bain, L. Teare et al., "Cannabis for dyskinesia in Parkinson disease: a randomized double-blind crossover study," *Neurology*, vol. 63, no. 7, pp. 1245–1250, 2004.
- [80] A. F. Carpentier, A. M. Bonnet, M. Vidailhet, and Y. Agid, "Improvement of levodopa-induced dyskinesia by propranolol in Parkinson's disease," *Neurology*, vol. 46, no. 6, pp. 1548–1551, 1996.
- [81] K. Buck, P. Voehringer, and B. Ferger, "The  $\alpha 2$  adrenoceptor antagonist idazoxan alleviates L-DOPA-induced dyskinesia by reduction of striatal dopamine levels: an in vivo microdialysis study in 6-hydroxydopamine-lesioned rats," *Journal of Neurochemistry*, vol. 112, no. 2, pp. 444–452, 2010.
- [82] R. Grondin, A. H. Tahar, V. D. Doan, P. Ladure, and P. J. Bédard, "Noradrenoceptor antagonism with idazoxan improves L-dopa-induced dyskinesias in MPTP monkeys," *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 361, no. 2, pp. 181–186, 2000.
- [83] O. Rascol, I. Arnulf, H. Peyro-Saint Paul et al., "Idazoxan, an alpha-2 antagonist, and L-DOPA-induced dyskinesias in patients with Parkinson's disease," *Movement Disorders*, vol. 16, no. 4, pp. 708–713, 2001.
- [84] M. D. Gottwald and M. J. Aminoff, "Therapies for dopaminergic-induced dyskinesias in Parkinson disease," *Annals of Neurology*, vol. 69, no. 6, pp. 919–927, 2011.
- [85] T. Müller, "New small molecules for the treatment of Parkinson's disease," *Expert Opinion on Investigational Drugs*, vol. 19, no. 9, pp. 1077–1086, 2010.
- [86] D. J. Brooks, S. Papapetropoulos, F. Vandenhende et al., "An open-label, positron emission tomography study to assess adenosine A2A brain receptor occupancy of vipadenant (BIIB014) at steady-state levels in healthy male volunteers," *Clinical Neuropharmacology*, vol. 33, no. 2, pp. 55–60, 2010.
- [87] J. Jankovic, "Are adenosine antagonists, such as istradefylline, caffeine, and chocolate, useful in the treatment of Parkinson's disease?" *Annals of Neurology*, vol. 63, no. 3, pp. 267–269, 2008.
- [88] E. Pourcher, H. H. Fernandez, M. Stacy, A. Mori, R. Ballerini, and P. Chaikin, "Istradefylline for Parkinson's disease patients experiencing motor fluctuations: results of the KW-6002-US-018 study," *Parkinsonism and Related Disorders*, vol. 18, no. 2, pp. 178–184, 2012.
- [89] H. H. Fernandez, D. R. Greeley, R. M. Zweig, J. Wojcieszek, A. Mori, and N. M. Sussman, "Istradefylline as monotherapy for Parkinson disease: results of the 6002-US-051 trial," *Parkinsonism and Related Disorders*, vol. 16, no. 1, pp. 16–20, 2010.
- [90] R. A. Hauser, L. M. Shulman, J. M. Trugman et al., "Study of istradefylline in patients with Parkinson's disease on levodopa with motor fluctuations," *Movement Disorders*, vol. 23, no. 15, pp. 2177–2185, 2008.

- [91] M. Stacy, D. Silver, T. Mendis et al., "A 12-week, placebo-controlled study (6002-US-006) of istradefylline in Parkinson disease," *Neurology*, vol. 70, no. 23, pp. 2233–2240, 2008.
- [92] P. A. LeWitt, M. Guttman, J. W. Tetrud et al., "Adenosine A2A receptor antagonist istradefylline (KW-6002) reduces off time in Parkinson's disease: a double-blind, randomized, multicenter clinical trial (6002-US-005)," *Annals of Neurology*, vol. 63, no. 3, pp. 295–302, 2008.
- [93] R. A. Hauser, M. Cantillon, E. Pourcher et al., "Preladenant in patients with Parkinson's disease and motor fluctuations: a phase 2, double-blind, randomised trial," *The Lancet Neurology*, vol. 10, no. 3, pp. 221–229, 2011.
- [94] M. Contin, P. Martinelli, F. Albani et al., "Kinetic-dynamic monitoring of levetiracetam effects in patients with Parkinson disease and levodopa-induced dyskinesias," *Clinical Neuropharmacology*, vol. 30, no. 2, pp. 122–124, 2007.
- [95] K. E. Lyons and R. Pahwa, "Efficacy and tolerability of levetiracetam in Parkinson disease patients with levodopa-induced dyskinesia," *Clinical Neuropharmacology*, vol. 29, no. 3, pp. 148–153, 2006.
- [96] M. Wolz, M. Löhle, K. Strecker et al., "Levetiracetam for levodopa-induced dyskinesia in Parkinson's disease: a randomized, double-blind, placebo-controlled trial," *Journal of Neural Transmission*, vol. 117, no. 11, pp. 1279–1286, 2010.
- [97] K. K. Wong, J. E. Alty, A. G. Goy, S. Raghav, D. C. Reutens, and P. A. Kempster, "A randomized, double-blind, placebo-controlled trial of levetiracetam for dyskinesia in Parkinson's disease," *Movement Disorders*, vol. 26, no. 8, pp. 1552–1555, 2011.
- [98] T. A. Zesiewicz, K. L. Sullivan, J. L. Maldonado, W. O. Tatum, and R. A. Hauser, "Open-label pilot study of levetiracetam (Keppra) for the treatment of levodopa-induced dyskinesias in Parkinson's disease," *Movement Disorders*, vol. 20, no. 9, pp. 1205–1209, 2005.
- [99] J. Bronzova, C. Sampaio, R. A. Hauser et al., "Double-blind study of pardopruxon, a new partial dopamine agonist, in early Parkinson's disease," *Movement Disorders*, vol. 25, no. 6, pp. 738–746, 2010.
- [100] R. A. Hauser, J. Bronzova, C. Sampaio et al., "Safety and tolerability of pardopruxon, a new partial dopamine agonist, in a randomized, controlled study of patients with advanced Parkinson's disease for the pardopruxon study group," *European Neurology*, vol. 62, no. 1, pp. 40–48, 2009.
- [101] O. Rascol, J. Bronzova, R. A. Hauser et al., "Pardopruxon as adjunct therapy to levodopa in patients with Parkinson's disease experiencing motor fluctuations: results of a double-blind, randomized, placebo-controlled, trial," *Parkinsonism and Related Disorders*, vol. 18, no. 4, pp. 370–376, 2012.
- [102] F. Stocchi, G. Arnold, M. Onofrij et al., "Improvement of motor function in early Parkinson disease by safinamide," *Neurology*, vol. 63, no. 4, pp. 746–748, 2004.
- [103] D. Grosset and A. Grosset, "The movement disorder society—14th international congress of Parkinson's disease and movement disorders," *IDrugs*, vol. 13, no. 8, pp. 539–542, 2010.
- [104] D. Berg, J. Godau, C. Trenkwalder et al., "AFQ056 treatment of levodopa-induced dyskinesias: results of 2 randomized controlled trials," *Movement Disorders*, vol. 26, no. 7, pp. 1243–1250, 2011.
- [105] F. Fregni, D. K. Simon, A. Wu, and A. Pascual-Leone, "Non-invasive brain stimulation for Parkinson's disease: a systematic review and meta-analysis of the literature," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 76, no. 12, pp. 1614–1623, 2005.
- [106] T. N. Chase, F. Baronti, G. Fabbrini, I. J. Heuser, J. L. Juncos, and M. M. Mouradian, "Rationale for continuous dopaminomimetic therapy of Parkinson's disease," *Neurology*, vol. 39, supplement 2, no. 11, pp. 7–19, 1989.
- [107] R. Horowski, C. D. Marsden, and J. A. Obeso, "Continuous dopaminergic stimulation: state of the art and outlook," *Journal of Neural Transmission, Supplement*, vol. 27, pp. 249–252, 1988.
- [108] J. A. Obeso, F. Grandas, M. T. Herrero, and R. Horowski, "The role of pulsatile versus continuous dopamine receptor stimulation for functional recovery in Parkinson's disease," *European Journal of Neuroscience*, vol. 6, no. 6, pp. 889–897, 1994.
- [109] M. M. Mouradian, I. J. E. Heuser, F. Baronti, and T. N. Chase, "Modification of central dopaminergic mechanisms by continuous levodopa therapy for advanced Parkinson's disease," *Annals of Neurology*, vol. 27, no. 1, pp. 18–23, 1990.
- [110] A. Antonini, I. U. Isaías, M. Canesi et al., "Duodenal levodopa infusion for advanced Parkinson's disease: 12-month treatment outcome," *Movement Disorders*, vol. 22, no. 8, pp. 1145–1149, 2007.
- [111] D. Devos, "Patient profile, indications, efficacy and safety of duodenal levodopa infusion in advanced Parkinson's disease," *Movement Disorders*, vol. 24, no. 7, pp. 993–1000, 2009.
- [112] D. Nyholm, T. Lewander, A. Johansson, P. A. LeWitt, C. Lundqvist, and S. M. Aquilonius, "Enteral levodopa/carbidopa infusion in advanced Parkinson disease: long-term exposure," *Clinical Neuropharmacology*, vol. 31, no. 2, pp. 63–73, 2008.
- [113] D. Nyholm, A. I. M. Nilsson Remahl, N. Dizdar et al., "Duodenal levodopa infusion monotherapy vs oral polypharmacy in advanced Parkinson disease," *Neurology*, vol. 64, no. 2, pp. 216–223, 2005.
- [114] T. Müller, K. Renger, and W. Kuhn, "Levodopa-associated increase of homocysteine levels and sural axonal neurodegeneration," *Archives of Neurology*, vol. 61, no. 5, pp. 657–660, 2004.
- [115] M. Onofrij, L. Bonanni, G. Cossu, D. Manca, F. Stocchi, and A. Thomas, "Emergencies in parkinsonism: akinetic crisis, life-threatening dyskinesias, and polyneuropathy during L-Dopa gel treatment," *Parkinsonism and Related Disorders*, vol. 15, supplement 3, pp. S233–S236, 2009.
- [116] G. Block, C. Liss, S. Reines et al., "Comparison of immediate-release and controlled release carbidopa/levodopa in Parkinson's disease. A multicenter 5-year study," *European Neurology*, vol. 37, no. 1, pp. 23–27, 1997.
- [117] F. Stocchi, O. Rascol, K. Kiebert et al., "Initiating levodopa/carbidopa therapy with and without entacapone in early Parkinson disease: the STRIDE-PD study," *Annals of Neurology*, vol. 68, no. 1, pp. 18–27, 2010.
- [118] A. J. Stoessl, "Continuous dopaminergic therapy in Parkinson disease: time to stride back?" *Annals of Neurology*, vol. 68, no. 1, pp. 3–5, 2010.
- [119] R. A. Hauser, A. L. Ellenbogen, L. V. Metman et al., "Cross-over comparison of IPX066 and a standard levodopa formulation in advanced Parkinson's disease," *Movement Disorders*, vol. 26, no. 12, pp. 2246–2252, 2011.
- [120] F. Stocchi, S. Ruggieri, L. Vacca, and C. W. Olanow, "Prospective randomized trial of lisuride infusion versus oral levodopa in patients with Parkinson's disease," *Brain*, vol. 125, pp. 2058–2066, 2002.

- [121] J. A. Obeso, M. R. Luquin, and J. M. Martinez-Lage, "Lisuride infusion pump: a device for the treatment of motor fluctuations in Parkinson's disease," *The Lancet*, vol. 1, no. 8479, pp. 467–470, 1986.
- [122] J. Vaamonde, M. R. Luquin, and J. A. Obeso, "Subcutaneous lisuride infusion in Parkinson's disease. Response to chronic administration in 34 patients," *Brain*, vol. 114, pp. 601–617, 1991.
- [123] A. J. Manson, K. Turner, and A. J. Lees, "Apomorphine monotherapy in the treatment of refractory motor complications of Parkinson's disease: long-term follow-up study of 64 patients," *Movement Disorders*, vol. 17, no. 6, pp. 1235–1241, 2002.
- [124] P. J. García Ruiz, Á. S. Ignacio, B. A. Pensado et al., "Efficacy of long-term continuous subcutaneous apomorphine infusion in advanced Parkinson's disease with motor fluctuations: a multicenter study," *Movement Disorders*, vol. 23, no. 8, pp. 1130–1136, 2008.
- [125] R. Katzenschlager, A. Hughes, A. Evans et al., "Continuous subcutaneous apomorphine therapy improves dyskinesias in Parkinson's disease: a prospective study using single-dose challenges," *Movement Disorders*, vol. 20, no. 2, pp. 151–157, 2005.
- [126] C. Ohye, T. Maeda, and H. Narabayashi, "Physiologically defined VIM nucleus. Its special reference to control of tremor," *Applied Neurophysiology*, vol. 39, no. 3-4, pp. 285–295, 1976.
- [127] F. A. Lenz, H. C. Kwan, R. L. Martin, R. R. Tasker, J. O. Dostrovsky, and Y. E. Lenz, "Single unit analysis of the human ventral thalamic nuclear group. Tremor-related activity in functionally identified cells," *Brain*, vol. 117, pp. 531–543, 1994.
- [128] R. R. Tasker and Z. H. T. Kiss, "The role of the thalamus in functional neurosurgery," *Neurosurgery Clinics of North America*, vol. 6, no. 1, pp. 73–104, 1995.
- [129] J. M. Van Buren, C. L. Li, and D. Y. Shapiro, "A qualitative and quantitative evaluation of parkinsonians three to six years following thalamotomy," *Confinia Neurologica*, vol. 35, no. 4, pp. 202–235, 1973.
- [130] J. Husby and A. G. Korsgaard, "Proceedings: late results of thalamotomy in Parkinsonism with and without the influence of levodopa," *Acta Neurochirurgica*, vol. 31, no. 3-4, p. 260, 1975.
- [131] P. J. Derome, C. P. Jedynak, A. Visot, and O. Delalande, "Treatment of abnormal movements by thalamic lesions," *Revista de Neurologia*, vol. 142, no. 4, pp. 391–397, 1986.
- [132] Y. Nagaseki, T. Shibasaki, and T. Hirai, "Long-term follow-up results of selective VIM-thalamotomy," *Journal of Neurosurgery*, vol. 65, no. 3, pp. 296–302, 1986.
- [133] M. W. Fox, J. E. Ahlskog, and P. J. Kelly, "Stereotactic ventrolateralis thalamotomy for medically refractory tremor in post-levodopa era Parkinson's disease patients," *Journal of Neurosurgery*, vol. 75, no. 5, pp. 723–730, 1991.
- [134] J. Jankovic, F. Cardoso, R. G. Grossman, W. J. Hamilton, R. R. Tasker, and P. J. Kelly, "Outcome after stereotactic thalamotomy for parkinsonian, essential, and other types of tremor," *Neurosurgery*, vol. 37, no. 4, pp. 680–687, 1995.
- [135] P. J. Kelly and F. J. Gillingham, "The long-term results of stereotaxic surgery and L-dopa therapy in patients with Parkinson's disease. A 10-year follow-up study," *Journal of Neurosurgery*, vol. 53, no. 3, pp. 332–327, 1980.
- [136] T. Miyamoto, H. Bekku, E. Moriyama, and S. Tsuchida, "Present role of stereotactic thalamotomy for Parkinsonism. Retrospective analysis of operative results and thalamic lesions in computed tomograms," *Applied Neurophysiology*, vol. 48, no. 1–6, pp. 294–304, 1985.
- [137] N. Diederich, C. G. Goetz, G. T. Stebbins et al., "Blinded evaluation confirms long-term asymmetric effect of unilateral thalamotomy or subthalamotomy on tremor in Parkinson's disease," *Neurology*, vol. 42, no. 7, pp. 1311–1314, 1992.
- [138] H. Narabayashi, F. Yokochi, and Y. Nakajima, "Levodopa-induced dyskinesia and thalamotomy," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 47, no. 8, pp. 831–839, 1984.
- [139] R. D. Page, M. A. Sambrook, and A. R. Crossman, "Thalamotomy for the alleviation of levodopa-induced dyskinesia: experimental studies in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-treated Parkinsonian monkey," *Neuroscience*, vol. 55, no. 1, pp. 147–165, 1993.
- [140] A. L. Benabid, P. Pollak, M. Hommel, J. M. Gaio, J. de Rougemont, and J. Perret, "Treatment of Parkinson tremor by chronic stimulation of the ventral intermediate nucleus of the thalamus," *Revista de Neurologia*, vol. 145, no. 4, pp. 320–323, 1989.
- [141] A. L. Benabid, P. Pollak, E. Seigneuret, D. Hoffmann, E. Gay, and J. Perret, "Chronic VIM thalamic stimulation in Parkinson's disease, essential tremor and extra-pyramidal dyskinesias," *Acta Neurochirurgica, Supplement*, vol. 58, pp. 39–44, 1993.
- [142] P. Limousin, J. D. Speelman, F. Gielen, and M. Janssens, "Multicentre European study of thalamic stimulation in parkinsonian and essential tremor," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 66, no. 3, pp. 289–296, 1999.
- [143] M. I. Hariz, P. Krack, F. Alesch et al., "Multicentre European study of thalamic stimulation for parkinsonian tremor: a 6 year follow-up," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 79, no. 6, pp. 694–699, 2008.
- [144] R. R. Tasker, M. Munz, F. S. Junn et al., "Deep brain stimulation and thalamotomy for tremor compared," *Acta Neurochirurgica, Supplement*, vol. 68, pp. 49–53, 1997.
- [145] S. Blond, D. Caparros-Lefebvre, F. Parker et al., "Control of tremor and involuntary movement disorders by chronic stereotactic stimulation of the ventral intermediate thalamic nucleus," *Journal of Neurosurgery*, vol. 77, no. 1, pp. 62–68, 1992.
- [146] D. Caparros-Lefebvre, S. Blond, P. Vermersch, N. Pecheux, J. D. Guieu, and H. Petit, "Chronic thalamic stimulation improves tremor and levodopa induced dyskinesias in Parkinson's disease," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 56, no. 3, pp. 268–273, 1993.
- [147] D. Caparros-Lefebvre, S. Blond, M. P. Feltin, P. Pollak, and A. L. Benabid, "Improvement of levodopa induced dyskinesias by thalamic deep brain stimulation is related to slight variation in electrode placement: possible involvement of the centre median and parafascicularis complex," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 67, no. 3, pp. 308–314, 1999.
- [148] J. L. Lanciego, M. C. Rodríguez-Oroz, F. J. Blesa et al., "Lesion of the centromedian thalamic nucleus in MPTP-treated monkeys," *Movement Disorders*, vol. 23, no. 5, pp. 708–715, 2008.
- [149] L. V. Laitinen, A. T. Bergenheim, and M. I. Hariz, "Leksell's posteroventral pallidotomy in the treatment of Parkinson's disease," *Journal of Neurosurgery*, vol. 76, no. 1, pp. 53–61, 1992.
- [150] M. I. Hariz and A. T. Bergenheim, "A 10-year follow-up review of patients who underwent Leksell's posteroventral

- pallidotomy for Parkinson disease," *Journal of Neurosurgery*, vol. 94, no. 4, pp. 552–558, 2001.
- [151] M. S. Baron, J. L. Vitek, R. A. Bakay et al., "Treatment of advanced Parkinson's disease by unilateral posterior GPI pallidotomy: 4-year results of a pilot study," *Movement Disorders*, vol. 15, no. 2, pp. 230–237, 2000.
- [152] J. A. Obeso, C. W. Olanow, M. C. Rodriguez-Oroz, P. Krack, R. Kumar, and A. E. Lang, "Deep-brain stimulation of the subthalamic nucleus or the pars interna of the globus pallidus in Parkinson's disease," *The New England Journal of Medicine*, vol. 345, no. 13, pp. 956–963, 2001.
- [153] M. C. Rodriguez-Oroz, J. A. Obeso, A. E. Lang et al., "Bilateral deep brain stimulation in Parkinson's disease: a multicentre study with 4 years follow-up," *Brain*, vol. 128, pp. 2240–2249, 2005.
- [154] J. Volkmann, N. Allert, J. Voges, V. Sturm, A. Schnitzler, and H. J. Freund, "Long-term results of bilateral pallidal stimulation in Parkinson's disease," *Annals of Neurology*, vol. 55, no. 6, pp. 871–875, 2004.
- [155] E. Moro, A. M. Lozano, P. Pollak et al., "Long-term results of a multicenter study on subthalamic and pallidal stimulation in Parkinson's disease," *Movement Disorders*, vol. 25, no. 5, pp. 578–586, 2010.
- [156] L. Alvarez, R. Macias, N. Pavón et al., "Therapeutic efficacy of unilateral subthalamotomy in Parkinson's disease: results in 89 patients followed for up to 36 months," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 80, no. 9, pp. 979–985, 2009.
- [157] P. Krack, P. Pollak, P. Limousin, A. Benazzouz, G. Deuschl, and A. L. Benabid, "From off-period dystonia to peak-dose chorea. The clinical spectrum of varying subthalamic nucleus activity," *Brain*, vol. 122, pp. 1133–1146, 1999.
- [158] H. Stolze, S. Klebe, M. Poepping et al., "Effects of bilateral subthalamic nucleus stimulation on parkinsonian gait," *Neurology*, vol. 57, no. 1, pp. 144–146, 2001.
- [159] T. Simuni, J. L. Jaggi, H. Mulholland et al., "Bilateral stimulation of the subthalamic nucleus in patients with Parkinson disease: a study of efficacy and safety," *Journal of Neurosurgery*, vol. 96, no. 4, pp. 666–672, 2002.
- [160] P. Krack, A. Batir, N. Van Blercom et al., "Five-year follow-up of bilateral stimulation of the subthalamic nucleus in advanced Parkinson's disease," *The New England Journal of Medicine*, vol. 349, no. 20, pp. 1925–1934, 2003.
- [161] M. C. Rodriguez-Oroz, I. Zamerbide, J. Gyridi, M. R. Palmero, and J. A. Obeso, "Efficacy of deep brain stimulation of the subthalamic nucleus in Parkinson's disease 4 years after surgery: double blind and label evaluation," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 75, no. 10, pp. 1382–1385, 2004.
- [162] G. Deuschl, C. Schade-Brittinger, P. Krack et al., "A randomized trial of deep-brain stimulation for Parkinson's disease," *The New England Journal of Medicine*, vol. 355, no. 9, pp. 896–908, 2006.
- [163] C. Hamani, E. Richter, J. M. Schwalb, and A. M. Lozano, "Bilateral subthalamic nucleus stimulation for Parkinson's disease: a systematic review of the clinical literature," *Neurosurgery*, vol. 56, no. 6, pp. 1313–1324, 2005.
- [164] K. A. Follett, "Comparison of pallidal and subthalamic deep brain stimulation for the treatment of levodopa-induced dyskinesias," *Neurosurg Focus*, vol. 17, no. 1, p. E3, 2004.
- [165] P. Limousin, P. Krack, P. Pollak et al., "Electrical stimulation of the subthalamic nucleus in advanced Parkinsonian's disease," *The New England Journal of Medicine*, vol. 339, no. 16, pp. 1105–1111, 1998.
- [166] E. B. Montgomery and K. B. Baker, "Mechanisms of deep brain stimulation and future technical developments," *Neurological Research*, vol. 22, no. 3, pp. 259–266, 2000.
- [167] F. J. G. Vingerhoets, J. G. Villemure, P. Temperli, C. Pollo, E. Pralong, and J. Ghika, "Subthalamic DBS replaces levodopa in Parkinson's disease: two-year follow-up," *Neurology*, vol. 58, no. 3, pp. 396–401, 2002.
- [168] H. Russmann, J. Ghika, J. G. Villemure et al., "Subthalamic nucleus deep brain stimulation in Parkinson disease patients over age 70 years," *Neurology*, vol. 63, no. 10, pp. 1952–1954, 2004.
- [169] J. L. Molinuevo, F. Valldeoriola, E. Tolosa et al., "Levodopa withdrawal after bilateral subthalamic nucleus stimulation in advanced Parkinson disease," *Archives of Neurology*, vol. 57, no. 7, pp. 983–988, 2000.
- [170] R. Figueiras-Mendez, F. Marin-Zarza, J. A. Molina et al., "Subthalamic nucleus stimulation improves directly levodopa induced dyskinesias in Parkinson's disease," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 66, no. 4, pp. 549–550, 1999.
- [171] M. C. Rodriguez-Oroz, A. Gorospe, J. Guridi et al., "Bilateral deep brain stimulation of the subthalamic nucleus in Parkinson's disease," *Neurology*, vol. 55, supplement 6, no. 12, pp. S45–S51, 2000.
- [172] J. A. Obeso, M. Rodriguez-Oroz, C. Marin et al., "The origin of motor fluctuations in Parkinson's disease: importance of dopaminergic innervation and basal ganglia circuits," *Neurology*, vol. 62, supplement 1, no. 1, pp. S17–S30, 2004.
- [173] J. A. Saint-Cyr, T. Hoque, L. C. M. Pereira et al., "Localization of clinically effective stimulating electrodes in the human subthalamic nucleus on magnetic resonance imaging," *Journal of Neurosurgery*, vol. 97, no. 5, pp. 1152–1166, 2002.
- [174] J. Voges, J. Volkmann, N. Allert et al., "Bilateral high-frequency stimulation in the subthalamic nucleus for the treatment of Parkinson disease: correlation of therapeutic effect with anatomical electrode position," *Journal of Neurosurgery*, vol. 96, no. 2, pp. 269–279, 2002.
- [175] W. Hamel, U. Fietzek, A. Morsnowski et al., "Deep brain stimulation of the subthalamic nucleus in Parkinson's disease: evaluation of active electrode contacts," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 74, no. 8, pp. 1036–1046, 2003.
- [176] J. Guridi, J. A. Obeso, M. C. Rodriguez-Oroz, A. A. Lozano, and M. Manrique, "L-dopa-induced dyskinesia and stereotactic surgery for Parkinson's disease," *Neurosurgery*, vol. 62, no. 2, pp. 311–323, 2008.

## Review Article

# A<sub>2A</sub> Receptor Antagonism and Dyskinesia in Parkinson's Disease

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Dyskinesia, a major complication of treatment of Parkinson's disease (PD), involves two phases: induction, which is responsible for dyskinesia onset, and expression, which underlies its clinical manifestation. The unique cellular and regional distribution of adenosine A<sub>2A</sub> receptors in basal ganglia areas that are richly innervated by dopamine, and their antagonistic role towards dopamine receptor stimulation, have positioned A<sub>2A</sub> receptor antagonists as an attractive nondopaminergic target to improve the motor deficits that characterize PD. In this paper, we describe the biochemical characteristics of A<sub>2A</sub> receptors and the effects of adenosine A<sub>2A</sub> antagonists in rodent and primate models of PD on L-DOPA-induced dyskinesia, together with relevant biomarker studies. We also review clinical trials of A<sub>2A</sub> antagonists as adjuncts to L-DOPA in PD patients with motor fluctuations. These studies have generally demonstrated that the addition of an A<sub>2A</sub> antagonist to a stable L-DOPA regimen reduces OFF time and mildly increases dyskinesia. However, limited clinical data suggest that the addition of an A<sub>2A</sub> antagonist along with a reduction of L-DOPA might maintain anti-Parkinsonian benefit and reduce dyskinesia. Whether A<sub>2A</sub> antagonists might reduce the development of dyskinesia has not yet been tested clinically.

## 1. Adenosine A<sub>2A</sub> Receptor Localization and Biochemistry

Adenosine A<sub>2A</sub> receptors are present in medium to high concentrations in several basal ganglia (BG) nuclei and may therefore be capable of influencing motor activity by acting at different BG levels. This feature renders A<sub>2A</sub> receptors particularly attractive for modulation of dopamine receptor functions in a disease such as Parkinson's disease (PD), which is caused by degeneration of dopaminergic neurons in the nigrostriatal pathway, but associated with changes at several receptor levels. An interesting peculiarity of A<sub>2A</sub> receptors is their selective localization in the indirect striatonigral GABAergic pathway, which contains enkephalin (ENK) and which is known to lead to inhibition of motor behavior [1, 2].

A<sub>2A</sub> receptors are positively coupled to adenylate cyclase and, either at the level of second messengers or through the formation of receptor heterodimers, negatively influence dopamine D<sub>2</sub> receptor activity [3–6]. On the basis of this

anatomical and functional organization, A<sub>2A</sub> receptors acting in concert with D<sub>2</sub> and D<sub>1</sub> receptors are capable of affecting planning and execution of movements [7, 8]. Moreover, the low levels of A<sub>2A</sub> receptors expressed in brain areas other than the BG are at the basis of the low incidence of nonmotor side effects observed in clinical trials so far performed [9]. A<sub>2A</sub> receptors, however, are expressed in some peripheral organs and blood cells, underlying the importance of evaluating these elements in clinical trials testing the efficacy of A<sub>2A</sub> receptor antagonists in PD [10, 11].

Interestingly, an abnormal increase in A<sub>2A</sub> signaling, in the striatum of 6-hydroxydopamine- (6-OHDA-) lesioned rats, and in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine- (MPTP-) treated primates, as well as in PD patients chronically treated with L-DOPA [12–14], might produce a prevailing tone of A<sub>2A</sub> receptors, the activation of which inhibits motor activity. Therefore, blockade of the A<sub>2A</sub> receptor inhibitory tone could be one of the factors underlying the positive effects produced by A<sub>2A</sub> antagonists in PD.

## 2. Adenosine A<sub>2A</sub> Receptor Antagonists in Animal Models of Dyskinesia

**2.1. Behavioral Studies.** Preclinical behavioral investigations suggest that A<sub>2A</sub> antagonists may be of interest in the management of dyskinesia in PD. The first preclinical evidence suggesting that A<sub>2A</sub> antagonists may be utilized in patients rendered dyskinetic by L-DOPA was obtained in 6-OHDA unilaterally lesioned rats subchronically treated with L-DOPA [17]. In this paradigm, the repeated administration of L-DOPA causes a progressive, sensitized, increase in contraversive turning behavior, which is thought to reproduce some aspects of the abnormal motor responses induced by the prolonged treatment with L-DOPA [17–19]. Of great interest, sensitization in contraversive turning behavior did not take place when L-DOPA was administered at a low dose in association with an A<sub>2A</sub> antagonist [17, 20]. Subsequent studies utilizing a full effective L-DOPA dose in rats with established dyskinesia [21] did not report any benefit, since L-DOPA treatment alone or in combination with an A<sub>2A</sub> antagonist presented the same degree of dyskinesia. These results demonstrated that A<sub>2A</sub> antagonists are not antidyskinetic drugs; however, in this model, they did not worsen existing dyskinesia while increasing the efficacy of L-DOPA on motor symptoms.

Studies in MPTP-treated primates, the best experimental model of PD and PD-associated dyskinesia, have confirmed the beneficial effects of blockade of A<sub>2A</sub> receptors. A<sub>2A</sub> antagonists were found not to be prodyskinetic drugs, since their administration to Parkinsonian primates with established dyskinesia induced by L-DOPA relieved motor impairment and did not worsen dyskinesia [22–24]. Moreover, an attenuation of dyskinesia induced by long-term apomorphine was observed when the drug was administered in combination with an A<sub>2A</sub> antagonist [25], suggesting that A<sub>2A</sub> antagonists might lower the dyskinetic potential of dopamine-replacement therapy in specific conditions. The previous coadministration of an A<sub>2A</sub> antagonist was also found to delay the onset of severe dyskinesia when the same primates were maintained on apomorphine alone [25].

**2.2. Biochemical Studies.** Regarding the mechanisms underlying dyskinesia and the effects of A<sub>2A</sub> antagonists in experimental models of dyskinesia, it seems likely that these drugs interfere with the neuroplastic changes induced by dopamine-replacement therapy in the dopamine-denervated BG (Figure 1). Studies in 6-OHDA-lesioned rats demonstrate that striatal dopamine denervation is associated with persistent modifications in the levels of the neuropeptides dynorphin (DYN) and ENK, as well as the enzyme glutamic acid decarboxylase 67 (GAD-67) [21, 26–28] (Figure 1). Moreover, it was observed that chronic treatment with L-DOPA, which induces a dyskinetic-like motor response, further contributes to these biochemical changes [21, 26, 27] (Figure 1). Importantly, the coadministration of an A<sub>2A</sub> antagonist, besides resulting in a stable motor response, attenuated the effects of chronic L-DOPA treatment on ENK and GAD-67 [21, 26, 27]. It has to be acknowledged that, as of

today, no evidence supports a direct role of DYN, ENK, and GAD-67 in dyskinesia. Nevertheless, changes in the expression of neuropeptides are a marker of the activity of striatal neurons [26]. Therefore, it can be suggested that A<sub>2A</sub> antagonists modulate the effects of L-DOPA and mitigate the neuroplastic changes this drug induces in the striatum. These effects could arise from the opposite functional interactions involving adenosine A<sub>2A</sub> and dopamine D<sub>1</sub> and D<sub>2</sub> receptors [29]. These interactions, by amplifying dopaminergic signaling, would regulate the activity of striatal output neurons in conditions of dopamine denervation and nonphysiological stimulation of dopamine receptors (Figure 1).

It has to be considered that A<sub>2A</sub> receptor antagonists, in addition to their potential effects on biochemical and functional changes induced by dopamine-replacement therapy, potentiate the motor-activating effects of L-DOPA and dopaminergic agonists, allowing their use at lower, nondyskinetic doses [7]. Hence, the sparing of dopaminomimetic drugs in combination with an A<sub>2A</sub> antagonist may contribute to the attenuation, or delay, of the maladaptive modifications in striatal function which underlie dyskinesia.

**2.3. Role of Glutamate Transmission.** Besides the facilitation of dopamine transmission, other mechanisms have been proposed to underlie, or at least participate in, the effects of A<sub>2A</sub> receptor antagonists observed in experimental models of dyskinesia. Neuroanatomical studies demonstrate that striatal A<sub>2A</sub> receptors are highly expressed at the postsynaptic level in asymmetric synapses, where they can interact with glutamate receptors [30]. Glutamate receptors are thought to participate in the pathophysiology of dyskinesia [31] and, interestingly, chronic administration of L-DOPA to 6-OHDA-lesioned rats was reported to induce a hyperphosphorylation state of the  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid (AMPA) receptor [25, 32]. This effect was found to be significantly attenuated when L-DOPA was administered in combination with an A<sub>2A</sub> antagonist [25, 32]. Evidence also exists that A<sub>2A</sub> receptors may regulate the conductance of N-methyl-D-aspartate (NMDA) receptors [33]. This may have important implications for dyskinesia, since NMDA receptors play a major role in neuroplasticity phenomena [34, 35], including those which take place in motor circuits, and may underlie abnormal motor responses to dopamine-replacement therapy used in PD.

Interactions between A<sub>2A</sub> receptors and type 5 metabotropic glutamate receptors (mGlu5) have also been reported [36–38]. In the light of the evidence showing that antagonism of mGlu5 receptors may reduce dyskinesia in MPTP-lesioned primates treated with L-DOPA [39], it is possible to envision that combined antagonism on the two receptors might contribute to the beneficial effects of A<sub>2A</sub> antagonists on dyskinesia. Additional mechanisms involved in the modulation of therapy-induced abnormal motor responses by A<sub>2A</sub> antagonists could include interaction with nondopaminergic and nonglutamatergic receptors, such as cannabinoid and serotonin receptors, and regulation of neurotransmitter release [40–42]. In this regard, it has to be recalled that A<sub>2A</sub> receptors powerfully modulate extracellular concentrations

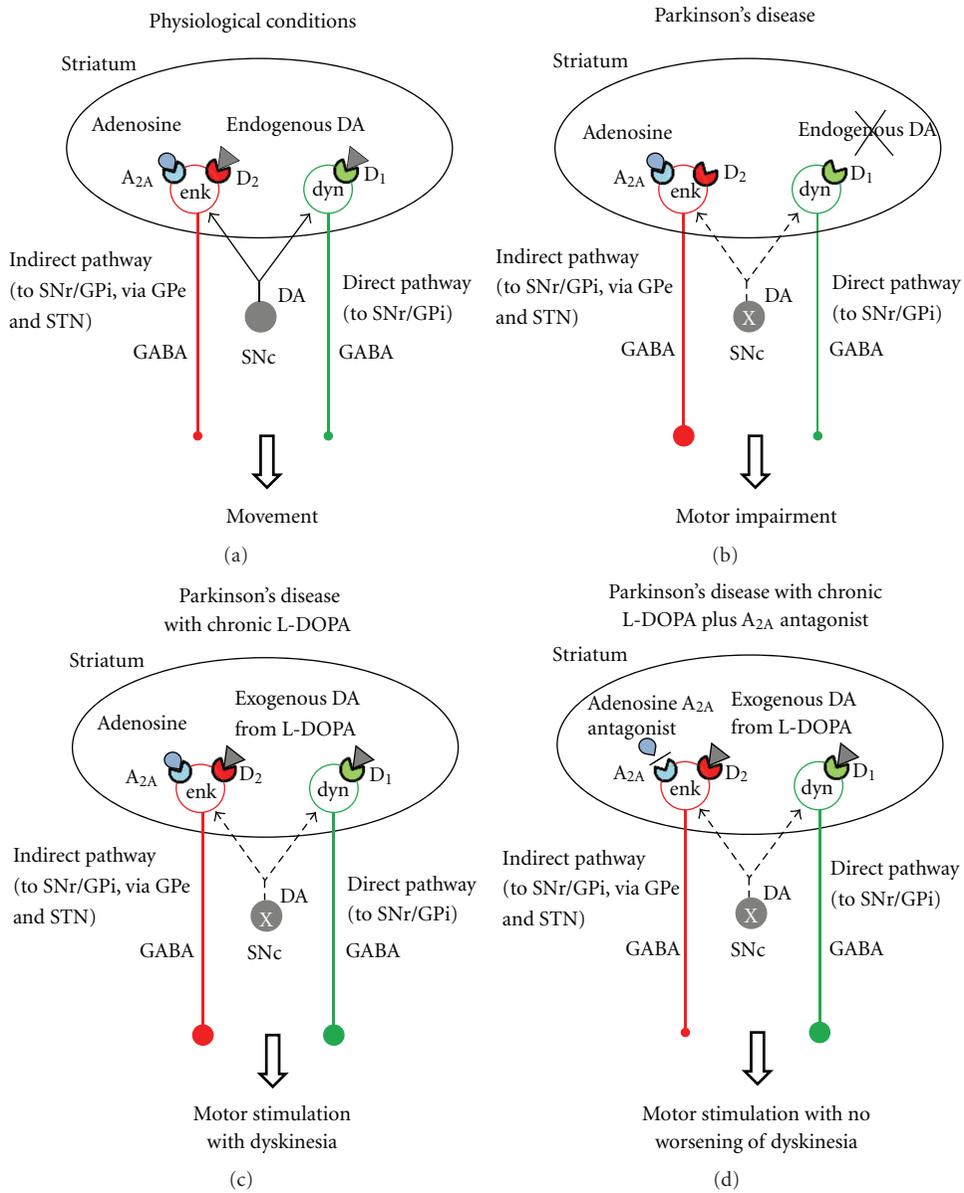


FIGURE 1: Role of A<sub>2A</sub> receptors on modifications in the activity of the striatal efferent pathways. Under physiological conditions (a), striatal neurons receive dopaminergic inputs from the substantia nigra *pars compacta* (SNc). Endogenous dopamine (DA) activates the neurons belonging to the so-called direct pathway (in green), which send GABAergic projections to the substantia nigra *pars reticulata*/globus pallidus *pars interna* (SNr/GPi) and express D<sub>1</sub> stimulatory dopamine receptors, together with the neuropeptide dynorphin (dyn). At the same time, dopamine also depresses the neurons belonging to the so-called indirect pathway (in red) which send GABAergic projections to the SNr/GPi via globus pallidus *pars externa* (GPe) and subthalamic nucleus (STN) and express D<sub>2</sub> inhibitory dopamine receptors and the neuropeptide enkephalin (enk). Adenosine A<sub>2A</sub> receptors stimulate the indirect pathway where they are selectively expressed, and their activation negatively modulates the function of D<sub>2</sub> receptors. A balanced level of activity of the direct and indirect pathways is at the basis of the correct processing of motor information and movement execution. In Parkinson's disease (b), the degeneration of the neurons located in the SNc leads to a drop in the dopaminergic input to the striatum. This results in a reduced activation of the direct pathway and in a disinhibition of the indirect pathway, which is associated with the elevation of A<sub>2A</sub> receptor transmission. Such unbalanced activity of the striatal output pathways is at the basis of the motor impairment observed in Parkinson's disease (b). Administration of L-DOPA restores the compromised dopaminergic tone since it stimulates the direct pathway and inhibits the indirect one (not shown). However, chronic treatment with L-DOPA (c) leads to the overactivation of the direct pathway, which together with the increase of A<sub>2A</sub> receptor activity [12, 15, 16] and enhanced indirect pathway transmission is at the basis of L-DOPA-induced dyskinesia and loss of efficacy. The addition of an A<sub>2A</sub> antagonist to L-DOPA (d) although not counteracting the overactivity of the direct pathway (dyskinesia) stabilizes the activity of the indirect pathway, resulting in motor stimulation, potentially without a worsening of dyskinesia.

of glutamate [43, 44], the excessive increase of which plays a role in the abnormal functioning of BG existing in PD and in neuroplasticity phenomena.

### 3. Biomarkers and Neuroimaging Studies Involving the A<sub>2A</sub> Receptor

A crucial need in the translation from preclinical studies to clinical trials is the availability of reliable biomarkers, which would give the opportunity of monitoring the effects of the compound on its biological target—the adenosine receptor—in addition to evaluating its clinical efficacy. In this field, substantial contributions have been made by neuroimaging studies, while biological findings in peripheral tissues have opened interesting perspectives.

**3.1. Neuroimaging Studies in Humans.** Neuroimaging techniques, based on positron emission tomography (PET), have been recently used to analyze A<sub>2A</sub> receptor distribution in the human brain, either in normal subjects or in PD patients exposed to L-DOPA; in this latter case, the purpose was to draw potential *in vivo* correlations between changes in A<sub>2A</sub> receptor availability and the presence of L-DOPA-induced dyskinesias.

In 2007, Mishina et al. examined the distribution of A<sub>2A</sub> receptors in the brain of 5 normal subjects using PET tracer [7-methyl-<sup>11</sup>C]-(E)-8-(3,4,5-trimethoxystyryl)-1,3,7-trimethylxanthine ([<sup>11</sup>C]TMSX) [45]. Various brain regions were examined, including the cerebellum, brainstem, thalamus, head of caudate nucleus, anterior and posterior putamen, frontal lobe, temporal lobe, parietal lobe, occipital lobe, and posterior cingulate gyrus. Results showed the highest levels of A<sub>2A</sub> receptor binding in the putamen, followed by the caudate nucleus and thalamus, while the lowest levels were detected in the cerebral cortex. Using a different A<sub>2A</sub> receptor-specific radiotracer, [<sup>11</sup>C]SCH442416, Brooks et al. assessed binding of vipadenant (3-(4-amino-3-methylbenzyl)-7-(2-furyl)-3H-[1, 2, 3]triazolo[4, 5-d]pyrimidine-5-amine), a selective nonxanthine A<sub>2A</sub> receptor antagonist synthesized by Vernalis Plc (also known as BIIB014 or V2006) [15]. Displacement of the PET tracer by increasing doses of vipadenant (2.5–100 mg/day for 10 or 11 days) was investigated in various brain regions—including the putamen, caudate nucleus, nucleus accumbens, thalamus of both hemispheres, and cerebellum—of 15 healthy volunteers. The estimated receptor occupancy of vipadenant in the brain varied from 74% to 94% at the lowest daily dose (2.5 mg), with the highest value being observed in the putamen and the lowest value in the cerebellum. Saturation was reached in all regions at the highest dose administered (100 mg). It is noteworthy that double-blind, placebo-controlled, phase 2 clinical trials with vipadenant have been conducted in PD patients, showing modest anti-PD activity, until a review of preclinical toxicology studies, conducted by Vernalis Plc, led to discontinuation of this drug in July 2010 (<http://www.vernalis.com/media-centre/latest-releases/2010-releases/584/>).

Recently, Mishina et al., using PET with [<sup>11</sup>C]TMSX, measured the binding ability of striatal A<sub>2A</sub> receptors in 9 untreated PD patients, 7 PD patients with dyskinesia, and 6 age-matched control subjects [16]. They found that the distribution volume ratio of A<sub>2A</sub> receptors in the putamen was larger in patients with L-DOPA-induced dyskinesias than in control subjects and that L-DOPA treatment tended to increase the presence of A<sub>2A</sub> receptors in the putamen.

Further information on the relationship between A<sub>2A</sub> receptors and L-DOPA-induced dyskinesias has been provided by Ramlackhansingh et al., who investigated adenosine A<sub>2A</sub> receptor availability in the caudate and putamen of PD patients with (*n* = 6) and without L-DOPA-induced dyskinesias (*n* = 6) and in age-matched healthy controls (*n* = 6) [46]. In line with previous studies [12], they found that A<sub>2A</sub> receptor binding was higher in the caudate and putamen of PD patients with L-DOPA-induced dyskinesias, with respect to both PD patients without L-DOPA-induced dyskinesias and controls, thereby lending further support to the view that A<sub>2A</sub> antagonists may prove beneficial in the management of motor complications associated with L-DOPA treatment. It is worth mentioning that although their cohort was small and the power was probably too limited to detect a difference, Ramlackhansingh et al. did not find a correlation between striatal [<sup>11</sup>C]SCH442416 uptake and dyskinesia severity.

An additional study tested the hypothesis that blockade of striatal A<sub>2A</sub> receptors, caused by the selective antagonist SYN115, a benzothiazole derivative, may reduce the inhibitory output of the striatofugal indirect pathway [47]. For this purpose, the authors used a perfusion magnetic resonance imaging (MRI) technique, which gives a functional measure of the cerebral blood flow (CBF) reflecting neuronal activity. The study was conducted during a randomized, double-blind, placebo-controlled, crossover study with SYN115 in 21 PD patients on L-DOPA. The results showed that SYN115 produced a dose-dependent decrease in thalamic CBF, which the authors deemed consistent with reduced pallid-thalamic inhibition via the indirect pathway [47].

**3.2. Peripheral Expression of Adenosine Receptors.** To our knowledge, only one paper has reported the characterization of adenosine receptors in peripheral tissues (peripheral blood cells) of human Parkinsonian subjects [48]. In this study, Varani et al. investigated affinity and density of A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub>, and A<sub>3A</sub> receptors in lymphocyte and neutrophil membranes from PD patients and healthy control subjects; they also analyzed A<sub>2A</sub> receptors density in autoptic samples of putamen from PD patients and control subjects. They found that A<sub>2A</sub> receptors were significantly different between PD patients and controls, in terms of affinity and density, while no changes seemed to affect A<sub>1</sub>, A<sub>2B</sub>, or A<sub>3A</sub> receptors. In particular, increased density of A<sub>2A</sub> receptors, coupled with decreased affinity, was detected in lymphocyte and neutrophil membranes of PD patients, with respect to control subjects. This finding was associated with a reduction in the mRNA of A<sub>2A</sub> receptors, while no changes were observed in the mRNAs of the other adenosine receptor subtypes

investigated. The postmortem study confirmed this result, showing increased A<sub>2A</sub> receptor density in the putamen of PD patients [48].

#### 4. Clinical Trials of A<sub>2A</sub> Receptor Antagonists

**4.1. Istradefylline.** Clinical trials of A<sub>2A</sub> antagonists in patients with motor complications have focused on reductions in OFF time rather than changes in dyskinesia. Istradefylline was the first A<sub>2A</sub> receptor antagonist to enter clinical trials seeking an indication in PD. Bara-Jimenez et al. conducted an early proof-of-principle study using intravenous L-DOPA infusions in 15 moderate to advanced PD patients with motor fluctuations, 6 of whom had L-DOPA-induced peak-dose dyskinesia [49]. Twelve subjects were randomized to istradefylline, 3 to placebo, and 1 dropped out. Istradefylline 40 or 80 mg had no effect on Parkinsonian signs or dyskinesia when added to an optimal L-DOPA infusion. However, when added to a low-dose L-DOPA infusion, istradefylline 40 mg improved Unified Parkinson's Disease Rating Scale (UPDRS) motor scores by 24% ( $P < 0.05$ ) and istradefylline 80 mg improved motor scores by 36% ( $P < 0.05$ ). The anti-Parkinsonian response to a low-dose L-DOPA infusion plus istradefylline 80 mg was similar to an optimal-dose L-DOPA infusion. Notably, the severity of dyskinesia with a low-dose L-DOPA infusion plus istradefylline 80 mg was 45% less ( $P < 0.05$ ) than with an optimal-dose L-DOPA infusion. This suggests that by lowering the L-DOPA dose and adding istradefylline, one might be able to maintain anti-Parkinsonian benefit and reduce dyskinesia, a paradigm that has not been studied in clinical trials using oral L-DOPA preparations.

Istradefylline was then studied in a 12-week, randomized, placebo-controlled exploratory trial in which patients with *both* motor fluctuations and dyskinesia were randomized to the addition of placebo ( $n = 29$ ), istradefylline in ascending doses up to 20 mg/day ( $n = 26$ ), or istradefylline in ascending doses up to 40 mg/day ( $n = 28$ ) [50]. Anti-Parkinsonian medications were kept unchanged except that the total daily L-DOPA dose could be reduced, if necessary, to ameliorate L-DOPA-related adverse events. Over the course of the study, there were no significant changes in daily L-DOPA doses comparing istradefylline and placebo groups ( $P = 0.96$ ). Diary results showed that the combined istradefylline groups experienced a reduction in OFF time of 1.2 hours, whereas the placebo group experienced an increase in OFF time of 0.5 hours ( $P < 0.004$ ). Multiple assessments of change in dyskinesia did not demonstrate significant differences between the placebo and istradefylline groups, including Goetz dyskinesia scale scores (−0.2 versus −0.1,  $P = 0.3$ ), Parkinson dyskinesia scale scores (−1.4 versus −1.3,  $P = 0.9$ ), and UPDRS items 32–34 (−0.03 versus −0.4,  $P = 0.8$ ). However, diary results indicated that ON time with dyskinesia was significantly more increased with istradefylline than placebo (0.6 hours versus −1.5 hours,  $P = 0.001$ ). Troublesome dyskinesia was not included as a diary category in this study. As an adverse event, increased dyskinesia was reported by 13.8% of placebo patients and 16.7% of istradefylline patients.

This is an important study in that it is the largest study of an A<sub>2A</sub> receptor antagonist in a population of patients, all of whom have L-DOPA-induced dyskinesia. In addition, dyskinesia was most thoroughly assessed by multiple scales. Clearly, the addition of istradefylline to a stable antiparkinson regimen did not reduce dyskinesia, nor was there a very substantial increase. Perhaps the most parsimonious interpretation is that overall severity of dyskinesia was essentially unchanged, but much or all of the reduction in OFF time was replaced by an increase in ON time with dyskinesia.

Subsequent trials in moderate to advanced PD patients all included subjects with motor fluctuations, some of whom had dyskinesia and some of whom did not. These included two phase 2 istradefylline trials. In one, istradefylline 40 mg/day reduced OFF time compared with placebo by 1.2 hours ( $P = 0.005$ ) [51]. ON time with dyskinesia increased by 1.0 hour more in the istradefylline group than the placebo group ( $P = 0.035$ ). Of this differential increase in ON time with dyskinesia, approximately 0.8 hours were ON time with nontroublesome dyskinesia ( $P = 0.065$ ), and 0.2 hours were ON time with troublesome dyskinesia ( $P = 0.347$ ). Dyskinesia was reported as an adverse event in 15.2% of placebo subjects and 30.2% of istradefylline subjects. In the other phase 2 study [52], istradefylline 20 mg/day reduced OFF time by 0.64 hours, and istradefylline 60 mg/day reduced OFF time by 0.77 hours (overall  $P$  value = 0.065). Compared with placebo, istradefylline 20 mg/day increased ON time with dyskinesia by 0.54 hours, and ON time with troublesome dyskinesia by 0.06 hours; istradefylline 60 mg/day increased ON time with dyskinesia by 0.23 hours and ON time with troublesome dyskinesia by 0.04 hours. Dyskinesia was reported as an adverse event in 14.3% of placebo subjects, 23.9% of istradefylline 20 mg/day subjects, and 22.6% of istradefylline 60 mg/day subjects.

In a phase 3 study, istradefylline 20 mg/day reduced OFF time compared with placebo 0.7 hours ( $P = 0.03$ ) [53]. Increases in dyskinesia were similar in placebo and istradefylline groups (ON time with dyskinesia: 0.5 versus 0.7 hours,  $P = 0.57$ ; ON time with nontroublesome dyskinesia: 0.4 versus 0.4 hours,  $P = 0.82$ ; ON time with troublesome dyskinesia: 0.2 versus 0.3 hours,  $P = 0.48$ ). However, dyskinesia was reported as an adverse event in 22.6% of istradefylline subjects compared with 12.2% of placebo subjects.

In a phase 3 study in Japan [54], istradefylline 20 mg/day reduced OFF time compared with placebo by 0.65 hours ( $P = 0.013$ ) and 40 mg reduced OFF time compared with placebo by 0.92 hours ( $P < 0.001$ ). Compared with placebo, istradefylline 40 mg/day significantly increased ON time with troublesome dyskinesia (0.35 hours,  $P = 0.011$ ). As an adverse event, dyskinesia was reported in 2.5% of placebo subjects, 8.5% of istradefylline 20 mg/day subjects, and 6.4% of istradefylline 40 mg/day subjects.

Thus, in most of the clinical trials, the addition of istradefylline was associated with some increase in ON time with dyskinesia, and dyskinesia was reported as an adverse event more commonly in istradefylline- than placebo-treated subjects.

A recent population pharmacokinetic-pharmacodynamic study analyzed data from 1798 patients participating in

6 phase 2/3 istradefylline trials [55]. The analysis predicted a maximum probability of experiencing dyskinesia as an adverse event sometime during a study as 15.4% for placebo, 22.5% for istradefylline 20 mg/day, 24.1% for istradefylline 40 mg/day, and 24.3% for istradefylline 60 mg/day.

Thus, clinical data to date do not provide evidence for an antidyskinetic effect of istradefylline but rather suggest that istradefylline mildly increases dyskinesia in a dose-dependent fashion. Results vary slightly from trial to trial and may depend, in part, on the percentage of subjects with dyskinesia at baseline and the severity of their dyskinesia. Other potential factors may include concomitant medications such as amantadine and dietary intake of caffeine, a nonspecific adenosine antagonist, although these factors have not been systematically evaluated.

**4.2. Preladenant.** Preladenant was evaluated in a phase 2, 12-week, dose-finding study of PD patients experiencing motor fluctuations [56]. In this study, patients were randomized to preladenant 1, 2, 5, or 10 mg twice daily (BID) or matching placebo. OFF time was significantly reduced compared with placebo in subjects randomized to preladenant 5 mg BID (1.0 hours,  $P = 0.0486$ ) and preladenant 10 mg BID (1.2 hours,  $P = 0.019$ ). In the 5 mg BID group, compared with placebo, ON time with dyskinesia was increased 0.9 hours ( $P = 0.185$ ), ON time with nontroublesome dyskinesia was increased by 1.0 hour ( $P = 0.064$ ), and ON time with troublesome dyskinesia was decreased by 0.1 hours ( $P = 0.812$ ). In the preladenant 10 mg BID group, compared with placebo, ON time with dyskinesia was increased by 1.3 hours ( $P = 0.054$ ), ON time with nontroublesome dyskinesia was increased by 1.1 hours ( $P = 0.047$ ), and ON time with troublesome dyskinesia was increased by 0.2 hours ( $P = 0.540$ ). These results appear to be similar to some of the istradefylline findings in which much of the reduction in OFF time was replaced by ON time with nontroublesome dyskinesia. Dyskinesia was reported as an adverse event by 13% of placebo subjects, by 9% of preladenant 5 mg BID subjects, and by 13% of preladenant 10 mg BID subjects. This result may be different from what has been observed with istradefylline where dyskinesia was rather consistently reported more frequently as an adverse event in istradefylline-compared with placebo-treated groups. Thus, preliminary results suggest that like istradefylline, preladenant does not reduce dyskinesia, but it remains to be seen whether preladenant causes less dyskinesia than istradefylline, as suggested by these adverse event results.

To our knowledge, there have been no clinical trials evaluating whether an  $A_{2A}$  receptor antagonist can reduce the development of dyskinesia when administered in early disease concomitant with the introduction of dopaminergic therapy. Based on animal model data, this remains an important avenue for future investigation. Similarly, we are not aware of clinical trials of patients with L-DOPA-induced dyskinesia to determine whether lowering the L-DOPA dose and adding an  $A_{2A}$  receptor antagonist will allow maintenance of the anti-Parkinsonian response with reduction of dyskinesia.

## 5. Conclusions

The management of PD is most complex in the treatment of late, complicated PD, when the response to L-DOPA is associated with dyskinesia. From the studies described in the present paper, it is suggested that the management of the first (uncomplicated) phase has important consequences on the induction of dyskinesia that characterize the second (complicated) phase.

Preclinical studies suggest that  $A_{2A}$  antagonists might reduce the development of dyskinesia, but this has not yet been tested clinically. In PD patients, once dyskinesias are established, adding an  $A_{2A}$  antagonist to a stable dopaminergic therapeutic regimen does not appear to provide an antidyskinetic response, and most clinical trials have suggested a mild increase in dyskinesia in association with a reduction in OFF time. Limited clinical data suggest the possibility that in PD patients with established dyskinesia, one might be able to maintain the anti-Parkinsonian response and reduce dyskinesia by adding an  $A_{2A}$  antagonist and lowering the L-DOPA dose, but this remains to be proven. Thus, critical aspects of the potential benefits of  $A_{2A}$  antagonists with regard to dyskinesia are yet to be evaluated.

## Abbreviations

AMPA:	$\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid
BG:	Basal ganglia
BID:	Bis in die, twice a day
CBF:	Cerebral blood flow
PD:	Parkinson's disease
6-OHDA:	6-Hydroxydopamine
MPTP:	1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine
DYN:	Dynorphin
ENK:	Enkephalin
GAD-67:	Glutamic acid decarboxylase 67
mGLU5:	Type 5 metabotropic glutamate receptors 5
MRI:	Magnetic resonance imaging
NMDA:	N-Methyl-D-aspartate
PET:	Positron emission tomography
UPDRS:	Unified Parkinson's disease rating scale.

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## References

- [1] J. S. Fink, D. R. Weaver, S. A. Rivkees et al., "Molecular cloning of the rat  $A_2$  adenosine receptor: selective co-expression with  $D_2$  dopamine receptors in rat striatum," *Molecular Brain Research*, vol. 14, no. 3, pp. 186–195, 1992.
- [2] S. N. Schiffmann, F. Libert, G. Vassart, and J. J. Vanderhaeghen, "Distribution of adenosine  $A_2$  receptor mRNA in the human brain," *Neuroscience Letters*, vol. 130, no. 2, pp. 177–181, 1991.
- [3] M. Canals, D. Marcellino, F. Fanelli et al., "Adenosine  $A_{2A}$ -dopamine  $D_2$  receptor-receptor heteromerization: qualitative

- and quantitative assessment by fluorescence and bioluminescence energy transfer," *Journal of Biological Chemistry*, vol. 278, no. 47, pp. 46741–46749, 2003.
- [4] K. Fuxe, L. F. Agnati, K. Jacobsen et al., "Receptor heteromerization in adenosine A<sub>2A</sub> receptor signaling: relevance for striatal function and Parkinson's disease," *Neurology*, vol. 61, no. 11, pp. S19–S23, 2003.
  - [5] J. Hillion, M. Canals, M. Torvinen et al., "Coaggregation, coinertalization, and codesensitization of adenosine A<sub>2A</sub> receptors and dopamine D<sub>2</sub> receptors," *Journal of Biological Chemistry*, vol. 277, no. 20, pp. 18091–18097, 2002.
  - [6] J. L. Short, C. Ledent, E. Borrelli, J. Drago, and A. J. Lawrence, "Genetic interdependence of adenosine and dopamine receptors: evidence from receptor knockout mice," *Neuroscience*, vol. 139, no. 2, pp. 661–670, 2006.
  - [7] M. Morelli, T. Di Paolo, J. Wardas, F. Calon, D. Xiao, and M. A. Schwarzschild, "Role of adenosine A<sub>2A</sub> receptors in parkinsonian motor impairment and L-DOPA-induced motor complications," *Progress in Neurobiology*, vol. 83, no. 5, pp. 293–309, 2007.
  - [8] A. Pinna, S. Pontis, F. Borsini, and M. Morelli, "Adenosine A<sub>2A</sub> receptor antagonists improve deficits in initiation of movement and sensory motor integration in the unilateral 6-hydroxydopamine rat model of Parkinson's disease," *Synapse*, vol. 61, no. 8, pp. 606–614, 2007.
  - [9] R. A. Hauser and M. A. Schwarzschild, "Adenosine A<sub>2A</sub> receptor antagonists for Parkinson's disease rationale, therapeutic potential and clinical experience," *Drugs and Aging*, vol. 22, no. 6, pp. 471–482, 2005.
  - [10] T. H. Adair, "Growth regulation of the vascular system: an emerging role for adenosine," *American Journal of Physiology*, vol. 289, no. 2, pp. R283–R296, 2005.
  - [11] D. Lukashov, A. Ohta, S. Apasov, J. F. Chen, and M. Sitkovsky, "Cutting edge: physiologic attenuation of proinflammatory transcription by the Gs protein-coupled A<sub>2A</sub> adenosine receptor in vivo," *Journal of Immunology*, vol. 173, no. 1, pp. 21–24, 2004.
  - [12] F. Calon, M. Dridi, O. Hornykiewicz, P. J. Bédard, A. H. Rajput, and T. Di Paolo, "Increased adenosine A<sub>2A</sub> receptors in the brain of Parkinson's disease patients with dyskinesias," *Brain*, vol. 127, no. 5, pp. 1075–1084, 2004.
  - [13] A. Pinna, C. Corsi, A. R. Carta, V. Valentini, F. Pedata, and M. Morelli, "Modification of adenosine extracellular levels and adenosine A<sub>2A</sub> receptor mRNA by dopamine denervation," *European Journal of Pharmacology*, vol. 446, no. 1-3, pp. 75–82, 2002.
  - [14] M. Tomiyama, T. Kimura, T. Maeda, H. Tanaka, K. Kannari, and M. A. Baba, "Upregulation of Striatal Adenosine A<sub>2A</sub> Receptor mRNA in 6-Hydroxydopamine-Lesioned Rats Intermittently Treated with L-DOPA," *Synapse*, vol. 52, no. 3, pp. 218–222, 2004.
  - [15] D. J. Brooks, S. Papapetropoulos, F. Vandenhende et al., "An open-label, positron emission tomography study to assess adenosine A<sub>2A</sub> brain receptor occupancy of vipadenant (BIIB014) at steady-state levels in healthy male volunteers," *Clinical Neuropharmacology*, vol. 33, no. 2, pp. 55–60, 2010.
  - [16] M. Mishina, K. Ishiwata, M. Naganawa et al., "Adenosine A<sub>2A</sub> receptors measured with [11C]TMSX pet in the striata of parkinson's disease patients," *PLoS ONE*, vol. 6, no. 2, Article ID e17338, 2011.
  - [17] A. Pinna, S. Fenu, and M. Morelli, "Motor stimulant effects of the adenosine A<sub>2A</sub> receptor antagonist SCH 58261 do not develop tolerance after repeated treatment in 6-hydroxydopamine-lesioned rats," *Synapse*, vol. 39, no. 3, pp. 233–238, 2001.
  - [18] B. Henry, A. R. Crossman, and J. M. Brotchie, "Characterization of enhanced behavioral responses to L-DOPA following repeated administration in the 6-hydroxydopamine-lesioned rat model of Parkinson's disease," *Experimental Neurology*, vol. 151, no. 2, pp. 334–342, 1998.
  - [19] A. Pinna, S. Pontis, and M. Morelli, "Expression of dyskinetic movements and turning behaviour in subchronic L-DOPA 6-hydroxydopamine-treated rats is influenced by the testing environment," *Behavioural Brain Research*, vol. 171, no. 1, pp. 175–178, 2006.
  - [20] E. Tronci, N. Simola, F. Borsini et al., "Characterization of the antiparkinsonian effects of the new adenosine A<sub>2A</sub> receptor antagonist ST1535: acute and subchronic studies in rats," *European Journal of Pharmacology*, vol. 566, no. 1–3, pp. 94–102, 2007.
  - [21] M. Lundblad, E. Vaudano, and M. A. Cenci, "Cellular and behavioural effects of the adenosine A<sub>2A</sub> receptor antagonist KW-6002 in a rat model of L-DOPA-induced dyskinesia," *Journal of Neurochemistry*, vol. 84, no. 6, pp. 1398–1410, 2003.
  - [22] R. Grondin, P. J. Bédard, A. H. Tahar, L. Grégoire, A. Mori, and H. Kase, "Antiparkinsonian effect of a new selective adenosine A(2A) receptor antagonist in MPTP-treated monkeys," *Neurology*, vol. 52, no. 8, pp. 1673–1677, 1999.
  - [23] R. A. Hodgson, P. J. Bedard, G. B. Varty et al., "Preladenant, a selective A<sub>2A</sub> receptor antagonist, is active in primate models of movement disorders," *Experimental Neurology*, vol. 225, no. 2, pp. 384–390, 2010.
  - [24] T. Kanda, M. J. Jackson, L. A. Smith et al., "Combined use of the adenosine A<sub>2A</sub> antagonist KW-6002 with L-DOPA or with selective D<sub>1</sub> or D<sub>2</sub> dopamine agonists increases antiparkinsonian activity but not dyskinesia in MPTP-treated monkeys," *Experimental Neurology*, vol. 162, no. 2, pp. 321–327, 2000.
  - [25] F. Bibbiani, J. D. Oh, J. P. Petzer et al., "A<sub>2A</sub> antagonist prevents dopamine agonist-induced motor complications in animal models of Parkinson's disease," *Experimental Neurology*, vol. 184, no. 1, pp. 285–294, 2003.
  - [26] C. R. Gerfen, T. M. Engber, L. C. Mahan et al., "D<sub>1</sub> and D<sub>2</sub> dopamine receptor-regulated gene expression of striatonigral and striatopallidal neurons," *Science*, vol. 250, no. 4986, pp. 1429–1432, 1990.
  - [27] A. R. Carta, A. Pinna, O. Cauli, and M. Morelli, "Differential regulation of GAD67, enkephalin and dynorphin mRNAs by chronic-intermittent L-dopa and A<sub>2A</sub> receptor blockade plus L-dopa in dopamine-denervated rats," *Synapse*, vol. 44, no. 3, pp. 166–174, 2002.
  - [28] B. Y. Zeng, R. K. B. Pearce, G. M. MacKenzie, and P. Jenner, "Alterations in preproenkephalin and adenosine-2a receptor mRNA, but not preprotachykinin mRNA correlate with occurrence of dyskinesia in normal monkeys chronically treated with L-DOPA," *European Journal of Neuroscience*, vol. 12, no. 3, pp. 1096–1104, 2000.
  - [29] S. Ferré, B. B. Fredholm, M. Morelli, P. Popoli, and K. Fuxe, "Adenosine-dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia," *Trends in Neurosciences*, vol. 20, no. 10, pp. 482–487, 1997.
  - [30] D. L. Rosin, B. D. Hettinger, A. Lee, and J. Linden, "Anatomy of adenosine A<sub>2A</sub> receptors in brain: morphological substrates for integration of striatal function," *Neurology*, vol. 61, no. 11, pp. S12–S18, 2003.

- [31] F. Gardoni, V. Ghiglieri, M. D. Luca, and P. Calabresi, "Assemblies of glutamate receptor subunits with post-synaptic density proteins and their alterations in Parkinson's disease," *Progress in Brain Research*, vol. 183, no. C, pp. 169–182, 2010.
- [32] T. N. Chase, F. Bibbiani, W. Bara-Jimenez, T. Dimitrova, and J. D. Oh-Lee, "Translating A<sub>2A</sub> antagonist KW6002 from animal models to parkinsonian patients," *Neurology*, vol. 61, no. 11, pp. S107–S111, 2003.
- [33] K. Wirkner, Z. Gerevich, T. Krause et al., "Adenosine A<sub>2A</sub> receptor-induced inhibition of NMDA and GABA A receptor-mediated synaptic currents in a subpopulation of rat striatal neurons," *Neuropharmacology*, vol. 46, no. 7, pp. 994–1007, 2004.
- [34] P. Calabresi, F. Galletti, E. Saggese, V. Ghiglieri, and B. Picconi, "Neuronal networks and synaptic plasticity in Parkinson's disease: beyond motor deficits," *Parkinsonism and Related Disorders*, vol. 13, no. 3, pp. S259–S262, 2007.
- [35] A. E. Kelley, M. E. Andrzejewski, A. E. Baldwin, P. J. Hernandez, and W. E. Pratt, "Glutamate-mediated plasticity in corticostriatal networks: role in adaptive motor learning," *Annals of the New York Academy of Sciences*, vol. 1003, pp. 159–168, 2003.
- [36] S. Ferré, M. Karcz-Kubicha, B. T. Hope et al., "Synergistic interaction between adenosine A<sub>2A</sub> and glutamate mGlu5 receptors: implications for striatal neuronal function," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 18, pp. 11940–11945, 2002.
- [37] A. Kachroo, L. R. Orlando, D. K. Grandy, J. F. Chen, A. B. Young, and M. A. Schwarzschild, "Interactions between metabotropic glutamate 5 and adenosine A<sub>2A</sub> receptors in normal and parkinsonian mice," *Journal of Neuroscience*, vol. 25, no. 45, pp. 10414–10419, 2005.
- [38] S. Lopez, N. Turle-Lorenzo, T. H. Johnston et al., "Functional interaction between adenosine A<sub>2A</sub> and group III metabotropic glutamate receptors to reduce parkinsonian symptoms in rats," *Neuropharmacology*, vol. 55, no. 4, pp. 483–490, 2008.
- [39] T. H. Johnston, S. H. Fox, M. J. McIlldowie, M. J. Piggott, and J. M. Brotchie, "Reduction of L-DOPA-induced dyskinesia by the selective metabotropic glutamate receptor 5 antagonist 3-[(2-methyl-1,3-thiazol-4-yl)ethynyl]pyridine in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-lesioned macaque model of Parkinson's disease," *Journal of Pharmacology and Experimental Therapeutics*, vol. 333, no. 3, pp. 865–873, 2010.
- [40] P. Carriba, O. Ortiz, K. Patkar et al., "Striatal adenosine A<sub>2A</sub> and cannabinoid CB1 receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids," *Neuropsychopharmacology*, vol. 32, no. 11, pp. 2249–2259, 2007.
- [41] S. Ferré, C. Lluís, Z. Justinova et al., "Adenosine-cannabinoid receptor interactions. Implications for striatal function," *British Journal of Pharmacology*, vol. 160, no. 3, pp. 443–453, 2010.
- [42] S. Łukasiewicz, E. Błasiak, A. Faron-Górecka et al., "Fluorescence studies of homooligomerization of adenosine A<sub>2A</sub> and serotonin 5-HT<sub>1A</sub> receptors reveal the specificity of receptor interactions in the plasma membrane," *Pharmacological Reports*, vol. 59, no. 4, pp. 379–392, 2007.
- [43] F. Ciruela, V. Casadó, R. J. Rodrigues et al., "Presynaptic control of striatal glutamatergic neurotransmission by adenosine A<sub>1</sub>-A<sub>2A</sub> receptor heteromers," *Journal of Neuroscience*, vol. 26, no. 7, pp. 2080–2087, 2006.
- [44] P. Popoli, C. Frank, M. T. Tebano et al., "Modulation of glutamate release and excitotoxicity by adenosine A<sub>2A</sub> receptors," *Neurology*, vol. 61, no. 11, pp. S69–S71, 2003.
- [45] M. Mishina, K. Ishiwata, Y. Kimura et al., "Evaluation of distribution of adenosine A<sub>2A</sub> receptors in normal human brain measured with [11C]TMSX PET," *Synapse*, vol. 61, no. 9, pp. 778–784, 2007.
- [46] A. F. Ramlackhansingh, S. K. Bose, I. Ahmed, F. E. Turkheimer, N. Pavese, and D. J. Brooks, "Adenosine 2A receptor availability in dyskinetic and nondyskinetic patients with Parkinson disease," *Neurology*, vol. 76, no. 21, pp. 1811–1816, 2011.
- [47] K. J. Black, J. M. Koller, M. C. Campbell, D. A. Gusnard, and S. I. Bandak, "Quantification of indirect pathway inhibition by the adenosine A<sub>2A</sub> antagonist SYN115 in Parkinson disease," *Journal of Neuroscience*, vol. 30, no. 48, pp. 16284–16292, 2010.
- [48] K. Varani, F. Vincenzi, A. Tosi et al., "A<sub>2A</sub> adenosine receptor overexpression and functionality, as well as TNF- $\alpha$  levels, correlate with motor symptoms in Parkinson's disease," *FASEB Journal*, vol. 24, no. 2, pp. 587–598, 2010.
- [49] W. Bara-Jimenez, A. Sherzai, T. Dimitrova et al., "Adenosine A<sub>2A</sub> receptor antagonist treatment of Parkinson's disease," *Neurology*, vol. 61, no. 3, pp. 293–296, 2003.
- [50] R. A. Hauser, J. P. Hubble, and D. D. Truong, "Randomized trial of the adenosine A<sub>2A</sub> receptor antagonist istradefylline in advanced PD," *Neurology*, vol. 61, no. 3, pp. 297–303, 2003.
- [51] P. A. LeWitt, M. Guttman, J. W. Tetrud et al., "Adenosine A<sub>2A</sub> receptor antagonist istradefylline (KW-6002) reduces off time in Parkinson's disease: a double-blind, randomized, multicenter clinical trial (6002-US-005)," *Annals of Neurology*, vol. 63, no. 3, pp. 295–302, 2008.
- [52] M. Stacy, D. Silver, T. Mendis et al., "A 12-week, placebo-controlled study (6002-US-006) of istradefylline in Parkinson disease," *Neurology*, vol. 70, no. 23, pp. 2233–2240, 2008.
- [53] R. A. Hauser, L. M. Shulman, J. M. Trugman et al., "Study of istradefylline in patients with Parkinson's disease on levodopa with motor fluctuations," *Movement Disorders*, vol. 23, no. 15, pp. 2177–2185, 2008.
- [54] Y. Mizuno, K. Hasegawa, T. Kondo et al., "Clinical efficacy of istradefylline (KW-6002) in Parkinson's disease: a randomized, controlled study," *Movement Disorders*, vol. 25, no. 10, pp. 1437–1443, 2010.
- [55] W. Knebel, N. Rao, T. Uchimura et al., "Population pharmacokinetic analysis of istradefylline in healthy subjects and in patients with parkinson's disease," *Journal of Clinical Pharmacology*, vol. 51, no. 1, pp. 40–52, 2011.
- [56] R. A. Hauser, M. Cantillon, E. Pourcher et al., "Preladenant in patients with Parkinson's disease and motor fluctuations: a phase 2, double-blind, randomised trial," *The Lancet Neurology*, vol. 10, no. 3, pp. 221–229, 2011.

## Review Article

# Role of Serotonin Neurons in L-DOPA- and Graft-Induced Dyskinesia in a Rat Model of Parkinson's Disease

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L-DOPA, the most effective drug to treat motor symptoms of Parkinson's disease, causes abnormal involuntary movements, limiting its use in advanced stages of the disease. An increasing body of evidence points to the serotonin system as a key player in the appearance of L-DOPA-induced dyskinesia (LID). In fact, exogenously administered L-DOPA can be taken up by serotonin neurons, converted to dopamine and released as a false transmitter, contributing to pulsatile stimulation of striatal dopamine receptors. Accordingly, destruction of serotonin fibers or silencing serotonin neurons by serotonin agonists could counteract LID in animal models. Recent clinical work has also shown that serotonin neurons are present in the caudate/putamen of patients grafted with embryonic ventral mesencephalic cells, producing intense serotonin hyperinnervation. These patients experience graft-induced dyskinesia (GID), a type of dyskinesia phenotypically similar to the one induced by L-DOPA but independent from its administration. Interestingly, the 5-HT<sub>1A</sub> receptor agonist buspirone has been shown to suppress GID in these patients, suggesting that serotonin neurons might be involved in the etiology of GID as for LID. In this paper we will discuss the experimental and clinical evidence supporting the involvement of the serotonin system in both LID and GID.

## 1. Introduction

Parkinson's disease (PD) is the second most common neurodegenerative disease and is characterized by loss of dopamine (DA) neurons in the substantia nigra. The cell loss results in decreased activation of striatal DA receptors, thus causing motor impairments, such as tremor, rigidity, bradykinesia, and postural instability. The DA precursor L-3,4-dihydroxyphenylalanine (L-DOPA) represents the most effective drug to alleviate the motor symptoms. Although this medication is very efficient during the first few years of administration, its efficacy gradually diminishes overtime, and uncontrolled excessive movements, known as dyskinesia, appear as a side effect after a variable number of years in most of patients, limiting the use of L-DOPA in advanced stages of the disease.

A better understanding of the mechanisms underlying the appearance of dyskinesia has been achieved during recent years using animal models of L-DOPA-induced dyskinesia (LID). In fact, abnormal involuntary movements (AIMs)

develop in response to sub-chronic L-DOPA treatment in 6-hydroxydopamine (6-OHDA)-lesioned rats and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated monkeys, resembling peak-dose dyskinesia seen in patients [1–4]. Using these models, a number of alterations have been identified at the level of striatal neurons of dyskinetic subjects, such as abnormal trafficking of DA D<sub>1</sub> and N-methyl-D-aspartate (NMDA) receptors [5, 6], leading to alterations in key striatal signaling pathways.

More recently, the serotonin system has emerged as a putative player in the induction of fluctuations in synaptic DA levels following administration of L-DOPA in animal models of PD, which cause pulsatile stimulation of DA receptors and promote the maladaptive changes that characterize the parkinsonian dyskinetic brain [7–11]. The emerging role of serotonin neurons in LID has also prompted researchers to investigate a possible involvement of these neurons in the appearance of *off-drug* dyskinesia that has emerged in a subset of PD patients following transplantation of fetal

ventral mesencephalic DA neuroblasts, namely, graft-induced dyskinesia (GID) [12, 13]. In this paper, we will discuss the recent experimental and clinical evidence supporting a role of serotonin neurons both in LID and GID.

## 2. The Role of Serotonin Neurons in the Induction of L-DOPA-Induced Dyskinesia

Progression of DA neuron degeneration represents the first risk factor for development of LID in patients. In fact, the efficacy of L-DOPA in providing its therapeutic effect during the first years of administration is conceivably due to the ability of spared DA neurons to take up exogenously administered L-DOPA, convert it to DA, and release it into the synaptic cleft, but also to regulate synaptic DA levels via the D<sub>2</sub> autoreceptor and DA transporter (DAT). When the disease is in its early stage, sufficient DA terminals remain in the striatum to efficiently mediate a feedback-controlled mechanism of release. The ability of the spared DA terminals to prevent development of AIMs upon L-DOPA treatment is well demonstrated in a recent report by Ulusoy et al. [14]. In this study, a significant DA deficiency was established in rats by viral vector delivery of short hairpin RNA for the tyrosine hydroxylase (TH) enzyme, without affecting cell survival. Animals were then made dyskinetic by subchronic treatment with the direct DA agonist apomorphine; however, when treated with L-DOPA, the same rats appeared to be fully resistant to the induction of LID, despite apomorphine treatment had already promoted postsynaptic alterations, such as increased striatal FosB expression.

The ability of the presynaptic DA compartment to prevent excessive DA receptor stimulation, and aberrant downstream signaling, even in presence of supersensitive striatal DA receptors is also confirmed in transplantation studies. In fact, ventral mesencephalic DA graft, which reconstitutes the presynaptic buffering capacity into the lesioned and maladapted striatum, normalizes the response of L-DOPA-primed dyskinetic rats to L-DOPA administration [7]. In light with these results, it is conceivable to think that the efficacy of L-DOPA during the first years of administration is also due to the presence of a sufficient number of DA neurons that can buffer the exogenously administered L-DOPA, and provide a source of regulated DA release. However, with the progression of DA degeneration, this buffering capacity, and thus the ability to provide physiological level of DA receptor stimulation, is progressively lost. In this situation, the serotonin neurons come to play a major role in L-DOPA-derived DA production and release, as they possess both the aromatic amino acid decarboxylase enzyme (AADC) and the vesicular monoamine transporter (VMAT). Unlike DA neurons, though, serotonin neurons cannot regulate the extracellular levels of DA due to the lack of the autoregulatory loop, hence, causing un-controlled DA release following L-DOPA administration. DA released from serotonin neurons will therefore act in concert with the intermittent nature of the orally administered L-DOPA to cause pulsatile stimulation of DA receptors, and thus, changes in downstream signaling pathways at striatal neurons. In support of this view, removal of serotonin innervation by toxin lesion was

shown to produce about 80% reduction in L-DOPA-derived striatal extracellular DA levels [15], and to induce a near-to-complete suppression of LID in dyskinetic rats [8, 16]. Accordingly, pharmacological silencing of serotonin neuron activity by 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptor agonists has been shown to reduce L-DOPA-derived extracellular DA levels [17], and to suppress LID in rats [8] as well as in MPTP-treated monkeys [18]. In addition, chronic administration of the 5-HT<sub>1</sub> agonists was able to prevent the development of dyskinesia and upregulation of FosB expression in the striatum of 6-OHDA-lesioned rats [18], thus linking dysregulated DA release from serotonin terminals with the induction of a well-known striatal marker of dyskinesia. Interestingly, simultaneous activation of 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptors was found to trigger a potent synergistic effect in the suppression of LID in both rats and monkeys [8, 18]. In fact, LID was nearly fully abolished at doses of combined agonists that were ineffective when given individually. This finding has now led to the initiation of a first double-blind, proof-of-concept clinical trial employing a mixed 5-HT<sub>1A/1B</sub> receptor agonist in dyskinetic patients.

In confirming the interaction between exogenously administered L-DOPA and serotonin neurons, Navailles and coworkers have recently demonstrated that serotonin neuron-dependent DA release takes place, upon chronic L-DOPA treatment in 6-OHDA-lesioned rats, not only in the striatum but also in other brain areas receiving sufficient serotonin innervation, such as substantia nigra, hippocampus, and prefrontal cortex [19]. Moreover, L-DOPA administration appears to result in reduced striatal serotonin tissue content in 6-OHDA-lesioned rats [8]. The latter result supports the existence of a competition between serotonin and DA for serotonergic vesicles, which causes serotonin depletion. Thus, an increasing body of experimental evidence points to the serotonin system as a key player in the appearance of LID.

Progressive reduction of L-DOPA-derived extracellular DA levels upon chronic L-DOPA treatment has been recently found in 6-OHDA-lesioned rats [20, 21], leading some researcher to question the role of serotonin neurons in the appearance of LID [21]. Nevertheless, we believe that the potent inhibitory effect of 5,7-dihydroxytryptamine (5,7-DHT) lesion on both development and expression of dyskinesia in 6-OHDA-lesioned rats [8], together with the striking suppression of LID induced by low doses of 5-HT<sub>1A</sub> + 5-HT<sub>1B</sub> receptor agonists both in rats and macaques [18] provided unquestionable evidence supporting an important role of serotonin neurons, at least in animal models of PD. It should be taken into account that during chronic administration of L-DOPA, postsynaptic DA receptors become supersensitive; thus, the dyskinetic response to L-DOPA might be maintained by lower extracellular DA levels once postsynaptic alterations have been already induced. The relevance for the human disease of the progressive reduction of extracellular DA levels seen upon chronic L-DOPA in the rat 6-OHDA model remains to be established, as L-DOPA-derived synaptic DA levels were shown to increase with progression of the disease in a positron emission tomography (PET) imaging study in PD patients [22].

Interestingly, Rylander and coworkers have recently shown marked serotonin hyperinnervation in the lesioned striatum of parkinsonian dyskinetic subjects across different species, including patients, thus raising the possibility that serotonin neurons play a relevant role in the emergence of LID also in the patients [10, 11, 23]. This study suggested that L-DOPA treatment may be able to provoke sprouting of serotonin axon terminals and change their morphology, hence, possibly enhancing the fluctuations in extracellular DA concentration, consistent with findings of de la Fuente-Fernandez and coworkers in their PET imaging study [22, 24].

In further experimental support for a detrimental effect of serotonin neurons on LID, grafted serotonin neurons, which induced an intense hyperinnervation of the grafted striatum, exacerbated LID in both partial and complete DA lesioned rats [7, 25], raising the possibility that inclusion of these cells in the grafted tissue may have detrimental effects on LID in grafted PD patients.

### 3. The Role of Serotonin Neurons in the Modulation of Graft-Induced Dyskinesia

Transplantation of fetal ventral mesencephalic neurons is a therapeutic approach to PD that has already been tested in clinical trials [26–30]. While the variability in the outcome of these studies halted further investigations, the presence of highly responsive patients provided proof-of-concept that this therapeutic intervention can be significantly beneficial. Indeed, there is now general agreement that the reasons accounting for the observed variability rely on the lack of standardization of the cell preparations, surgical procedures, as well as on the selection of patients and presence or absence of postsurgical immunosuppressive treatment [31]. However, another element that has contributed to raise concern about fetal transplantation is the appearance in a subset of grafted patients of *off-drug* dyskinesia [29, 31–33], a form of involuntary movements that is phenotypically similar to the one induced by L-DOPA but independent from its administration. The recent findings on the role of serotonin neurons in the induction of LID has led to hypothesize that serotonin neurons included in the graft may also play a role in GID [8, 16, 18].

In fact, serotonin neuroblasts are located in close vicinity of the DA ones in the fetal ventral mesencephalic area that is dissected for transplantation. Accordingly, about 50% of grafted cells were found to be serotonin neurons in a post-mortem analysis of the caudate-putamen of grafted patients [34].

In possible support of this hypothesis, in a recent PET study, Politis and colleagues have found intense serotonergic hyperinnervation in the striatum of grafted patients showing GID [12, 13]. Interestingly, administration of the partial 5-HT<sub>1A</sub> agonist buspirone suppressed GID in all tested patients. An involvement of the serotonin neurons in GID is further supported by the high serotonin transporter (SERT)/DAT ratio found in one GID patient compared to both healthy age-matched control and non-grafted PD patients [12]. Thus, it is postulated that serotonin terminals may take

up DA released by the graft through SERT, and release DA, as a false transmitter, away from the uptake site, in striatal regions lacking sufficient DA innervation, thus, leading to activation of supersensitive DA receptors. However, it should be acknowledged that these clinical observations have been made in a very few subjects, and further evidence should be provided. In particular, it would be important to investigate the state of the striatal serotonin innervation also in patients free of GID.

Although spontaneous GID, that is dyskinesia in the absence of any drug treatment, is inconsistent in grafted rodents [35, 36], it does appear after administration of a low dose of amphetamine [35, 37], which is known to evoke massive DA release from grafted DA neurons [38]. These abnormal movements can be scored with the same scale used for LID [35, 37], and are now widely used as a convenient and reproducible model of GID [7, 39–42]. While appearance of GID in this rat model is clearly dependent on the presence of an adequate number of DA neurons in the graft, we have recently found a bidirectional modulatory effect of endogenous serotonin neurons on GID. In fact, reduction of serotonin neuron activity, by a combination of 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptor agonists, produced a significant reduction of GID, while increased serotonin neuron release by fenfluramine exacerbated GID [43]. Strikingly, administration of a low dose of buspirone (1 mg/kg) completely suppressed GID, as seen in grafted patients. Interestingly, removal of the endogenous serotonin innervation by specific toxin lesions appeared to abolish the anti-GID effect of the selective 5-HT<sub>1</sub> agonists, both in serotonin-containing and serotonin-free grafts, suggesting that the modulatory effect on GID may be due to the endogenous rather than the graft-derived serotonin neurons, at least in our experimental conditions. By contrast, neither removal of the endogenous serotonin innervation nor pretreatment with the selective 5-HT<sub>1A</sub> antagonist WAY100135 reduced the anti-GID efficacy of buspirone. In fact, buspirone is also known to possess antagonistic properties on D<sub>2</sub> receptors [44–47]. In support for a D<sub>2</sub>-mediated effect of buspirone, similar anti-GID effect was induced by a low dose of the selective D<sub>2</sub> receptor antagonist eticlopride (0.03 mg/kg). Thus, our data support a modulatory role of the endogenous serotonin neurons on expression of GID, as well as a peculiar role of D<sub>2</sub> receptors. Indeed, both buspirone and eticlopride were ineffective against LID at doses fully suppressing GID [43].

Further work is required to understand whether inclusion of serotonin neurons in the graft can be detrimental for appearance of GID, although current experimental data do not support this hypothesis.

### 4. Conclusion

Overall, loss of DA in basal ganglia circuits and DA replacement by chronic L-DOPA administration result in complex alterations in the parkinsonian brain, that affect several systems and key signaling proteins, most of which remain poorly understood. DA released as a false transmitter from serotonin neurons appears to play a key role in initiating these events, at least in animal models. Serotonin neurons,

therefore, represent an intriguing pharmacological target to treat already established LID and/or to prevent the events leading to the appearance of LID from taking place. Recent evidence suggests that serotonin neurons may also participate to the induction of dyskinesia seen in the *off-state* in grafted PD patients. The upcoming new clinical trial, funded by the European Community, employing fetal ventral mesencephalic cells will answer the question whether optimization of the surgical procedures and preparation of the grafted material, including exclusion of serotonin neuroblasts, will improve clinical outcome and avoid appearance of GID.

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## References

- [1] E. Bézard, S. Ferry, U. Mach et al., "Attenuation of levodopa-induced dyskinesia by normalizing dopamine D3 receptor function," *Nature Medicine*, vol. 9, no. 6, pp. 762–767, 2003.
- [2] M. P. Hill, E. Bézard, S. G. McGuire et al., "Novel antiepileptic drug levetiracetam decreases dyskinesia elicited by L-dopa and ropinirole in the MPTP-lesioned marmoset," *Movement Disorders*, vol. 18, no. 11, pp. 1301–1305, 2003.
- [3] A. Hsu, D. M. Togasaki, E. Bézard et al., "Effect of the D3 dopamine receptor partial agonist BP897 [N-[4-(4-(2-methoxyphenyl)piperazinyl)butyl]-2-naphthamide] on L-3,4-dihydroxyphenylalanine-induced dyskinesias and parkinsonism in squirrel monkeys," *Journal of Pharmacology and Experimental Therapeutics*, vol. 311, no. 2, pp. 770–777, 2004.
- [4] M. Lundblad, M. Andersson, C. Winkler, D. Kirik, N. Wierup, and M. A. Cenci, "Pharmacological validation of behavioural measures of akinesia and dyskinesia in a rat model of Parkinson's disease," *European Journal of Neuroscience*, vol. 15, no. 1, pp. 120–132, 2002.
- [5] A. Berthet, G. Porras, E. Doudnikoff et al., "Pharmacological analysis demonstrates dramatic alteration of D<sub>1</sub> dopamine receptor neuronal distribution in the rat analog of L-DOPA-induced dyskinesia," *Journal of Neuroscience*, vol. 29, no. 15, pp. 4829–4835, 2009.
- [6] C. Guigoni, E. Doudnikoff, Q. Li, B. Bloch, and E. Bézard, "Altered D<sub>1</sub> dopamine receptor trafficking in parkinsonian and dyskinetic non-human primates," *Neurobiology of Disease*, vol. 26, no. 2, pp. 452–463, 2007.
- [7] T. Carlsson, M. Carta, C. Winkler, A. Björklund, and D. Kirik, "Serotonin neuron transplants exacerbate L-DOPA-induced dyskinesias in a rat model of Parkinson's disease," *Journal of Neuroscience*, vol. 27, no. 30, pp. 8011–8022, 2007.
- [8] M. Carta, T. Carlsson, D. Kirik, and A. Björklund, "Dopamine released from 5-HT terminals is the cause of L-DOPA-induced dyskinesia in parkinsonian rats," *Brain*, vol. 130, no. 7, pp. 1819–1833, 2007.
- [9] A. Muñoz, T. Carlsson, E. Tronci, D. Kirik, A. Björklund, and M. Carta, "Serotonin neuron-dependent and -independent reduction of dyskinesia by 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptor agonists in the rat Parkinson model," *Experimental Neurology*, vol. 219, no. 1, pp. 298–307, 2009.
- [10] D. Rylander, M. Parent, S. S. O-Sullivan et al., "Maladaptive plasticity of serotonin axon terminals in levodopa-induced dyskinesia," *Annals of Neurology*, vol. 68, no. 5, pp. 619–628, 2010.
- [11] B. Y. Zeng, M. M. Iravani, M. J. Jackson, S. Rose, A. Parent, and P. Jenner, "Morphological changes in serotonergic neurites in the striatum and globus pallidus in levodopa primed MPTP treated common marmosets with dyskinesia," *Neurobiology of Disease*, vol. 40, no. 3, pp. 599–607, 2010.
- [12] M. Politis, W. H. Oertel, K. Wu et al., "Graft-induced dyskinesias in Parkinson's disease: high striatal serotonin/dopamine transporter ratio," *Movement Disorders*, vol. 26, no. 11, pp. 1997–2003, 2011.
- [13] M. Politis, K. Wu, C. Loane et al., "Serotonergic neurons mediate dyskinesia side effects in Parkinson's patients with neural transplants," *Science Translational Medicine*, vol. 2, no. 38, Article ID 38ra46, 2010.
- [14] A. Ulusoy, G. Sahin, and D. Kirik, "Presynaptic dopaminergic compartment determines the susceptibility to L-DOPA-induced dyskinesia in rats," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 29, pp. 13159–13164, 2010.
- [15] H. Tanaka, K. Kannari, T. Maeda, M. Tomiyama, T. Suda, and M. Matsunaga, "Role of serotonergic neuron in L-DOPA-derived extracellular dopamine in the striatum of 6-OHDA-lesioned rats," *NeuroReport*, vol. 10, no. 3, pp. 631–634, 1999.
- [16] K. L. Eskow, K. B. Dupre, C. J. Barnum, S. O. Dickinson, J. Y. Park, and C. Bishop, "The role of the dorsal raphe nucleus in the development, expression, and treatment of L-dopa-induced dyskinesia in hemiparkinsonian rats," *Synapse*, vol. 63, no. 7, pp. 610–620, 2009.
- [17] H. S. Lindgren, D. R. Andersson, S. Lagerkvist, H. Nissbrandt, and M. A. Cenci, "L-DOPA-induced dopamine efflux in the striatum and the substantia nigra in a rat model of Parkinson's disease: temporal and quantitative relationship to the expression of dyskinesia," *Journal of Neurochemistry*, vol. 112, no. 6, pp. 1465–1476, 2010.
- [18] A. Muñoz, Q. Li, F. Gardoni et al., "Combined 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptor agonists for the treatment of L-DOPA-induced dyskinesia," *Brain*, vol. 131, no. 12, pp. 3380–3394, 2008.
- [19] S. Navailles, A. Benazzouz, B. Bioulac, C. Gross, and P. De Deurwaerdère, "High-frequency stimulation of the subthalamic nucleus and L-3,4-dihydroxyphenylalanine inhibit *in vivo* serotonin release in the prefrontal cortex and hippocampus in a rat model of Parkinson's disease," *Journal of Neuroscience*, vol. 30, no. 6, pp. 2356–2364, 2010.
- [20] S. Navailles, B. Bioulac, C. Gross, and P. De Deurwaerdère, "Chronic L-DOPA therapy alters central serotonergic function and L-DOPA-induced dopamine release in a region-dependent manner in a rat model of Parkinson's disease," *Neurobiology of Disease*, vol. 41, no. 2, pp. 585–590, 2011.
- [21] N. Nevalainen, S. Af Bjerken, M. Lundblad, G. A. Gerhardt, and I. Stromberg, "Dopamine release from serotonergic nerve fibers is reduced in L-DOPA-induced dyskinesia," *Journal of Neurochemistry*, vol. 118, pp. 12–23, 2011.
- [22] R. de La Fuente-Fernández, V. Sossi, Z. Huang et al., "Levodopa-induced changes in synaptic dopamine levels increase with progression of Parkinson's disease: implications for dyskinesias," *Brain*, vol. 127, no. 12, pp. 2747–2754, 2004.
- [23] S. Gil, C. Park, J. Lee, and H. Koh, "The roles of striatal serotonin and l-amino-acid decarboxylase on l-DOPA-induced dyskinesia in a hemiparkinsonian rat model," *Cellular and Molecular Neurobiology*, vol. 30, no. 6, pp. 817–825, 2010.

- [24] R. de La Fuente-Fernández, M. Schulzer, E. Mak, D. B. Calne, and A. J. Stoessl, "Presynaptic mechanisms of motor fluctuations in Parkinson's disease: a probabilistic model," *Brain*, vol. 127, no. 4, pp. 888–899, 2004.
- [25] T. Carlsson, M. Carta, A. Muñoz et al., "Impact of grafted serotonin and dopamine neurons on development of L-DOPA-induced dyskinesias in parkinsonian rats is determined by the extent of dopamine neuron degeneration," *Brain*, vol. 132, no. 2, pp. 319–335, 2009.
- [26] C. R. Freed, R. E. Breeze, N. L. Rosenberg et al., "Survival of implanted fetal dopamine cells and neurologic improvement 12 to 46 months after transplantation for Parkinson's disease," *The New England Journal of Medicine*, vol. 327, no. 22, pp. 1549–1555, 1992.
- [27] O. Lindvall, G. Sawle, H. Widner et al., "Evidence for long-term survival and function of dopaminergic grafts in progressive Parkinson's disease," *Annals of Neurology*, vol. 35, no. 2, pp. 172–180, 1994.
- [28] O. Lindvall, H. Widner, S. Rehncrona et al., "Transplantation of fetal dopamine neurons in Parkinson's disease: one-year clinical and neurophysiological observations in two patients with putaminal implants," *Annals of Neurology*, vol. 31, no. 2, pp. 155–165, 1992.
- [29] C. W. Olanow, C. G. Goetz, J. H. Kordower et al., "A double-blind controlled trial of bilateral fetal nigral transplantation in Parkinson's disease," *Annals of Neurology*, vol. 54, no. 3, pp. 403–414, 2003.
- [30] P. Piccini, D. J. Brooks, A. Björklund et al., "Dopamine release from nigral transplants visualized *in vivo* in a Parkinson's patient," *Nature Neuroscience*, vol. 2, no. 12, pp. 1137–1140, 1999.
- [31] E. L. Lane, A. Björklund, S. B. Dunnett, and C. Winkler, "Neural grafting in Parkinson's disease. Unraveling the mechanisms underlying graft-induced dyskinesia," *Progress in Brain Research*, vol. 184, pp. 295–309, 2010.
- [32] C. R. Freed, P. E. Greene, R. E. Breeze et al., "Transplantation of embryonic dopamine neurons for severe Parkinson's disease," *The New England Journal of Medicine*, vol. 344, no. 10, pp. 710–719, 2001.
- [33] P. Hagell, P. Piccini, A. Björklund et al., "Dyskinesias following neural transplantation in Parkinson's disease," *Nature Neuroscience*, vol. 5, no. 7, pp. 627–628, 2002.
- [34] I. Mendez, A. Viñuela, A. Astradsson et al., "Dopamine neurons implanted into people with Parkinson's disease survive without pathology for 14 years," *Nature Medicine*, vol. 14, no. 5, pp. 507–509, 2008.
- [35] E. L. Lane, C. Winkler, P. Brundin, and M. A. Cenci, "The impact of graft size on the development of dyskinesia following intrastriatal grafting of embryonic dopamine neurons in the rat," *Neurobiology of Disease*, vol. 22, no. 2, pp. 334–345, 2006.
- [36] A. Viñuela, P. J. Hallett, C. Reske-Nielsen et al., "Implanted reuptake-deficient or wild-type dopaminergic neurons improve on L-dopa dyskinesias without OFF-dyskinesias in a rat model of Parkinson's disease," *Brain*, vol. 131, no. 12, pp. 3361–3379, 2008.
- [37] T. Carlsson, C. Winkler, M. Lundblad, M. A. Cenci, A. Björklund, and D. Kirik, "Graft placement and uneven pattern of reinnervation in the striatum is important for development of graft-induced dyskinesia," *Neurobiology of Disease*, vol. 21, no. 3, pp. 657–668, 2006.
- [38] T. Zetterstrom, M. Herrera-Marschitz, and U. Ungerstedt, "Simultaneous measurement of dopamine release and rotational behaviour in 6-hydroxydopamine denervated rats using intracerebral dialysis," *Brain Research*, vol. 376, no. 1, pp. 1–7, 1986.
- [39] E. L. Lane, L. Vercammen, M. A. Cenci, and P. Brundin, "Priming for L-DOPA-induced abnormal involuntary movements increases the severity of amphetamine-induced dyskinesia in grafted rats," *Experimental Neurology*, vol. 219, no. 1, pp. 355–358, 2009.
- [40] E. L. Lane, D. Soulet, L. Vercammen, M. A. Cenci, and P. Brundin, "Neuroinflammation in the generation of post-transplantation dyskinesia in Parkinson's disease," *Neurobiology of Disease*, vol. 32, no. 2, pp. 220–228, 2008.
- [41] E. L. Lane, P. Brundin, and M. A. Cenci, "Amphetamine-induced abnormal movements occur independently of both transplant- and host-derived serotonin innervation following neural grafting in a rat model of Parkinson's disease," *Neurobiology of Disease*, vol. 35, no. 1, pp. 42–51, 2009.
- [42] J. Garcia, T. Carlsson, M. Döbrössy, G. Nikkiah, and C. Winkler, "Extent of pre-operative L-DOPA-induced dyskinesia predicts the severity of graft-induced dyskinesia after fetal dopamine cell transplantation," *Experimental Neurology*, vol. 232, pp. 270–279, 2011.
- [43] E. Shin, J. Garcia, C. Winkler, A. Björklund, and M. Carta, "Serotonergic and dopaminergic mechanisms in graft-induced dyskinesia in a rat model of Parkinson's disease," *Neurobiology of Disease*. In press.
- [44] A. S. Eison and D. L. Temple, "Buspirone: review of its pharmacology and current perspectives on its mechanism of action," *American Journal of Medicine*, vol. 80, no. 3, pp. 1–9, 1986.
- [45] B. A. McMillen, R. T. Matthews, and M. K. Sanghera, "Dopamine receptor antagonism by the novel anti-anxiety drug, buspirone," *Journal of Neuroscience*, vol. 3, no. 4, pp. 733–738, 1983.
- [46] H. J. Rijnders and J. L. Slangen, "The discriminative stimulus properties of buspirone involve dopamine-2 receptor antagonist activity," *Psychopharmacology*, vol. 111, no. 1, pp. 55–61, 1993.
- [47] J. Scuvee-Moreau, I. Giesbers, and A. Dresse, "Electrophysiological and microiontophoretic studies with buspirone: influence on the firing rate of central monoaminergic neurons and their responsiveness to dopamine, clonidine or GABA," *Archives Internationales de Physiologie et de Biochimie*, vol. 95, no. 5, pp. 439–446, 1987.

## Review Article

# Clinical Aspects and Management of Levodopa-Induced Dyskinesia

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In Parkinson's disease, one of the most troublesome dilemmas is the treatment of levodopa-induced dyskinesia. After a few years, chronic treatment with levodopa is associated with the development of dyskinesias. Strategies to delay or to reduce dyskinesias are based on the change of levodopa dosing or the early use of dopamine agonists. Dopamine agonists with different pharmacological profile are available. Our paper was aimed to analyse the clinical impact and the management of dyskinesias with dopamine agonists.

## 1. Introduction

Four decades after its introduction, levodopa remains the most effective agent to improve motor symptoms in PD, but chronic use is associated with the emergence of motor fluctuations, defined as a loss of clinical benefit before the next levodopa dose (wearing off), abnormal involuntary movements (dystonia, chorea, and athetosis—collectively referred to as dyskinesia) [1, 2], and nonmotor complications, as behavioural and cognitive changes [3]. Levodopa is initially well tolerated in most of the cases and allows a substantial improvement of motor performances despite its erratic pharmacokinetics [1, 4]. With the disease progression, therapeutic window of levodopa narrows, and the duration of each dose shortens. Motor fluctuations usually precede dyskinesias [5], and it has been observed that the development of one is a risk factor for the development of the other [5].

Although more commonly associated with levodopa, dyskinesias can also occur with dopamine agonist monotherapy [6–8]. The development of dyskinesia in some patients treated with dopamine agonists that have relatively long half-lives (ropinirole, 6 h; pramipexole, 8 h) or very long half-lives (cabergoline, 68 h) suggests that, to some extent, even

dopamine stimulation provided in a continuous fashion can cause dyskinesias.

## 2. Epidemiology and Clinical Aspects of Motor Complications

The three most important risk factors positively associated with increased occurrence of dyskinesias are younger age at disease onset [9, 10], longer disease duration [11, 12], and longer duration of pulsatile dopaminergic treatment (typically, levodopa) [13, 14]. The first two factors are interrelated and almost all patients with early-onset PD [15] develop dyskinesias, whereas they are less frequent in patients with late-onset PD [16]. PD patients with early disease onset have a high probability to carry mutations for monogenic PD forms, and therefore, early onset and genetic predisposition are two overlapping and possibly interrelated risk factors. Other risk factors associated with increased risk of dyskinesias are female gender [17, 18] and the occurrence of specific polymorphisms for dopamine receptors or dopamine transporters [19–21].

Dyskinesias more commonly appear as choreiform, but in some cases, they may resemble dystonia, myoclonus, or

other movement disorders. Peak-dose dyskinesias are the most common type of dyskinesia. They occur during peaks of levodopa-derived dopamine in the brain, when the patient is otherwise experiencing a beneficial response (the “on” state). Peak-dose dyskinesias worsen with increases in dopaminergic dose and lessen with its reductions. In certain cases, dyskinesias seem to appear with a more particular pattern, as dyskinesia-improvement-dyskinesia. This is termed diphasic dyskinesia, and it tends to occur when levodopa-derived dopamine concentrations are increasing or decreasing, whereas the clinical condition of the patient turns “on” and “off” [22]. Diphasic dyskinesias are typically displayed with large-amplitude stereotypic, rhythmic, and repetitive movements, more often of the legs, that may be associated with Parkinsonian features in other body regions. In extreme cases, patients treated with levodopa can cycle between “on” periods, which are complicated by disabling dyskinesias, and “off” periods in which Parkinsonism is uncontrolled and the patient is akinetic and frozen.

Motor complications occur in about 50% of patients with PD who have been in therapy with levodopa for more than 5 years, and in almost 100% of patients with young-onset disease [23, 24]. Achieving an acceptable clinical control once these motor fluctuations have appeared is usually a relatively simple matter, nearing together the levodopa doses or adding medications that reduce “off” time. However, when a patient develops peak-dose dyskinesias too, it becomes difficult to smooth the clinical response. Although for many patients, dyskinesias are not disabling, they create a barrier to adequate treatment of fluctuations and Parkinsonian symptoms.

### 3. Pathophysiology of Dyskinesias

A primary condition in LID pathophysiology is the presence of dopaminergic cell loss in substantia nigra [25–27]. The nonappearance of dyskinesia in normal humans chronically treated with levodopa (i.e., mistaken diagnosis [28]) and its rapid emergence in PD patients either with late diagnosis or a young onset, where denervation is high at diagnosis [15, 29, 30], heavily support this theory. Moreover, the progression of nigral denervation seems to be closely related with the lowering of the dyskinesia onset threshold in MPTP-exposed primates [31]. Nonetheless, denervation cannot be the unique factor responsible for dyskinesia, whereas not all patients with advanced illness and extensive nigral denervation develop dyskinesia when treated with levodopa [32, 33]. Thus, a chronic dopaminergic stimulation on a denervated substantia nigra induces a process of sensitisation such that each following administration modifies the response to subsequent dopaminergic treatments. This process, called priming, increases over time of treatment the chance of eliciting dyskinesias and, once dyskinesias have been established, their severity. The priming process, which is responsible for the insidious evolution of dyskinesias over time of treatment, is associated with changes in receptors for dopamine or other neurotransmitters [34, 35]. A crucial role has been postulated for both dopamine receptors and NMDA glutamate receptors in the induction of priming; this mechanism could

be regarded as an increased responsiveness of postsynaptic striatal dopamine receptors (mainly D<sub>1</sub>-like), which are activated in conjunction with glutamatergic inputs [1]. Dyskinesias are probably generated by a persistent enhancement of the responsiveness of striatal medium-sized spiny neurones to dopaminergic treatment. This is an aftermath of dopamine depletion and is associated with overexpression of specific components of the signal transduction machinery. If protracted, this condition may ultimately lead to long-term changes in gene expression, which will permanently affect the function of striatal medium spiny neurones [36]. Following priming, the development of dyskinesias largely depends on two additional factors, the pulsatile administration of levodopa (or another short-acting dopaminergic agent) and the severity of dopaminergic denervation in the striatum. The latter plays an important role in setting the threshold required in developing dyskinesias [37]. A direct relationship between the severity of striatal denervation and the time required to develop dyskinesias has been demonstrated in PD patients [38] and has been indirectly confirmed by the finding that patients with dopa-responsive dystonia, who have Parkinsonism without nigrostriatal denervation, uncommonly develop dyskinesias [39].

In early PD patients, levodopa-derived dopamine is packaged into synaptic vesicles by vesicular monoamine transporter 2 (VMAT-2), stored, and released in both tonic and phasic bursts in response to impulse flow [40, 41], in order to preserve dopamine receptors from levodopa plasma concentration fluctuations and, therefore, to maintain physiological dopaminergic transmission [42, 43]. With the progression of the disease, and the striatal dopaminergic cell loss, the formation of dopamine from levodopa and its storage capacity are increasingly compromised, and the response to levodopa becomes dominated by its pharmacokinetic characteristics and general bioavailability [4]. Thus, in advanced PD, peak concentrations of drug in plasma become coincidental with the expression of dyskinesia. As observed in animal models, the continuous release of dopamine leads to improvements in motor function and, together, to a marked reduction in the expression of involuntary movements [44]. These studies support the clinical findings that the continuous intravenous or intraduodenal administration of levodopa or the continuous subcutaneous or intravenous infusion of apomorphine results in improved motor response but also with a marked reduction of dyskinesia [45, 46].

Other mechanisms are involved to explain the underlying cause and expression of dyskinesia. Although dopamine agonists when used as monotherapy in early PD are associated to a lower incidence of dyskinesia, involuntary movements are still observed, reflecting some kind of activity at the postsynaptic dopamine receptor level, as dopamine agonists are not dependent on the presence of presynaptic terminals.

Subtle changes in D<sub>1</sub> and D<sub>2</sub> receptor density as well as the complex interaction between receptor activation and synaptic plasticity [1] have been proposed as playing significant roles in dyskinesia induction and expression. Although the exact molecular mechanisms of LID still remain to be fully elucidated, exaggerated signalling of the striatal D<sub>1</sub> [47–49], the reduction of the modulating function of D<sub>2</sub>/D<sub>3</sub>

TABLE 1: Pharmacological characteristics of dopamine agonists.

	Pramipexole	Ropinirole	Rotigotine	Pergolide	Bromocriptine	Cabergoline	Apomorphine	Lisuride
D <sub>1</sub>	0	0	+	+	–	0/+	+++	–
D <sub>2</sub>	+++	+++	+++	++++	++	+++	++	++++
D <sub>3</sub>	++++	++++	++++	+++	++	++	++++	+++
Type	Nonergot	Nonergot	Nonergot	Ergoline	Ergoline	Ergoline	Morphine deriv.	Ergoline
Routes	os	os	td	os	os	os	sc	sc
Metabolism	—	Hepatic	Hepatic	Hepatic	?	Hepatic	Hepatic	Hepatic
Elimin.	Urine	Urine	Urine/fecal	Urine/fecal	Fecal	Fecal/urine	Urine/fecal	Urine/fecal
Half-life (h)	8–12	5–6	5–7	27	12–14	63–69	40 min.	2

td: transdermal; sc: subcutaneous.

receptors [42, 43, 50–52], and the interaction between D<sub>2</sub> and A2A adenosine receptors [53] have been implicated in both rodents and primates, suggesting that a normalisation of signalling may be beneficial in the treatment of dyskinesia.

In clinical practice, postsynaptic mechanisms can be partially explained by the dopamine agonists capability to prime for involuntary movements. Switching from a chronic dopamine agonist administration that usually results in a low expression and intensity of dyskinesia to an equivalent dose of levodopa in fact immediately results in the appearance of dramatic involuntary movements [54, 55]. These findings suggest that dopamine agonists principally prime for, but less commonly express dyskinesia. Vice versa, when considering the expression of dyskinesia in patients with PD with a history of levodopa exposure, switching to a dopamine agonist after the introduction of levodopa, established dyskinesia still occurred [56]. Moreover, patients receiving a combination of levodopa and the dopamine D<sub>2</sub>/D<sub>3</sub> agonist pramipexole showed a level of dyskinesia that was greater than the additive effect of the individual drug [57]. Once established, dopamine agonists produce the same pattern of dyskinesia although its intensity is reduced, suggesting that agonists do not express dyskinesia to the same extent as levodopa [54, 55, 58]. Both the lower priming for dyskinesia and the lower expression of involuntary movements by dopamine agonists may be a reflection of their more specific pharmacology compared to levodopa.

#### 4. Reducing or Delaying Parkinsonian Dyskinesias

Any type of exogenous dopaminergic stimulation in a denervated striatum can cause dyskinesias [59], but pulsatile stimulation produced by short-acting drugs (as typically occurs with levodopa) particularly favours their occurrence [60]. The expression LID is still currently used, although levodopa is not the only drug causing dyskinesias in PD patients [61]. Based on published series, it has been estimated that PD patients treated for less than 5 years have a 11% risk of developing dyskinesias, those treated for 6–9 years have a risk of 32%, whereas patients treated for more than 10 years have a risk of 89% [13].

Levodopa, however, seems to be the most important factor in inducing dyskinesia expression in chronically treated

PD patients; therefore, it appears that the benefit of initial treatment with a dopamine agonist in lowering the incidence of dyskinesias is related to the ability of the agonist to delay the need for levodopa [12, 62]. Moreover, experimental data suggest that the administration of long-acting dopamine agonists results in significantly less dyskinesia than does levodopa [63, 64] and other short-acting agents administered in a pulsatile fashion [65]. However, once a long-acting agonist is administered to animals already primed to exhibit dyskinesias with levodopa, the resultant dyskinesias are comparable to those seen in the levodopa group [63]. Clinical studies randomly assigning patients to initial treatment with a dopamine agonist or levodopa have shown a lower risk for dyskinesias in the groups treated with pramipexole [7], ropinirole [8, 12], bromocriptine [66, 67], pergolide [68], and cabergoline [6]; nevertheless, once levodopa was added, the rate of development of dyskinesias was similar in both groups.

One therapeutic strategy that has been tried in this sense is to use higher doses of a dopamine agonist to reduce both the total daily levodopa dose and its frequency [69] or to gradually substitute a dopamine agonist for levodopa [70]. Unfortunately, these strategies are unsatisfactory and typically reduce dyskinesias at the expense of less control of Parkinsonian symptoms. Indeed, the evidence that early levodopa exposure adversely affects the course of disease and leads to disabling dyskinesias and motor fluctuations constituted the rationale for the initial treatment with dopamine agonist.

#### 5. Different Profile and Efficacy of Dopamine Agonists in Reducing Dyskinesia

In order to create a valid alternative to levodopa, and with the aim of eliminating its related complications, many different drugs acting on dopaminergic receptors have been developed and studied during the last years. They have different metabolism, plasma half-life, affinity to receptors subtypes, excretion, and routes of administration (Table 1). Moreover, these drugs have different efficacies on reducing the incidence of dyskinesia, improving motor symptoms, and reducing the daily levodopa dose (Table 2, Figure 1).

Initially dopamine agonists have been used as adjuvant therapy to improve levodopa-induced complications, but

TABLE 2: Adjuvant therapy versus placebo.

	Pramipexole	Ropinirole	Pergolide*	Bromocriptine	Cabergoline
Off-time reduction (h/day)	-1.81	-0.93	-1.60	-1.78	-1.29
LEDD red (mg/day)	-114.82	-119.81	-183.90	-52.17	-149.60
UPDRS ADL reduction (pts)					-1.78
UPDRS III reduction (pts)		-4.80			-1.74
Incidence of dyskinesia (OR)	2.63	3.21	4.64	2.52	1.44

\*Based on data from just one trial [71].

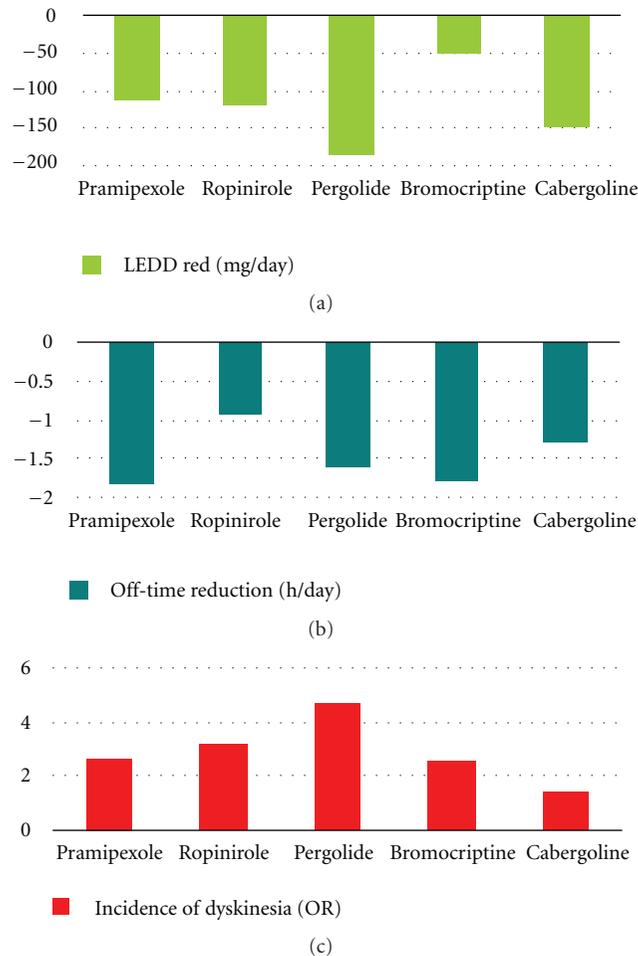


FIGURE 1: Effects of dopamine agonists on (a) reducing off time, (b) reducing levodopa daily dose, and (c) inducing dyskinesia.

once their effects on delaying the need for levodopa have been demonstrated, they have often been prescribed before the introduction of levodopa. Patients receiving dopamine agonists rather than levodopa as initial monotherapy showed a reduced risk for developing dyskinesias [7, 8, 12, 62, 72–76] (Table 3).

**5.1. Dopamine Agonists Monotherapy and the Risk of Dyskinesia.** The CALM-PD trial (Comparison of the Agonist Pramipexole versus Levodopa on Motor Complications of Parkinson's Disease) was a randomised controlled trial

evaluating the risk of developing dyskinesias in patients with early PD initially treated with either pramipexole or levodopa. After 24 months, pramipexole-treated patients were receiving pramipexole plus levodopa, compared with levodopa alone. A minority of pramipexole-treated patients reached the endpoint of time to first occurrence of wearing off, dyskinesias, or on-off motor fluctuations (27.8% versus 50.7%,  $P < 0.001$ ). Moreover, a significantly lower incidence of dyskinesias (9.9% versus 30.7%,  $P < 0.001$ ) also has been demonstrated in patients in the pramipexole group. However, after a mean 6-year followup, >90% of patients were receiving levodopa therapy regardless of their initial treatment assignment. Compared to those taking pramipexole, patients initially treated with levodopa had significantly more dyskinesias (20.4% versus 36.8%), but there was no difference between groups in the incidence of disabling or painful dyskinesias [62, 74]. Interestingly, 5 subjects taking pramipexole developed dyskinesias before the supplemental levodopa, and 4 of them had no prior levodopa exposure [73]. No significant difference in the Lang-Fahn activities of daily living dyskinesia score was observed (1.3 versus 1.1 with pramipexole,  $P < 0.06$ ) [7, 62, 72–74].

In a randomised, double-blind 5-year study of patients with early PD, the risk of developing dyskinesias after initial monotherapy with ropinirole was less than with levodopabenserazide (hazard ratio (HR), 2.82 (1.78, 4.44);  $P < 0.001$ ) [8]. However, many of these patients eventually required supplemental levodopa to control the symptoms of the disease [8, 12]. When patients receiving ropinirole monotherapy required the addition of levodopa, the risk for developing dyskinesias increased and then did not differ significantly from that associated with levodopa alone [12]. The use of ropinirole as monotherapy, with only later addition of levodopa, delayed the onset of dyskinesias by up to 3 years, although it was associated with a higher incidence of neuropsychiatric complications than levodopa monotherapy.

Apomorphine, a subcutaneous nonergolinic dopaminergic agent, has been studied in 2 retrospective chronic monotherapy trials in which no oral anti-parkinsonian therapies were permitted from the time the pump was turned on in the morning until it was turned off in the evening [77, 78]. The mean maximum reduction of dyskinesia per patient was 64% ( $P < 0.005$ ), and the mean time to achieve maximal dyskinesia improvement was 12.1 months.

Lisuride, another subcutaneously administered dopamine agonist, given as a continuous daytime infusion via pump, has been utilised as a strategy for minimising dyskinesias in 40 patients with advanced, levodopa-responsive PD

TABLE 3: Series on adjuvant therapy with dopamine agonists\*. In italic, dyskinesia evaluation.

Author	Duration	Characteristics of participants	Interventions	Primary outcomes	Secondary outcomes
Poewe et al. [79]	(6 months)	N: 302; MFs. Mean duration of PD: 8.5 y	<i>Pramipexole (n = 201) versus placebo (n = 101)</i>	<i>Disability; motor complications; on/off time</i>	SE
<i>Pahwa et al. [80, 81]; Sethi et al. [82, 83]; Stacy et al. [84]; Stocchi et al. [85, 86]</i>	(24 weeks)	N: 393; MFs. Mean duration of PD: 8.6 y	<i>Ropinirole (24-h) (n = 202) versus placebo (n = 191)</i>	<i>Disability; patient-rated QoL; on/off time; levodopa dose</i>	SE <i>Depression sleep scales</i>
Oertel et al. [87]; Pogarell et al. [88]	(32 weeks)	N: 363 (354 analysed); MFs. Mean duration of PD: 7.8 y	<i>Pramipexole (n = 180) versus placebo (n = 183)</i>	<i>Disability; off time; levodopa dose</i>	SE
Wong et al. [89]	(15 weeks)	N: 150; Mean duration of PD: 4.4 y	<i>Pramipexole (n = 73) versus placebo (n = 77)</i>	Disability; off time	SE
<i>Musch and Bonura [90]</i>	(24 weeks)	N: 218; on levodopa. Mean duration of PD: NA	<i>Cabergoline (n = 145) versus placebo (n = 73)</i>	<i>Disability; off time; levodopa dose</i>	SE
<i>Pinter et al. [91]</i>	(11 weeks)	N: 78; MFs. Mean duration of PD: 8.2 y	<i>Pramipexole (n = 34) versus placebo (n = 44)</i>	<i>Disability; off time; levodopa dose</i>	SE
<i>Wermuth [92]</i>	(12 weeks)	N: 69; MFs. Mean duration of PD: 10 y (range: 3–27 y)	<i>Pramipexole (n = 36) versus placebo (n = 33)</i>	<i>Disability; motor complications; off time; levodopa dose</i>	SE
<i>Lieberman et al. [93]; Weiner et al. [94]</i>	(32 weeks)	N: 360; MFs. Mean duration of PD: 9.2 y	<i>Pramipexole (n = 181) versus placebo (n = 179)</i>	<i>Disability; on/off time; levodopa dose</i>	SE
<i>Guttman [95]</i>	(9 months)	N: 247; MFs. Mean duration of PD: 7 y (range: 0.67–36 y)	<i>Pramipexole (n = 79) versus bromocriptine (n = 84) versus placebo (n = 83)</i>	<i>Disability; off time</i>	SE
<i>Kreider et al. [96]; Lieberman et al. [97]</i>	(6 months)	N: 149; predictable MFs. Mean duration of PD: 9 y	<i>Ropinirole (n = 95) versus placebo (n = 54)</i>	<i>Disability; motor complications; off time; levodopa dose</i>	SE
<i>Rascol et al. [98, 99]</i>	(12 weeks)	N: 46; not optimally controlled with levodopa. Mean duration of PD: 8 y	<i>Ropinirole (n = 23) versus placebo (n = 23)</i>	<i>Disability; motor complications; off time</i>	SE

TABLE 3: Continued.

Author	Duration	Characteristics of participants	Interventions	Primary outcomes	Secondary outcomes
Steiger et al. [100]	(6 months)	N: 37; MFs. Mean duration of PD: 12.8 y (range: 3–33 y)	Cabergoline (n = 19) versus placebo (n = 18)	Disability; motor complications; off time; levodopa dose	SE
Hutton et al. [101]; Lieberman and Hutton [102]; Schoenfelder et al. [103]	(24 weeks)	N: 188; MFs. Mean duration of PD: 10.6 y (range: 2–30 y)	Cabergoline (n = 123) versus placebo (n = 65)	Disability; on/off time; levodopa dose	SE
Olanow et al. [104]	(6 months)	N: 376; MFs. Mean duration of PD: 10.9 y	Pergolide (n = 189) versus placebo (n = 187)	Disability; motor complications; off time; levodopa dose	SE
Temlett et al. [105]	(5 weeks)	N: 44 (40 analysed); Mean duration of PD: 13.4 y	Bromocriptine (n = 22) versus placebo (n = 18)	Levodopa dose	SE
Toyokura et al. [106]	(8 weeks)	N: 222; not optimally controlled with levodopa. Mean duration of PD: 6.6 y	Bromocriptine (n = 114) versus placebo (n = 108)	Motor complications; on/off time	SE
Schneider and Fischer [107]	(4 weeks)	N: 40; not optimally controlled with levodopa. Mean duration of PD: 9.1 y	Bromocriptine (n = 20) versus placebo (n = 20)	On/off time; levodopa dose	
Jansen [108]	(5 months)	N: 23; not optimally controlled with levodopa. Mean duration of PD: 8.7 y	Bromocriptine (n = 12) versus placebo (n = 11)	Disability	

\* Performed on PD patients, parallel groups, double blind.  
MFs: motor fluctuations; SE: side effects.

characterised by motor fluctuations and dyskinesias [109]. After 4 years, the lisuride-treated patients had improved their baseline dyskinesia scores (measured by AIMS) by 49% ( $P < 0.0001$ ), whereas the levodopa-treated patients had worsened their scores by 59% ( $P < 0.0001$ ).

**5.2. Long-Acting Dopamine Agonists and the Risk of Dyskinesia.** In animal-model studies, the long-acting dopamine agonists have been demonstrated to prevent or reduce the onset time for LIDs. In a study of monkeys with MPTP-induced parkinsonism, small doses of subcutaneously administered cabergoline, a  $D_2$ -selective dopamine agonist with a relatively long half-life, were added as adjuvant therapy to orally administered levodopa/benserazide (100/25 mg) for 1 month, showing significantly lower dyskinesia scores (sum for all body segments) than when levodopa/benserazide was given alone for 1 month ( $P < 0.01$ ).

A report on the effect of cabergoline compared to levodopa showed a reduced incidence of dyskinesias [110]. Nevertheless, more recently, an increased incidence of dyskinesia and confusion in patients treated with bromocriptine was reported [111].

**5.3. Differences among Drugs in Adjuvant Therapy.** A recent systematic meta-analysis, which performs indirect comparisons among three classes of drugs, including nondopaminergic agents as catechol-O-methyl transferase inhibitors (COMTIs) or monoamine oxidase type B inhibitors (MAO-BIs), used as add-on (adjuvant) treatment to levodopa therapy in PD patients with motor complications, suggests that dopamine agonists may provide more effective symptomatic control [112].

**5.3.1. Off-Time Reduction.** There is no (or little) evidence of a difference across the different dopamine agonists for the overall reduction in off-time [pramipexole ( $-1.81$  hours/day, CI  $-2.19$  to  $-1.43$ ;  $P < 0.00001$ ); bromocriptine ( $-1.78$  hours/day, CI  $-2.91$  to  $-0.65$ ;  $P = 0.002$ ); pergolide ( $-1.60$  hours/day, CI  $-2.57$  to  $-0.63$ ;  $P = 0.001$ ); cabergoline ( $-1.29$  hours/day, CI  $-1.89$  to  $-0.69$ ;  $P < 0.0001$ ); ropinirole ( $-0.93$  hours/day, CI  $-1.53$  to  $-0.33$ ;  $P = 0.002$ )] [112].

**5.3.2. Levodopa Daily Dose Reduction.** The largest reduction was with pergolide ( $-183.90$  mg/day, CI  $-259.09$  to  $-72.71$ ;  $P = 0.001$ ), though this was based on data from just one trial [71]. Cabergoline reduced the required levodopa dose by  $149.60$  mg/day (CI  $-208.79$  to  $-90.41$ ;  $P < 0.00001$ ), ropinirole by  $119.81$  mg/day (CI  $-150.63$  to  $-89.00$ ;  $P < 0.00001$ ), pramipexole by  $114.82$  mg/day (CI  $-143.01$  to  $-86.64$ ;  $P < 0.00001$ ), and bromocriptine by  $52.17$  mg/day (CI  $-95.16$  to  $-9.18$ ;  $P = 0.02$ ) [112].

**5.3.3. UPDRS Scores Improvement.** The agonist pramipexole appeared to produce larger improvements for UPDRS motor score ( $-6.31$  points, CI  $-7.69$  to  $-4.93$ ;  $P < 0.00001$ ) compared to ropinirole (UPDRS motor:  $-4.80$  points, CI  $-7.32$  to  $-2.28$ ;  $P = 0.0002$ ) and cabergoline (UPDRS motor:  $-1.74$  points, CI  $-3.78$  to  $0.30$ ;  $P = 0.09$ ) [112].

**5.3.4. Incidence of Dyskinesia.** The analysis included 6476 participants, which represented 85% of the 7590 randomised participants included in the meta-analysis. Compared to placebo, the incidence of dyskinesia was increased with adjuvant therapy. The incidence of dyskinesia was greatest with pergolide (OR 4.64, CI 3.09 to 6.97;  $P < 0.00001$ ), although the data were obtained from just one trial [71], followed by ropinirole (OR 3.21, CI 1.98 to 5.21;  $P < 0.00001$ ), pramipexole (OR 2.63, CI 2.01 to 3.42;  $P < 0.00001$ ), bromocriptine (OR 2.52, CI 1.42 to 4.48;  $P = 0.002$ ), and cabergoline (OR 1.44, CI 0.96 to 2.16;  $P = 0.08$ ) [112].

Though this meta-analysis indirectly compares several series on dopaminergic agents as adjuvant treatment, the need of large randomised studies that directly compare different agents administered as monotherapy with patient-rated overall quality of life and health economic measures as primary outcomes is recommended.

## 6. Alternative Treatments to Reduce Dyskinesia

As seen earlier, the primary therapeutic strategy for managing LIDs in PD patients is to delay their occurrence through delaying the introduction of levodopa therapy administering dopaminergic agents.

Once dyskinesias have occurred, other strategies should be attempted: (1) substitution of immediate release for controlled-release levodopa. The immediate-release preparation is easier to adjust, as onset of its effects is sooner, and duration of action (and dyskinesias) is shorter than with controlled-release preparations. For the same reason, agents that prolong the half-life of levodopa, such as entacapone, should be stopped; (2) discontinuation of other therapy that may embitter dyskinesias, as dopamine agonists or other factors delaying dopamine degradation as selegiline and rasagiline; (3) incrementation of the number of administrations of levodopa, in lower doses; (4) addition of an antidyskinetic agent as amantadine, an NMDA receptor antagonist. Diphasic dyskinesias that may manifest at the beginning and the end of a dosing cycle should be managed by utilising more frequent doses of levodopa, and the therapy should be sewed on the patient [113].

**6.1. Amantadine.** The NMDA receptor-binding and neurotoxic effects of excessive glutamate have led to the hypothesis that an NMDA antagonist may have antidyskinetic effects and reduce the severity of LIDs. Amantadine has been studied as adjuvant treatment in levodopa-treated patients experiencing motor complications, including dyskinesias, with the aim of reducing these effects without worsening Parkinsonian symptoms [114–117]. Three randomized placebo-controlled crossover clinical studies in a group of 53 PD patients showed a reduction (up to 60%) in the severity of LIDs after challenge with acute levodopa administration, without impacting the beneficial effects of levodopa on motor function.

**6.2. Clozapine.** It is an atypical antipsychotic that has been assessed for the treatment of drug-induced psychosis in PD.

It may also be effective in decreasing dyskinesias [70], and a few studies have focused on its antidyskinetic effect [118, 119].

**6.3. Intraduodenal Levodopa.** It provides direct delivery of levodopa to the duodenum and jejunum. The method involves insertion of a permanent access tube in the abdominal wall by percutaneous endoscopic gastrostomy. Several clinical studies have been conducted using this approach, demonstrating significant reductions in “off” time and dyskinesia after 6 months. It may be an option for patients with marked fluctuations and dyskinesia in whom deep-brain stimulation (DBS) is contraindicated or not possible due to advanced age, or it may provide an alternative to DBS.

**6.4. Surgical Treatment.** Patients with PD who may benefit from surgery include those who have substantial dyskinesias unresponsive to medication adjustments, are levodopa responsive, do not have dementia, and do not have neuropsychiatric impairment [80]. DBS is the most frequently performed surgery for PD in North America [80]. In patients with advanced PD, DBS of the globus pallidus interna (GPI) or the subthalamic nucleus (STN) has been shown to reduce dyskinesia severity by up to 89% [120, 121] and to reduce the duration of dyskinesias by 86% [122]. It provides significant improvement in Parkinsonian motor features and allows a reduction of dyskinesias, in part through the subsequent reduction of levodopa [123, 124].

## References

- [1] P. Calabresi, M. D. Filippo, V. Ghiglieri, N. Tambasco, and B. Picconi, “Levodopa-induced dyskinesias in patients with Parkinson’s disease: filling the bench-to bedside gap,” *The Lancet Neurology*, vol. 9, no. 11, pp. 1106–1117, 2010.
- [2] F. Stocchi, M. Tagliati, and C. W. Olanow, “Treatment of levodopa-induced motor complications,” *Movement Disorders*, vol. 23, supplement 3, pp. S599–S612, 2008.
- [3] N. Tambasco, V. Belcastro, A. Gallina, A. Castrìo, P. Calabresi, and A. Rossi, “Levodopa-induced breathing, cognitive and behavioral changes in Parkinson’s disease,” *Journal of Neurology*, pp. 2296–2299, 2011.
- [4] J. G. Nutt, “Pharmacokinetics and pharmacodynamics of levodopa,” *Movement Disorders*, vol. 23, supplement 3, pp. S580–S584, 2008.
- [5] R. A. Hauser, M. P. McDermott, and S. Messing, “Factors associated with the development of motor fluctuations and dyskinesias in Parkinson disease,” *Archives of Neurology*, vol. 63, no. 12, pp. 1756–1760, 2006.
- [6] F. Bracco, A. Battaglia, C. Chouza et al., “The long-acting dopamine receptor agonist cabergoline in early Parkinson’s disease: final results of a 5-year, double-blind, levodopa-controlled study,” *CNS Drugs*, vol. 18, no. 11, pp. 733–746, 2004.
- [7] The Parkinson Study Group, “Pramipexole vs levodopa as initial treatment for Parkinson Disease: a 4-year randomized controlled trial,” *Archives of Neurology*, vol. 61, no. 7, pp. 1044–1053, 2004.
- [8] O. Rascol, D. J. Brooks, A. D. Korczyn, P. P. De Deyn, C. E. Clarke, and A. E. Lang, “A five-year study of the incidence of dyskinesia in patients with early Parkinson’s disease who were treated with ropinirole or levodopa,” *The New England Journal of Medicine*, vol. 342, no. 20, pp. 1484–1491, 2000.
- [9] V. Kostic, S. Przedborski, E. Flaster, and N. Sternic, “Early development of levodopa-induced dyskinesias and response fluctuations in young-onset Parkinson’s disease,” *Neurology*, vol. 41, no. 2 I, pp. 202–205, 1991.
- [10] A. Schrag, A. Hovris, D. Morley, N. Quinn, and M. Jahanshahi, “Young- versus older-onset Parkinson’s disease: impact of disease and psychosocial consequences,” *Movement Disorders*, vol. 18, no. 11, pp. 1250–1256, 2003.
- [11] The Parkinson Study Group, “Impact of deprenyl and tocopherol treatment on Parkinson’s disease in DATATOP subjects not requiring levodopa,” *Annals of Neurology*, vol. 39, no. 1, pp. 29–36, 1996.
- [12] O. Rascol, D. J. Brooks, A. D. Korczyn et al., “Development of dyskinesias in a 5-year trial and ropinirole and L-dopa,” *Movement Disorders*, vol. 21, no. 11, pp. 1844–1850, 2006.
- [13] G. Fabbrini, J. M. Brotchie, F. Grandas, M. Nomoto, and C. G. Goetz, “Levodopa-induced dyskinesias,” *Movement Disorders*, vol. 22, no. 10, pp. 1379–1389, 2007.
- [14] A. Schrag and N. Quinn, “Dyskinesias and motor fluctuations in Parkinson’s disease: a community-based study,” *Brain*, vol. 123, no. 11, pp. 2297–2305, 2000.
- [15] A. Schrag and J. M. Schott, “Epidemiological, clinical, and genetic characteristics of early-onset parkinsonism,” *The Lancet Neurology*, vol. 5, no. 4, pp. 355–363, 2006.
- [16] N. Kumar, J. A. Van Gerpen, J. H. Bower, and J. E. Ahlskog, “Levodopa-dyskinesia incidence by age of Parkinson’s disease onset,” *Movement Disorders*, vol. 20, no. 3, pp. 342–344, 2005.
- [17] K. E. Lyons, J. P. Hubble, A. I. Tröster, R. Pahwa, and W. C. Koller, “Gender differences in Parkinson’s disease,” *Clinical Neuropharmacology*, vol. 21, no. 2, pp. 118–121, 1998.
- [18] M. Zappia, G. Annesi, G. Nicoletti et al., “Sex differences in clinical and genetic determinants of levodopa peak-dose dyskinesias in Parkinson disease: an exploratory study,” *Archives of Neurology*, vol. 62, no. 4, pp. 601–605, 2005.
- [19] Y. Gilgun-Sherki, R. Djaldetti, E. Melamed, and D. Offen, “Polymorphism in candidate genes: implications for the risk and treatment of idiopathic Parkinson’s disease,” *Pharmacogenomics Journal*, vol. 4, no. 5, pp. 291–306, 2004.
- [20] R. Kaiser, A. Hofer, A. Grapengiesser et al., “L-Dopa-induced adverse effects in PD and dopamine transporter gene polymorphism,” *Neurology*, vol. 60, no. 11, pp. 1750–1755, 2003.
- [21] R. L. Oliveri, G. Annesi, M. Zappia et al., “Dopamine D<sub>2</sub> receptor gene polymorphism and the risk of levodopa-induced dyskinesias in PD,” *Neurology*, vol. 53, no. 7, pp. 1425–1430, 1999.
- [22] W. C. Koller, W. Tse, Olanow et al., “Unmet medical needs in Parkinson’s disease,” *Neurology*, vol. 62, supplement 1, pp. S1–S8, 2004.
- [23] S. Fahn, “The spectrum of levodopa-induced dyskinesias,” *Annals of Neurology*, vol. 47, supplement 1, pp. S2–S11, 2000.
- [24] L. I. Globe, “Young-onset Parkinson’s disease: a clinical review,” *Neurology*, vol. 41, pp. 168–173, 1991.
- [25] S. Boyce, C. E. Clarke, R. Luquin et al., “Induction of chorea and dystonia in parkinsonian primates,” *Movement Disorders*, vol. 5, no. 1, pp. 3–7, 1990.
- [26] F. Durif, “Treating and preventing levodopa-induced dyskinesias: current and future strategies,” *Drugs and Aging*, vol. 14, no. 5, pp. 337–345, 1999.
- [27] T. Boraud, E. Bezard, B. Bioulac, and C. E. Gross, “Dopamine agonist-induced dyskinesias are correlated to both firing pattern and frequency alterations of pallidal neurones in the MPTP-treated monkey,” *Brain*, vol. 124, pp. 546–557, 2001.

- [28] J. Jankovic, A. H. Rajput, M. P. McDermott, and D. P. Perl, "The evolution of diagnosis in early Parkinson disease," *Archives of Neurology*, vol. 57, no. 3, pp. 369–372, 2000.
- [29] A. Schrag, Y. Ben-Shlomo, R. Brown, C. D. Marsden, and N. Quinn, "Young-onset Parkinson's disease revisited—clinical features, natural history, and mortality," *Movement Disorders*, vol. 13, no. 6, pp. 885–894, 1998.
- [30] V. Sossi, R. De La Fuente-Fernández, M. Schulzer, J. Adams, and J. Stoessl, "Age-related differences in levodopa dynamics in Parkinson's: implications for motor complications," *Brain*, vol. 129, pp. 1050–1058, 2006.
- [31] D. A. Di Monte, A. McCormack, G. Petzinger, A. M. Janson, M. Quik, and W. J. Langston, "Relationship among nigrostriatal denervation, parkinsonism, and dyskinesias in the MPTP primate model," *Movement Disorders*, vol. 15, no. 3, pp. 459–466, 2000.
- [32] G. Linazasoro, "New ideas on the origin of L-dopa-induced dyskinesias: age, genes and neural plasticity," *Trends in Pharmacological Sciences*, vol. 26, no. 8, pp. 391–397, 2005.
- [33] G. Linazasoro, N. Van Blercom, A. Bergaretxe, I. Fernández Manchola, E. Laborda, and J. A. Ruiz Ortega, "Levodopa-induced dyskinesias in parkinson disease are independent of the extent of striatal dopaminergic denervation: a pharmacological and SPECT study," *Clinical Neuropharmacology*, vol. 32, no. 6, pp. 326–329, 2009.
- [34] C. R. Gerfen, T. M. Engber, L. C. Mahan et al., "D1 and D2 dopamine receptor-regulated gene expression of striatonigral and striatopallidal neurons," *Science*, vol. 250, no. 4986, pp. 1429–1432, 1990.
- [35] J. E. Nash and J. M. Brotchie, "A common signaling pathway for striatal NMDA and adenosine A(2a) receptors: implications for the treatment of Parkinson's disease," *Journal of Neuroscience*, vol. 20, no. 20, pp. 7782–7789, 2000.
- [36] E. Santini, E. Valjent, and G. Fisone, "Parkinson's disease: levodopa-induced dyskinesia and signal transduction," *FEBS Journal*, vol. 275, no. 7, pp. 1392–1399, 2008.
- [37] S. Boyce, N. M. J. Rupniak, M. J. Steventon, and S. D. Iversen, "Nigrostriatal damage is required for induction of dyskinesias by L-DOPA in squirrel monkeys," *Clinical Neuropharmacology*, vol. 13, no. 5, pp. 448–458, 1990.
- [38] A. Kumar, S. Mann, V. Sossi et al., "[11C]DTBZ-PET correlates of levodopa responses in asymmetric Parkinson's disease," *Brain*, vol. 126, no. 12, pp. 2648–2655, 2003.
- [39] R. de la Fuente-Fernández, "Drug-induced motor complications in dopa-responsive dystonia: implications for the pathogenesis of dyskinesias and motor fluctuations," *Clinical Neuropharmacology*, vol. 22, no. 4, pp. 216–219, 1999.
- [40] A. E. Fleckenstein, T. J. Volz, and G. R. Hanson, "Psychostimulant-induced alterations in vesicular monoamine transporter-2 function: neurotoxic and therapeutic implications," *Neuropharmacology*, vol. 56, supplement 1, pp. 133–138, 2009.
- [41] J. K. Dreyer, K. F. Herrik, R. W. Berg, and J. D. Hounsgaard, "Influence of phasic and tonic dopamine release on receptor activation," *Journal of Neuroscience*, vol. 30, no. 42, pp. 14273–14283, 2010.
- [42] J. A. Obeso, C. W. Olanow, and J. G. Nutt, "Levodopa motor complications in Parkinson's disease," *Trends in Neurosciences*, vol. 23, no. 10, pp. S2–S7, 2000.
- [43] J. A. Obeso, M. C. Rodriguez-Oroz, M. Rodriguez, M. R. DeLong, and C. W. Olanow, "Pathophysiology of levodopa-induced dyskinesias in Parkinson's disease: problems with the current model," *Annals of Neurology*, vol. 47, supplement 1, pp. S22–S34, 2000.
- [44] T. Björklund, T. Carlsson, E. A. Cederfjäll, M. Carta, and D. Kirik, "Optimized adeno-associated viral vector-mediated striatal DOPA delivery restores sensorimotor function and prevents dyskinesias in a model of advanced Parkinson's disease," *Brain*, vol. 133, pp. 496–511, 2010.
- [45] M. M. Mouradian, J. L. Juncos, G. Fabbrini, and T. N. Chase, "Motor fluctuations in Parkinson's disease: pathogenetic and therapeutic studies," *Annals of Neurology*, vol. 22, no. 4, pp. 475–479, 1987.
- [46] F. Stocchi, "The therapeutic concept of continuous dopaminergic stimulation (CDS) in the treatment of Parkinson's disease," *Parkinsonism and Related Disorders*, vol. 15, supplement 3, pp. S68–S71, 2009.
- [47] E. Bezard, J. M. Brotchie, and C. E. Gross, "Pathophysiology of levodopa-induced dyskinesia: potential for new therapies," *Nature Reviews Neuroscience*, vol. 2, no. 8, pp. 577–588, 2001.
- [48] M. A. Cenci, C. S. Lee, and A. Björklund, "L-DOPA-induced dyskinesia in the rat is associated with striatal overexpression of prodynorphin- and glutamic acid decarboxylase mRNA," *European Journal of Neuroscience*, vol. 10, no. 8, pp. 2694–2706, 1998.
- [49] C. Fiorentini, M. C. Rizzetti, C. Busi et al., "Loss of synaptic D1 dopamine/N-methyl-D-aspartate glutamate receptor complexes in L-DOPA-induced dyskinesia in the rat," *Molecular Pharmacology*, vol. 69, no. 3, pp. 805–812, 2006.
- [50] C. Missale, S. R. Nash, S. W. Robinson, M. Jaber, and M. G. Caron, "Dopamine receptors: from structure to function," *Physiological Reviews*, vol. 78, no. 1, pp. 189–225, 1998.
- [51] C. Le Moine and B. Bloch, "D<sub>1</sub> and D<sub>2</sub> dopamine receptor gene expression in the rat striatum: sensitive cRNA probes demonstrate prominent segregation of D1 and D2 mRNAs in distinct neuronal populations of the dorsal and ventral striatum," *Journal of Comparative Neurology*, vol. 355, no. 3, pp. 418–426, 1995.
- [52] M. Morelli and G. DiChiara, "Agonist-induced homologous and heterologous sensitization to D-1- and D-2-dependent contraversive turning," *European Journal of Pharmacology*, vol. 141, no. 1, pp. 101–107, 1987.
- [53] D. Xiao, E. Bastia, Y. H. Xu et al., "Forebrain adenosine A<sub>2A</sub> receptors contribute to L-3,4-dihydroxyphenylalanine-induced dyskinesia in hemiparkinsonian mice," *Journal of Neuroscience*, vol. 26, no. 52, pp. 13548–13555, 2006.
- [54] L. A. Smith, M. J. Jackson, L. Johnston et al., "Switching from levodopa to the long-acting dopamine D<sub>2</sub>/D<sub>3</sub> agonist piribedil reduces the expression of dyskinesia while maintaining effective motor activity in MPTP-treated primates," *Clinical Neuropharmacology*, vol. 29, no. 3, pp. 112–125, 2006.
- [55] M. J. Jackson, L. A. Smith, G. Al-Barghouthy, S. Rose, and P. Jenner, "Decreased expression of l-dopa-induced dyskinesia by switching to ropinirole in MPTP-treated common marmosets," *Experimental Neurology*, vol. 204, no. 1, pp. 162–170, 2007.
- [56] G. C. Cotzias, P. S. Papavasiliou, and E. S. Tolosa, "Treatment of Parkinson's disease with aporphines. Possible role of growth hormone," *The New England Journal of Medicine*, vol. 294, no. 11, pp. 567–572, 1976.
- [57] M. A. Brodsky, B. S. Park, and J. G. Nutt, "Effects of a dopamine agonist on the pharmacodynamics of levodopa in parkinson disease," *Archives of Neurology*, vol. 67, no. 1, pp. 27–32, 2010.
- [58] K. A. Stockwell, D. K. A. Scheller, L. A. Smith et al., "Continuous rotigotine administration reduces dyskinesia resulting from pulsatile treatment with rotigotine or l-DOPA in

- MPTP-treated common marmosets," *Experimental Neurology*, vol. 221, no. 1, pp. 79–85, 2010.
- [59] G. Linazasoro, "Pathophysiology of motor complications in Parkinson disease: postsynaptic mechanisms are crucial," *Archives of Neurology*, vol. 64, no. 1, pp. 137–140, 2007.
- [60] C. W. Olanow and J. A. Obeso, "Preventing levodopa-induced dyskinesias," *Annals of Neurology*, vol. 47, no. 4, pp. S167–S178, 2000.
- [61] L. V. Metman, S. Konitsiotis, and T. N. Chase, "Pathophysiology of motor response complications in Parkinson's disease: hypotheses on the why, where, and what," *Movement Disorders*, vol. 15, no. 1, pp. 3–8, 2000.
- [62] R. Constantinescu, M. Romer, M. P. McDermott, C. Kamp, and K. Kiebert, "Impact of pramipexole on the onset of levodopa-related dyskinesias," *Movement Disorders*, vol. 22, no. 9, pp. 1317–1319, 2007.
- [63] R. K. B. Pearce, T. Banerji, P. Jenner, and C. D. Marsden, "De novo administration of ropinirole and bromocriptine induces less dyskinesia than levodopa in the MPTP-treated marmoset," *Movement Disorders*, vol. 13, no. 2, pp. 234–241, 1998.
- [64] E. C. Maratos, M. J. Jackson, R. K. B. Pearce, and P. Jenner, "Antiparkinsonian activity and dyskinesia risk of ropinirole and levodopa combination therapy in drug naïve MPTP-lesioned common marmosets (*Callithrix jacchus*)," *Movement Disorders*, vol. 16, no. 4, pp. 631–641, 2001.
- [65] P. J. Blanchet, F. Calon, M. Morissette et al., "Regulation of dopamine receptors and motor behavior following pulsatile and continuous dopaminergic replacement strategies in the MPTP primate model," *Advances in Neurology*, vol. 86, pp. 337–344, 2001.
- [66] M. A. Hely, J. G. L. Morris, W. G. J. Reid et al., "The Sydney multicentre study of Parkinson's disease: a randomised, prospective five year study comparing low dose bromocriptine with low dose levodopa-carbidopa," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 57, no. 8, pp. 903–910, 1994.
- [67] A. J. Lees, R. Katzenschlager, J. Head, and Y. Ben-Shlomo, "Ten-year follow-up of three different initial treatments in de-novo PD: a randomized trial," *Neurology*, vol. 57, no. 9, pp. 1687–1694, 2001.
- [68] W. H. Oertel, E. Wolters, C. Sampaio et al., "Pergolide versus levodopa monotherapy in early Parkinson's disease patients: The PELMOPET study," *Movement Disorders*, vol. 21, no. 3, pp. 343–353, 2006.
- [69] J. A. Obeso, M. C. Rodriguez-Oroz, and I. Zamarbide, "Clinical features, pathophysiology, and management of motor complications in Parkinson's disease," in *Principles of Treatment in Parkinson's Disease*, A. H. V. Schapira and C. W. Olanow, Eds., pp. 99–112, Butterworth Heinemann Elsevier, Philadelphia, Pa, USA, 2005.
- [70] M. J. Nirenberg and S. Fahn, "The role of levodopa and catechol-o-methyltransferase inhibitors," in *Principles of Treatment in Parkinson's Disease*, A. H. V. Schapira and C. W. Olanow, Eds., pp. 3–16, Butterworth Heinemann Elsevier, Philadelphia, Pa, USA, 2005.
- [71] C. W. Olanow, S. Fahn, M. Muentner et al., "A multicenter double-blind placebo-controlled trial of pergolide as an adjunct to sinemet in Parkinson's disease," *Movement Disorders*, vol. 9, no. 1, pp. 40–47, 1994.
- [72] Parkinson Study Group, "A randomized controlled trial comparing pramipexole with levodopa in early Parkinson's disease: design and methods of the CALM-PD study," *Clinical Neuropharmacology*, vol. 23, no. 1, pp. 34–44, 2000.
- [73] Parkinson Study Group, "Pramipexole versus levodopa as initial treatment for Parkinson disease: a randomized controlled trial," *The Journal of the American Medical Association*, vol. 284, no. 15, pp. 1931–1938, 2000.
- [74] Parkinson Study Group CALM Cohort Investigators, "Long-term effect of initiating pramipexole versus levodopa in early Parkinson disease," *Archives of Neurology*, vol. 66, pp. 563–570, 2009.
- [75] R. A. Hauser, O. Rascol, A. D. Korczyn et al., "Ten-year follow-up of Parkinson's disease patients randomized to initial therapy with ropinirole or levodopa," *Movement Disorders*, vol. 22, no. 16, pp. 2409–2417, 2007.
- [76] R. L. Watts, K. E. Lyons, R. Pahwa et al., "Onset of dyskinesia with adjunct ropinirole prolonged-release or additional levodopa in early Parkinson's disease," *Movement Disorders*, vol. 25, no. 7, pp. 858–866, 2010.
- [77] A. Colzi, K. Turner, and A. J. Lees, "Continuous subcutaneous waking day apomorphine in the long term treatment of levodopa induced interdose dyskinesias in Parkinson's disease," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 64, no. 5, pp. 573–576, 1998.
- [78] A. J. Manson, K. Turner, and A. J. Lees, "Apomorphine monotherapy in the treatment of refractory motor complications of Parkinson's disease: long-term follow-up study of 64 patients," *Movement Disorders*, vol. 17, no. 6, pp. 1235–1241, 2002.
- [79] W. H. Poewe, O. Rascol, N. Quinn et al., "Efficacy of pramipexole and transdermal rotigotine in advanced Parkinson's disease: a double-blind, double-dummy, randomised controlled trial," *The Lancet Neurology*, vol. 6, no. 6, pp. 513–520, 2007.
- [80] R. Pahwa, M. A. Stacy, L. W. Elmer, and S. H. Isaacson, "Ropinirole 24-hour prolonged release provides efficacy as early as week 2 when used as adjunctive therapy to levodopa in patients with advanced Parkinson's disease," *Movement Disorders*, vol. 21, supplement 15, p. S595, 2006.
- [81] R. Pahwa, M. A. Stacy, S. A. Factor et al., "Ropinirole 24-hour prolonged release: randomized, controlled study in advanced Parkinson disease," *Neurology*, vol. 68, no. 14, pp. 1108–1115, 2007.
- [82] K. D. Sethi, R. A. Hauser, and N. L. Earl, "Ropinirole 24-hour prolonged release improves disease-specific and global symptoms when used as adjunctive therapy to levodopa in patients with advanced Parkinson's disease," *Movement Disorders*, vol. 21, supplement 15, p. S570, 2006.
- [83] K. D. Sethi, F. Stocchi, and L. Giorgi, "Ropinirole 24-hour prolonged release in advanced Parkinson's disease: relationship between treatment response and disease severity," *Movement Disorders*, vol. 22, supplement 16, p. S92, 2007.
- [84] M. A. Stacy, R. Pahwa, and N. L. Earl, "Ropinirole 24-hour prolonged release reduces "off" time and the dose of levodopa needed when used as adjunctive therapy in patients with advanced Parkinson's disease," *Movement Disorders*, vol. 21, supplement 15, p. S596, 2006.
- [85] F. Stocchi, B. P. Hersh, N. L. Earl, and B. L. Scott, "Safety and tolerability of ropinirole 24-hour prolonged release in patients with early and advanced Parkinson's disease," *Movement Disorders*, vol. 21, supplement 15, p. S572, 2006.
- [86] F. Stocchi, N. P. Stover, and L. Giorgi, "Ropinirole 24-hour prolonged release as adjunct to levodopa in patients with advanced Parkinson's disease—efficacy according to baseline depression score," *Movement Disorders*, vol. 22, supplement 16, p. S90, 2007.

- [87] J. C. Möller, W. H. Oertel, J. Köster, G. Pezzoli, and L. Provinciali, "Long-term efficacy and safety of pramipexole in advanced Parkinson's disease: results from a European multicenter trial," *Movement Disorders*, vol. 20, no. 5, pp. 602–610, 2005.
- [88] G. König, O. Pogarell, J. C. Möller, M. Delf, and W. H. Oertel, "Pramipexole, a nonergot dopamine agonist, is effective against rest tremor in intermediate to advanced Parkinson's disease," *Clinical Neuropharmacology*, vol. 22, no. 5, pp. 301–305, 1999.
- [89] K. S. Wong, C. S. Lu, D. E. Shan, C. C. Yang, T. H. Tsoi, and V. Mok, "Efficacy, safety, and tolerability of pramipexole in untreated and levodopa-treated patients with Parkinson's disease," *Journal of the Neurological Sciences*, vol. 216, no. 1, pp. 81–87, 2003.
- [90] B. Musch and L. Bonura, "Cabergoline once a day as adjunctive therapy to levodopa in Parkinson's disease," *Movement Disorders*, vol. 15, supplement 3, 2000.
- [91] M. M. Pinter, O. Pogarell, and W. H. Oertel, "Efficacy, safety, and tolerance of the non-ergoline dopamine agonist pramipexole in the treatment of advanced Parkinson's disease: a double blind, placebo controlled, randomised, multicentre study," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 66, no. 4, pp. 436–441, 1999.
- [92] L. Wermuth, "A double-blind, placebo-controlled, randomized, multi-center study of pramipexole in advanced Parkinson's disease," *European Journal of Neurology*, vol. 5, no. 3, pp. 235–242, 1998.
- [93] A. Lieberman, A. Ranhosky, and D. Korts, "Clinical evaluation of pramipexole in advanced Parkinson's disease: results of a double-blind, placebo-controlled, parallel-group study," *Neurology*, vol. 49, no. 1, pp. 162–168, 1997.
- [94] W. J. Weiner, S. A. Factor, J. Jankovic et al., "The long-term safety and efficacy of pramipexole in advanced Parkinson's disease," *Parkinsonism and Related Disorders*, vol. 7, no. 2, pp. 115–120, 2001.
- [95] M. Guttman, "Double-blind comparison of pramipexole and bromocriptine treatment with placebo in advanced Parkinson's disease," *Neurology*, vol. 49, no. 4, pp. 1060–1065, 1997.
- [96] M. Kreider, S. Knox, D. Gardiner, and D. Wheadon, "A multicenter double-blind study of ropinirole as an adjunct to levodopa in Parkinson's disease," *Neurology*, vol. 46: A475, 1996.
- [97] A. Lieberman, C. W. Olanow, K. Sethi et al., "A multicenter trial of ropinirole as adjunct treatment for Parkinson's disease," *Neurology*, vol. 51, no. 4, pp. 1057–1062, 1998.
- [98] O. Rascol, A. J. Lees, J. M. Senard et al., "A placebo-controlled study of ropinirole, a new D2 agonist, in the treatment of motor fluctuations of L-DOPA-treated parkinsonian patients," *Advances in Neurology*, vol. 69, pp. 531–534, 1996.
- [99] O. Rascol, A. J. Lees, J. M. Senard, Z. Pirtosek, J. L. Montastruc, and D. Fuell, "Ropinirole in the treatment of levodopa-induced motor fluctuations in patients with Parkinson's disease," *Clinical Neuropharmacology*, vol. 19, no. 3, pp. 234–245, 1996.
- [100] M. J. Steiger, T. El-Debas, T. Anderson, L. J. Findley, and C. D. Marsden, "Double-blind study of the activity and tolerability of cabergoline versus placebo in parkinsonians with motor fluctuations," *Journal of Neurology*, vol. 243, no. 1, pp. 68–72, 1996.
- [101] J. T. Hutton, W. C. Koller, J. E. Ahlskog et al., "Multicenter, placebo-controlled trial of cabergoline taken once daily in the treatment of Parkinson's disease," *Neurology*, vol. 46, no. 4, pp. 1062–1065, 1996.
- [102] A. N. Lieberman and J. T. Hutton, "Levodopa sparing effect of cabergoline compared to placebo in patients affected by motor fluctuations, under levodopa therapy," *Movement Disorders*, vol. 11, supplement 1, p. 269, 1996.
- [103] J. Schoenfelder, J. Simons, D. E. Souder, and J. Bianchine, "A placebo-controlled study of the safety and efficacy of cabergoline in the treatment of Parkinson's disease," *Functional Capacity Evaluation Reports 21336/731i*, 1993, Clinical Reference 42.
- [104] C. W. Olanow, S. Fahn, M. Muentner et al., "A multicenter double-blind placebo-controlled trial of pergolide as an adjunct to sinemet® in Parkinson's disease," *Movement Disorders*, vol. 9, no. 1, pp. 40–47, 1994.
- [105] J. A. Temlett, A. Ming, M. Saling et al., "Adjunctive therapy with bromocriptine in Parkinson's disease," *South African Medical Journal*, vol. 78, no. 11, pp. 680–685, 1990.
- [106] Y. Toyokura, Y. Mizuno, and M. Kase, "Effects of bromocriptine on Parkinsonism. A nation-wide collaborative double-blind study," *Acta Neurologica Scandinavica*, vol. 72, no. 2, pp. 157–170, 1985.
- [107] E. Schneider and P. A. Fischer, "Bromocriptine in the treatment of progressive stages of Parkinson's disease," *Deutsche Medizinische Wochenschrift*, vol. 107, no. 5, pp. 175–179, 1982.
- [108] E. N. H. Jansen, "Bromocriptine in levodopa response-losing parkinsonism. A double blind study," *European Neurology*, vol. 17, no. 2, pp. 92–99, 1978.
- [109] F. Stocchi, S. Ruggieri, L. Vacca, and C. W. Olanow, "Prospective randomized trial of lisuride infusion versus oral levodopa in patients with Parkinson's disease," *Brain*, vol. 125, no. 9, pp. 2058–2066, 2002.
- [110] U. K. Rinne, F. Bracco, C. Chouza et al., "Early treatment of Parkinson's disease with cabergoline delays the onset of motor complications. Results of a double-blind levodopa controlled trial," *Drugs*, vol. 55, supplement 1, pp. 23–30, 1998.
- [111] C. E. Clarke and K. D. Deane, "Cabergoline versus bromocriptine for levodopa-induced complications in Parkinson's disease," *Cochrane Database of Systematic Reviews*, no. 1, Article ID CD001519, 2001.
- [112] R. Stowe, N. Ives, C. E. Clarke et al., "Evaluation of the efficacy and safety of adjuvant treatment to levodopa therapy in Parkinson's disease patients with motor complications," *Cochrane Database of Systematic Reviews*, vol. 7, p. CD007166, 2010.
- [113] J. Jankovic, "Motor fluctuations and dyskinesias in Parkinson's disease: clinical manifestations," *Movement Disorders*, vol. 20, supplement 11, pp. S11–S16, 2005.
- [114] L. V. Metman, P. Del Dotto, P. van den Munchkof, J. Fang, M. M. Mouradian, and T. N. Chase, "Amantadine as treatment for dyskinesias in Parkinson's disease," *Neurology*, vol. 50, no. 5, pp. 1323–1326, 1998.
- [115] E. Lugging, G. K. Wenning, S. Bosch et al., "Beneficial effects of amantadine on levodopa-induced dyskinesias in Parkinson's disease," *Movement Disorders*, vol. 15, pp. 873–878, 2000.
- [116] B. J. Snow, L. Macdonald, D. Mcauley, and W. Wallis, "The effect of amantadine on levodopa-induced dyskinesias in Parkinson's disease: a double-blind, placebo-controlled study," *Clinical Neuropharmacology*, vol. 23, no. 2, pp. 82–85, 2000.
- [117] N. J. Crosby, K. H. Deane, and C. E. Clarke, "Amantadine for dyskinesia in Parkinson's disease," *Cochrane Database of Systematic Reviews*, no. 2, p. CD003467, 2003.

- [118] F. Durif, B. Debilly, M. Galitzky et al., "Clozapine improves dyskinesias in Parkinson disease A double-blind, placebo-controlled study," *Neurology*, vol. 62, no. 3, pp. 381–388, 2004.
- [119] F. Pierelli, A. Adipietro, G. Soldati, F. Fattapposta, G. Pozzessere, and C. Scoppetta, "Low dosage clozapine effects on levodopa induced dyskinesias in parkinsonian patients," *Acta Neurologica Scandinavica*, vol. 97, no. 5, pp. 295–299, 1998.
- [120] V. C. Anderson, K. J. Burchiel, P. Hogarth, J. Favre, and J. P. Hammerstad, "Pallidal vs subthalamic nucleus deep brain stimulation in Parkinson disease," *Archives of Neurology*, vol. 62, no. 4, pp. 554–560, 2005.
- [121] B. Ford, L. Winfield, S. L. Pullman et al., "Subthalamic nucleus stimulation in advanced Parkinson's disease: blinded assessments at one year follow up," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 75, no. 9, pp. 1255–1259, 2004.
- [122] K. Østergaard, N. Sunde, and E. Dupont, "Effects of bilateral stimulation of the subthalamic nucleus in patients with severe Parkinson's disease and motor fluctuations," *Movement Disorders*, vol. 17, no. 4, pp. 693–700, 2002.
- [123] E. Moro, M. Scerrati, L. M. A. Romito, R. Roselli, P. Tonali, and A. Albanese, "Chronic subthalamic nucleus stimulation reduces medication requirements in Parkinson's disease," *Neurology*, vol. 53, no. 1, pp. 85–90, 1999.
- [124] F. J. G. Vingerhoets, J. G. Villemure, P. Temperli, C. Pollo, E. Pralong, and J. Ghika, "Subthalamic DBS replaces levodopa in Parkinson's disease: two-year follow-up," *Neurology*, vol. 58, no. 3, pp. 396–401, 2002.

## Clinical Study

# Intensive Rehabilitation Treatment in Parkinsonian Patients with Dyskinesias: A Preliminary Study with 6-Month Followup

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A major adverse effect of levodopa therapy is the development of dyskinesia, which affects 30–40% of chronically treated Parkinsonian patients. We hypothesized that our rehabilitation protocol might allow a reduction in levodopa dosage without worsening motor performances, thus reducing frequency and severity of dyskinesias. Ten Parkinsonian patients underwent a 4-week intensive rehabilitation treatment (IRT). Patients were evaluated at baseline, at the end of the rehabilitation treatment and at 6-month followup. Outcome measures were the Unified Parkinson's Disease Rating Scale Sections II, III, and IV (UPDRS II, III, IV) and the Abnormal Involuntary Movement Scale (AIMS). At the end of the IRT, levodopa dosage was significantly reduced ( $P = 0.0035$ ), passing from  $1016 \pm 327$  to  $777 \pm 333$  mg/day. All outcome variables improved significantly ( $P < 0.0005$  all) by the end of IRT. At followup, all variables still maintained better values with respect to admission ( $P < 0.02$  all). In particular AIMS score improved passing from  $11.90 \pm 6.5$  at admission to  $3.10 \pm 2.3$  at discharge and to  $4.20 \pm 2.7$  at followup. Our results suggest that it is possible to act on dyskinesias in Parkinsonian patients with properly designed rehabilitation protocols. Intensive rehabilitation treatment, whose acute beneficial effects are maintained over time, might be considered a valid noninvasive therapeutic support for Parkinsonian patients suffering from dyskinesia, allowing a reduction in drugs dosage and related adverse effects.

## 1. Introduction

A variety of drugs have been developed in the last fifty years and are currently used to control the disability related to Parkinson's disease (PD): levodopa, dopamine-agonists, monoaminoxidase B inhibitors, catechol-*O*-methyltransferase inhibitors.

A major limiting factor in levodopa therapy is the development of motor complications, in particular dyskinesia, which affects 30–40% of chronically treated PD patients [1].

Dyskinesias can improve by reducing the dopaminergic therapy, but it is usually cumbersome to decrease the levodopa dosage since this reduction elicits a worsening of motor symptoms: an increased bradykinesia, an increased "off time," a reduction of motor performance, and autonomy in daily activities.

In the last decade, a considerable number of studies have shown that exercise is effective in improving gait, balance, freezing, and motor performance in PD. In particular, recent studies on animals allow hypothesizing a direct action of physical activity on the mechanisms responsible for dyskinesias [2, 3].

In this study we present preliminary data on the effectiveness of intensive rehabilitation treatment (IRT) in PD patients with dyskinesias and on the persistence over time of its beneficial effects.

## 2. Methods

**2.1. Study Population.** Patients were screened from among those consecutively admitted to the movement disorder ambulatories of the Rehabilitation Institute of Montescano.

Eligibility criteria for patients were (a) diagnosis of “clinically probable” idiopathic Parkinson’s disease according to Gelb et al. [4], (b) development of dyskinesias in the last 3 years and a history of several failed attempts to improve dyskinesia by reducing or modifying drug dosage, (c) ability to walk without any physical assistance, (d) no cognitive impairment (mini-mental state examination score  $\geq 26$ ), (e) no comorbidity unrelated to Parkinson’s disease, (f) no vestibular/visual dysfunction limiting locomotion or balance, and (h) antiparkinsonian medications stable for  $>4$  weeks.

Ten eligible patients were invited to be admitted to the Rehabilitation Institute of Montescano for a 4-week intensive rehabilitation treatment.

Patients were examined by the same neurologist expert in movement disorders, in the morning, one hour after they had taken the first dose of levodopa, at baseline, at the end of the rehabilitation treatment, and at 6-month followup. The neurologist was blinded with respect to the study design for the entire period.

The outcome measures used were the Unified Parkinson’s Disease Rating Scale Sections II, III, and IV (UPDRS II, III, IV) [5] and the Abnormal Involuntary Movement Scale (AIMS) [6].

Patients were treated with different drugs (levodopa, I-COMT, I-MAOB, or dopamine agonist), and we evaluated the drug dosage as levodopa equivalent (mg/day).

The study was approved by the local ethics committee, and all subjects gave their informed written consent before participation.

**2.2. Intervention.** IRT consisted of a 4-week cycle of physiotherapy that entailed three daily sessions (two, not consecutive, in the morning and one in the afternoon), 5 days a week. The global duration of each session, including recovery periods, was about one hour. The first session comprised cardiovascular warm-up activities, relaxation exercises, muscle-stretching exercises (scapular muscle group, hip flexor, hamstring and gastrocnemius muscles), exercises to improve the range of motion of spinal, pelvic, and scapular joints, exercises to improve the functionality of the abdominal muscles, and postural changes in the supine position.

The second session comprised exercises to improve balance and gait using a stabilometric platform with a visual cue (patients were asked to follow a circular pathway on the screen by using a cursor sensitive to their feet movements on the platform) and treadmill plus (treadmill training with both a visual and an auditory cue) [7]. The last session was a session of occupational therapy aimed at improving autonomy in daily living activities: transferring from sitting position to standing position, rolling from supine position to sitting position and from sitting to supine, dressing, use of tools, and exercises to improve hand functionality and skills (e.g., using screws and bolts). Moreover, patients spent 20 minutes every day in front of a mirror in order to control involuntary and exaggerated movements.

TABLE 1: Outcome variables at admission, at discharge after a 4-week intensive rehabilitation treatment, and at 6-month followup.

Variable	Admission	Discharge	6-month followup
UPDRS_II	14.30 $\pm$ 4.7	9.40 $\pm$ 5.1	9.40 $\pm$ 3.0
UPDRS_III	20.00 $\pm$ 4.9	14.10 $\pm$ 4.3	11.60 $\pm$ 4.1
UPDRS_IV	7.50 $\pm$ 3.7	1.70 $\pm$ 1.6	2.60 $\pm$ 2.1
AIMS	11.90 $\pm$ 6.5	3.10 $\pm$ 2.3	4.20 $\pm$ 2.7
Levodopa equivalent (mg/day)	1016 $\pm$ 327	777 $\pm$ 333	777 $\pm$ 333

UPDRS: Unified Parkinson’s Disease Rating Scale (Sections II, III, and IV).  
AIMS: Abnormal Involuntary Movement Scale.

During the follow-up period, patients were invited to continue some simple exercises learnt during hospitalization period.

**2.3. Statistical Analysis.** Descriptive statistics are given as mean  $\pm$  SD. The Shapiro-Wilk statistic was used to test the normality of the distribution of all variables.

The effect of treatment on each outcome variable and the persistence over the 6-month follow-up period were assessed by repeated measurements analysis of variance with three repeated measurements: admission, discharge, and 6-month followup. Pairwise comparisons (discharge versus admission and 6-month followup versus admission) were carried out by contrast analysis in repeated measurements analysis of variance. A  $P$  value  $< 0.05$  was considered statistically significant. All analyses were carried out using the SAS/STAT statistical package, release 9.2 (SAS Institute Inc., Cary, NC, USA).

### 3. Results

All 10 patients (aged  $70 \pm 8$  years, duration of the disease  $11.4 \pm 2.4$  years) completed the intensive rehabilitation treatment and the 6-month follow-up control. The characteristics of patients at admission, discharge and at the follow-up time are reported in Table 1.

At the end of IRT, levodopa-equivalent dosage was significantly reduced ( $P = 0.0035$ ), passing from  $1016 \pm 327$  to  $777 \pm 333$  mg/day. At follow-up the levodopa-equivalent dosage was unchanged.

All outcome variables improved significantly by the end of the rehabilitation treatment ( $P = 0.0003$ ,  $P < 0.0001$ ,  $P < 0.0001$  and  $P = 0.0005$  for UPDRS\_II, UPDRS\_III, UPDRS\_IV and AIMS, resp.). At followup, all variables still maintained better values with respect to admission ( $P = 0.0176$ ,  $P < 0.0001$ ,  $P < 0.0001$  and  $P = 0.0026$ , resp.).

### 4. Conclusion

In this study we investigated the efficacy of IRT in PD patients with dyskinesias and the persistence over time of the beneficial effects of this treatment. We found a statistically and clinically significant improvement in all

outcome variables after the 4-week rehabilitation period, which was largely preserved even after a 6-month period.

The improvement in UPDRS II and III observed in this study is in accordance with our previous studies, in which we demonstrated that IRT acts slowing the disease progression in Parkinsonian patients in a very long followup [8]. The patients continued to perform the recommended exercises during the follow-up period and this may explain the persistence of the beneficial effects obtained during hospitalization. Moreover, the simple reduction of intensity and duration of dyskinesias during the day leads the patients to improve their motor performance and autonomy during activity of daily life.

Our results suggest that it is possible to act on dyskinesias in Parkinsonian patients with an IRT. Several preclinical investigations carried out in animal models of PD have demonstrated that an overload of redundant motor information is stored in the basal ganglia motor circuits of dopamine-denervated animals.

In particular, the striatum receives the most important glutamatergic innervation, is the site of interaction glutamate/dopamine, is the source of the inhibitory outputs, and is involved in the generation of motor fluctuation linked to L-dopa treatment [2]. In animal models, after denervation, the striatal plasticity is lost, but the chronic L-dopa treatment is able to restore the long term potentiation (LTP) of synaptic transmission [9, 10].

The reversal of synaptic strength from the potentiated state to pre-LTP levels is named depotentiation, and this process represents the synaptic process of erasing unnecessary motor information. In Parkinsonian animal models treated with L-dopa which show dyskinesias movement, the synaptic depotentiation is lost [2]. The inability of corticostriatal synapses to depotentiate might represent the cellular basis of dyskinesias.

The execution of movements plays a fundamental role in determining the outcome of subsequent motor responses elicited by dopamine receptor stimulation [11]. Exaggerated movements in response to a stimulation of dopaminergic receptors, such as those occurring during dyskinesia, might consequently convey erroneous information to the motor striate circuits. Therefore, when concomitant, competing correct movements are performed (as during rehabilitation treatment), the manifestation of abnormal dyskinetic movements may be attenuated.

This study, therefore, suggests the possibility that the competition between a correct motor behaviour and an abnormal motor response may depend on the balance between the trace memory of the two.

Another possible explanation may be related to a neurorestorative strategy. The effects of intensive exercise in promoting cell proliferation and neuronal differentiation in animal models are reported in a large cohort of studies.

In animals with cerebral lesions produced by 6-hydroxydopamine (6-OHDA) or 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), the intensive use of a treadmill or running wheels led to improvement in motor performance as compared to animals that did not use these devices. Both in unilateral and bilateral models of PD, intensive

treadmill exercise produced improvement in motor symptoms, which was related to a reduction in the neurochemical deficit: preservation of both tyrosine hydroxylase-positive fibres in the striatum and substantia nigra, as well as of vesicular monoamine transporter and dopamine transporter levels [12–17]. Increased dopamine availability, especially within the dorsolateral striatum, has been found in an MPTP mice model after intensive exercise with a motorized treadmill [18]. Overall, these findings show that intensive exercise exerts beneficial effects on dopamine transmission in parkinsonian mice models.

These neuroplastic effects of intensive exercise are probably related to increased expression of a variety of neurotrophic factors. In particular, brain-derived neurotrophic factor (BDNF) and glia-derived neurotrophic factor (GDNF) are the most likely growth factors involved in this process. BDNF is a key component of a number of aspects of neuroplasticity: neurogenesis, synaptogenesis, and cell survival [19, 20], while GDNF has been shown to promote the survival and differentiation of dopamine neurons and to maintain the survival of adult catecholaminergic neurons in mice [21, 22]. Tajiri et al. [17] have recently shown that rat models of PD performing intensive treadmill exercise experience upregulation of BDNF and GDNF in the striatum in comparison to rats that do not exercise. These findings are consistent with the findings of another study by Lau et al. [23], who showed that intensive treadmill exercise raises the level of endogenous BDNF and GDNF in the substantia nigra and striatum.

In conclusion, our findings suggest that properly designed intensive multidisciplinary rehabilitation treatment using treadmill should be considered as a valid noninvasive therapeutic support for patients who show dyskinesias.

## References

- [1] J. Jankovic, "Motor fluctuations and dyskinesias in Parkinson's disease: clinical manifestations," *Movement Disorders*, vol. 20, no. 11, pp. S11–S16, 2005.
- [2] B. Picconi, D. Centonze, K. Håkansson et al., "Loss of bidirectional striatal synaptic plasticity in L-DOPA-induced dyskinesia," *Nature Neuroscience*, vol. 6, no. 5, pp. 501–506, 2003.
- [3] A. Pisani, D. Centonze, G. Bernardi, and P. Calabresi, "Striatal synaptic plasticity: implications for motor learning and Parkinson's disease," *Movement Disorders*, vol. 20, no. 4, pp. 395–402, 2005.
- [4] D. J. Gelb, E. Oliver, and S. Gilman, "Diagnostic criteria for Parkinson disease," *Archives of Neurology*, vol. 56, no. 1, pp. 33–39, 1999.
- [5] S. Fahn and R. L. Elton, "Unified Parkinson's disease rating scale," in *Recent Developments in Parkinson's Disease*, S. Fahn, C. D. Marsden, D. Calne, and M. Goldstein, Eds., vol. 2, pp. 153–163, Macmillan Health Care Information, Florham Park, NJ, USA, 1987.
- [6] G. M. Simpson, J. H. Lee, B. Zoubok, and G. Gardos, "A rating scale for tardive dyskinesia," *Psychopharmacology*, vol. 64, no. 2, pp. 171–179, 1979.
- [7] G. Frazzitta, R. Maestri, D. Uccellini, G. Bertotti, and P. Abelli, "Rehabilitation treatment of gait in patients with Parkinson's

- disease with freezing: a comparison between two physical therapy protocols using visual and auditory cues with or without treadmill training," *Movement Disorders*, vol. 24, no. 8, pp. 1139–1143, 2009.
- [8] G. Frazzitta, G. Bertotti, G. Riboldazzi et al., "Effectiveness of intensive inpatient rehabilitation treatment on disease progression in Parkinsonian patients: a randomized controlled trial with 1-year follow-up," *Neurorehabil Neural Repair*, vol. 26, pp. 144–150, 2012.
- [9] D. Centonze, B. Picconi, P. Gubellini, G. Bernardi, and P. Calabresi, "Dopaminergic control of synaptic plasticity in the dorsal striatum," *European Journal of Neuroscience*, vol. 13, no. 6, pp. 1071–1077, 2001.
- [10] D. Centonze, P. Gubellini, B. Picconi, P. Calabresi, P. Giacomini, and G. Bernardi, "Unilateral dopamine denervation blocks corticostriatal LTP," *Journal of Neurophysiology*, vol. 82, no. 6, pp. 3575–3579, 1999.
- [11] N. Simola, G. Di Chiara, W. M. U. Daniels, T. Schallert, and M. Morelli, "Priming of rotational behavior by a dopamine receptor agonist in hemiparkinsonian rats: movement-dependent induction," *Neuroscience*, vol. 158, no. 4, pp. 1625–1631, 2009.
- [12] C. R. Freed and B. K. Yamamoto, "Regional brain dopamine metabolism: a marker for the speed, direction, and posture of moving animals," *Science*, vol. 229, no. 4708, pp. 62–65, 1985.
- [13] P. G. MacRae, W. W. Spirduso, G. D. Cartee, R. P. Farrar, and R. E. Wilcox, "Endurance training effects on striatal D2 dopamine receptor binding and striatal dopamine metabolite levels," *Neuroscience Letters*, vol. 79, no. 1-2, pp. 138–144, 1987.
- [14] I. Liste, M. J. Guerra, H. J. Caruncho, and J. L. Labandeira-Garcia, "Treadmill running induces striatal Fos expression via NMDA glutamate and dopamine receptors," *Experimental Brain Research*, vol. 115, no. 3, pp. 458–468, 1997.
- [15] G. J. Wang, N. D. Volkow, J. S. Fowler et al., "PET studies of the effects of aerobic exercise on human striatal dopamine release," *Journal of Nuclear Medicine*, vol. 41, no. 8, pp. 1352–1356, 2000.
- [16] J. L. Tillerson, W. M. Caudle, M. E. Reverón, and G. W. Miller, "Exercise induces behavioral recovery and attenuates neurochemical deficits in rodent models of Parkinson's disease," *Neuroscience*, vol. 119, no. 3, pp. 899–911, 2003.
- [17] N. Tajiri, T. Yasuhara, T. Shingo et al., "Exercise exerts neuroprotective effects on Parkinson's disease model of rats," *Brain Research*, vol. 1310, pp. 200–207, 2010.
- [18] G. M. Petzinger, B. E. Fisher, J. E. Van Leeuwen et al., "Enhancing neuroplasticity in the basal ganglia: the role of exercise in Parkinson's disease," *Movement Disorders*, vol. 25, no. 1, pp. S141–S145, 2010.
- [19] C. W. Cotman, N. C. Berchtold, and L. A. Christie, "Exercise builds brain health: key roles of growth factor cascades and inflammation," *Trends in Neurosciences*, vol. 30, no. 9, pp. 464–472, 2007.
- [20] M. P. Mattson, S. Maudsley, and B. Martin, "A neural signaling triumvirate that influences ageing and age-related disease: insulin/IGF-1, BDNF and serotonin," *Ageing Research Reviews*, vol. 3, no. 4, pp. 445–464, 2004.
- [21] L. F. Lin, D. H. Doherty, J. D. Lile, S. Bektesh, and F. Collins, "GDNF: a glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons," *Science*, vol. 260, no. 5111, pp. 1130–1132, 1993.
- [22] A. Pascual, M. Hidalgo-Figueroa, J. I. Piruat, C. O. Pintado, R. Gómez-Díaz, and J. López-Barneo, "Absolute requirement of GDNF for adult catecholaminergic neuron survival," *Nature Neuroscience*, vol. 11, no. 7, pp. 755–761, 2008.
- [23] Y. S. Lau, G. Patki, K. Das-Panja, W. D. Le, and S. O. Ahmad, "Neuroprotective effects and mechanisms of exercise in a chronic mouse model of Parkinson's disease with moderate neurodegeneration," *European Journal of Neuroscience*, vol. 33, no. 7, pp. 1264–1274, 2011.

## Review Article

# Understanding and Prevention of “Therapy-” Induced Dyskinesias

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L-dopa is the most effective, currently available treatment for Parkinson's disease (PD), but it leads to the development of involuntary movements known as L-dopa-induced dyskinesia (LID) in the majority of patients after long-term use. Both gene and cell therapy approaches are the subject of multiple ongoing studies as potential ways of relieving symptoms of PD without the complication of dyskinesia. However, the spectre of dyskinesia in the absence of L-dopa, the so-called “off-phase” or graft-induced dyskinesia (GID), remains a major obstacle particularly in the further development of cell therapy in PD, but it is also a concern for proponents of gene therapy approaches. LID results from nonphysiological dopamine release, supersensitivity of dopamine receptors, and consequent abnormal signalling through mechanisms of synaptic plasticity. Restoration of physiological circuitry within the basal ganglia loops is ultimately the aim of all cell and gene therapy approaches but each using distinctive strategies and accompanied by risks of exacerbation of LID or development of “off-phase”/GID. In this paper we discuss the details of what is understood regarding the development of dyskinesias with relevance to cell and gene therapy and potential strategies to minimize their occurrence.

## 1. Introduction

L-dopa is the most effective treatment for Parkinson's disease currently available and for many patients can provide effective relief of symptoms for many years after diagnosis. In most patients, L-dopa treatment leads to a “honeymoon” period during which the motor symptoms are well controlled. However, after 5 years of treatment, approximately 40% of patients will develop fluctuations in symptom control in response to the drug, as well as involuntary movements known as “L-dopa-induced dyskinesias” (LID) [1]. These complications affect as many as 89% of PD patients after 10 years of L-dopa treatment [2]. LID can be seen during “peak dose” periods, during “off” medication periods or in a “biphasic” pattern as L-dopa levels rise and fall following oral intake. For this reason, other strategies have been developed to try and restore normal function of the basal ganglia circuitry. Deep brain stimulation (DBS) of the subthalamic nucleus (STN) or globus pallidus pars interna (GPi) have

both been shown to be highly successful ways of controlling symptoms of PD. Dyskinesia is generally reduced following STN DBS as a result of reduction in L-dopa dose. GPi DBS is a highly effective way of reducing LID, but many patients remain reliant on frequent high doses of L-dopa to maintain control of PD symptoms [3, 4].

To improve upon the limitations of currently available therapies, studies are being performed to assess the role of either gene therapy or cell therapy to provide PD symptom relief without the complication of dyskinesia. Cell therapy trials have been seriously hindered by reports of dyskinesia occurring in the absence of L-dopa—the so-called “off-phase” or graft-induced dyskinesia (GID) [5, 6]. There are also theoretical concerns that such “off-phase” dyskinesias might limit the ability of gene therapy to lead to an effective PD therapy. As a prelude to discussing the origin of “therapy-” induced dyskinesia, and strategies to minimize or control them, a discussion of our current understanding of LID is required.

## 2. Origin of LID

Risk factors for the development of LID include younger age [7, 8], dose of L-dopa [9], pattern of L-dopa administration [9–12], and stage of disease [13–15]. The neural mechanisms that underlie LID in PD are not completely understood; however, the study of basal ganglia anatomy and physiology in the normal and dopamine-depleted states has been of great help. In PD, the degeneration of the dopaminergic neurons of the substantia nigra compacta (SNc) compromises the equilibrium between the direct (D1 receptor) and indirect (D2 receptor) pathways resulting in abnormal GPi hyperactivity. Initially, the clinical features of PD were thought to follow simple increases in the “rate” of activity of the GPi, which through inhibition of the motor thalamus acted as a brake to activity in the supplementary motor area. It was further considered that excessive levodopa stimulation might induce dyskinesia by reduction of the inhibition of thalamocortical neurons resulting in an overactivity of motor cortical areas [16].

This model, is however, inconsistent with several clinical and experimental findings. During LID in the nonhuman primate model of PD, there is decreased rather than increased metabolic activity in the ventral anterior (VA) and ventrolateral (VL) thalamic nuclei, regions of the thalamus that receive input from the GPi [17]. Furthermore, among patients with PD and LID, creation of a lesion within the GPi (pallidotomy) is associated with a reduction in LID rather than a deterioration in LID that would have been predicted by the previously described “rate model” [18].

The pathophysiological changes that underlie the development of LID must therefore be far more complex. Recordings taken from PD patients undergoing DBS surgery have revealed that during periods of LID there is an increase in 4–10 Hz activity in the contralateral STN, suggesting that there is an abnormal pattern of oscillatory activity throughout the basal ganglia [19]. The cause of this oscillatory activity is likely to be multifactorial involving both pre and post synaptic components.

**2.1. Dysfunctional Dopamine Release.** The surviving dopaminergic neurons in the progressively denervated striatum, sprout branches that successfully compensate for the neurodegenerative process until ~60% of neurons, are lost. Until this point, administration of L-dopa does not alter the concentration of striatal dopamine, but beyond the 60% deficit, the concentration of dopamine in the striatum increases 3-fold after L-dopa administration [20]. While L-dopa administration continues to enhance dopamine synthesis and release by the surviving dopaminergic neurons, L-dopa is also decarboxylated and released as dopamine by serotonergic terminals, noradrenergic neurons, striatal capillaries, and nonaminergic striatal interneurons [20–23]. These terminals do not store and release dopamine in a regulated way, thus leading to nonphysiological dopamine receptor stimulation [24].

The role of serotonergic neurons in the development of LID has been the subject of particular study. In rats with 6-hydroxy dopamine lesions, approximately 80% of the peak

dopamine (DA) efflux measured in the striatum following the administration of L-dopa originates from serotonergic neurons [25–28]. This nonphysiological DA release is highly dyskinesigenic; indeed recent evidence shows that dyskinetic rodents have increased numbers of serotonergic terminals and sprouting of serotonin axon varicosities stimulated by L-dopa exposure, leading to larger swings of extracellular DA release [29].

**2.2. Dopamine Receptor Supersensitivity.** Under conditions of chronic denervation, dopamine receptors develop supersensitivity, involving an increased expression of receptors on the postsynaptic membrane of medium spiny neurons [30, 31]. D1 receptor supersensitivity has been shown to have a direct relationship with LID severity [32]. Persistent stimulation of dopamine receptors normally leads to their desensitisation and induces receptor internalisation, and it is hypothesised that, in LID, this desensitisation and internalisation process is impaired [33–35]. In a rodent model of LID, it seems that D1 receptors become “anchored” on the plasma membrane of medium spiny neurons due to crosstalk with D3 receptors following chronic administration of L-dopa [34]. Consistent with this it has been shown that the use of a D3 antagonist restores normal levels of D1 receptors on the plasma membrane and has been associated with reduction in LID in both the rodent and primate models [36, 37]. However, it is clear that LID does not occur solely as a result of abnormal D1 receptor expression or sensitivity alone since D2 selective agonists can also provoke dyskinesia [38].

**2.3. Synaptic Plasticity.** Physiological dopaminergic input from nigrostriatal neurons onto the striatal medium spiny neurons plays an important role in the potentiation and depotentiation of synapses of the corticostriatal pathway. Repetitive stimulation can cause either a long-lasting increase in synaptic strength known as long-term potentiation (LTP), or an enduring decrease known as long-term depression (LTD), a phenomenon known as synaptic plasticity [39, 40]. It is this process that allows deletion of unwanted or unnecessary connections and strengthening of desirable motor programs.

Disruption of normal synaptic plasticity is strongly linked to the appearance of dyskinesia [41–45]. It is hypothesized that the disrupted motor control underlying dyskinesia is attributable to specific changes occurring along the dopamine D1 receptor/protein kinase A/dopamine and cyclic AMP-regulated phosphoprotein-32 (D1/PKA/DARPP-32) intracellular signalling pathway leading to the loss of synaptic depotentiation at corticostriatal synapses and to the development of nonphysiological motor circuits within the basal ganglia [39].

L-dopa can restore normal synaptic plasticity among individuals free of dyskinesia, but not when dyskinesias are already developed [46]. It has been proposed that patients with LID have lost the ability to depotentiate synapses normally, that is, they have lost the mechanism that underlies “synaptic forgetting,” resulting in pathological storage of information that would normally be erased, leading to the development of abnormal motor patterns, that is,

LID [42, 47]. Possible consequences of this process include increased phosphorylation of N-methyl-D-aspartate (NMDA) and alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) receptor subunits [48] and increased striatal dynorphin mRNA levels [49]. Amelioration of abnormal glutamatergic NMDA transmission likely explains the beneficial effects of Amantadine [50–52] on reversal of LID.

### 3. Graft-Induced Dyskinesia

Graft-induced dyskinesia (GID) was first brought to widespread attention following the publication of the randomised sham-surgery-controlled trials of fetal dopaminergic cell transplantation [5, 6]. GIDs are characterized by the presence of both hyperkinesias and dystonic postures occurring in the “off phase,” generally considered to be greater than 12 hours following the last dose of L-dopa. In the Freed trial, “off phase” GID was seen in 5/33 (15%) of transplanted patients, more than 1 year after transplant [5]. The second sham-surgery-randomised-controlled trial (Olanow trial) reported the presence of “off-phase” GID in 13/23 (56.5%) of the grafted patients, 6–12 months after transplantation [6]. In 3 of the patients from the Olanow trial the GIDs were disabling enough to require further neurosurgical intervention [6]. In contrast to the worrying appearance of “off-phase” GID, neither trial reported any increase in L-dopa-induced or “on-phase” dyskinesia.

Patients receiving open label transplants that predated the Freed and Olanow trials had also experienced GID [53–55] but these had caused only mild to moderate disability to the patients even with follow-up extending to 11 years [55]. Speculation regarding the origin of “off-phase” dyskinesias has included the possibility of excessive dopamine production by the grafts, dopamine receptor supersensitivity away from “islands” of grafted cells, individual factors related to the patient and their PD phenotype, a relationship with immune rejection, or contamination of the grafts with cells of a nondopaminergic phenotype.

**3.1. Excess Dopamine Release?** Initial hypotheses were that GID occurred due to excessive dopamine release by the grafts. This has not, however, been supported by functional imaging of the patients in the Olanow trial or the Hagell patient series [55–57]. Dyskinesia severity was not related to the magnitude of graft-derived dopaminergic reinnervation, as judged by 18F-dopa positron emission tomography (PET), indicating that “off-phase” dyskinesias probably do not result from excessive growth of grafted dopaminergic neurons. Furthermore, the severity of GID was not correlated with improvement in the “off” UPDRS motor scores [55, 58]. The fact that GIDs have not been seen in patients without any benefit from their transplants suggests that a functional graft is necessary for GID development although their appearance does not simply relate to excess dopamine release.

**3.2. Imbalanced Dopamine Release?** It was further proposed that islands of excessive dopaminergic activity might relate to GID [59]. Indeed the patients with the best functional outcome after transplantation exhibited no dopaminergic

denervation in areas outside the grafted areas, either preoperatively or at 1 or 2 years postoperatively. Comparing PET signal among patients with and without GID, there was a greater increase of putaminal 18F-Dopa uptake seen in the posterodorsal zone of GID patients. However, the GID group also showed a relative increase ventrally not seen at all in the GID-negative patients suggesting that unbalanced increases in dopaminergic function might complicate the outcome of neuronal transplantation for parkinsonism. The implantation of dopamine cells into the ventrocaudal putamen, may also contribute to the unusual distribution of GID, in which the face, neck, and arms tend to be the most clinically involved. These data are corroborated by animal models showing that dyskinesias occur following transplantation of cells to a particular prodyskinetic subregion of the putamen [49].

**3.3. Patient Phenotype?** In the same clinical studies, the severity of GID was not found to correlate with the severity of pretransplant L-dopa-induced dyskinesias (LID). However, a negative correlation was seen between the severity of post-operative “off-phase” GID and preoperative putaminal 18F-Dopa uptake [55, 57]. This finding indicates that the manifestation of “off-phase” dyskinesias after grafting, similar to that of L-dopa-induced “on-phase” dyskinesias [60, 61] might be related to the baseline severity of striatal dopaminergic denervation.

**3.4. Tissue Storage?** In the Freed trial, cells were stored for up to 4 weeks before transplantation, In the Hagell series, the appearance of GID was reported in patients with grafts stored for 1–8 days. However, any hypothetical relationship between tissue storage and GID hypothesis was not supported by the Olanow trial, in which no grafts were stored for >48 hours.

**3.5. Immunosuppression?** In the Olanow trial, the initial significant improvement in the grafted patients compared with sham-operated cases [6] was lost following withdrawal of immune suppression after 6 months. Also in two patients who came to autopsy, the grafts were surrounded by activated microglia suggesting an immune response [6]. Such inflammatory reactions could lead to reduced graft survival and functional deterioration [62, 63]. GIDs develop slowly over time and appear to be most pronounced in patients that have received no immune-suppression [5] or only mild immunosuppressive treatment [56]. There has been speculation, therefore, that such an ongoing inflammatory/immune process could affect the way the grafted DA neurons release and/or handle DA at the synaptic level, which in turn may constitute a triggering factor for the induction of dyskinesias [64].

**3.6. Serotonergic Contamination?** It has now been shown that in 2 patients experiencing GID, there was excessive serotonergic innervation in their brains following transplant, (252–285% higher than comparable advanced PD patients). This was measured using functional imaging with ((11)C)-3-amino-4-(2-dimethylaminomethyl-phenyl-sulfanyl) benzonitrile positron emission tomography (C<sup>11</sup>-DASB PET). Importantly this excessive serotonergic

innervation was seen restricted to the areas of their grafts. There is mounting evidence that serotonergic neurons contaminating the original graft release dopamine in an uncontrolled manner and then lack the ability to reuptake DA and buffer extracellular DA levels leading to GID [65].

#### 4. The Relationship between LID and GID

The clinical similarity between LID and GID suggests that similar pathophysiological mechanisms may underlie the development of both. Animal models strongly support the suggestion that the severity of LID in animals that have received putaminal grafts are related to the number of serotonergic neurons contained within the graft, as well as the severity of the dopaminergic lesion. Among animals receiving serotonin grafts [66], there was no impact on either motor asymmetry in the amphetamine-induced rotation test or spontaneous forelimb use in the cylinder test, but in contrast there was a progressive worsening of LID. Other studies have also confirmed that removal of serotonin innervation, or dampening of serotonin neuron activity by agonist drugs, results in a near complete blockade of LID in 6-OHDA lesioned rats [27].

It would appear that the relative abundance of dopaminergic compared to serotonergic neurons (whether host or grafted) is a critical factor in LID development after graft [67, 68]. It appears that, in the presence of a severe dopaminergic deficit, dopamine released from serotonergic neurons may trigger severe dyskinetic responses, but provided ~10–20% of the dopamine innervation remained intact, the grafted serotonin neurons have limited detrimental effect on dyskinesia severity. This has been corroborated independently showing that serotonergic neurons are not detrimental, provided sufficient dopamine neurons remain in the graft [69].

The appearance of LID after graft may occur via (i) dysregulated DA release from serotonergic neurons themselves, (ii) lack of reuptake of released DA due to insufficient DA neurons, (iii) inhibition of the dopamine transporter (DAT) on DA neurons by 5HT release thus preventing DA reuptake by DA neurons, and (iv) abnormal dopamine receptor supersensitivity (i.e., a postsynaptic component) as evidenced by increases in apomorphine-induced rotations [28, 68].

Despite these consistent findings of LID appearance following serotonergic contamination of cell grafts, “off-phase” dyskinesias (GID) do not appear to occur in animal models of PD undergoing cell grafts with the exception of very mild abnormal movements occurring in 2 studies [70, 71]. This is of major importance since there is little or no relationship between the change in LID following transplantation in patients (which tends to improve) and the development of GID. Given that the patients exhibiting GID are those that had the most severe dopaminergic deficits, in the absence of exogenous L-dopa administration, it seems likely that GID appearance must relate to dopamine production from the graft itself. Dopamine released into the extracellular space can be taken up by serotonergic neurons via the serotonin transporter and subsequently be rereleased as a false transmitter [72]. Furthermore, serotonin release

can block the dopamine transporter and add to abnormal accumulation of dopamine in the synaptic cleft and onset of dyskinesia [65, 73]. Whether serotonergic neurons have a role in the development of “off-phase” GID simply by maintaining postsynaptic dopamine receptors in a supersensitive state has not yet been confirmed. Any relevance of abnormal synaptic plasticity as a secondary or parallel process in the development of GID has also yet to be comprehensively studied.

#### 5. Gene Therapy for PD and Its Effects on Dyskinesia

Gene therapy represents an exciting new prospect for the treatment of patients with PD. This technology exploits the properties of viral vectors to invade host cells and incorporate DNA into the host genome. Appropriate modification of viral vectors allows control over which genes are incorporated and within PD research the main focus has been predominantly on genes encoding growth factors or enzymes involved in dopamine synthesis [74, 75]. Gene transfer offers a practical means of solving the problems associated with implanted hardware while still providing a continuous and selective delivery system of the desired gene/protein at the targeted site.

There are 4 gene therapy programs that have already reached the stage of clinical trial evaluation.

**5.1. AAV2-Neurturin.** AAV2-Neurturin, an analogue of glial-cell-derived neurotrophic factor (GDNF), has been developed in an attempt to provide trophic support to neurons/glia and thus manipulate the progression of PD. Both (GDNF) and Neurturin enhance dopaminergic neuron survival and nigrostriatal function in animal models of PD. Both factors provide protection from 6-OHDA-induced degeneration in rats [76] and provide neuroprotection in parkinsonian monkeys [77]. In a clinical trial of 58 patients at 12 months, delivery of neurturin to the striatum failed to show any change in the primary outcome measure (off-medication part III-UPDRS) [78]. There was not any change in dyskinesia scores in neither the Neurturin nor the sham surgery groups. Retrograde transport to the substantia nigra pars compacta (SNc) was lower (or possibly slower) than expected and therefore a follow-up evaluation is underway targeting both the striatum and SNc. It seems unlikely that this approach would lead to worsening of dyskinesia severity, indeed beneficial effects on dopaminergic number, survival or function should lead in theory to improvement in LID. Nevertheless, any disproportionate neurotrophic action on serotonergic neurons might in theory lead to provocation of dyskinesia, and these should be specifically sought during clinical evaluation of patients receiving this treatment.

**5.2. AAV2-GAD.** Gene therapy consisting of insertion of the glutamic acid decarboxylase gene (*GAD*) into the neurons of the STN offers an alternative therapeutic strategy. Inspired by the effectiveness of STN DBS, the hypothesis emerged that expression of GAD (the rate-limiting enzyme for GABA production) would inhibit overactivity in the STN and would

improve off-medication UPDRS motor score. This strategy has been effective in the rat model of PD [79]. It is known that STN DBS can itself provoke “off-medication dyskinesia” but this is usually a transient phenomenon when it occurs and may be relieved by adjustment of the stimulation amplitude. Since the mechanism of action of STN DBS remains controversial, it remains theoretically possible that GAD gene therapy might also provoke “off-phase” dyskinesia. However in the results published to date in a double blind evaluation, no increase in dyskinesias was reported, while a modest improvement in PD severity was observed [80]. Open label follow up of these patients will allow further quantitative estimates of the extent to which LID may improve or deteriorate following this approach.

**5.3. AAV-hAADC.** Bilateral intraputamin infusion of AAV-hAADC (adeno-associated virus-human aromatic L-amino acid decarboxylase), the enzyme that converts L-dopa into dopamine, aims to improve the conversion of exogenously administered L-dopa. Since this therapy remains dependent on exogenous administration of L-dopa, “off-phase” dyskinesia is unlikely to occur. However, striatal transfection with AAV-hAADC has been shown to increase LID in primate parkinsonian models if delivered in a nonhomogeneous way [81], reminiscent of the experience of “off-phase” graft-induced dyskinesia in cell therapy experiments. Despite these theoretical concerns, data published to date have shown an increase in on-time and reduction in off-time without any increase in LID [75]. Two patients had a transient increase in mild LID, but none experienced “troublesome LID,” which were reduced for the group as a whole [75].

**5.4. LV-TH-GCH1-AADC-ProSavin.** The ProSavin approach incorporates all 3 enzymes required for dopamine biosynthesis (tyrosine hydroxylase (TH), AADC, and GTP cyclohydrolase L1(GCH1)), with the aim of transfection of nondopaminergic cells so that they may produce and release dopamine endogenously. Again the theoretical concern is that transfected neurons may synthesize dopamine but may not be able to store and release this dopamine in a physiological manner, and an increase in dyskinesia may follow. The pilot phases of this program are using a dose escalation strategy, with careful evaluation of efficacy and dyskinesia severity at each stage, before proceeding with dose increases in subsequent patient cohorts. So far, there have been no concerns regarding “off-phase” dyskinesias. Further addition of the vesicular monoamine transporter 2 (VMAT2) gene to allow dopamine transport by transfected cells has not shown any advantage over the ProSavin approach in laboratory experiments [82].

An additional gene therapy program that has not yet reached clinical trial stage aims to deliver continuous dopamine therapy using AAV-TH-GCH1. By omitting the AADC enzyme, transfected cells would be able to synthesize L-dopa but rely on endogenous AADC activity to produce dopamine. This reduces the risk of uncontrolled dopamine production that cannot be stored or transported physiologically and aims to deliver “continuous dopaminergic stimulation (CDS),” to mimic the advantages observed using other

methods of CDS [12]. In rodent models, this therapy has been shown to allow resistance to LID development [83].

All of the current gene therapy approaches, if successful, could permit a reduction of L-dopa dose and, therefore, achieve greater control of LID. No major concerns regarding “off-phase” dyskinesias have been reported among patients exposed to PD gene therapy. However, dose escalation to try and achieve greater efficacy from these approaches will necessitate continued vigilance with regard to dyskinesia emergence.

## 6. Preventing “Therapy-” Induced Dyskinesias

Our knowledge regarding the underlying causes of LID is growing, and it is clear that the pathophysiological processes are complex, depending on the number of intact dopaminergic terminals, the chronicity, and pattern of administration of L-dopa replacement as well as the possibility of genetic variability in pathways controlling receptor supersensitivity/internalization and synaptic plasticity. There is converging evidence to implicate nondopaminergic neurons, in particular serotonergic neurons releasing dopamine in a nonphysiological manner, as a major contributory factor in LID and also GID.

This knowledge is vital in trying to minimize the likelihood of “off-phase” GID developing in PD patients participating in future trials of fetal cell therapy. Careful attention must be paid to selecting the optimal patients phenotype with respect to the severity of their PD and by implication the number of surviving dopaminergic terminals at the time of transplantation. Patients with advanced dopaminergic degeneration will be at greater risk of developing dyskinesia from grafts that contain an excessive number of serotonergic neurons. Patients with less advanced dopaminergic cells loss should be more tolerant of a greater number of serotonin contaminating cells. The relative extent of dopamine/serotonin striatal innervations can be revealed using preoperative functional imaging, to quantify the extent and severity of both dopaminergic and serotonergic innervations in the striatum.

The perioperative and intraoperative details are also extremely important. It is possible to manipulate the number of contaminating serotonergic neurons within grafts, through optimisation of the dissection margins in the ventral mesencephalon of the fetal tissue, during graft harvesting. Furthermore, clinical and animal data both suggest that surgical targeting should avoid the ventral putamen. While the type and duration of immunosuppressive regimes may or may not have major relevance for GID development, it is likely that overall cell survival is improved by maintaining immunosuppression for longer than the 6 months adopted in previous cell therapy trials.

**6.1. Additional Use of Nondopaminergic Medications.** The use of serotonin (5HT-1A) agonists as antidyskinetic agents is not new. Buspirone, a (5-hydroxytryptamine (5HT 1A) receptor agonist, has previously showed beneficial effects lessening the severity of LID [84], by dampening transmitter release from serotonergic neurons through activation of

inhibitory 5HT-1A autoreceptors without any worsening of extrapyramidal symptoms. Beneficial effects seen among patients with GID have also been reported [65]. Sarizotan (also a 5-HT1A receptor agonist but with additional high affinity for D3 and D4 receptors) showed encouraging results in an open-label evaluation [85]; however, in a blinded trial the effects of Sarizotan on LID were disappointing [86]. While the mechanism(s) remain unclear, one likely explanation for the possible beneficial effect of serotonin receptor agonists on LID is that stimulating 5HT1A receptors diminish dysregulated release of dopamine from raphe-striatal serotonergic neurons. However, it has also been shown that 5HT1A agonist drugs alleviate dyskinesias provoked by direct D1 receptor agonists, suggesting a further interaction between D1 and 5-HT1A receptors [87].

The beneficial effects of Amantadine on LID through its action on NMDA receptors have prompted further study of agents acting on the corticostriatal glutamatergic input evaluating the effects of metabotropic glutamate receptor (mGluR) antagonists and Adenosine A2A receptor antagonists. AFQ056 is a potent, selective mGluR5 antagonist that shows antidyskinetic effects in a rodent PD model and has been shown to have significant antidyskinetic effects in 2 small blinded trials [88]. Istradefylline (an A2A antagonist) has been shown to improve motor function and reduce the development of LID in nonhuman primates [89]; indeed rodent A2a knockout animals do not develop LID [90]. In patients with PD, however, multiple randomised trials have failed to show beneficial effects of Istradefylline on LID severity [80, 91].  $\alpha$ 2Adrenergic agonists modulate the activity of the direct striatopallidal pathway and have been shown to reduce LID in PD patients but their use in PD patients has been limited by development of side effects [92, 93]. Given the demonstration of beneficial effects in animal models [36, 37], a further emerging possibility might be the use of D3 antagonists to allow D1 receptor internalisation, with the aim of reducing the postsynaptic "supersensitive" state [94]; however, this approach has yet to be evaluated in patients.

## 7. Conclusions

The current and future gene therapy and cell therapy programs represent a great source of optimism for patients with PD. However, PD is a heterogeneous disease and only a subset of patients are likely to benefit—perhaps the subgroup of patients with young onset PD that remain with a predominantly motor deficit for many years [95]. In these individuals, LID is a major problem and the success of cell or gene therapy will depend on providing relief of PD "off" symptoms without "off-phase"/therapy-induced dyskinesia.

Dyskinesia development has multiple determinants, and therefore multiple potential opportunities exist to intervene and prevent their occurrence. Some of these processes may be relevant solely as a secondary consequence of nonphysiological dopamine release (presynaptic component) from serotonergic or other neurons, or as a downstream effect of chronic dopamine receptor supersensitivity (postsynaptic component). Consequent downstream changes in synaptic plasticity may account for abnormal oscillatory firing patterns,

throughout the basal ganglia circuitry. L-dopa itself (presumably when released physiologically) has been identified as having a role in the restoration of normal synaptic plasticity, as evidenced from recordings of patients undergoing high-frequency stimulation of the substantia nigra pars reticulata, in both on- and off-medication states [96, 97]. Whether other pharmacological options such as Buspirone, AFQ056, or D3 receptor antagonists can be exploited to relieve off-medication GID needs further study.

In parallel with approaches to relieve LID and GID, our understanding of the importance of the ratio of serotonin and dopaminergic neurons allows optimisation of future interventions and accompanying trial design. Our greater understanding of the causes of dyskinesia, either L-dopa related or graft and gene therapy induced, represents a considerable step towards ensuring the success of future gene and cell therapy programs.

## Conflict of Interests

There is no actual or potential conflict of interests in relation to this paper.

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## References

- [1] J. E. Ahlskog and M. D. Muentzer, "Frequency of levodopa-related dyskinesias and motor fluctuations as estimated from the cumulative literature," *Movement Disorders*, vol. 16, no. 3, pp. 448–458, 2001.
- [2] A. Schrag and N. Quinn, "Dyskinesias and motor fluctuations in Parkinson's disease: a community-based study," *Brain*, vol. 123, part 11, pp. 2297–2305, 2000.
- [3] V. C. Anderson, K. J. Burchiel, P. Hogarth, J. Favre, and J. P. Hammerstad, "Pallidal vs subthalamic nucleus deep brain stimulation in Parkinson disease," *Archives of Neurology*, vol. 62, no. 4, pp. 554–560, 2005.
- [4] K. J. Burchiel, V. C. Anderson, J. Favre, and J. P. Hammerstad, "Comparison of pallidal and subthalamic nucleus deep brain stimulation for advanced Parkinson's disease: results of a randomized, blinded pilot study," *Neurosurgery*, vol. 45, no. 6, pp. 1375–1384, 1999.
- [5] C. R. Freed, P. E. Greene, R. E. Breeze et al., "Transplantation of embryonic dopamine neurons for severe Parkinson's disease," *New England Journal of Medicine*, vol. 344, no. 10, pp. 710–719, 2001.
- [6] C. W. Olanow, C. G. Goetz, J. H. Kordower et al., "A double-blind controlled trial of bilateral fetal nigral transplantation

- in Parkinson's disease," *Annals of Neurology*, vol. 54, no. 3, pp. 403–414, 2003.
- [7] C. B. Lücking, A. Dürr, V. Bonifati et al., "Association between early-onset Parkinson's disease and mutations in the parkin gene," *New England Journal of Medicine*, vol. 342, no. 21, pp. 1560–1567, 2000.
  - [8] N. L. Khan, E. Graham, P. Critchley et al., "Parkin disease: a phenotypic study of a large case series," *Brain*, vol. 126, no. 6, pp. 1279–1292, 2003.
  - [9] L. A. Smith, M. J. Jackson, M. J. Hansard, E. Maratos, and P. Jenner, "Effect of pulsatile administration of levodopa on dyskinesia induction in drug-naïve MPTP-treated common marmosets: effect of dose, frequency of administration, and brain exposure," *Movement Disorders*, vol. 18, no. 5, pp. 487–495, 2003.
  - [10] J. G. Nutt, J. A. Obeso, and F. Stocchi, "Continuous dopamine-receptor stimulation in advanced Parkinson's disease," *Trends in Neurosciences*, vol. 23, no. 10, pp. S109–S115, 2000.
  - [11] M. M. Mouradian, J. L. Juncos, G. Fabbrini, and T. N. Chase, "Motor fluctuations in Parkinson's disease: pathogenetic and therapeutic studies," *Annals of Neurology*, vol. 22, no. 4, pp. 475–479, 1987.
  - [12] D. Nyholm, H. Askmark, C. Gomes-Trolin et al., "Optimizing levodopa pharmacokinetics: intestinal infusion versus oral sustained-release tablets," *Clinical Neuropharmacology*, vol. 26, no. 3, pp. 156–163, 2003.
  - [13] E. Bezard, J. M. Brotchie, and C. E. Gross, "Pathophysiology of levodopa-induced dyskinesia: potential for new therapies," *Nature Reviews Neuroscience*, vol. 2, no. 8, pp. 577–588, 2001.
  - [14] M. Rodríguez, P. Barroso-Chinea, P. Abdala, J. Obeso, and T. González-Hernández, "Dopamine cell degeneration induced by intraventricular administration of 6-hydroxydopamine in the rat: similarities with cell loss in Parkinson's disease," *Experimental Neurology*, vol. 169, no. 1, pp. 163–181, 2001.
  - [15] M. Kuoppamäki, G. Al-Barghouthy, M. J. Jackson, L. A. Smith, N. Quinn, and P. Jenner, "L-dopa dose and the duration and severity of dyskinesia in primed MPTP-treated primates," *Journal of Neural Transmission*, vol. 114, no. 9, pp. 1147–1153, 2007.
  - [16] T. Wichmann and M. R. DeLong, "Pathophysiology of Parkinson's disease: the MPTP primate model of the human disorder," *Annals of the New York Academy of Sciences*, vol. 991, pp. 199–213, 2003.
  - [17] I. J. Mitchell, S. Boyce, M. A. Sambrook, and A. R. Crossman, "A 2-deoxyglucose study of the effects of dopamine agonists on the Parkinsonian primate brain. Implications for the neural mechanisms that mediate dopamine agonist-induced dyskinesia," *Brain*, vol. 115, part 3, pp. 809–824, 1992.
  - [18] L. V. Laitinen, A. T. Bergenheim, and M. I. Hariz, "Ventral posterolateral pallidotomy can abolish all parkinsonian symptoms," *Stereotactic and Functional Neurosurgery*, vol. 58, no. 1–4, pp. 14–21, 1992.
  - [19] F. Alonso-Frech, I. Zamarbide, M. Alegre et al., "Slow oscillatory activity and levodopa-induced dyskinesias in Parkinson's disease," *Brain*, vol. 129, no. 7, pp. 1748–1757, 2006.
  - [20] J. Lee, W. M. Zhu, D. Stanic et al., "Sprouting of dopamine terminals and altered dopamine release and uptake in Parkinsonian dyskinesia," *Brain*, vol. 131, no. 6, pp. 1574–1587, 2008.
  - [21] E. Melamed, F. Hefti, and D. J. Pettibone, "Aromatic L-amino acid decarboxylase in rat corpus striatum: Implications for action of L-dopa in parkinsonism," *Neurology*, vol. 31, no. 6, pp. 651–655, 1981.
  - [22] K. Y. Ng, T. N. Chase, R. W. Colburn, and I. J. Kopin, "L-Dopa-induced release of cerebral monoamines," *Science*, vol. 170, no. 3953, pp. 76–77, 1970.
  - [23] M. Carta, T. Carlsson, A. Muñoz, D. Kirik, and A. Björklund, "Involvement of the serotonin system in L-dopa-induced dyskinesias," *Parkinsonism and Related Disorders*, vol. 14, supplement 2, pp. S154–S158, 2008.
  - [24] M. Carta, T. Carlsson, A. Muñoz, D. Kirik, and A. Björklund, "Serotonin-dopamine interaction in the induction and maintenance of L-DOPA-induced dyskinesias," *Progress in Brain Research*, vol. 172, pp. 465–478, 2008.
  - [25] H. Tanaka, K. Kannari, T. Maeda, M. Tomiyama, T. Suda, and M. Matsunaga, "Role of serotonergic neuron in L-DOPA-derived extracellular dopamine in the striatum of 6-OHDA-lesioned rats," *Neuroreport*, vol. 10, no. 3, pp. 631–634, 1999.
  - [26] H. S. Lindgren, D. R. Andersson, S. Lagerkvist, H. Nissbrandt, and M. A. Cenci, "L-DOPA-induced dopamine efflux in the striatum and the substantia nigra in a rat model of Parkinson's disease: temporal and quantitative relationship to the expression of dyskinesia," *Journal of Neurochemistry*, vol. 112, no. 6, pp. 1465–1476, 2010.
  - [27] M. Carta, T. Carlsson, D. Kirik, and A. Björklund, "Dopamine released from 5-HT terminals is the cause of L-DOPA-induced dyskinesia in parkinsonian rats," *Brain*, vol. 130, no. 7, pp. 1819–1833, 2007.
  - [28] M. A. Cenci and M. Lundblad, "Post-versus presynaptic plasticity in L-DOPA-induced dyskinesia," *Journal of Neurochemistry*, vol. 99, no. 2, pp. 381–392, 2006.
  - [29] D. Rylander, M. Parent, S. S. O'Sullivan et al., "Maladaptive plasticity of serotonin axon terminals in levodopa-induced dyskinesia," *Annals of Neurology*, vol. 68, no. 5, pp. 619–628, 2010.
  - [30] T. Lee, P. Seeman, A. Rajput, I. J. Farley, and O. Hornykiewicz, "Receptor basis for dopaminergic supersensitivity in Parkinson's disease," *Nature*, vol. 273, no. 5657, pp. 59–61, 1978.
  - [31] H. Shinotoh, K. Hirayama, and Y. Tateno, "Dopamine D1 and D2 receptors in Parkinson's disease and striatonigral degeneration determined by PET," *Advances in Neurology*, vol. 60, pp. 488–493, 1993.
  - [32] I. Aubert, C. Guigoni, K. Häkansson et al., "Increased D<sub>1</sub> dopamine receptor signaling in levodopa-induced dyskinesia," *Annals of Neurology*, vol. 57, no. 1, pp. 17–26, 2005.
  - [33] C. Guigoni and E. Bezard, "Involvement of canonical and non-canonical D1 dopamine receptor signalling pathways in L-dopa-induced dyskinesia," *Parkinsonism and Related Disorders*, vol. 15, supplement 3, pp. S64–S67, 2009.
  - [34] A. Berthet, G. Porras, E. Doudnikoff et al., "Pharmacological analysis demonstrates dramatic alteration of D<sub>1</sub> dopamine receptor neuronal distribution in the rat analog of L-DOPA-induced dyskinesia," *Journal of Neuroscience*, vol. 29, no. 15, pp. 4829–4835, 2009.
  - [35] C. Guigoni, E. Doudnikoff, Q. Li, B. Bloch, and E. Bezard, "Altered D<sub>1</sub> dopamine receptor trafficking in parkinsonian and dyskinetic non-human primates," *Neurobiology of Disease*, vol. 26, no. 2, pp. 452–463, 2007.
  - [36] E. Bézard, S. Ferry, U. Mach et al., "Attenuation of levodopa-induced dyskinesia by normalizing dopamine D<sub>3</sub> receptor function," *Nature Medicine*, vol. 9, no. 6, pp. 762–767, 2003.
  - [37] N. P. Visanji, S. H. Fox, T. Johnston, G. Reyes, M. J. Millan, and J. M. Brotchie, "Dopamine D<sub>3</sub> receptor stimulation underlies the development of L-DOPA-induced dyskinesia in animal models of Parkinson's disease," *Neurobiology of Disease*, vol. 35, no. 2, pp. 184–192, 2009.

- [38] R. A. Hauser, O. Rascol, A. D. Korczyn et al., "Ten-year follow-up of Parkinson's disease patients randomized to initial therapy with ropinirole or levodopa," *Movement Disorders*, vol. 22, no. 16, pp. 2409–2417, 2007.
- [39] P. Calabresi, M. di Filippo, V. Ghiglieri, and B. Picconi, "Molecular mechanism underlying levodopa-induced dyskinesia," *Movement Disorders*, vol. 23, supplement 3, pp. S570–S579, 2008.
- [40] M. di Filippo, B. Picconi, M. Tantucci et al., "Short-term and long-term plasticity at corticostriatal synapses: implications for learning and memory," *Behavioural Brain Research*, vol. 199, no. 1, pp. 108–118, 2009.
- [41] G. Linazasoro, "New ideas on the origin of L-dopa-induced dyskinesias: age, genes and neural plasticity," *Trends in Pharmacological Sciences*, vol. 26, no. 8, pp. 391–397, 2005.
- [42] B. Picconi, D. Centonze, K. Håkansson et al., "Loss of bidirectional striatal synaptic plasticity in L-DOPA-induced dyskinesia," *Nature Neuroscience*, vol. 6, no. 5, pp. 501–506, 2003.
- [43] P. Calabresi, F. Galletti, E. Saggese, V. Ghiglieri, and B. Picconi, "Neuronal networks and synaptic plasticity in Parkinson's disease: beyond motor deficits," *Parkinsonism and Related Disorders*, vol. 13, supplement 3, pp. S259–S262, 2007.
- [44] T. N. Chase, "Striatal plasticity and extrapyramidal motor dysfunction," *Parkinsonism and Related Disorders*, vol. 10, no. 5, pp. 305–313, 2004.
- [45] Y. Ueki, T. Mima, M. A. Kotb et al., "Altered plasticity of the human motor cortex in Parkinson's disease," *Annals of Neurology*, vol. 59, no. 1, pp. 60–71, 2006.
- [46] F. Morgante, A. J. Espay, C. Gunraj, A. E. Lang, and R. Chen, "Motor cortex plasticity in Parkinson's disease and levodopa-induced dyskinesias," *Brain*, vol. 129, no. 4, pp. 1059–1069, 2006.
- [47] B. Picconi, V. Paillé, V. Ghiglieri et al., "L-DOPA dosage is critically involved in dyskinesia via loss of synaptic depotentiation," *Neurobiology of Disease*, vol. 29, no. 2, pp. 327–335, 2008.
- [48] T. N. Chase, F. Bibbiani, and J. D. Oh, "Striatal glutamatergic mechanisms and extrapyramidal movement disorders," *Neurotoxicity Research*, vol. 5, no. 1-2, pp. 139–145, 2003.
- [49] M. A. Cenci, C. S. Lee, and A. Björklund, "L-DOPA-induced dyskinesia in the rat is associated with striatal overexpression of prodynorphin- and glutamic acid decarboxylase mRNA," *European Journal of Neuroscience*, vol. 10, no. 8, pp. 2694–2706, 1998.
- [50] E. Luginger, G. K. Wenning, S. Bösch, and W. Poewe, "Beneficial effects of amantadine on L-dopa-induced dyskinesias in Parkinson's disease," *Movement Disorders*, vol. 15, pp. 873–878, 2000.
- [51] L. Verhagen Metman, P. del Dotto, P. van den Munckhof, J. Fang, M. M. Mouradian, and T. N. Chase, "Amantadine as treatment for dyskinesias and motor fluctuations in Parkinson's disease," *Neurology*, vol. 50, no. 5, pp. 1323–1326, 1998.
- [52] L. V. Metman, P. del Dotto, K. LePoole, S. Konitsiotis, J. Fang, and T. N. Chase, "Amantadine for levodopa-induced dyskinesias: a 1-year follow-up study," *Archives of Neurology*, vol. 56, no. 11, pp. 1383–1386, 1999.
- [53] J. H. Kordower, T. B. Freeman, E. Y. Chen et al., "Fetal nigral grafts survive and mediate clinical benefit in a patient with Parkinson's disease," *Movement Disorders*, vol. 13, no. 3, pp. 383–393, 1998.
- [54] O. Lindvall, "Update on fetal transplantation: the Swedish experience," *Movement Disorders*, vol. 13, supplement 1, pp. 83–87, 1998.
- [55] P. Hagell, P. Piccini, A. Björklund et al., "Dyskinesias following neural transplantation in Parkinson's disease," *Nature Neuroscience*, vol. 5, no. 7, pp. 627–628, 2002.
- [56] C. W. Olanow, J. M. Gracies, C. G. Goetz et al., "Clinical pattern and risk factors for dyskinesias following fetal nigral transplantation in parkinson's disease: a double blind video-based analysis," *Movement Disorders*, vol. 24, no. 3, pp. 336–343, 2009.
- [57] P. Piccini, N. Pavese, P. Hagell et al., "Factors affecting the clinical outcome after neural transplantation in Parkinson's disease," *Brain*, vol. 128, no. 12, pp. 2977–2986, 2005.
- [58] R. A. Hauser, T. B. Freeman, B. J. Snow et al., "Long-term evaluation of bilateral fetal nigral transplantation in Parkinson disease," *Archives of Neurology*, vol. 56, no. 2, pp. 179–187, 1999.
- [59] Y. Ma, A. Feigin, V. Dhawan et al., "Dyskinesia after fetal cell transplantation for parkinsonism: a PET study," *Annals of Neurology*, vol. 52, no. 5, pp. 628–634, 2002.
- [60] S. Boyce, N. M. J. Rupniak, M. J. Steventon, and S. D. Iversen, "Nigrostriatal damage is required for induction of dyskinesias by L-DOPA in squirrel monkeys," *Clinical Neuropharmacology*, vol. 13, no. 5, pp. 448–458, 1990.
- [61] C. S. Lee, M. A. Cenci, M. Schulzer, and A. Björklund, "Embryonic ventral mesencephalic grafts improve levodopa-induced dyskinesia in a rat model of Parkinson's disease," *Brain*, vol. 123, part 7, pp. 1365–1379, 2000.
- [62] J. L. Hudson, A. Hoffman, I. Strömberg, B. J. Hoffer, and J. W. Moorhead, "Allogeneic grafts of fetal dopamine neurons: behavioral indices of immunological interactions," *Neuroscience Letters*, vol. 171, no. 1-2, pp. 32–36, 1994.
- [63] M. Shinoda, J. L. Hudson, I. Stromberg, B. J. Hoffer, J. W. Moorhead, and L. Olson, "Allogeneic grafts of fetal dopamine neurons: immunological reactions following active and adoptive immunizations," *Brain Research*, vol. 680, no. 1-2, pp. 180–195, 1995.
- [64] P. Hagell and M. A. Cenci, "Dyskinesias and dopamine cell replacement in Parkinson's disease: a clinical perspective," *Brain Research Bulletin*, vol. 68, no. 1-2, pp. 4–15, 2005.
- [65] M. Politis, W. H. Oertel, K. Wu et al., "Graft-induced dyskinesias in Parkinson's disease: high striatal serotonin/dopamine transporter ratio," *Movement Disorders*, vol. 26, no. 11, pp. 1997–2003, 2011.
- [66] T. Carlsson, M. Carta, C. Winkler, A. Björklund, and D. Kirik, "Serotonin neuron transplants exacerbate L-DOPA-induced dyskinesias in a rat model of Parkinson's disease," *Journal of Neuroscience*, vol. 27, no. 30, pp. 8011–8022, 2007.
- [67] T. Carlsson, M. Carta, A. Muñoz et al., "Impact of grafted serotonin and dopamine neurons on development of L-DOPA-induced dyskinesias in parkinsonian rats is determined by the extent of dopamine neuron degeneration," *Brain*, vol. 132, no. 2, pp. 319–335, 2009.
- [68] M. Carta, T. Carlsson, A. Muñoz, D. Kirik, and A. Björklund, "Role of serotonin neurons in the induction of levodopa—and graft—induced dyskinesias in Parkinson's disease," *Movement Disorders*, vol. 25, 1, pp. S174–S179, 2010.
- [69] J. García, T. Carlsson, M. Döbrössy, G. Nikkha, and C. Winkler, "Impact of dopamine to serotonin cell ratio in transplants on behavioral recovery and L-DOPA-induced dyskinesia," *Neurobiology of Disease*, vol. 43, no. 3, pp. 576–587, 2011.
- [70] E. L. Lane, C. Winkler, P. Brundin, and M. A. Cenci, "The impact of graft size on the development of dyskinesia following intrastratial grafting of embryonic dopamine neurons in the rat," *Neurobiology of Disease*, vol. 22, no. 2, pp. 334–345, 2006.

- [71] A. Vinuela, P. J. Hallett, C. Reske-Nielsen et al., "Implanted reuptake-deficient or wild-type dopaminergic neurons improve on L-dopa dyskinesias without OFF-dyskinesias in a rat model of Parkinson's disease," *Brain*, vol. 131, no. 12, pp. 3361–3379, 2008.
- [72] K. Kannari, H. Shen, A. Arai, M. Tomiyama, and M. Baba, "Reuptake of L-DOPA-derived extracellular dopamine in the striatum with dopaminergic denervation via serotonin transporters," *Neuroscience Letters*, vol. 402, no. 1-2, pp. 62–65, 2006.
- [73] H. Sershen, A. Hashim, and A. Lajtha, "Serotonin-mediated striatal dopamine release involves the dopamine uptake site and the serotonin receptor," *Brain Research Bulletin*, vol. 53, no. 3, pp. 353–357, 2000.
- [74] W. J. Marks Jr., J. L. Ostrem, L. Verhagen et al., "Safety and tolerability of intraputamenal delivery of CERE-120 (adeno-associated virus serotype 2-neurturin) to patients with idiopathic Parkinson's disease: an open-label, phase I trial," *The Lancet Neurology*, vol. 7, no. 5, pp. 400–408, 2008.
- [75] C. W. Christine, P. A. Starr, P. S. Larson et al., "Safety and tolerability of putamenal AADC gene therapy for Parkinson disease," *Neurology*, vol. 73, no. 20, pp. 1662–1669, 2009.
- [76] B. A. Horger, M. C. Nishimura, M. P. Armanini et al., "Neurturin exerts potent actions on survival and function of midbrain dopaminergic neurons," *Journal of Neuroscience*, vol. 18, no. 13, pp. 4929–4937, 1998.
- [77] J. H. Kordower, C. D. Herzog, B. Dass et al., "Delivery of neurturin by AAV2 (CERE-120)-mediated gene transfer provides structural and functional neuroprotection and neurorestoration in MPTP-treated monkeys," *Annals of Neurology*, vol. 60, no. 6, pp. 706–715, 2006.
- [78] W. J. Marks Jr., R. T. Bartus, J. Siffert et al., "Gene delivery of AAV2-neurturin for Parkinson's disease: a double-blind, randomised, controlled trial," *The Lancet Neurology*, vol. 9, no. 12, pp. 1164–1172, 2010.
- [79] J. Luo, M. G. Kaplitt, H. L. Fitzsimons et al., "Subthalamic GAD gene therapy in a Parkinson's disease rat model," *Science*, vol. 298, no. 5592, pp. 425–429, 2002.
- [80] P. A. LeWitt, A. R. Rezai, M. A. Leehey et al., "AAV2-GAD gene therapy for advanced Parkinson's disease: a double-blind, sham-surgery controlled, randomised trial," *The Lancet Neurology*, vol. 10, no. 4, pp. 309–319, 2011.
- [81] K. S. Bankiewicz, M. Daadi, P. Pivrotto et al., "Focal striatal dopamine may potentiate dyskinesias in parkinsonian monkeys," *Experimental Neurology*, vol. 197, no. 2, pp. 363–372, 2006.
- [82] M. Sun, G. R. Zhang, L. Kong et al., "Correction of a rat model of Parkinson's disease by coexpression of tyrosine hydroxylase and aromatic amino acid decarboxylase from a helper virus-free herpes simplex virus type 1 vector," *Human Gene Therapy*, vol. 14, no. 5, pp. 415–424, 2003.
- [83] T. Björklund, T. Carlsson, E. A. Cederfjäll, M. Carta, and D. Kirik, "Optimized adeno-associated viral vector-mediated striatal DOPA delivery restores sensorimotor function and prevents dyskinesias in a model of advanced Parkinson's disease," *Brain*, vol. 133, no. 2, pp. 496–511, 2010.
- [84] V. Bonifati, E. Fabrizio, R. Cipriani, N. Vanacore, and G. Meco, "Buspirone in levodopa-induced dyskinesias," *Clinical Neuropharmacology*, vol. 17, no. 1, pp. 73–82, 1994.
- [85] C. W. Olanow, P. Damier, C. G. Goetz et al., "Multicenter, open-label, trial of sarizotan in Parkinson disease patients with levodopa-induced dyskinesias (the SPLENDID study)," *Clinical Neuropharmacology*, vol. 27, no. 2, pp. 58–62, 2004.
- [86] C. G. Goetz, P. Damier, C. Hicking et al., "Sarizotan as a treatment for dyskinesias in Parkinson's disease: a double-blind placebo-controlled trial," *Movement Disorders*, vol. 22, no. 2, pp. 179–186, 2007.
- [87] K. B. Dupre, K. L. Eskow, G. Negron, and C. Bishop, "The differential effects of 5-HT<sub>1A</sub> receptor stimulation on dopamine receptor-mediated abnormal involuntary movements and rotations in the primed hemiparkinsonian rat," *Brain Research*, vol. 1158, no. 1, pp. 135–143, 2007.
- [88] D. Berg, J. Godau, C. Trenkwalder et al., "AFQ056 treatment of levodopa-induced dyskinesias: results of 2 randomized controlled trials," *Movement Disorders*, vol. 26, no. 7, pp. 1243–1250, 2011.
- [89] F. Bibbiani, J. D. Oh, J. P. Petzer et al., "A<sub>2A</sub> antagonist prevents dopamine agonist-induced motor complications in animal models of Parkinson's disease," *Experimental Neurology*, vol. 184, no. 1, pp. 285–294, 2003.
- [90] S. Fredduzzi, R. Moratalla, A. Monopoli et al., "Persistent behavioral sensitization to chronic L-DOPA requires A<sub>2A</sub> adenosine receptors," *Journal of Neuroscience*, vol. 22, no. 3, pp. 1054–1062, 2002.
- [91] R. A. Hauser, L. M. Shulman, J. M. Trugman et al., "Study of istradefylline in patients with Parkinson's disease on levodopa with motor fluctuations," *Movement Disorders*, vol. 23, no. 15, pp. 2177–2185, 2008.
- [92] A. J. Manson, E. Iakovidou, and A. J. Lees, "Idazoxan is ineffective for levodopa-induced dyskinesias in Parkinson's disease," *Movement Disorders*, vol. 15, no. 2, pp. 336–337, 2000.
- [93] T. N. Chase, "Alpha-2A adrenergic antagonists effects in advanced Parkinson's disease. Rome, Italy: Movement Disorders Congress," *Movement Disorders*, vol. 15, no. 2, pp. 336–337, 2000.
- [94] A. Berthet and E. Bezard, "Dopamine receptors and L-dopa-induced dyskinesia," *Parkinsonism and Related Disorders*, vol. 15, supplement 4, pp. S8–S12, 2009.
- [95] T. Foltynie, C. Brayne, and R. A. Barker, "The heterogeneity of idiopathic Parkinson's disease," *Journal of Neurology*, vol. 249, no. 2, pp. 138–145, 2002.
- [96] I. A. Prescott, J. O. Dostrovsky, E. Moro, M. Hodaie, A. M. Lozano, and W. D. Hutchison, "Levodopa enhances synaptic plasticity in the substantia nigra pars reticulata of Parkinson's disease patients," *Brain*, vol. 132, no. 2, pp. 309–318, 2009.
- [97] P. Calabresi, N. B. Mercuri, and M. di Filippo, "Synaptic plasticity, dopamine and Parkinson's disease: one step ahead," *Brain*, vol. 132, no. 2, pp. 285–287, 2009.

## Review Article

# Corticostriatal Plastic Changes in Experimental L-DOPA-Induced Dyskinesia

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In Parkinson's disease (PD), alteration of dopamine- (DA-) dependent striatal functions and pulsatile stimulation of DA receptors caused by the discontinuous administration of levodopa (L-DOPA) lead to a complex cascade of events affecting the postsynaptic striatal neurons that might account for the appearance of L-DOPA-induced dyskinesia (LID). Experimental models of LID have been widely used and extensively characterized in rodents and electrophysiological studies provided remarkable insights into the inner mechanisms underlying L-DOPA-induced corticostriatal plastic changes. Here we provide an overview of recent findings that represent a further step into the comprehension of mechanisms underlying maladaptive changes of basal ganglia functions in response to L-DOPA and associated to development of LID.

## 1. Introduction

In Parkinson's disease (PD), degeneration of dopaminergic neurons of the *substantia nigra* causes critical reduction in dopamine (DA) levels in the target areas. The subsequent abnormal DA receptor stimulation exerts its main effects in the striatum, the principal input structure of basal ganglia-thalamo-cortical network, producing changes in input integration that lead to imbalance between direct and indirect striatofugal pathways and dysfunctional changes in basal ganglia output.

Impairment in the induction of the two forms of striatal synaptic plasticity, the long-term depression (LTD) and the long-term potentiation (LTP), has been found to correlate with DA depletion and onset of symptoms in experimental models of PD. DA depletion initially affects LTP and then, when symptoms are fully manifested, also LTD is impaired [1].

The resulting motor symptoms are effectively treated with a replacement therapy that uses the DA precursor L-3,4-dihydroxyphenylalanine (L-DOPA) to rescue striatal DA-dependent neuronal activity. However, L-DOPA treatment

does not arrest disease progression and, with time, neuronal degeneration advances and leads to the emergence of a complex pattern of alterations that involves other basal ganglia nuclei, causing symptoms that are refractory to conventional therapy. In addition, the initial excellent antiparkinsonian effects of L-DOPA are lost in the long run, and the route of drug administration utilized in the clinical practice leads to a pulsatile stimulation of DA receptors that causes a broader neuronal destabilization. Therefore, new motor complications unavoidably develop, resulting in L-DOPA-induced dyskinesia (LID), a very disabling long-term side effect of L-DOPA therapy associated with the loss of corticostriatal bidirectional plasticity [2].

The expression of an aberrant plasticity following chronic L-DOPA treatment has been also demonstrated in PD patients [3–5], further supporting the notion that a treatment with a drug able to ameliorate disease symptoms can be associated with the recovery of a selective form of synaptic plasticity.

This review provides an overview of papers that contributed to characterize the plastic changes occurring at striatal synapses in experimental models of LID. After a

description of the main forms of DA-dependent synaptic plasticity at glutamatergic corticostriatal synapses, we will introduce seminal studies focusing on the plastic changes observed in dyskinetic models. We will then review the most recent papers that further explored mechanisms underlying L-DOPA-induced changes in experimental PD models and discuss recent findings that, in our opinion, represent new promising avenues to future electrophysiological studies on dyskinetic animals.

## 2. DA-Dependent Synaptic Plasticity at Corticostriatal Synapses

At corticostriatal synapses, repetitive cortical activation can induce either LTD or LTP in the striatal medium spiny neurons (MSNs), depending on the level of membrane depolarization, the subtype of glutamate receptor activated [6–8], and the interneuronal subtypes involved in the induction process [9]. Unique characteristic of striatal neurons is that DA critically regulates both the induction and the maintenance of neuroplasticity via DA D<sub>1</sub>-like (D<sub>1</sub>) and D<sub>2</sub>-like (D<sub>2</sub>) receptors activation. Specifically, DA acting on D<sub>1</sub> receptors cooperates to the induction of LTP, whereas activation of both D<sub>1</sub> and D<sub>2</sub> receptors is required for LTD [2, 10, 11].

Electrophysiological studies in corticostriatal slices from 6-hydroxydopamine- (6-OHDA-) lesioned parkinsonian rats have shed light on the pivotal role that DA exerts in modulating glutamatergic transmission and synaptic plasticity within the striatum [12].

A complete DA denervation abolishes both forms of corticostriatal plasticity [11, 13] that can be restored by treatment with either DA receptor agonists or the DA precursor L-DOPA [2, 11, 14].

We have recently shown that distinct degrees of DA denervation influence the two forms of plasticity in different ways, as full DA denervation blocks the induction of both LTP and LTD, while partial DA depletion allows LTP induction but selectively alters its maintenance, leaving LTD induction unaffected [1].

A third form of striatal plasticity, distinct from LTD, called synaptic depotentiation, results from the reversal of an established LTP by the application of a low-frequency stimulation (LFS) of corticostriatal fibers [2, 15]. This form of plasticity critically relies on glutamatergic N-methyl-D-aspartate (NMDA) receptor activation [16] and striatal endogenous tone of acetylcholine [17]. During LTP, protein kinase A (PKA), a downstream effector of DA D<sub>1</sub> receptors, phosphorylates and activates DA- and cAMP-regulated phosphoprotein of 32 kDa (DARPP-32), a potent inhibitor of protein phosphatase 1 (PP-1). PP-1 dephosphorylates several downstream targets of PKA, thereby amplifying behavioral responses produced by activation of cAMP signalling [18–20], and it is necessary for depotentiation, as this form of plasticity is blocked by application of PP-1 inhibitors.

DA and glutamate receptors functional interaction in the striatum has been shown to regulate locomotion, positive reinforcement, attention, and working memory.

In particular, activation of D<sub>1</sub> receptors is needed for the correct integration of cortical glutamatergic signals to the striatum [21]. In striatal MSNs, D<sub>1</sub> receptors are located within dendritic spines, where they colocalize with NMDA receptors [22, 23] regulate the rapid trafficking of NMDA receptor subunits [24] and the potentiation of NMDA responses [25], leading to activity-dependent adaptive changes [10] and also to the activation of excitotoxic pathways. Among the signalling cascades regulating D<sub>1</sub> receptor-dependent enhancement of NMDA responses in the striatum, the most important involves PKA- and DARPP-32-regulated phosphorylation of NMDA receptor NR1 subunits [26].

D<sub>1</sub> DA receptor stimulation also enhances phosphorylation of the alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) glutamate receptor subunit GluR1 at the PKA site, increases surface expression of AMPA receptors, and facilitates their synaptic insertion in several brain areas [27, 28].

Besides the concurrent activation of glutamatergic and dopaminergic receptors, activity-dependent plasticity of glutamatergic synapses at MSNs is also modulated by other signalling pathways like endocannabinoids, adenosine (presynaptically), and metabotropic glutamate (pre- and postsynaptic) receptors [29] and by striatal interneurons [30, 31], which represent a minority of total striatal population but play a crucial role in the modulation of basal ganglia function, contributing to the processing of corticostriatal information [9, 13, 32]. In particular, two interneuronal subtypes have been suggested to play a critical role in the pathogenesis of LID: the large-aspiny cholinergic interneurons and the nitric-oxide-synthase- (NOS-) positive interneurons.

The cholinergic interneurons, which represent the main source of acetylcholine within the striatum [33], play a permissive role in corticostriatal synaptic plasticity by modulating the striatal cholinergic tone [9, 34]. These interneurons respond to cortical stimulations with long lasting changes of synaptic efficacy [17, 35] and are important sites of interaction among DA, adenosine, and endocannabinoid receptor signalling systems [36], further supporting the idea that cholinergic interneuronal activity contributes to striatal-dependent learning and motor habit formation.

The NOS-immunoreactive neurons represent, along with the cholinergic interneurons, the other interneuronal subtype that plays an important role in the induction of LTD [13, 31, 34]. These interneurons express mRNA encoding for ionotropic glutamate receptors that appear to be coupled to nitric oxide production [37–40]. Nitric oxide activates soluble guanylyl cyclase (sGC), which in turn induces increases of intracellular cyclic guanosine monophosphate (cGMP) levels to activate the protein kinase G (PKG) [41–43], whose levels are regulated by the action of phosphodiesterases (PDEs), a family of enzymes responsible for the conversion of cGMP to GMP. Accordingly, pharmacological LTD can occur in MSNs following the application of phosphodiesterases inhibitors [13], as a consequence of increased cGMP levels. In fact, the amount of this nucleotide is crucial for the activity of PKG and DARPP-32, which in turn control the phosphorylation

of AMPA receptor, a main player in the induction of LTD [10].

In summary, the integrative action exerted by striatal projection neurons on the converging information arising from the cortex, the nigral DA neurons and the striatal interneurons, shapes the activity of neurons throughout the entire basal ganglia circuitry.

### 3. L-DOPA-Induced Plastic Changes at Glutamatergic Synapses

The effects of L-DOPA administration in the DA-depleted striatum have been extensively studied in experimental models of LID, leading to the concept that a combination of presynaptic and postsynaptic maladaptive changes is needed for the parkinsonian animals to develop dyskinesia [3, 44, 45].

During progressive degeneration of nigrostriatal terminals, sprouting of DA terminals and reduced DA uptake contribute to preserve DA striatal levels [46], and increase in glutamate transmission is observed in corticostriatal pathway [47–51] as well as in basal ganglia output nuclei [52, 53].

However, such presynaptic adaptive changes together with changed presynaptic and postsynaptic DA receptor sensitivity and density lead to an altered substrate in which L-DOPA exerts its actions. Thus, initially, L-DOPA is converted into DA, stored in synaptic vesicles, and released by surviving DA-releasing terminals. However, when degeneration advances, DA catabolism and uptake are reduced and decarboxylation of L-DOPA to DA and release occur in non-dopaminergic cells [54, 55], causing a failure in the buffering of DA levels.

The consequent large fluctuations in extracellular DA concentrations, mainly relying on the drug-dosing cycle, contribute to the establishment of further morphological and functional changes at both pre- and postsynaptic levels.

During chronic treatment with L-DOPA, several postsynaptic pathways downstream DA and glutamate receptors activation are progressively dysregulated, causing a loss of control of phosphorylation cascades with increase of phosphorylated striatal substrates such as NMDA receptor subunits [56, 57], AMPA receptor subunits [58], and extracellular signal-regulated kinase (ERK)1/2 [44, 58–60]. One crucial pathway that has been extensively investigated is the signalling activated by D<sub>1</sub> receptor stimulation [61]. In the DA-depleted striatum, in fact, chronic L-DOPA treatment, through stimulation of sensitized D<sub>1</sub> receptors causes hyperactivation of PKA and increased striatal phosphorylation of DARPP-32 at the threonine-34 residue [58, 62]. As above mentioned, this protein plays a pivotal role in the synaptic alterations caused by unphysiological stimulation of DA D<sub>1</sub> receptors. In fact, DARPP-32 is a potent inhibitor of PP-1 activity, which in turn is necessary to depotentiate the synapse.

A critical link between abnormal involuntary movements (AIMs), resembling human dyskinesia, and loss of bidirectional synaptic plasticity at corticostriatal synapses of dyskinetic rats has been firstly provided by our group

[2, 63]. In the unilateral 6-OHDA model of PD, chronic treatment with either high or low doses of L-DOPA is able to restore LTP expression. However, in a consistent number of treated animals, the corticostriatal glutamatergic signalling undergoes further adaptive changes and AIMs develop [2, 64, 65]. Hyperphosphorylation of DARPP-32 at the threonine-34 residue occurs selectively in animals developing dyskinetic behavior and is associated to the loss of capability to depotentiate the corticostriatal synapse [2]. Moreover, in dyskinetic animals, prolonged L-DOPA treatment remarkably reduces synaptic D<sub>1</sub>/NMDA receptor complexes without changing their interaction [23]. However, further complex molecular alterations take place at glutamatergic synapse that are strictly correlated to abnormal synaptic plasticity and motor behavior in L-DOPA-treated dyskinetic rats [2, 16]. Specifically, levels of NR2A subunit are higher in dyskinetic animals compared to nondyskinetic ones, and this effect is paralleled by decreased levels of NR2B subunit, which are found increased in extrasynaptic sites [16]. Such redistribution of NMDA receptor subunits is associated with alterations in the binding of NMDA receptor subunits with their cargo proteins, in particular, SAP-97 and SAP-102 [16]. Impairment of the physiological trafficking of NMDA receptor subunits from the reticulum toward the postsynaptic density may, therefore, determine the enhancement of NMDA receptor signalling in dyskinesia.

Accordingly, pharmacological manipulation aimed at reducing synaptic localization of NR<sub>2B</sub>, and consequently increasing NR<sub>2A</sub>/NR<sub>2B</sub> ratio at synaptic sites, causes in nondyskinetic subjects a worsening of motor symptoms with appearance of dyskinetic behaviours [16]. Intracerebral administration of a cell-permeable peptide (TAT2B), able to alter the NR<sub>2B</sub> synaptic localization by perturbing its binding with scaffolding proteins, causes loss of depotentiation that correlated with AIMs in nondyskinetic animals [16].

Taken together, these findings support the notion that abnormal activation of PKA and concomitant hyperphosphorylation of DARPP-32 observed in experimental models of LID are two of the main causes of changes in the state of phosphorylation state of target effector proteins, with consequent profound repercussion on the excitability and plasticity of striatal MSNs.

### 4. Novel Insights into L-DOPA-Induced Changes in Corticostriatal Synaptic Plasticity

Three new studies have investigated further on the mechanisms underlying the loss of synaptic scaling down at corticostriatal synapses.

Gardoni and coworkers have recently shown that pharmacological manipulations interfering with the interactions between NMDA receptor subunits and their scaffolding proteins, responsible for their trafficking and correct assembly at synaptic membranes, prevents the unbalance of NR<sub>2A</sub>/NR<sub>2B</sub> subunit ratio by reducing the synaptic localization of NR<sub>2A</sub> subunit. Systemic coadministration of the cell-permeable

peptide TAT2A and L-DOPA reduces the percentage of animals developing dyskinesia [66]. However, once the AIMs are established, the administration of TAT2A fails to reduce incidence of dyskinesia, indicating that altered NMDA receptor composition has a critical role in initiating the dyskinetic phenotype. Moreover, these data support the concept that molecular disturbances of the glutamatergic synapse, initially caused by DA denervation, create a pathological substrate that induce and maintain the overworking synapse at an altered steady state that triggers the development of LID [2, 16].

A further advance in the characterization of bidirectional synaptic plasticity following L-DOPA therapy has been made in a recent study conducted by our group. Based on the evidence that striatal cGMP signalling is decreased in dyskinetic animals [67], we explored the possibility that LTD, which strictly relies on the nitric oxide-dependent activation of PKG, was altered following L-DOPA treatment. We found that MSNs recorded from L-DOPA-treated dyskinetic parkinsonian rats do not express activity-dependent LTD. Increase of cGMP levels by PDEs inhibitors leads to the activation of PKG, mimicking the action of nitric oxide released from NOS-positive neurons that represents a critical factor for LTD induction following HFS [13]. Accordingly, application of a low dose of PDEs inhibitor, unable to induce *per se* a pharmacological LTD in dyskinetic parkinsonian rats, is sufficient to rescue activity-dependent LTD in these animals.

Interestingly, application of PDEs inhibitors induces pharmacological LTD in both dyskinetic and nondyskinetic rats but not in untreated parkinsonian animals, indicating that the presence of endogenous striatal DA represents a critical condition also for the induction of this form of pharmacological plasticity. Local injection of these drugs into the striatum of dyskinetic rats rescues LTD and reduces the dyskinetic response to L-DOPA [62].

This phenomenon, together with the loss of depotentiation [2], is in line with the view that LID is caused by impaired control of striatal excitatory synapses with excessive increase of glutamatergic transmission.

Accordingly, the third study by Usiello and coworkers investigated the contribution of a basal hyperglutamatergic tone in the development of dyskinesia associated to altered DA-dependent bidirectional synaptic plasticity.

Using mutant mice lacking the D-Aspartate Oxidase (Ddo) enzyme (Ddo<sup>-/-</sup> mice), showing nonphysiological high levels of the excitatory free D-amino acids D-aspartate and NMDA [68], they found that a condition of persistent hyperstimulation of glutamatergic transmission results in an aberrant striatal synaptic plasticity. In the MSNs recorded from Ddo<sup>-/-</sup> mice, similar to what observed in dyskinetic animals, LFS protocol fails to reverse the synaptic transmission levels to those preceding LTP.

When subjected to 6-OHDA lesion, Ddo<sup>-/-</sup> mice display increased sensitivity to L-DOPA and early onset of dyskinetic behavior [69] further supporting the concept that increased glutamatergic release is a critical risk factor to develop LID.

## 5. New Promising Avenues to Further Investigate L-DOPA-Induced Corticostriatal Plastic Changes

In the recent past, new molecular targets for LID have been explored that may play a critical role in the synaptic alterations underlying plastic changes in the DA-depleted striatum exposed to long-term L-DOPA. An important contribution to the understanding of mechanisms involved in the development of dyskinesia has been provided by the evidence that not only ERK but also its downstream targets, including molecules involved in the regulation of protein translation and gene transcription [60, 70], are entailed in the dysregulation of phosphorylation cascades induced by L-DOPA. The group of Fisone and coworkers has recently demonstrated that abnormal activation of ERK is associated to increased signalling of mammalian target of rapamycin complex 1 (mTORC1) via inhibitory control of tuberous sclerosis complex (TSC) 1 and 2 that, in turn, suppresses activation of Ras homolog enriched in brain (Rheb), a highly conserved member of the Ras superfamily of G-proteins, ultimately responsible for mTORC1 activity. Coadministration of L-DOPA and rapamycin, a selective allosteric inhibitor of mTOR complex, diminishes the development of LID without interfering with the therapeutic effects of L-DOPA [56]. Recently, it has been shown that besides Rheb, another small G protein, the Ras homologue enriched in striatum (Rhes), is critically involved in the pathological upregulation of mTORC1 during LID [71]. These data further strengthen the hypothesis of an involvement of mTORC1 signalling in LID, as Rhes knockout mice show reduced dyskinesia in response to L-DOPA, but the therapeutic improvement of limb motion remains unchanged. Interestingly, a role of mTORC1 in synaptic plasticity has been recently put forward [72]. Relevant to corticostriatal pathway, it has been shown that inhibition of mTORC complexes is able to block a pathological form of persistent LTP associated to increased glutamatergic signalling and neurodegeneration [73].

Taken together, these data suggest that enhanced mRNA translation, leading to abnormal local protein synthesis in the cytoplasm, may participate in the development of aberrant enhancement of synaptic strength, as observed in LID.

Another intriguing aspect that has been recently investigated is the capability of L-DOPA to exert its action through nondopaminergic systems. Indeed, as PD progresses, degeneration of nigrostriatal terminals also advances, and L-DOPA is converted in DA, stored, and released also from other cellular elements within the striatum, including the serotonin (5-HT) terminals [54, 74, 75]. This action might have both beneficial and detrimental consequences in that it allows L-DOPA to maintain DA levels in the virtual absence of dopaminergic neurons but it also enhances the non-physiological DA receptor stimulation as the feedback control of DA release is absent in the 5-HT system. This might have important implications for corticostriatal synaptic plasticity as unregulated DA transmission may induce further adaptive rearrangement of DA/glutamatergic

ionotropic receptors interactions at postsynaptic sites that would critically affect the bidirectional synaptic plasticity.

The hypothesis of the involvement of 5-HT terminals in LID has gained support from recent evidence showing that lesion of the 5-HT system by 5,7-dihydroxytryptamine [75] or pharmacological manipulation of serotonergic transmission [54, 74] significantly reduces L-DOPA-induced increase of extracellular DA levels in the striatum and abolishes dyskinetic movements in parkinsonian rats chronically treated with L-DOPA [54]. However, decrease of corticostriatal glutamate release could be another mechanism underlying additional antidyskinetic effect [76–78].

A potent synergistic interaction between 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptors in counteracting the induction of dyskinetic movements has also been demonstrated in the 1-methyl-4-phenyl 1,2,3,6-tetrahydropyridine- (MPTP-) treated macaques, in which administration of 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> agonists reduces the upregulated levels of FosB, the main postsynaptic striatal marker for LID [79, 80].

Most recently, it has been demonstrated that profound structural changes are associated to the capability of serotonergic terminals to release DA as “false transmitter.” Cenci and coworkers provided evidence that L-DOPA treatment induces the sprouting of 5-HT axon terminals (increased number of synaptic contacts between 5-HT terminals and striatal neurons) [55]. This specific morphological feature positively correlates with the severity of dyskinesia as shown by increased binding levels of the plasma membrane 5-HT transporter in both experimental models (rodents and nonhuman primates) and in PD patients subjected to L-DOPA therapy. Such increase was correlated with the dyskinetic score and paralleled by the upregulation of brain-derived neurotrophic factor (BDNF) expression [55, 81], which exerts complex functional and structural actions within the striatum.

These results are consistent with the evidence that increased concentrations of striatal BDNF are associated with LID [82] although the role of this neurotrophin in LID development is still under debate [83].

A link between BDNF and LID is also suggested by the fact that striatal BDNF is regulated by the activity of another nondopaminergic pathway involved in the development of LID, the striatal purinergic system. Indeed striatal adenosine, through A<sub>2A</sub> receptors, has been suggested to play a pivotal role in the regulation of BDNF function and levels in the brain [84, 85] and it has been also implicated in the development of LID [86].

Presynaptically, A<sub>2A</sub> receptors act to finely tune glutamate release from corticostriatal terminals and they are also present postsynaptically on striatopallidal MSNs of the indirect pathway that express DA D<sub>2</sub> receptors.

In control condition, concomitant activation of DA D<sub>2</sub> receptors and blockade of A<sub>2A</sub> adenosine receptors is able to decrease striatal glutamatergic transmission [87]. This interaction is made possible by a retrograde action of endocannabinoids released by postsynaptic MSNs and acting on CB1 cannabinoid receptors located on glutamatergic terminals [36] suggesting that the convergence of DA D<sub>2</sub> and A<sub>2A</sub> signalling systems on the endocannabinoids

pathway represents a potent feedback mechanism to control glutamatergic transmission in the striatum. While in control condition, concurrent activation of D<sub>2</sub> and blockade of A<sub>2A</sub> are necessary to reduce glutamate release via an endocannabinoid-dependent mechanism, in DA-depleted animals, D<sub>2</sub> receptor agonism alone is able to reduce glutamatergic transmission due to D<sub>2</sub> receptor sensitization. This effect could be further enhanced by A<sub>2A</sub> receptor antagonists providing a solid experimental support for the combined use of D<sub>2</sub> receptor agonists and A<sub>2A</sub> receptor antagonists in clinical settings. In fact, alterations in A<sub>2A</sub> receptor expression and signalling have been extensively observed in PD patients undergoing L-DOPA therapy and in experimental models of LID and A<sub>2A</sub> antagonists have proven to be effective in clinical and preclinical studies [86].

Notably, striatal cholinergic interneurons, coexpressing D<sub>2</sub> and A<sub>2A</sub> receptors, are also interested in this pharmacological modulation, since concomitant activation of D<sub>2</sub> DA receptors and blockade of A<sub>2A</sub> receptors reduces the firing rate of this neuronal subtype and muscarinic M<sub>1</sub> receptor antagonism blocks the D<sub>2</sub>/A<sub>2A</sub> receptor-mediated modulation of excitatory transmission in both D<sub>2</sub>- and D<sub>1</sub>-expressing MSNs [36]. These results are in agreement with previous studies showing altered acetylcholine signalling in DA-denervated striatum [88] resulting in a loss of feedback control of acetylcholine release [89]. Striatal acetylcholine levels critically determine the direction of synaptic plasticity at corticostriatal synapses with low levels of acetylcholine facilitating LTD and high levels facilitating LTP [90].

Taken together, these data suggest a strong involvement of the striatal cholinergic interneurons in LID pathogenesis. A recent paper [91] shows that in animals lacking the transcription factor Pitx3, modeling PD, chronic L-DOPA enhances baseline and DA-induced firing rate in striatal cholinergic interneurons. This effect is seen also in 6-OHDA-lesioned mice and is associated with increased phospho-ERK immunoreactivity in this neuronal population as inhibition of ERK is able to restore firing rate at control values [91]. In both the unilateral lesion and the genetic models, chronic L-DOPA caused development of LID that was attenuated by administration of dicyclomine, a muscarinic antagonist, without affecting L-DOPA's beneficial antiparkinsonian action.

These findings provide new lines of evidence that L-DOPA exerts its widespread action at multiple levels in the functional organization of the striatum (Figure 1). However, a clear-cut definition of a scenario comprising the various maladaptive changes is made difficult by the fact that striatal response to DA-denervation and subsequent DA replacement may vary between the two distinct populations of striatal projecting neurons, the striatopallidal and the striatonigral MSNs, with the latter population being more consistently involved in LID induction, as suggested by some recent reports [61, 70, 92]. A recent *in vivo* electrophysiological study has given substantial foundation to the distinction between direct and indirect pathways suggesting that a range of different dysfunctional changes in these two populations of projecting neurons may concur to the induction of LID. One interesting aspect that comes out from this paper is

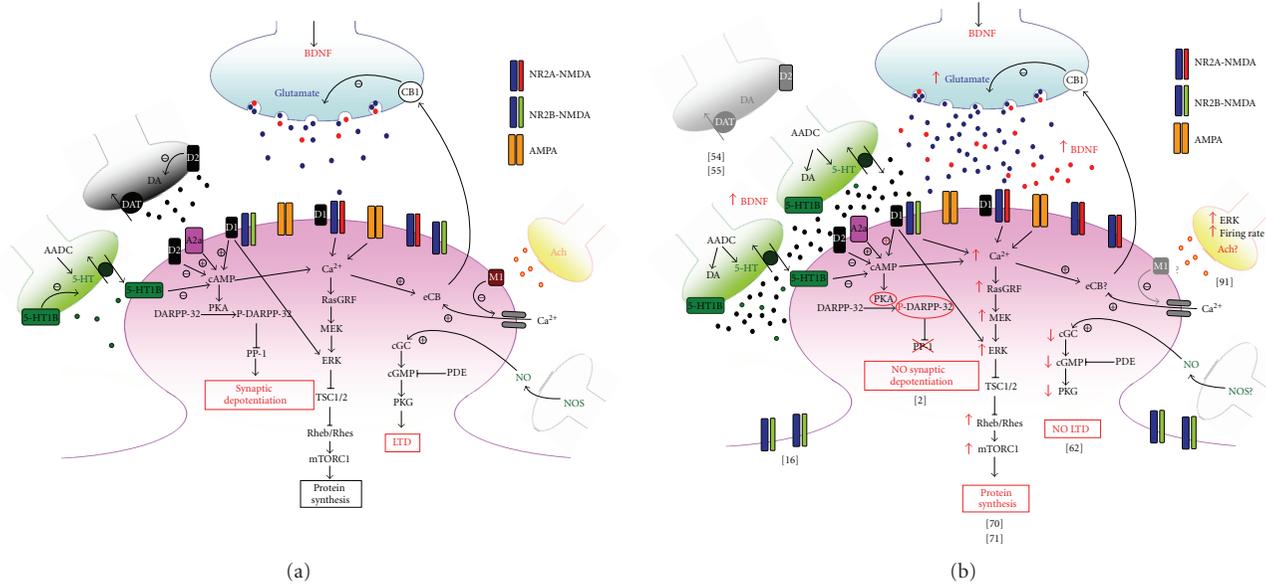


FIGURE 1: (a) In control condition, dopamine (DA) transmission is regulated by feedback control of release from nigrostriatal terminals (black) through D2 autoreceptors and uptake processes. DA binds striatal postsynaptic D1 receptors inducing the formation of cAMP, which in turn favours the activation of PKA, able to phosphorylate and activate DA- and cAMP-regulated phosphoprotein of 32 KDa (DARPP-32) and extracellular signal-regulated kinase (ERK). Once phosphorylated, DARPP-32 is able to inhibit protein phosphatase 1 (PP-1). Glutamate (blue) and BDNF (red) are released from corticostriatal terminals into the striatum. Glutamate release is regulated by endocannabinoids (eCB) activated by increases in intracellular calcium ( $Ca^{2+}$ ) concentrations through adenosine A2a and muscarinic M1 receptors activation, among other mechanisms, and retrogradely released by postsynaptic striatal neurons. Once released, glutamate activates metabotropic as well as NMDA and AMPA ionotropic receptors, whose activity and surface expression at postsynaptic membrane is also regulated by D1 receptors. In serotonergic afferents, 5-hydroxytryptophan is converted to serotonin (5-HT) (green) by Aromatic-L-Amino Acid Decarboxylase (AADC) and released into the striatum. Cholinergic and nitric oxide synthase (NOS)-positive interneurons cooperate to induction of corticostriatal LTP and LTD. (b) In dyskinetic state L-DOPA is converted to DA by AADC and released from serotonergic terminals in unregulated manner. Higher levels of striatal BDNF may support morphological changes in serotonergic neurons. Excess of DA abnormally stimulates D1 pathway with hyperphosphorylation of ERK and uncontrolled activation of PKA that results in hyperphosphorylation of DARPP32, which persistently blocks PP-1 causing loss of synaptic depotentiation. Abnormal D1 receptor stimulation is associated to increased intracellular  $Ca^{2+}$  levels and dysregulation of NMDAR subunit composition with reduction of NR2B-containing NMDAR at synaptic sites, leading to increase in NR2A/NR2B ratio that has been suggested to have a role in the loss of depotentiation. Hyperactivation of ERK through convergent altered signalling pathways brings to increased inhibition of tuberous sclerosis complex (TSC)1/2, and consequent disinhibition of Rheb/Rhes, leading to excessive increase of signalling of mTORC1 that, in turn, exerts its long term effects through changes in protein synthesis. After chronic L-DOPA, cholinergic interneurons show increased phospho-ERK immunoreactivity and higher firing rates with increased release of acetylcholine (Ach). Striatal cGMP signalling is decreased and corticostriatal LTD, which strictly relies on the nitric-oxide- (NO-) dependent activation of protein kinase G (PKG) is abolished in dyskinetic state.

that also striatopallidal neurons present specific alterations of synaptic plasticity in response to L-DOPA, although the study leaves open unresolved questions regarding the relevance of these findings for *in vivo* behavior [93].

Besides the distinct contribution of direct and indirect pathways to LID, several lines of evidence support the idea that also striatal regional compartmentalization matters in the response to L-DOPA. Within the striatum, it is possible to distinguish two compartmentalizations, whose activation can be modulated by striatal interneurons: the matrix, including the direct and indirect pathway MSNs that form parts of sensorimotor and associative circuits, and the striosomes, which contain MSNs that receive input from parts of limbic cortex and project directly or indirectly to the dopamine-releasing neurons of the *substantia nigra pars compacta*.

An interesting recent review has strengthened this idea, discussing the role of imbalances between striatal striosome

and matrix functions in relation to neurodegenerative disorders, including LID [94]. Findings in support of this idea may have important implications in the perspective of considering PD and LID as network disorders that cause a range of motor and nonmotor symptoms.

## 6. Concluding Remarks

We have discussed seminal and recent papers that explored the mechanisms underlying the establishment of aberrant forms of synaptic plasticity at glutamatergic corticostriatal synapses in LID experimental models. We have also provided an overview of recent studies dealing with novel aspects of the multifaceted L-DOPA effect. Taken together, all the reviewed studies strongly support the notion of a failure of the principal scaling down mechanisms at corticostriatal synapses as a major mechanism in the development of LID.

The scenario emerging from these findings is predictive of a more complex pattern of altered plasticity that involves structural and functional changes within the striatal circuitry and opens new perspectives for future electrophysiological investigations.

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## References

- [1] V. Paillé, B. Picconi, V. Bagetta et al., "Distinct levels of dopamine denervation differentially alter striatal synaptic plasticity and NMDA receptor subunit composition," *Journal of Neuroscience*, vol. 30, no. 42, pp. 14182–14193, 2010.
- [2] B. Picconi, D. Centonze, K. Håkansson et al., "Loss of bidirectional striatal synaptic plasticity in L-DOPA-induced dyskinesia," *Nature Neuroscience*, vol. 6, no. 5, pp. 501–506, 2003.
- [3] P. Calabresi, M. D. Filippo, V. Ghiglieri, N. Tambasco, and B. Picconi, "Levodopa-induced dyskinesias in patients with Parkinson's disease: filling the bench-to-bedside gap," *The Lancet Neurology*, vol. 9, no. 11, pp. 1106–1117, 2010.
- [4] F. Morgante, A. J. Espay, C. Gunraj, A. E. Lang, and R. Chen, "Motor cortex plasticity in Parkinson's disease and levodopa-induced dyskinesias," *Brain*, vol. 129, no. 4, pp. 1059–1069, 2006.
- [5] I. A. Prescott, J. O. Dostrovsky, E. Moro, M. Hodaie, A. M. Lozano, and W. D. Hutchison, "Levodopa enhances synaptic plasticity in the substantia nigra pars reticulata of Parkinson's disease patients," *Brain*, vol. 132, no. 2, pp. 309–318, 2009.
- [6] D. M. Lovinger, E. C. Tyler, and A. Merritt, "Short- and long-term synaptic depression in rat neostriatum," *Journal of Neurophysiology*, vol. 70, no. 5, pp. 1937–1949, 1993.
- [7] P. Calabresi, R. Maj, A. Pisani, N. B. Mercuri, and G. Bernardi, "Long-term synaptic depression in the striatum: physiological and pharmacological characterization," *Journal of Neuroscience*, vol. 12, no. 11, pp. 4224–4233, 1992.
- [8] P. Calabresi, A. Pisani, N. B. Mercuri, and G. Bernardi, "Long-term potentiation in the striatum is unmasked by removing the voltage-dependent magnesium block of NMDA receptor channels," *European Journal of Neuroscience*, vol. 4, no. 10, pp. 929–935, 1992.
- [9] P. Calabresi, B. Picconi, A. Tozzi, and M. Di Filippo, "Dopamine-mediated regulation of corticostriatal synaptic plasticity," *Trends in Neurosciences*, vol. 30, no. 5, pp. 211–219, 2007.
- [10] P. Calabresi, D. Centonze, and G. Bernardi, "Electrophysiology of dopamine in normal and denervated striatal neurons," *Trends in Neurosciences*, vol. 23, no. 10, supplement 1, pp. S57–S63, 2000.
- [11] P. Calabresi, R. Maj, N. B. Mercuri, and G. Bernardi, "Coactivation of D1 and D2 dopamine receptors is required for long-term synaptic depression in the striatum," *Neuroscience Letters*, vol. 142, no. 1, pp. 95–99, 1992.
- [12] V. Bagetta, V. Ghiglieri, C. Sgobio, P. Calabresi, and B. Picconi, "Synaptic dysfunction in Parkinson's disease," *Biochemical Society Transactions*, vol. 38, no. 2, pp. 493–497, 2010.
- [13] D. Centonze, P. Gubellini, G. Bernardi, and P. Calabresi, "Permissive role of interneurons in corticostriatal synaptic plasticity," *Brain Research Reviews*, vol. 31, no. 1, pp. 1–5, 1999.
- [14] B. Picconi, F. Gardoni, D. Centonze et al., "Abnormal Ca<sup>2+</sup>-calmodulin-dependent protein kinase II function mediates synaptic and motor deficits in experimental parkinsonism," *Journal of Neuroscience*, vol. 24, no. 23, pp. 5283–5291, 2004.
- [15] T. J. O'Dell and E. R. Kandel, "Low-frequency stimulation erases LTP through an NMDA receptor-mediated activation of protein phosphatases," *Learning Memory*, vol. 1, no. 2, pp. 129–139, 1994.
- [16] F. Gardoni, B. Picconi, V. Ghiglieri et al., "A critical interaction between NR2B and MAGUK in L-DOPA induced dyskinesia," *Journal of Neuroscience*, vol. 26, no. 11, pp. 2914–2922, 2006.
- [17] B. Picconi, E. Passino, C. Sgobio et al., "Plastic and behavioral abnormalities in experimental Huntington's disease: a crucial role for cholinergic interneurons," *Neurobiology of Disease*, vol. 22, no. 1, pp. 143–152, 2006.
- [18] A. Borgkvist and G. Fisone, "Psychoactive drugs and regulation of the cAMP/PKA/DARPP-32 cascade in striatal medium spiny neurons," *Neuroscience and Biobehavioral Reviews*, vol. 31, no. 1, pp. 79–88, 2007.
- [19] A. A. Fienberg, N. Hiroi, P. G. Mermelstein et al., "DARPP-32: regulator of the efficacy of dopaminergic neurotransmission," *Science*, vol. 281, no. 5378, pp. 838–842, 1998.
- [20] P. Greengard, "The neurobiology of dopamine signaling," *Bioscience Reports*, vol. 21, no. 3, pp. 247–269, 2001.
- [21] F. Gardoni, V. Ghiglieri, M. D. Luca, and P. Calabresi, "Assemblies of glutamate receptor subunits with post-synaptic density proteins and their alterations in Parkinson's disease," *Progress in Brain Research*, vol. 183, no. C, pp. 169–182, 2010.
- [22] C. Fiorentini, F. Gardoni, P. Spano, M. Di Luca, and C. Missale, "Regulation of dopamine D1 receptor trafficking and desensitization by oligomerization with glutamate N-methyl-D-aspartate receptors," *Journal of Biological Chemistry*, vol. 278, no. 22, pp. 20196–20202, 2003.
- [23] C. Fiorentini, M. C. Rizzetti, C. Busi et al., "Loss of synaptic D1 dopamine/N-methyl-D-aspartate glutamate receptor complexes in L-DOPA-induced dyskinesia in the rat," *Molecular Pharmacology*, vol. 69, no. 3, pp. 805–812, 2006.
- [24] P. J. Hallett, R. Spoelgen, B. T. Hyman, D. G. Standaert, and A. W. Dunah, "Dopamine D1 activation potentiates striatal NMDA receptors by tyrosine phosphorylation-dependent subunit trafficking," *Journal of Neuroscience*, vol. 26, no. 17, pp. 4690–4700, 2006.
- [25] C. Cepeda, N. A. Buchwald, and M. S. Levine, "Neuromodulatory actions of dopamine in the neostriatum are dependent upon the excitatory amino acid receptor subtypes activated," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 20, pp. 9576–9580, 1993.
- [26] G. L. Snyder, A. A. Fienberg, R. L. Haganir, and P. Greengard, "A dopamine/D1 receptor/protein kinase A/dopamine- and cAMP-regulated phosphoprotein (Mr 32 kDa)/protein phosphatase-1 pathway regulates dephosphorylation of the NMDA receptor," *Journal of Neuroscience*, vol. 18, no. 24, pp. 10297–10303, 1998.
- [27] X. Sun, Y. Zhao, and M. E. Wolf, "Dopamine receptor stimulation modulates AMPA receptor synaptic insertion in prefrontal cortex neurons," *Journal of Neuroscience*, vol. 25, no. 32, pp. 7342–7351, 2005.
- [28] C. Gao, X. Sun, and M. E. Wolf, "Activation of D1 dopamine receptors increases surface expression of AMPA receptors and

- facilitates their synaptic incorporation in cultured hippocampal neurons," *Journal of Neurochemistry*, vol. 98, no. 5, pp. 1664–1677, 2006.
- [29] P. Gubellini, A. Pisani, D. Centonze, G. Bernardi, and P. Calabresi, "Metabotropic glutamate receptors and striatal synaptic plasticity: implications for neurological diseases," *Progress in Neurobiology*, vol. 74, no. 5, pp. 271–300, 2004.
- [30] Y. Kawaguchi, "Physiological, morphological, and histochemical characterization of three classes of interneurons in rat neostriatum," *Journal of Neuroscience*, vol. 13, no. 11, pp. 4908–4923, 1993.
- [31] Y. Kawaguchi, C. J. Wilson, S. J. Augood, and P. C. Emson, "Striatal interneurons: chemical, physiological and morphological characterization," *Trends in Neurosciences*, vol. 18, no. 12, pp. 527–535, 1995.
- [32] D. Centonze, P. Gubellini, A. Pisani, G. Bernardi, and P. Calabresi, "Dopamine, acetylcholine, and nitric oxide systems interact to induce corticostriatal synaptic plasticity," *Reviews in the Neurosciences*, vol. 14, no. 3, pp. 207–216, 2003.
- [33] T. Suzuki, M. Miura, K. Y. Nishimura, and T. Aosaki, "Dopamine-dependent synaptic plasticity in the striatal cholinergic interneurons," *Journal of Neuroscience*, vol. 21, no. 17, pp. 6492–6501, 2001.
- [34] P. Calabresi, D. Centonze, P. Gubellini, A. Pisani, and G. Bernardi, "Acetylcholine-mediated modulation of striatal function," *Trends in Neurosciences*, vol. 23, no. 3, pp. 120–126, 2000.
- [35] E. Fino, J. M. Deniau, and L. Venance, "Cell-specific spike-timing-dependent plasticity in GABAergic and cholinergic interneurons in corticostriatal rat brain slices," *Journal of Physiology*, vol. 586, no. 1, pp. 265–282, 2008.
- [36] A. Tozzi, A. De Iure, M. Di Filippo et al., "The distinct role of medium spiny neurons and cholinergic interneurons in the D2/A2A receptor interaction in the striatum: implications for Parkinson's disease," *Journal of Neuroscience*, vol. 31, no. 5, pp. 1850–1862, 2011.
- [37] S. J. Augood, E. M. McGowan, and P. C. Emson, "Expression of *N*-methyl-D-aspartate receptor subunit NR1 messenger RNA by identified striatal somatostatin cells," *Neuroscience*, vol. 59, no. 1, pp. 7–12, 1994.
- [38] M. V. Catania, T. R. Tölle, and H. Monyer, "Differential expression of AMPA receptor subunits in NOS-positive neurons of cortex, striatum, and hippocampus," *Journal of Neuroscience*, vol. 15, no. 11, pp. 7046–7061, 1995.
- [39] P. Marin, M. Lafon-Cazal, and J. Bockaert, "A nitric oxide synthase activity selectively stimulated by NMDA receptors depends on protein kinase C activation in mouse striatal neurons," *European Journal of Neuroscience*, vol. 4, no. 5, pp. 425–432, 1992.
- [40] S. W. Weiss, D. S. Albers, M. J. Iadarola, T. M. Dawson, V. L. Dawson, and D. G. Standaert, "NMDAR1 glutamate receptor subunit isoforms in neostriatal, neocortical, and hippocampal nitric oxide synthase neurons," *Journal of Neuroscience*, vol. 18, no. 5, pp. 1725–1734, 1998.
- [41] H. Prast and A. Philippu, "Nitric oxide as modulator of neuronal function," *Progress in Neurobiology*, vol. 64, no. 1, pp. 51–68, 2001.
- [42] A. R. West, M. P. Galloway, and A. A. Grace, "Regulation of striatal dopamine neurotransmission by nitric oxide: effector pathways and signaling mechanisms," *Synapse*, vol. 44, no. 4, pp. 227–245, 2002.
- [43] P. Greengard, "The neurobiology of slow synaptic transmission," *Science*, vol. 294, no. 5544, pp. 1024–1030, 2001.
- [44] M. A. Cenci and C. Konradi, "Maladaptive striatal plasticity in L-DOPA-induced dyskinesia," *Progress in Brain Research*, vol. 183, no. C, pp. 209–233, 2010.
- [45] M. M. Iravani and P. Jenner, "Mechanisms underlying the onset and expression of levodopa-induced dyskinesia and their pharmacological manipulation," *Journal of Neural Transmission*, vol. 118, no. 12, pp. 1661–1690, 2011.
- [46] J. Lee, W. M. Zhu, D. Stanic et al., "Sprouting of dopamine terminals and altered dopamine release and uptake in Parkinsonian dyskinesia," *Brain*, vol. 131, no. 6, pp. 1574–1587, 2008.
- [47] P. Calabresi, N. B. Mercuri, G. Sancesario, and G. Bernardi, "Electrophysiology of dopamine-denervated striatal neurons. Implications for Parkinson's disease," *Brain*, vol. 116, no. 2, pp. 433–452, 1993.
- [48] C. A. Ingham, S. H. Hood, P. Taggart, and G. W. Arbuthnott, "Plasticity of synapses in the rat neostriatum after unilateral lesion of the nigrostriatal dopaminergic pathway," *Journal of Neuroscience*, vol. 18, no. 12, pp. 4732–4743, 1998.
- [49] N. Lindfors and U. Ungerstedt, "Bilateral regulation of glutamate tissue and extracellular levels in caudate-putamen by midbrain dopamine neurons," *Neuroscience Letters*, vol. 115, no. 2-3, pp. 248–252, 1990.
- [50] C. K. Meshul, N. Emre, C. M. Nakamura, C. Allen, M. K. Donohue, and J. F. Buckman, "Time-dependent changes in striatal glutamate synapses following a 6-hydroxydopamine lesion," *Neuroscience*, vol. 88, no. 1, pp. 1–16, 1999.
- [51] B. Picconi, D. Centonze, S. Rossi, G. Bernardi, and P. Calabresi, "Therapeutic doses of L-dopa reverse hypersensitivity of corticostriatal D2-dopamine receptors and glutamatergic overactivity in experimental parkinsonism," *Brain*, vol. 127, no. 7, pp. 1661–1669, 2004.
- [52] J. Abarca, K. Gysling, R. H. Roth, and G. Bustos, "Changes in extracellular levels of glutamate and aspartate in rat substantia nigra induced by dopamine receptor ligands: *in vivo* microdialysis studies," *Neurochemical Research*, vol. 20, no. 2, pp. 159–169, 1995.
- [53] C. S. Biggs and M. S. Starr, "Dopamine and glutamate control each other's release in the basal ganglia: a microdialysis study of the entopeduncular nucleus and substantia nigra," *Neuroscience and Biobehavioral Reviews*, vol. 21, no. 4, pp. 497–504, 1997.
- [54] M. Carta, T. Carlsson, D. Kirik, and A. Björklund, "Dopamine released from 5-HT terminals is the cause of L-DOPA-induced dyskinesia in parkinsonian rats," *Brain*, vol. 130, no. 7, pp. 1819–1833, 2007.
- [55] D. Rylander, M. Parent, S. S. O'Sullivan et al., "Maladaptive plasticity of serotonin axon terminals in levodopa-induced dyskinesia," *Annals of Neurology*, vol. 68, no. 5, pp. 619–628, 2010.
- [56] T. N. Chase and J. D. Oh, "Striatal mechanisms and pathogenesis of parkinsonian signs and motor complications," *Annals of Neurology*, vol. 47, Supplement 1, no. 4, pp. S122–S129, 2000.
- [57] A. W. Dunah, Y. Wang, R. P. Yasuda et al., "Alterations in subunit expression, composition, and phosphorylation of striatal *N*-methyl-D-aspartate glutamate receptors in a rat 6-hydroxydopamine model of Parkinson's disease," *Molecular Pharmacology*, vol. 57, no. 2, pp. 342–352, 2000.
- [58] E. Santini, E. Valjent, A. Usiello et al., "Critical involvement of cAMP/DARPP-32 and extracellular signal-regulated protein kinase signaling in L-DOPA-induced dyskinesia," *Neuroscience*, vol. 27, no. 26, pp. 6995–7005, 2007.
- [59] N. Pavón, A. B. Martín, A. Mendiáldua, and R. Moratalla, "ERK phosphorylation and FosB expression are associated

- with L-DOPA-induced dyskinesia in hemiparkinsonian mice," *Biological Psychiatry*, vol. 59, no. 1, pp. 64–74, 2006.
- [60] J. E. Westin, L. Vercammen, E. M. Strome, C. Konradi, and M. A. Cenci, "Spatiotemporal pattern of striatal ERK1/2 phosphorylation in a rat model of L-DOPA-induced dyskinesia and the role of dopamine D1 receptors," *Biological Psychiatry*, vol. 62, no. 7, pp. 800–810, 2007.
- [61] M. Feyder, A. Bonito-Oliva, and G. Fisone, "L-dopa-induced dyskinesia and abnormal signalling in striatal medium spiny neurons: focus on dopamine D1 receptor-mediated transmission," *Frontiers in Behavioral Neuroscience*, vol. 5, article 71, 2011.
- [62] B. Picconi, V. Bagetta, V. Ghiglieri et al., "Inhibition of phosphodiesterases rescues striatal long-term depression and reduces levodopa-induced dyskinesia," *Brain*, vol. 134, no. 2, pp. 375–387, 2011.
- [63] B. Picconi, V. Paillé, V. Ghiglieri et al., "L-DOPA dosage is critically involved in dyskinesia via loss of synaptic depotentiation," *Neurobiology of Disease*, vol. 29, no. 2, pp. 327–335, 2008.
- [64] M. A. Cenci, "Dopamine dysregulation of movement control in L-DOPA-induced dyskinesia," *Trends in Neurosciences*, vol. 30, no. 5, pp. 236–243, 2007.
- [65] M. Lundblad, B. Picconi, H. Lindgren, and M. A. Cenci, "A model of L-DOPA-induced dyskinesia in 6-hydroxydopamine lesioned mice: relation to motor and cellular parameters of nigrostriatal function," *Neurobiology of Disease*, vol. 16, no. 1, pp. 110–123, 2004.
- [66] F. Gardoni, C. Sgobio, V. Pendolino, P. Calabresi, M. Di Luca, and B. Picconi, "Targeting NR2A-containing NMDA receptors reduces L-DOPA-induced dyskinesias," *Neurobiology of Aging*. In press.
- [67] M. Giorgi, V. D'Angelo, Z. Esposito et al., "Lowered cAMP and cGMP signalling in the brain during levodopa-induced dyskinesias in hemiparkinsonian rats: new aspects in the pathogenetic mechanisms," *European Journal of Neuroscience*, vol. 28, no. 5, pp. 941–950, 2008.
- [68] F. Errico, M. T. Pirro, A. Affuso et al., "A physiological mechanism to regulate d-aspartic acid and NMDA levels in mammals revealed by d-aspartate oxidase deficient mice," *Gene*, vol. 374, no. 1–2, pp. 50–57, 2006.
- [69] F. Errico, A. Bonito-Oliva, V. Bagetta et al., "Higher free D-aspartate and N-methyl-D-aspartate levels prevent striatal depotentiation and anticipate L-DOPA-induced dyskinesia," *Experimental Neurology*, vol. 232, no. 2, pp. 240–250, 2011.
- [70] E. Santini, M. Heiman, P. Greengard, E. Valjent, and G. Fisone, "Inhibition of mTOR signaling in parkinson's disease prevents L-DOPA-induced dyskinesia," *Science Signaling*, vol. 2, no. 80, p. ra36, 2009.
- [71] S. Subramaniam, F. Napolitano, R. G. Mealer et al., "Rhes, a striatal-enriched small G protein, mediates mTOR signalling and L-DOPA-induced dyskinesia," *Nature Neuroscience*, vol. 15, no. 2, pp. 191–193, 2012.
- [72] C. A. Hoeffer and E. Klann, "mTOR signaling: at the crossroads of plasticity, memory and disease," *Trends in Neurosciences*, vol. 33, no. 2, pp. 67–75, 2010.
- [73] V. Ghiglieri, V. Pendolino, V. Bagetta, C. Sgobio, P. Calabresi, and B. Picconi, "mTOR inhibitor rapamycin suppresses striatal post-ischemic LTP," *Experimental Neurology*, vol. 226, no. 2, pp. 328–331, 2010.
- [74] K. Kannari, H. Yamato, H. Shen, M. Tomiyama, T. Suda, and M. Matsunaga, "Activation of 5-HT1A but not 5-HT1B receptors attenuates an increase in extracellular dopamine derived from exogenously administered L-DOPA in the striatum with nigrostriatal denervation," *Journal of Neurochemistry*, vol. 76, no. 5, pp. 1346–1353, 2001.
- [75] H. Tanaka, K. Kannari, T. Maeda, M. Tomiyama, T. Suda, and M. Matsunaga, "Role of serotonergic neuron in L-DOPA-derived extracellular dopamine in the striatum of 6-OHDA-lesioned rats," *NeuroReport*, vol. 10, no. 3, pp. 631–634, 1999.
- [76] K. B. Dupre, C. Y. Ostock, K. L. Eskow Jaunarajs et al., "Local modulation of striatal glutamate efflux by serotonin 1A receptor stimulation in dyskinetic, hemiparkinsonian rats," *Experimental Neurology*, vol. 229, no. 2, pp. 288–299, 2011.
- [77] P. Huot and J. M. Brotchie, "5-HT(1A) receptor stimulation and L-DOPA-induced dyskinesia in Parkinson's disease: bridging the gap between serotonergic and glutamatergic mechanisms," *Nature Neuroscience*, vol. 231, no. 2, pp. 195–198, 2011.
- [78] C. Y. Ostock, K. B. Dupre, K. L. Eskow Jaunarajs et al., "Role of the primary motor cortex in L-DOPA-induced dyskinesia and its modulation by 5-HT1A receptor stimulation," *Neuropharmacology*, vol. 61, no. 4, pp. 753–760, 2011.
- [79] M. Andersson, A. Hilbertson, and M. A. Cenci, "Striatal fosB expression is causally linked with L-DOPA-induced abnormal involuntary movements and the associated upregulation of striatal prodynorphin mRNA in a rat model of Parkinson's disease," *Neurobiology of Disease*, vol. 6, no. 6, pp. 461–474, 1999.
- [80] A. Muñoz, Q. Li, F. Gardoni et al., "Combined 5-HT1A and 5-HT1B receptor agonists for the treatment of L-DOPA-induced dyskinesia," *Brain*, vol. 131, no. 12, pp. 3380–3394, 2008.
- [81] D. Rylander, "The serotonin system: a potential target for anti-dyskinetic treatments and biomarker discovery," *Parkinsonism & Related Disorders*, vol. 18, Supplement 1, pp. S126–S128, 2012.
- [82] O. Guillin, J. Diaz, P. Carroll, N. Griffon, J. C. Schwartz, and P. Sokoloff, "BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization," *Nature*, vol. 411, no. 6833, pp. 86–89, 2001.
- [83] P. Samadi, M. Morissette, D. Lévesque, and T. Di Paolo, "BDNF levels are not related with levodopa-induced dyskinesias in MPTP monkeys," *Movement Disorders*, vol. 25, no. 1, pp. 116–121, 2010.
- [84] R. L. Potenza, M. T. Tebano, A. Martire et al., "Adenosine A2A receptors modulate BDNF both in normal conditions and in experimental models of Huntington's disease," *Purinergic Signalling*, vol. 3, no. 4, pp. 333–338, 2007.
- [85] M. T. Tebano, A. Martire, V. Chiodi, A. Ferrante, and P. Popoli, "Role of adenosine A<sub>2A</sub> receptors in modulating synaptic functions and brain levels of BDNF: a possible key mechanism in the pathophysiology of Huntington's disease," *TheScientificWorldJOURNAL*, vol. 10, pp. 1768–1782, 2010.
- [86] F. Blandini and M. T. Armentero, "New pharmacological avenues for the treatment of L-DOPA-induced dyskinesias in Parkinson's disease: targeting glutamate and adenosine receptors," *Expert Opinion on Investigational Drugs*, vol. 21, no. 2, pp. 153–168, 2012.
- [87] A. Tozzi, A. Tschertter, V. Belcastro et al., "Interaction of A2A adenosine and D2 dopamine receptors modulates corticostriatal glutamatergic transmission," *Neuropharmacology*, vol. 53, no. 6, pp. 783–789, 2007.
- [88] N. Kayadjanian, "Cortical and nigral deafferentation and striatal cholinergic markers in the rat dorsal striatum: different effects on the expression of mRNAs encoding choline acetyltransferase and muscarinic m1 and m4 receptors," *European Journal of Neuroscience*, vol. 11, no. 10, pp. 3659–3668, 1999.

- [89] J. Ding, J. N. Guzman, T. Tkatch et al., "RGS4-dependent attenuation of M4 autoreceptor function in striatal cholinergic interneurons following dopamine depletion," *Nature Neuroscience*, vol. 9, no. 6, pp. 832–842, 2006.
- [90] P. Bonsi, G. Martella, D. Cuomo et al., "Loss of muscarinic autoreceptor function impairs long-term depression but not long-term potentiation in the striatum," *Neuroscience*, vol. 28, no. 24, pp. 6258–6263, 2008.
- [91] Y. Ding, L. Won, J. P. Britt, S. A. O. Lim, D. S. McGehee, and U. J. Kang, "Enhanced striatal cholinergic neuronal activity mediates L-DOPA-induced dyskinesia in parkinsonian mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 2, pp. 840–845, 2011.
- [92] H. S. Bateup, E. Santini, W. Shen et al., "Distinct subclasses of medium spiny neurons differentially regulate striatal motor behaviors," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 33, pp. 14845–14850, 2010.
- [93] P. Belujon, D. J. Lodge, and A. A. Grace, "Aberrant striatal plasticity is specifically associated with dyskinesia following levodopa treatment," *Movement Disorders*, vol. 25, no. 11, pp. 1568–1576, 2010.
- [94] J. R. Crittenden and A. M. Graybiel, "Basal ganglia disorders associated with imbalances in the striatal striosome and matrix compartments," *Frontiers in Neuroanatomy*, vol. 5, article 59, 2011.

## Review Article

# Dyskinesias and Treatment with Pramipexole in Patients with Parkinson's Disease

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Dopamine agonists such as pramipexole (PPX) have first been proposed as adjunctive treatment to levodopa (L-DOPA) for patients with Parkinson's disease (PD) and then as a monotherapy alternative to alleviate dyskinesia. Treatment with PPX has overall been associated with improvement in parkinsonian symptoms. Although the majority of placebo-controlled studies demonstrated that dyskinesia was more prevalent in the PPX compared to the placebo groups, some studies did not detect any dyskinesia as a side effect of this medication. PPX was consistently associated with lower risk for developing dyskinesia compared to L-DOPA. Moreover, the presence of these symptoms in the placebo groups suggests involvement of non-PPX-related factors for developing dyskinesia. It is suggested that future research should aim at ascertaining whether cotherapy with L-DOPA, PPX dosage, and other patient characteristics are contributory factors for the development of PPX-related dyskinesia in patients with PD.

## 1. Dyskinesia in Parkinson's Disease

Parkinson's disease (PD) is a neurodegenerative disease characterised by motor (particularly tremor, rigidity, and bradykinesia) as well as cognitive and behavioural symptoms. The pathophysiology of PD has been related to the degeneration of nigrostriatal dopaminergic pathways [1], and this has allowed the treatment for PD to be targeted towards modulating dopamine (DA) neurotransmission (Figure 1).

Levodopa (L-DOPA) has long been the mainstay of PD treatment although over time patients on L-DOPA develop motor complications including dyskinesias, which are associated with the timing of drug administration. Dyskinesias are involuntary muscular contractions and include choreic, dystonic, myoclonic, and tic movements [2]. After 5 and 10 years of L-DOPA therapy, 91% and all of the participants in a longitudinal cohort ( $N = 99$ ), respectively, experienced dyskinesias [3]. Another study also identified that cumulative L-DOPA dosage was significantly associated with the development of dyskinesia [4].

Given that dyskinesias has been consistently shown to negatively affect patients' quality of life [5, 6], there is considerable debate on how to forestall its onset, including initial treatment with another class of drugs: the dopamine receptor agonists (DAAs) [7, 8]. Pramipexole (PPX) belongs to this drug class and is selective for the D<sub>2</sub>-like receptor subfamily, particularly the D<sub>3</sub> compared to the D<sub>2</sub> and D<sub>4</sub> subtypes [9]. Following the observation by Hauser et al. [4] that treatment with PPX was significantly associated with later onset of dyskinesia, we carried out a systematic literature review to examine the effects of PPX therapy on dyskinesic events in patients with PD.

## 2. Literature Search Methodology

This paper systematically reviews the existing evidence on the development of dyskinesia during PPX therapy for PD. We performed a literature search across the databases Medline, EMBASE and PsycInfo via the NHS Evidence tool (<http://www.library.nhs.uk>). We used the search terms

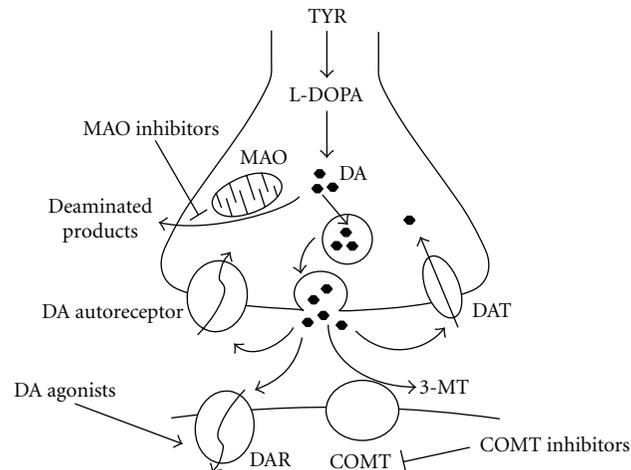


FIGURE 1: Treatment of Parkinson's disease: anti-Parkinson's medications modulate key stages of dopaminergic neurotransmission. Abbreviations: TYR: tyrosine, L-DOPA: L-3, 4-dihydroxyphenylalanine, DA: dopamine, MAO: monoamine oxidase, DAT: DA reuptake transporter, COMT: catechol-O-methyltransferase, 3-MT: 3-methoxytyramine, DAR: DA receptor. Anti-Parkinson's drugs are highlighted in bold. Pointed arrows indicate stimulatory, and closed arrows indicate inhibitory activity.

“Parkinson\*,” “dyskinesi\*,” and “pramipexole”. The Cochrane Library was also searched for randomised and double-blind human trials of PPX in patients with PD. We limited our search to papers published in English language.

### 3. Pramipexole-Placebo Comparisons

The majority of the studies on PPX included in this review were comparisons with placebo only (Table 1). Six out of ten of these studies found that dyskinesia in PPX-treated patients was prevalent, at a higher rate than in the placebo group. The incidence of dyskinesia was 7.0–61.3% and 3.0–40.8% in the PPX and placebo groups, respectively [10–15]. The differences in rates of dyskinesia were between 4.9 and 20.5%. Two of these studies had follow-up data. An extension to the Lieberman et al. [10] protocol by up to 50 months was carried out, in which both PPX and placebo groups were re-titrated onto open-label PPX [16]. Out of the sample ( $N = 306$ ), 61.1% reported dyskinesias, but there were no related discontinuations. Furthermore, UPDRS IV scores remained below the baseline values, indicating some improvements in these symptoms. Möller et al. [13] reported an open-label extension to their study of up to 57 months follow-up. Out of their cohort, 34.4% ( $N = 262$ ) developed dyskinesias. This led to study discontinuation in 2.3% of participants. Two studies reported the incidence of dyskinesia to be higher with the placebo than the PPX groups: 0.6% versus 0.0% [17] and 6.1% versus 5.6% [18]. However, it should be noted that the strength of such evidence is weak. One study [17] was specifically designed to assess the antiparkinsonian properties in early PD, rather than the potential dyskinesic effects of pramipexole. The other one [18] found 2 dyskinetic patients in a group of 33 patients treated with placebo and 2 dyskinetic patients in a group of 36 patients treated with PPX. Of note, a further study failed to identify participants who experienced dyskinesia with PPX treatment or placebo [19, 20].

In terms of neurological scales as a measure of dyskinesia, Lieberman et al. [10] reported that % change in UPDRS IV scores was significantly ( $P < .0001$ ) higher in the PPX group compared to the placebo group. Changes in the PDS were not statistically significant. These exact patterns in UPDRS IV and PDS scores were reproduced by Pinter et al. [11] and Möller et al. [13]. In these studies, however, the incidence of dyskinesia was higher with PPX treatment than placebo. Wermuth et al. [18] did not find significant changes in UPDRS IV or PDS scores. In a cross-over design with PPX and placebo, as well as L-DOPA infusion before and after the switch-over, Brodsky et al. [32] found that PPX treatment increased PDS scores to statistically significant levels ( $P = .05$ ). Furthermore, L-DOPA infusion also increased peak dyskinesia scores.

### 4. Studies with Active Comparators

**4.1. Pramipexole Only.** There have been two studies comparing different dosages and other two looking at different preparations of PPX. One of the earliest investigations on PPX was carried out by the Parkinson Study Group [21], which compared four different dosages of the drug (1.5, 3.0, 4.5, and 6.0 mg) against placebo. Another study by the same group compared different low-dose schedules of PPX [21]. Both did not detect any incidence of dyskinesia. Likewise, comparisons of immediate release (IR) and extended release (ER) PPX preparations failed to detect any incidence of dyskinesia symptoms in either treatment or placebo group [26, 27].

**4.2. Pramipexole versus Other Dopamine Agonists.** Other DAAs have been compared with PPX. Two studies reported a comparison with bromocriptine (BRC), showing that dyskinesias were found more often with DAAs compared with placebo. In terms of UPDRS IV and PDS scores, there were no significant changes in one trial [29], whereas, in

TABLE 1: Dyskinesia with pramipexole treatment: summary of double-blind randomised-controlled trials of pramipexole in Parkinson's disease.

Disease stage*	Study	Drug regimen (N): mean daily dose, mg (SD/range)	Incidence of dyskinesia (%), change in UPDRS IV or PDS (%)	Concomitant L-DOPA (% usage and/or mean dose, mg (SD/range))	Other concomitant APDs	Study duration	PPX group characteristics: H-Y stage, age, disease duration (mean (SD))
	Hubble et al. [20]	PPX (28), PL (27): 4.5	None	None	MAOBI	9 wks	1-3, 63.5 (12.3), 2.1 (2.5)
	PSG [21]	PPX (213), PL (151): fixed dose at 1.5, 3.0, 4.5, 6.0	None	PPX: 24.4%, PL: 27.5%	MAOBI	10 wks	1-4 (1.9, 0.6), 62.0 (10.9), 2.0 (1.6)
	Shannon et al. [17]	PPX (164): 3.8, PL (171): 0.375-4.5	PPX: 0%, PL: 0.6% (leading to discontinuation) PPX: 9.9%, L-DOPA: 30.7% (HR 0.33 (95% CI, 0.18-0.60); $P < .001$ )-166.7% (PPX), 182.6% after O/L L-DOPA	None	MAOBI	31 wks	1-3, 62.7, 1.8
	PSG [22] <sup>†</sup>	PPX (151): 1.5-4.5, L-DOPA (150): 300-600	PPX: 10.0%, PGL: 10.0%, PL: 0.0%	Part of study intervention	MAOBI, amantadine, AC	23.5 mos	1-3 (82.8% $\leq$ 2), 61.5 (10.1), 1.5 (1.4)
	Pogarell et al. [19]	PPX (44), PL (39): 0.375-4.5	None	PPX: 11%, PL: 13%	MAOBI and amantadine	11 wks	1-3 (72.7% $\leq$ 2), 62.0 (10.1), 6.5 (4.0)
	Navan et al. [23]	PPX, PGL, PL (10): 4.5	PPX: 10.0%, PGL: 10.0%, PL: 0.0%	PPX: 60%, 400 (200-600), PGL: 60%, 383 (300-700), PL: 40%, 550 (300-800)	MAOBI, amantadine, and AC	3 mos	1-2 (1), 66 (55-80), 4 (0.5-10)
Early	Wong et al. [12]	PPX (73), PL (77): 0.375-4.5	PPX: 12.3%, PL: 5.2%	PPX: 68.5%, PL: 70.1%	None	15 wks	2.2 (0.07), 58.8 (1.28), 4.5 (0.4)
	PSG [24] <sup>†</sup>	PPX (83): 2.78 (1.1), L-DOPA (100): 427 (112)	PPX: 24.5.5, L-DOPA: 54.0 (HR 0.37 (95% CI 0.25-0.56); $P < .001$ )	PPX: 434 (498), L-DOPA: 274 (442)	MAOBI, amantadine, AC	4 yrs	1-3 (79.5% $\leq$ 2), 61.1 (9.6), 1.4 (1.3)
	Navan et al. [25]	PPX (9): 3.09, PGL (8): 3.0, cross-over	PPX: 33.3%, PGL: 37.5%	52.9%; 544 (300-1000)	AC	12 wks-9 wks cross-over	1-2 (1.4), 68.4 (55-84), 3.9 (0.2-12.0)
	Barone et al. [15]	PPX (139): 2.18 (0.83), PL (148): 2.51 (1.66)	PPX: 7%, PL: 3%	PPX: 76%, PL: 74%	Amantadine, MAOBI, ACI, and ODD	12 wks	1-3 (79% $\geq$ 2), 67.4 (9.0), 4.0 (4.5)
	Hauser et al. [26]	PPX IR (103), PPX ER (106), PL (50): 0.375-4.5	None	PPX IR: 1.0%, PPX ER: 2.9%, PL: 14.0%	None	18 wks	1-3 (72.3% 2-3), 61.8 (8.9), 1.0 (1.3)
	Rascol et al. [27]	PPX IR (52), PPX ER (104): 1.5-4.5	None	PPX IR: 51.9%, PPX ER: 56.7%	MAOBI, COMTI, AC, and amantadine	4 wks IR + 9 wks IR/ER	1-3 (80.8% 1-2), 63.7 (9.1), 3.3 (2.0)
	PSG [28]	PPX 0.5 bd (81), 0.75 bd (73), 0.5 td (80), PL (77)	None	None	MAOBI, AC, and amantadine	12 wks	1-2.5 (89.7% 1-2), 63.3 (10.0), 2.6 (2.6)

TABLE 1: Continued.

Disease stage*	Study	Drug regimen (N): mean daily dose, mg (SD/range)	Incidence of dyskinesia (%), change in UPDRS IV or PDS (%)	Concomitant L-DOPA (% usage and/or mean dose, mg (SD/range))	Other concomitant APDs	Study duration	PPX group characteristics: H-Y stage, age, disease duration (mean (SD))
	Guttman et al. [29]	PPX (79): 3.4, BRC (84): 22.6, PL (83)	PPX: 40%, BRC: 45%, PL: 27%; NS changes in UPDRS IV and PDS	100%	AC, amantadine, and MAOBI	9 mos	2–4, 62.9 (10.0), 0.67–36
	Lieberman et al. [10]	PPX (181), PL (179): 0.375–4.5	PPX: 61.3%, PL: 40.8%; UPDRS IV PPX > PL ( $P < .0001$ ), PDS PPX > PL ( $P = NS$ )	100%; PPX: 843.4 (578.9), PL: 819.2 (466.1)	MAOBI and amantadine	31 wks	2–4 (3.0), 63.4, 9.4
	Wermuth et al. [18]	PPX (36): 4.59 (0.95), PL (33): 4.77	PPX: 5.6%, PL: 6.1%; NS change in PDS; NS change in UPDRS IV	PPX: 100% (61.1% > 600), PL: 100% (66.7% > 600)	MAOBI, ACI, and amantadine	11 wks	2–4 (91.7% 2–3), 63.2 (7.9), 10.1 (5.0)
	Pinter et al. [11]	PPX (34), PL (44): 0.2–5.0	PPX: 14.7%, PL: 4.5%; UPDRS IV PPX > PL ( $P = .0092$ ); PDS PPX < PL ( $P = NS$ )	26.5% (11.8% > 600), PL: 34.1% (18.2% > 600)	MAOBI and amantadine	11 wks	2–4 (79.4% $\geq 3$ ), 59.3 (8.3), 7.8 (4.3)
Advanced	Mizuno et al. [30]	PPX (102): 3.24 (1.33), BRC (105): 17.8 (5.8), PL (108)	PPX: 15.7%, BRC: 8.6%, PL: 5.6%; UPDRS IV PPX < PL ( $P = .006$ ), PPX $\approx$ BRC ( $P = NS$ )	PPX: 404.9 (275.2), BRC: 377.9 (237.8), PL: 422.4 (330.3)	ACI, amantadine, ODD, and MAOBI	12 wks	2.7 (0.7), 65.5 (9.5), 4.8 (4.2)
	Möller et al. [13]	PPX (168): 3.7, PL: 0.375–4.5	PPX: 30.0%, PL: 8.7%; UPDRS IV PPX > PL ( $P = .0114$ ), PDS ( $P = NS$ )	PPX: 637.7, PL: 648.8	MAOBI and AC	31 wks	1–4 (85.0% 2–3), 63.4, 7.6
	Poewe et al. [31]	PPX (200): 3.1 (1.2), RTG (201): 13.0 (3.5), PL (100)	PPX: 15%, RTG: 12%, PL: 3%; hrs “on” without troublesome dyskinesia PPX > PL ( $P = 0.429$ ), PPX < RTG ( $P = NS$ )	PPX: 813 (459), RTG: 795 (380), PL: 814 (398)	AC, COMTI, amantadine, and MAOBI	23 wks	2–4, 63.2 (9.7), 8.4 (4.7)
	PSG [14]	PPX (109), PL (35): 0.375–4.5	PPX: 21.1%, PL: 11.4%	100%; PPX: 278.9 (211.6), PL: 272.9 (204.1)	NR	10 wks	2–4 (2.5, 0.5), 64.8 (10.6), 6.1 (5.1)
	Brodsky et al. [32]	PPX/PL cross-over (13): 3.0	PPX $\uparrow$ PDS scores compared to baseline ( $P = .05$ ), $\uparrow$ peak scores with L-DOPA infusion	100%, 871.2 (448.6); + infusion at 5 + 10 wks	Unclear on which APDs	10 wks–5 wks cross-over	NR, 61.9 (8.0), 10.3 (4.3)

\* Studies were categorised according to the disease stage (early versus advanced, plus Hoehn and Yahr stage, where available).

Abbreviations: N: number of patients, SD: standard deviation, PSG: Parkinson Study Group, NR: not reported, HR: hazard ratio (risk ratio of developing dyskinesia per unit of time for patients assigned to PPX compared to risk ratio for L-DOPA), CI: confidence interval,  $\uparrow$  the report by PSG [22] is an extension of the PSG [31] protocol, H-Y stage: Hoehn-Yahr stage (a staging system to describe PD progression from 0 to 5 with stages 1.5 and 2.5 in the modified version, incorporated into the UPDRS), PPX: pramipexole, PL: placebo, BRC: bromocriptine, L-DOPA: levodopa (with or without carbidopa), PGI: pergolide, RTG: rotigotine, APDs: anti-Parkinson's drugs, MAOBI: monoamine oxidase-B inhibitors (e.g., selegiline), AC: anticholinergics (e.g., orphenadrine, benzhexol), ODD: other dopaminergic drugs (e.g., droxidopa), UPDRS IV: Unified Parkinson's Disease Rating Scale part IV (complications of therapy, higher scores indicate more severe dyskinesia), which also includes subitems on dyskinesia symptoms [33], PDS: Parkinson's Dyskinesia Scale (higher scores indicate more severe dyskinesia), which rates the severity of dyskinesia according to body regions [34].

the other, % UPDRS IV score changes were significantly lower in the PPX compared to the placebo group [30]. When comparing PPX and BRC, Guttman et al. [29] found that the incidence of dyskinesia between these treatment groups was approximately similar (5% difference). On the other hand, dyskinesia was found to be nearly twice as prevalent with PPX treatment compared to BRC [30].

Two studies compared pergolide (PGL) with PPX. One of these studies found that dyskinesia was equally prevalent with PPX and PGL ( $N = 1$ ) but was not reported in the placebo group [23]. Another trial by the same group with a cross-over design found an overall lower incidence of dyskinesia with PPX treatment than PGL: 33.3% versus 37.5% [25]. Transdermally administered rotigotine (RTG) was also compared with PPX [31]. The placebo group had less dyskinesia than the active treatment groups, with PPX having slightly higher incidence than RTG (3% versus 15% versus 12%). Furthermore, this group reported that participants in the PPX group had significantly more time in the day "on" without troublesome dyskinesias compared to placebo ( $P = .0429$ ), whereas the difference between PPX and RTG was not significant.

**4.3. Pramipexole versus L-DOPA.** Only one cohort was involved with a trial comparing PPX and L-DOPA. The first report was the two-year completion of initial treatment with PPX and L-DOPA, with open-label L-DOPA for emerging disability [22]. There was a significantly lower incidence of dyskinesia in the PPX compared to the L-DOPA group: 9.9% versus 30.7%, hazard ratio (HR) 0.33 (confidence interval (CI) 0.18–0.60),  $P < .001$ . Some of the participants who completed the two-year trial were also enrolled for a further two-year follow-up study, with the randomised and blinded protocol maintained [24]. This showed a similar pattern of the incidence of dyskinesia, which was significantly lower in the PPX ( $N = 83$ ) compared to the L-DOPA ( $N = 100$ ) group: 24.5% versus 54.0%, HR 0.37 (95% CI 0.25–0.56),  $P < .001$ . None of the PPX cohort withdrew from follow-up due to dyskinesia, whereas dyskinesia-related discontinuation was found in 2.0% of the L-DOPA group. Some of the patients in the 2- and 4-year trials were also recruited to the open-label extension study with mean follow-up of 6.0 (SD 0.2) years, the majority of whom were in H-Y stages 2 [35]. Both the initial PPX ( $N = 108$ ) and L-DOPA ( $N = 114$ ) groups showed overall reductions in the incidence of dyskinesia: 20.4% and 36.8%, respectively. Despite changes in the incidence of dyskinesia in the treatment groups compared to the previous reports of this cohort, L-DOPA treatment was associated with higher events. Furthermore, disability associated with dyskinesia in the PPX group was comparably lower than that in the L-DOPA group, which was at trend-level significance ( $P = .06$ ).

## 5. Are There Indicators for the Development of Dyskinesia with PPX Treatment?

The majority of placebo-controlled studies demonstrated that dyskinesias can develop during PPX treatment. The inci-

TABLE 2: Pharmacokinetic profiles of selected anti-Parkinson's drugs.

APD	Half-life (hrs)
PPX	8–12
L-DOPA/carbidopa	1–1.5
RTG	5–7
PGL	7–16
BRC	12–15

Abbreviations: APD: anti-Parkinson's drug, PPX: pramipexole, L-DOPA: levodopa, RTG: rotigotine, PGL: pergolide, BRC: bromocriptine.

dence of these events in the placebo groups ranged between 0 and 40.8%, and the differences compared to the PPX group ranged between 4.0 and 21.3%. Two studies demonstrated slightly more dyskinesia in the placebo compared to the PPX group [17, 18]. These results clearly indicate that there are non-PPX-related factors contributing to the development of dyskinesias in PD.

Treatment of L-DOPA has been proposed as an important factor for the development of dyskinesia in PD [4]. From the two trials and the long-term follow-up of their cohort [22, 24, 35], the PSG consistently demonstrated a higher incidence of dyskinesia associated with L-DOPA therapy. Furthermore, Brodsky et al. [32] showed that PPX treatment increased L-DOPA-related dyskinesias and increased the severity and duration of dyskinesia. They hypothesised that the observed effects in their study are beyond potential additive effects, given that DAAs rarely cause dyskinesia. The 2-hour infusion of therapeutic dose L-DOPA (1.0 mg/kg/hr) also produced more dyskinesia compared to subtherapeutic doses (0.5 mg/kg/hr), regardless of PPX treatment. These findings suggest that L-DOPA on its own can have some effect on dyskinesia events during PPX therapy. Although a study distinguished between L-DOPA and non-L-DOPA-treated participants, the report about adverse event-related withdrawals (including dyskinesia) in the L-DOPA group was not supplemented by information about what proportions were affected [12]. The effects of L-DOPA in other studies are also less clear because L-DOPA usage is reported for the whole sample and there is no differentiation of which patients were dyskinetic.

The literature also allows limited exploration from clinical studies whether continuous dopaminergic stimulation (CDS) is a protective factor for dyskinesia [36]. CDS is a proposed strategy to prevent fluctuations in DA transmission and therefore the development of dyskinesia [37]. Studies with PPX and other DAAs as active comparators show similar incidence of dyskinesia. In all studies with placebo comparisons, DAAs have consistently been associated with more dyskinesia events, suggesting the involvement of dopaminergic activation. In terms of the pharmacokinetic profile of PPX (Table 2), its longer half-life compared to L-DOPA makes it difficult to explain the higher incidence of dyskinesias with this treatment. In fact, the shorter half-life of L-DOPA compared to other antiparkinsonian medications (such as dopamine D2 receptor agonists) is regarded as a factor contributing to its pulsatile action and

ultimately to dyskinesia [38]. Moreover, studies on the ER preparation of PPX, which produces a continuous release of active ingredient over a twenty-four-hour period [39], did not find high rates of dyskinesia. Finally, there were no differences whether the DAA was ergoline (BRC, PGL) or non-ergoline-based (PPX, RTG). It has therefore been suggested that previous exposure to L-DOPA (i.e., priming) can lead to increased susceptibility to develop dyskinesias after exposure to drugs which would not otherwise have had this effect. Specifically, pulsatile activation of type D2 dopamine receptors is reported to be the principal factor in the triggering of dyskinesias and may well be involved in the priming phenomenon [40].

Dose-ranging studies of PPX allow some degree of examination of the hypothesis that PPX-related dyskinesia may be dosage dependent. One study examining PPX at doses of 1.5, 3.0, 4.5, and 6.0 mg did not report any incidence of dyskinesia in either treatment or placebo groups [21]. Another study of different schedules of low-dose PPX (0.5 bd, 0.75 bd, and 0.5 mg td) also did not report dyskinesic events in the participants [28]. These findings suggest that dyskinesia may not be PPX dose dependent. Exploration of this hypothesis with other studies is difficult because PPX dosages are reported as a mean or range of values for the whole sample. Therefore, it is not possible to determine what doses were administered to patients with dyskinesia.

Due to insufficient details in published reports, it has also been difficult to explore whether concomitant APD usage or patient characteristics (such as illness stage, age, and duration of disease) are associated with dyskinesia. APDs were either kept constant at baseline dosages or used as add-on therapy for emerging disability and the majority of studies allowed concomitant usage. Furthermore, patient characteristics were reported as mean or range values without differentiating which patients exhibited dyskinesia. However, it is interesting to note that patients with dyskinesias were generally older (early sixties and above). The patients in PPX-only studies in which there was no incidence of dyskinesia were newly diagnosed with PD.

In terms of methodological issues, the duration of the treatment phase (i.e., titration and maintenance) has overall been adequate to allow sufficient time in detecting or establishing a timeframe for the onset of dyskinesia. The sample size has also generally been statistically viable to allow detection of clinically relevant findings, including dyskinesic adverse events, although there have been some studies with small sample sizes. Most studies also maintained some equivalency with patient characteristics in their treatment groups by carrying out block randomisations and statistical tests before and after treatment to investigate related differences in outcome measures.

More problematic issues, however, may confound conclusions about the effects of PPX on dyskinesia. In particular, some studies only reported adverse events including dyskinesia that occurred above threshold incidence, for example,  $\geq 2\%$ ,  $\geq 5\%$ , or  $10\%$ . Thus, PPX-related dyskinesias may be underreported and contribute to the lower incidence in certain studies. Indeed, the studies which did not detect dyskinesia in their participants had thresholds of reporting

these events at  $\geq 5\%$  [26–28] and  $10\%$  [19]. There have also been studies in which patients already experiencing motor fluctuations including dyskinesia were enrolled for PPX treatment. These studies examined whether PPX is a suitable adjuvant medication for L-DOPA-related dyskinesia. However, from these studies it is difficult to establish whether PPX can contribute to the development of dyskinesias since it was not described whether the incidence of dyskinesia in treatment-emergent and relevant neurological scales were not used to measure effects on these symptoms. The reporting of dyskinesia in follow-up studies also does not allow the differentiation between patients with treatment-emergent events and those who continued to experience such symptoms. Additionally, the combination of ratings of dyskinesia and neurological scales to measure effects of PPX further confound conclusions without differentiating which patients had dyskinesic events. For example, the incidence of dyskinesia may be high in the PPX group even if the scales show significant improvement associated with PPX treatment [10, 11, 13].

## 6. Conclusions

We systematically reviewed the existing evidence on the use of PPX in PD with focus on the development of dyskinesia. Treatment with PPX has overall been associated with significantly better improvement in motor and daily function compared to placebo. The majority of placebo-controlled studies demonstrated that dyskinesias can develop during PPX treatment, as these symptoms were generally (except for two studies) more prevalent compared to the placebo groups. Therefore, the evidence in support of lower incidence of dyskinesia in combination with PPX is far from convincing. However, in six studies, no dyskinesia events were reported. Four of these six studies were PPX comparisons (i.e., dose-ranging or immediate versus extended release preparation). Active comparator studies showed that the incidence of dyskinesia events was quite similar relative to other DAAs. An L-DOPA comparison study in one cohort with long-term follow-up consistently showed that PPX treatment was associated with lower risk for developing dyskinesia. These symptoms have also been reported in the placebo groups (albeit at generally lower rates), suggesting the potential involvement of non-PPX-related factors in the development of dyskinesia. It is still to be established whether L-DOPA treatment, PPX dosage, and other patient characteristics such as age or disease stage can play a role as contributory factors. Elucidation of such factors is likely to optimise the efficacy of anti-Parkinson's treatment and its compliance.

## References

- [1] J. B. Schulz, "Update on the pathogenesis of Parkinson's disease," *Journal of Neurology*, vol. 255, no. 5, supplement, pp. 3–7, 2008.
- [2] Parkinson Study Group, "A randomised controlled trial comparing pramipexole with levodopa in early Parkinson's disease: design and methods of the CALM-PD Study," *Clinical Neuropharmacology*, vol. 23, no. 1, pp. 34–44, 2000.

- [3] A. Schrag, Y. Ben-Shlomo, R. Brown, C. D. Marsden, and N. Quinn, "Young-onset Parkinson's disease revisited - Clinical features, natural history, and mortality," *Movement Disorders*, vol. 13, no. 6, pp. 885–894, 1998.
- [4] R. A. Hauser, M. P. McDermott, and S. Messing, "Factors associated with the development of motor fluctuations and dyskinesias in Parkinson disease," *Archives of Neurology*, vol. 63, no. 12, pp. 1756–1760, 2006.
- [5] C. Marras, A. Lang, M. Krahn, G. Tomlinson, and G. Naglie, "Quality of life in early Parkinson's disease: impact of dyskinesia and motor fluctuations," *Movement Disorders*, vol. 19, no. 1, pp. 22–28, 2004.
- [6] S. Chapuis, L. Ouchchane, O. Metz, L. Gerbaud, and F. Durif, "Impact of the motor complications of Parkinson's disease on the quality of life," *Movement Disorders*, vol. 20, no. 2, pp. 224–230, 2005.
- [7] J. L. Montastruc, O. Rascol, and J. M. Senard, "Treatment of Parkinson's disease should begin with a dopamine agonist," *Movement Disorders*, vol. 14, no. 5, pp. 725–730, 1999.
- [8] W. J. Weiner, "The initial treatment of Parkinson's disease should begin with levodopa," *Movement Disorders*, vol. 14, no. 5, pp. 716–724, 1999.
- [9] R. Constantinescu, "Update on the use of pramipexole in the treatment of Parkinson's disease," *Neuropsychiatric Disease and Treatment*, vol. 4, no. 2, pp. 337–352, 2008.
- [10] A. Lieberman, A. Ranhosky, and D. Korts, "Clinical evaluation of pramipexole in advanced Parkinson's disease: results of a double-blind, placebo-controlled, parallel-group study," *Neurology*, vol. 49, no. 1, pp. 162–168, 1997.
- [11] M. M. Pinter, O. Pogarell, and W. H. Oertel, "Efficacy, safety, and tolerance of the non-ergoline dopamine agonist pramipexole in the treatment of advanced Parkinson's disease: a double blind, placebo controlled, randomised, multicentre study," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 66, no. 4, pp. 436–441, 1999.
- [12] K. S. Wong, C. S. Lu, D. E. Shan, C. C. Yang, T. H. Tsoi, and V. Mok, "Efficacy, safety, and tolerability of pramipexole in untreated and levodopa-treated patients with Parkinson's disease," *Journal of the Neurological Sciences*, vol. 216, no. 1, pp. 81–87, 2003.
- [13] J. C. Möller, W. H. Oertel, J. Köster, G. Pezzoli, and L. Provinciali, "Long-term efficacy and safety of pramipexole in advanced Parkinson's disease: results from a European multicenter trial," *Movement Disorders*, vol. 20, no. 5, pp. 602–610, 2005.
- [14] C. Tanner, C. Comella, C. Kamp et al., "Pramipexole in levodopa-treated Parkinson disease patients of African, Asian, and Hispanic heritage," *Clinical Neuropharmacology*, vol. 30, no. 2, pp. 72–85, 2007.
- [15] P. Bxarone, W. Poewe, S. Albrecht et al., "Pramipexole for the treatment of depressive symptoms in patients with Parkinson's disease: a randomised, double-blind, placebo-controlled trial," *The Lancet Neurology*, vol. 9, no. 6, pp. 573–580, 2010.
- [16] W. J. Weiner, S. A. Factor, J. Jankovic et al., "The long-term safety and efficacy of pramipexole in advanced Parkinson's disease," *Parkinsonism and Related Disorders*, vol. 7, no. 2, pp. 115–120, 2001.
- [17] K. M. Shannon, J. P. Bennett, and J. H. Friedman, "Efficacy of pramipexole, a novel dopamine agonist, as monotherapy in mild to moderate Parkinson's disease," *Neurology*, vol. 49, no. 3, pp. 724–728, 1997.
- [18] L. Wermuth and The Danish Pramipexole Study Group, "A double-blind, placebo-controlled, randomised, multi-centre study of pramipexole in advanced Parkinson's disease," *European Journal of Neurology*, vol. 5, pp. 235–242, 1998.
- [19] O. Pogarell, T. Gasser, J. J. van Hilten et al., "Pramipexole in patients with Parkinson's disease and marked drug resistant tremor: a randomised, double blind, placebo controlled multicentre study," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 72, no. 6, pp. 713–720, 2002.
- [20] J. P. Hubble, W. C. Koller, N. R. Cutler et al., "Pramipexole in patients with early Parkinson's disease," *Clinical Neuropharmacology*, vol. 18, no. 4, pp. 338–347, 1995.
- [21] K. Kieburtz, "Safety and efficacy of pramipexole in early Parkinson disease: a randomized dose-ranging study," *Journal of the American Medical Association*, vol. 278, no. 2, pp. 125–130, 1997.
- [22] R. Holloway, I. Shoulson, K. Kieburtz et al., "Pramipexole vs Levodopa as initial treatment for Parkinson disease: a randomized controlled trial," *Journal of the American Medical Association*, vol. 284, no. 15, pp. 1931–1938, 2000.
- [23] P. Navan, L. J. Findley, J. A. R. Jeffs, R. K. B. Pearce, and P. G. Bain, "Randomized, double-blind, 3-month parallel study of the effects of pramipexole, pergolide, and placebo on Parkinsonian tremor," *Movement Disorders*, vol. 18, no. 11, pp. 1324–1331, 2003.
- [24] Parkinson Study Group, "Pramipexole vs levodopa as initial treatment for Parkinson disease: a 4-year randomised controlled trial," *Archives of Neurology*, vol. 61, pp. 1044–1053, 2004.
- [25] P. Navan, L. J. Findley, M. B. Undy, R. K. B. Pearce, and P. G. Bain, "A randomly assigned double-blind cross-over study examining the relative anti-Parkinsonian tremor effects of pramipexole and pergolide," *European Journal of Neurology*, vol. 12, no. 1, pp. 1–8, 2005.
- [26] R. A. Hauser, A. H. Schapira, O. Rascol et al., "Randomized, double-blind, multicenter evaluation of pramipexole extended release once daily in early Parkinson's disease," *Movement Disorders*, vol. 25, no. 15, pp. 2542–2549, 2010.
- [27] O. Rascol, P. Barone, R. A. Hauser et al., "Efficacy, safety, and tolerability of overnight switching from immediate- to once daily extended-release pramipexole in early Parkinson's disease," *Movement Disorders*, vol. 25, no. 14, pp. 2326–2332, 2010.
- [28] Parkinson Study Group, "Twice-daily, low-dose pramipexole in early Parkinson's disease: a randomized, placebo-controlled trial," *Movement Disorders*, vol. 26, no. 1, pp. 37–44, 2011.
- [29] M. Guttman, "Double-blind comparison of pramipexole and bromocriptine treatment with placebo in advanced Parkinson's disease," *Neurology*, vol. 49, no. 4, pp. 1060–1065, 1997.
- [30] Y. Mizuno, N. Yanagisawa, S. Kuno et al., "Randomized, double-blind study of pramipexole with placebo and bromocriptine in advanced Parkinson's disease," *Movement Disorders*, vol. 18, no. 10, pp. 1149–1156, 2003.
- [31] W. H. Poewe, O. Rascol, N. Quinn et al., "Efficacy of pramipexole and transdermal rotigotine in advanced Parkinson's disease: a double-blind, double-dummy, randomised controlled trial," *The Lancet Neurology*, vol. 6, no. 6, pp. 513–520, 2007.
- [32] M. A. Brodsky, B. S. Park, and J. G. Nutt, "Effects of a dopamine agonist on the pharmacodynamics of levodopa in parkinson disease," *Archives of Neurology*, vol. 67, no. 1, pp. 27–32, 2010.
- [33] S. Fahn, R. L. Elton, and UPDRS Development Committee, "Unified Parkinson's disease rating scale," in *Recent Developments in Parkinson's Disease*, S. Fahn, C. D. Marsden, D. B. Calne, and M. Goldstein, Eds., pp. 153–163, Macmillan, Florham Park, NJ, USA, 1987.

- [34] W. Guy, "Abnormal involuntary movement scale," in *The ECDEU Assessment Manual for Psychopharmacology*, US Department of Health Education and Welfare, National Institute of Health Education and Welfare, and National Institute of Mental Health, Eds., pp. 534–537, US Government Printing Office, Washington, DC, USA, 1976.
- [35] R. Holloway, K. Marek, K. Biglan et al., "Long-term effect of initiating Pramipexole vs Levodopa in early Parkinson disease," *Archives of Neurology*, vol. 66, no. 5, pp. 563–570, 2009.
- [36] P. Jenner, "Avoidance of dyskinesia: preclinical evidence for continuous dopaminergic stimulation," *Neurology*, vol. 62, no. 1, pp. S47–S55, 2004.
- [37] P. Barone, "Clinical strategies to prevent and delay motor complications," *Neurology*, vol. 61, no. 6, supplement, pp. S12–S16, 2003.
- [38] L. A. Smith, M. J. Jackson, L. Johnston et al., "Switching from levodopa to the long-acting dopamine D2/D3 agonist piribedil reduces the expression of dyskinesia while maintaining effective motor activity in MPTP-treated primates," *Clinical Neuropharmacology*, vol. 29, no. 3, pp. 112–125, 2006.
- [39] W. Eisenreich, B. Sommer, S. Hartter, and W. H. Jost, "Pramipexole extended release: a novel treatment option in Parkinson's disease," *Parkinson's Disease*, vol. 2010, Article ID 612619, 7 pages, 2010.
- [40] P. Damier, L. Tremblay, J. Féger, and E. C. Hirsch, "Development of dyskinesias induced by treatment for parkinson's disease: potential role of first exposure to L-DOPA (priming)," *Revue Neurologique*, vol. 156, no. 3, pp. 224–235, 2000.

## Review Article

# Imbalanced Dopaminergic Transmission Mediated by Serotonergic Neurons in L-DOPA-Induced Dyskinesia

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L-DOPA-induced dyskinesias (LIDs) are one of the main motor side effects of L-DOPA therapy in Parkinson's disease. The review will consider the biochemical evidence indicating that the serotonergic neurons are involved in the dopaminergic effects of L-DOPA in the brain. The consequences are an ectopic and aberrant release of dopamine that follows the serotonergic innervation of the brain. After mid- to long-term treatment with L-DOPA, the pattern of L-DOPA-induced dopamine release is modified. In several brain regions, its effect is dramatically reduced while, in the striatum, its effect is quite preserved. LIDs could appear when the dopaminergic effects of L-DOPA fall in brain areas such as the cortex, enhancing the subcortical impact of dopamine and promoting aberrant motor responses. The consideration of the serotonergic system in the core mechanism of action of L-DOPA opens an important reserve of possible strategies to limit LIDs.

## 1. Introduction

Parkinson's disease is the second most devastating neurodegenerative disease affecting more than 6 million people worldwide and whose prevalence is expected to double within the next twenty years [1]. This neurological disorder is characterized by the progressive loss of mesencephalic dopaminergic (DA) neurons from the substantia nigra pars compacta and associated with numerous motor symptoms (bradykinesia, rigidity, and tremor) [2, 3]. L-DOPA, the precursor of DA, has been introduced in the mid 60's as a miracle pill to prevent the motor symptoms [4, 5]. However, upon chronic use of this medication, its efficacy slowly decreases leading to increase the doses of L-DOPA, which generate numerous side effects. After 5 to 10 years of L-DOPA treatment, Parkinsonian patients develop dyskinesias [6], which consist of stereotypical choreic or ballistic movements involving mostly the head, trunk, and limbs [7]. These abnormal involuntary movements are often more debilitating than the motor symptoms themselves.

Preclinical research has permitted validating animal models to study the mechanisms of L-DOPA-induced dysk-

inesias (LIDs). The most commonly used rat model that shows best face and predictive validity, has been developed by Cenci and collaborators [8, 9] by producing severe lesion of the nigrostriatal DA pathway in adult rats with the unilateral injection of 6-hydroxydopamine (6-OHDA) in the medial forebrain bundle [10, 11]. A chronic treatment with L-DOPA for 3 weeks at low therapeutic doses (6–10 mg/kg) induced axial, limb, and orolingual abnormal involuntary movements (ALO AIMS) of variable occurrence and severity in rats [9, 12]. Despite extensive research done to understand how these motor complications develop in the Parkinsonian brain, all hypotheses could not be fully validated and new insights in this field need to be pushed forward to further gain in understanding of LIDs. In the present review, we will focus on the literature showing a prominent role of serotonergic neurons (5-HT) in the mechanisms of action of L-DOPA and how these neurons may contribute to the development of LIDs. Specifically, we will try to develop a new hypothesis that LIDs appear when the effect of L-DOPA falls in brain areas such as the cortex, then enhancing the subcortical impact of DA at the risk to elicit LIDs.

## 2. Mechanism of Action of L-DOPA in 5-HT Neurons and Collateral Consequences

It has long been thought that the therapeutic benefit of L-DOPA may depend on its ability to restore DA extracellular levels in the striatum through spared DA neurons [13–15]. However, contradictory data have shown that the fewer DA neurons that are spared, the more pronounced is the release of DA induced by L-DOPA [16–21]. Furthermore, L-DOPA-induced DA release is not sensitive to DA autoregulatory processes (DA-D2 autoreceptor stimulation and DAT blockade) [19]. Other monoaminergic cells [22, 23], namely serotonergic (5-HT) neurons, that are able to convert L-DOPA into DA, store and induce an exocytotic release of DA, rather participate in the mechanism of action of L-DOPA [24].

**2.1. L-DOPA and 5-HT Neurons.** 5-HT neurons express the amino acid decarboxylase (AADC) that converts L-DOPA into DA and the vesicular membrane transporter VMAT2 that packages DA into exocytosis vesicles [25–28]. In line with these molecular features, 5-HT neurons have been shown for several years to release the newly synthesized DA from their cell bodies and terminals [25, 29, 30]. Indeed, 5-HT neurons are responsible for the TTX-sensitive, reserpine-sensitive, and DA drugs-insensitive release of DA induced by L-DOPA. The lesion of 5-HT neurons by the selective neurotoxin 5,7-DHT drastically reduces the increase in DA extracellular levels induced by a wide range of L-DOPA doses (3–100 mg/kg) [31, 32]. This effect is dependent on the extent of 5-HT denervation [31], which excludes the involvement of any other cellular system in the release of DA induced by L-DOPA. Furthermore, L-DOPA-induced DA release is sensitive to 5-HT autoregulatory mechanisms. Both the stimulation of 5-HT<sub>1A</sub> autoreceptors by the 5-HT<sub>1A</sub> agonist 8-OHDPAT [33] and the blockade of 5-HT transporters (SERT) by the selective serotonergic reuptake inhibitors (SSRI) fluoxetine [34] or citalopram [31] reduce the increase in L-DOPA-derived DA extracellular levels. These effects are thought to occur *via* the inhibition of 5-HT neuron activity [35–42]. Accordingly, it has been recently shown that high-frequency stimulation of the subthalamic nucleus, a surgical approach in Parkinson's disease able to inhibit 5-HT neuronal firing [43], also reduces L-DOPA-induced DA release [44].

5-HT neurons send a widespread innervation from the raphe nuclei to the entire forebrain including the striatum [46, 47]. Beyond the increase in striatal DA extracellular levels, L-DOPA also induces a massive rise in DA levels in the prefrontal cortex (PFC), the substantia nigra pars reticulata (SNr), and the hippocampus (HIPP) [31]. In all brain regions, L-DOPA-induced DA release is sensitive to 5-HT pharmacological manipulation and the lesion of 5-HT neurons [31, 44, 48]. This ectopic release of DA induced by L-DOPA *via* 5-HT neurons creates a new balance in DA chemistry throughout the Parkinsonian brain (Figure 1) [24, 31]. In physiological conditions, basal DA concentrations are more than 30 times higher in the striatum compared to other brain regions, in line with the restricted innervation

of mesencephalic DA neurons to striatal territories [5, 49]. While DA extracellular levels are from 4.6 to 7.8 fmol/uL, they are barely detectable depending on experimental conditions (below 0.2 fmol/uL) in the PFC, SNr, and HIPP although DA receptors are expressed [50]. In Parkinsonian conditions, the dose of L-DOPA required to “restore” similar DA concentrations in the DA-denervated striatum is about 12 mg/kg while it increases about 10 to 25 times DA concentrations in other brain regions (see Figure 1). Interestingly, L-DOPA at 3 mg/kg enhances DA levels to similar amounts (0.7 to 1.3 fmol) in the PFC, SNr, HIPP, and striatum. Therefore, huge amounts of DA can be released beyond the striatum [51] and may impact on DA receptors throughout the Parkinsonian brain. In keeping with the increased sensitivity of DA receptors that develops after DA denervation [52–54], such an imbalanced DA transmission between the striatum and other brain regions may participate in the emergence in both short-term benefits and long-term side effects of L-DOPA treatment (see Section 4).

**2.2. Chronic Impact of L-DOPA on DA Release Pattern in the Entire Forebrain.** The therapeutic efficacy of L-DOPA treatment decreases over time with the development of numerous side effects including L-DOPA-induced dyskinesias (LIDs). LIDs are thought to emerge as a consequence of the dysregulated release of DA as a “false neurotransmitter” from 5-HT neurons [12, 31, 33, 44, 48, 57–63]. Indeed, the inhibition of L-DOPA-induced DA release by 5-HT<sub>1</sub> autoreceptors stimulation [33, 48] and/or 5,7-DHT lesion [31, 32] is associated with a marked reduction in LIDs [48, 61]. However, these mechanisms have been described mostly in the striatum while other brain regions could be involved in the development of LIDs [60, 64–69]. Furthermore, the dose of L-DOPA used, even within the therapeutic range (3–12 mg/kg), represents a critical parameter to consider in the understanding of LIDs [70].

The occurrence and severity of LIDs in animals treated chronically with L-DOPA depend on numerous parameters, that is, the dose of L-DOPA, the site of 6-OHDA injection, the extent of DA lesion, and rat strain. About half of animals treated chronically with 3 mg/kg of L-DOPA develop LIDs. At 6 mg/kg, about 2/3 of the animals treated with L-DOPA display severe LIDs. At 12 mg/kg and above, almost all animals develop LIDs [9]. One consistent result observed after chronic L-DOPA treatment is that, whatever the dose, basal DA extracellular levels remain barely detectable in all brain regions. In our experimental conditions (12 mg/kg for 10 days), basal DA levels were below the detection limit in the striatum, SNr, PFC, and HIPP (Figure 1) [45]. In another study using a 14-day treatment with 6 mg/kg L-DOPA, baseline DA concentrations were reduced by 99% to 0.04 fmol/ $\mu$ L in the striatum compared to intact animals (4 fmol/ $\mu$ L) without changes in the SNr (0.1–0.2 fmol/ $\mu$ L) [48]. One study showed a slight increase in basal DA levels in the striatum (reaching about 0.6 fmol/ $\mu$ L) by using a higher dose (25 mg/kg) of L-DOPA administered twice a day [81].

Dynamics in the increase in DA release after each L-DOPA administration may, however, differ regarding the dose of L-DOPA used. Some authors have proposed that

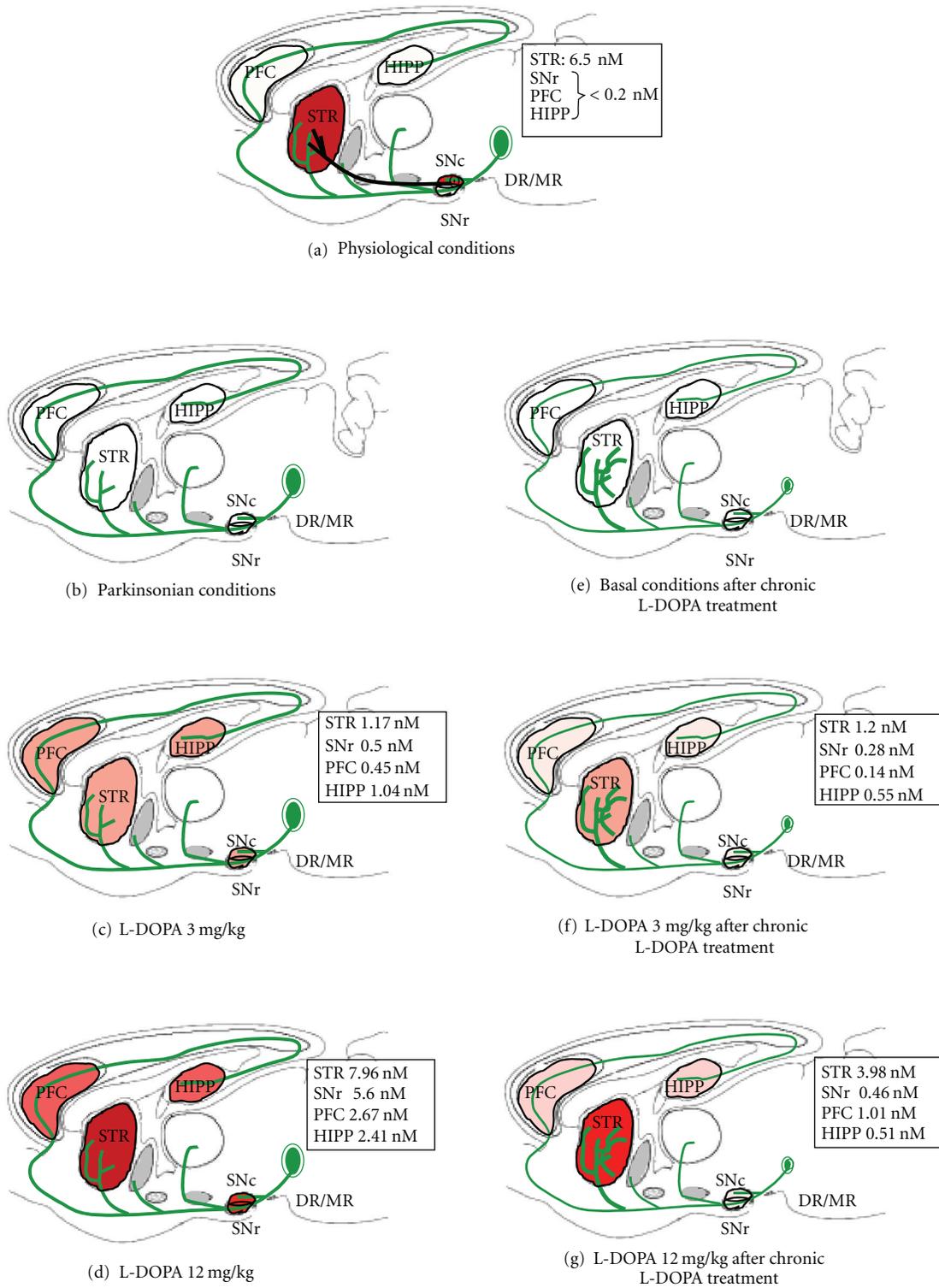


FIGURE 1: Serotonergic neurons are responsible for an imbalance of dopamine chemistry within brain regions in the Parkinsonian brain after acute and chronic L-DOPA treatment. Data taken from [31, 45].

LIDs may emerge as a consequence of abnormal fluctuations in synaptic DA levels induced by L-DOPA treatment in dyskinetic animals [48, 58, 59, 61, 73]. Larger increases in synaptic DA levels induced by L-DOPA have been proposed to be responsible for the emergence of peak-dose dyskinesia in PD patients [59]. Data obtained with a chronic L-DOPA treatment at 6 mg/kg have shown that the kinetics of DA release are different in animals developing LIDs or not [73]. Although a higher magnitude of DA release was observed in the striatum and SNr of dyskinetic animals compared to nondyskinetic animals [48], this has not been consistently observed [73]. In a recent report, our data have provided new evidence for reconsidering the mechanisms of L-DOPA within the Parkinsonian brain and the putative consequences in many side effects including LIDs [45]. We showed that after a chronic L-DOPA treatment at 12 mg/kg for 10 days, the reactivity of 5-HT neurons to an acute challenge at 3 or 12 mg/kg of L-DOPA was modified and resulted in a potent loss of efficacy of L-DOPA to increase DA release (Figure 1). Most importantly, our data could depict a new imbalance created by chronic L-DOPA treatment within the striatum and other brain regions. The capacity of 5-HT neurons to increase DA release in the SNr, HIPP, and PFC was drastically reduced (about 70 to 90%) while it was less affected in the striatum. Indeed, the increase in striatal DA release induced by 3 mg/kg of L-DOPA after a 12 mg/kg treatment for 10 days was similar to that induced by an acute administration of 3 mg/kg. At 12 mg/kg, the effect of L-DOPA was reduced by only 50% after chronic compared to acute treatment. It appears that different mechanisms may be processed in the striatum compared to other brain structures that may account for the relatively preserved striatal DA effect of L-DOPA. Some of these mechanisms may be directly related to the specific features and heterogeneity of 5-HT terminals within brain regions. The resulted imbalance between cortical versus subcortical brain regions in DA transmission may potentially participate in development of LIDs.

The following paragraph corresponds to the description of the Figure 1, (a) in physiological conditions, dopaminergic neurons originating from the substantia nigra pars compacta (SNc) densely innervate the striatum (STR) where basal dopamine (DA) concentrations range between 4.6 and 7.8 nM. In the prefrontal cortex (PFC), the hippocampus (HIPP), and the substantia nigra pars reticulata (SNr), basal DA concentrations are much lower (<0.2 nM). All these brain regions express DA receptors and are innervated by serotonergic neurons that originate from the dorsal and medial raphe nuclei (DR/MR). (b) In Parkinsonian conditions (i.e., unilateral 6-hydroxydopamine lesion in rats, 6-OHDA rats), the neurodegeneration of DA neurons leads to undetectable levels of DA in any brain region examined. (c) In the absence of DA neurons, L-DOPA is decarboxylated into DA, stored into exocytosis vesicles, and released in the extracellular space by serotonergic neurons. In such physiopathological condition, an acute administration of L-DOPA at the low therapeutic dose of 3 mg/kg induces a homogeneous increase in DA concentrations in all brain regions (see values in the square box). These concentrations

are 2, 2.5, and 5 times higher than in physiological conditions in the SNr, PFC, and HIPP, respectively, while they are 5 times lower in the STR. (d) An acute administration of L-DOPA at the moderate therapeutic dose of 12 mg/kg increases DA concentrations in the STR within the range of physiological values. Similar concentrations of DA are observed in the SNr and corresponded to >25 times the physiological concentrations. In the PFC and HIPP, DA concentrations are >10 times higher than in physiological conditions. (e) After a chronic L-DOPA treatment at a dose known to induce dyskinesias in all 6-OHDA rats (12 mg/kg/day for 10 days), basal DA concentrations remain below the detection limit in all brain regions. All biochemical 5-HT indexes (extracellular and tissue levels of 5-HT and its metabolite 5-HIAA) are decreased after chronic L-DOPA treatment, suggesting that 5-HT neurons suffer from chronic exposure to L-DOPA. Numerous data provide evidence for a 5-HT sprouting occurring specifically in the striatum [55, 56]. (f) After a chronic L-DOPA treatment (12 mg/kg/day for 10 days), a subsequent administration of 3 mg/kg L-DOPA is less efficient to increase DA release in the SNr, PFC, and HIPP compared to an acute administration of the same dose in L-DOPA-naïve 6-OHDA rats (see (c)). The ability of L-DOPA to increase DA levels is reduced by 43%, 68%, and 45% in the SNr, PFC, and HIPP, respectively. However, the efficacy of L-DOPA is not altered in the STR as DA levels reached similar concentrations in both L-DOPA-treated and L-DOPA-naïve 6-OHDA rats. (g) The ability of L-DOPA at 12 mg/kg to increase DA release is diminished in all brain regions after chronic L-DOPA treatment (12 mg/kg/day for 10 days). The highest loss of efficacy is observed in the SNr (-92%), then in the HIPP (-79%) and the PFC (62%). By comparison, the efficacy of L-DOPA remained mostly preserved in the STR (-50%), an effect that may be related to the striatal 5-HT hyperinnervation [55].

### 3. Changes in 5-HT Transmission Associated with L-DOPA Treatment

L-DOPA, by entering 5-HT neurons, mediates numerous changes in 5-HT neuron homeostasis [45]. The production of massive amounts of DA has tremendous impact on 5-HT function at the level of the metabolism, the activity, and the morphology of 5-HT neurons (Table 1). Changes in 5-HT indexes have been associated with the emergence of LIDs (Table 2). Such changes may represent critical indicators of the physiopathological state of the Parkinsonian brain that should be taken into consideration to better control 5-HT transmission and L-DOPA's side effects [24, 82, 83].

**3.1. Impact of L-DOPA on 5-HT Transmission.** Since the beginning of the 70's, numerous evidences started accumulating for an alteration of 5-HT neuron activity in response to L-DOPA (Table 1) [86–88]. The first report in 1970 by Ng et al. [25] showed that L-DOPA-enhanced [<sup>3</sup>H]-5-HT release from [<sup>3</sup>H]-5-HT preloaded midbrain slices. This increase in 5-HT release has been later confirmed *in vivo* by local administration of L-DOPA in the substantia nigra [71, 89].

TABLE 1: Changes in biochemical, morphological, and molecular 5-HT indexes in response to L-DOPA treatment.

Animal model	L-DOPA treatment	Biochemical 5-HT indexes	% of change	Reference
[ <sup>3</sup> H]-5-HT preloaded rat midbrain slices naive rats	10 $\mu$ M	[ <sup>3</sup> H]-5-HT release	+60%	[25, 29, 30]
6-OHDA rats	intra-SNr 5 $\mu$ M	ext 5-HT: STR and SNr	+55% in STR +102% in SNr	[71]
	3, 6, 12, 100 mg/kg/d ip	ext 5-HT: STR, SNr, HIPp, PFC	3: $\emptyset$ 6: STR/PFC $\emptyset$ , SNr -22%, HIPp -27%	[31] + unpublished observations
			12: STR/HIPp $\emptyset$ , SNr -17%, PFC -27%	
6-OHDA rats	12 mg/kg/d ip 14 d	tiss 5-HT: STR	100: STR/SNr/HIPp $\emptyset$ , PFC -28% 3-12: $\emptyset$	[31] [45]
		ext 5-HT and 5-HIAA: STR, SNr, HIPp, PFC	100: -73% 5-HT: STR -39%, SNr -45%, HIPp -29%, PFC -47%	
		tiss 5-HT and 5-HIAA: STR and CX	5-HIAA: STR -32%, SNr -58%, HIPp -44%, PFC -51% 5-HT: STR -48%, CX -63%, 5-HIAA: STR -67%, CX -73%	
6-OHDA rats	6 mg/kg/d ip 14 d	ext 5-HT and 5-HIAA: STR and SNr	5-HT: STR-LID +125%, STR-LND +75%; SNr-LID +104%, SNr-LND +108%	[48]
		tiss 5-HT and 5-HIAA: STR and SNr	5-HIAA: STR-LID -30%, STR-LND -73%; SNr-LID -28%, SNr-LND -37% 5-HT: STR-LID -32%, STR-LND -78%	
6-OHDA rats	6 mg/kg/d ip 14 d	tiss 5-HT: STR	5-HIAA: STR-LND -76% -48%	[61]
6-OHDA rats	6 mg/kg/d ip 21 d	tiss 5-HT: STR	+150%	[72]

TABLE 1: Continued.

Animal model	L-DOPA treatment	Morphological 5-HT indexes	% of change	Reference
6-OHDA rats	5 mg/kg/d ip 14 d	SERT immunoreactivity: STR	+266%	[73]
6-OHDA rats	6 mg/kg/d ip 21 d	5-HT immunoreactivity: STR	+70% in STR-LID ∅ in STR-LND	[74]
6-OHDA rats	6 and 50 mg/kg/d ip 14–21 d	SERT-binding density: STR and CX	<b>6:</b> STR +37.5%, CX +75%	
		5-HT immunoreactivity: number of varicosities, STR	<b>50:</b> STR +87.5%, CX +125%	
		5-HT immunoreactivity: synapse incidence in STR	<b>6:</b> +125%	
		SERT-binding density: PUT and GP	<b>50:</b> +200%	[55]
		TPH immunoreactivity: STR and GP	<b>6:</b> +155%	
MPTP monkeys	Modopar (4:1) 15–20 mg/kg po 6–8 m	SERT-binding density: PUT and GP	PUT-LID +72%, GP-LID +400%, LND ∅	[55]
MPTP monkeys	12.5 mg/kg/d po 1 m	TPH immunoreactivity: STR and GP	<b>STR:</b> increased number and size of varicosities and enlargement <b>GP:</b> enlargement in GPI/e + increased number of varicosities and length of fibres in GPe	[56]
Animal model	L-DOPA treatment	Molecular 5-HT indexes	% of change	Reference
6-OHDA mice and rats	(1) mice: 50 mg/kg/d ip 28 d (2) rat: 100 mg/kg 2×d ip 5 d (3) rat: 10 mg/kg/d ip 28 d	5-HT <sub>1B</sub> R binding: STR, GP and SNr	(1) STR +20%, GP ∅, SNr +30% (2) STR +17%, GP +38%, SNr +61% (3) STR +25%, GP ∅, SNr +55%	[75]
6-OHDA rats	100 mg/kg 2×d ip 5 d	5-HT <sub>1B</sub> R protein: STR	+33%	[75]
6-OHDA rats	100 mg/kg 2×d ip 5 d	5-HT <sub>2A</sub> R mRNA: STR	–57%	[76]
		5-HT <sub>2C</sub> R mRNA: STR and STN	∅	
MPTP monkeys	Modopar acute: 14.6 mg/kg po chronic: 14.6 mg/kg 2×d po 120 d Prolopa 100/25 mg/kg po 1 m	5-HT <sub>1A</sub> R-binding: STR, premotor-motor CX, HIPPO	acute: ∅ chronic: +140% in Caud matrix	[77]
MPTP monkeys PD patients (LIDs)		5-HT <sub>2A</sub> R binding: STR and PFC 5-HT <sub>2C</sub> R binding: SNr	+58% in DM Caud +108%	[78] [79, 80]

6-OHDA: 6-hydroxydopamine; MPTP: 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; ip: intraperitoneal; sc: subcutaneous; po: oral; d: day; m: month; 2×d: twice a day; *tiss*: tissue; *ext*: extracellular; 5-HT: serotonin; 5-HIAA: 5-hydroxyindolacetic acid; AADC: amino acid decarboxylase; SERT: serotonergic transporter; 5-HT<sub>1A</sub>R: serotonin 1A receptor; 5-HT<sub>1B</sub>R: serotonin 1B receptor; 5-HT<sub>2A</sub>R: serotonin 2A receptor; 5-HT<sub>2C</sub>R: serotonin 2C receptor; STR: striatum; CX: cortex; PFC: prefrontal cortex; HIPPO: hippocampus; SNr: substantia nigra pars reticulata; PUT: putamen; PFC: prefrontal cortex; STN: subthalamic nucleus; GPI/e: globus pallidus, internal/external part; DM Caud: dorsomedial caudate nucleus; LID: L-DOPA-treated dyskinetic animals; LND: L-DOPA-treated nondyskinetic animals; LIDs: L-DOPA-induced dyskinesias.

TABLE 2: Changes in biochemical, morphological, and molecular 5-HT indexes in dyskinetic (LID) *versus* nondyskinetic (LND) animals: correlation with dyskinesias (R).

Animal model	L-DOPA treatment	Biochemical 5-HT indexes	LID versus LND (R)	Reference
6-OHDA rats	6 mg/kg/d ip 14 d	<i>tiss</i> 5-HT: STR and CX	LID > LND R = 0.73 in CX	[84]
6-OHDA rats	12 mg/kg/d sc 5 d	<i>tiss</i> 5-HT (5,7-DHT): STR	R = 0.713	[85]
6-OHDA rats	6 mg/kg/d sc 14 d	<i>ext</i> and <i>tiss</i> 5-HT and 5-HIAA: STR and SNr	LID > LND in STR	[48]
6-OHDA rats	6 mg/kg/d ip 21 d	<i>tiss</i> 5-HT: STR	LID > LND R = -0.655	[72]
Animal model	L-DOPA treatment	Morphological 5-HT indexes	LID versus LND (R)	Reference
6-OHDA rats	5 mg/kg/d ip 14 d	SERT immunoreactivity: STR	LID > LND	[73]
6-OHDA rats	6 mg/kg/d ip 21 d	5-HT immunoreactivity and AADC levels: STR	LID > LND	[74]
6-OHDA rats	6 and 50 mg/kg/d ip 14–21 d	SERT-binding density: STR and CX	LID > LND, R = 0.796 in STR	[55]
MPTP monkeys	Modopar (4 : 1) 15–20 mg/kg po 6–8 m	SERT-binding density: PUT and GP	LID > LND in PUT	[55]
Animal model	L-DOPA treatment	Molecular 5-HT indexes	LID versus LND (R)	Reference
MPTP monkeys	Prolopa 100/25 mg/kg po 1 m	5-HT <sub>2A</sub> R-binding: STR and PFC	LID > LND in DM Caud and anterior cingulate gyrus	[78]

6-OHDA: 6-hydroxydopamine; MPTP: 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; ip: intraperitoneal; sc: subcutaneous; po: oral; d: day; m: month; *tiss*: tissue; *ext*: extracellular; 5-HT: serotonin; 5-HIAA: 5-hydroxyindolacetic acid; 5,7-DHT: 5,7-dihydroxytryptamine; AADC: amino acid decarboxylase; SERT: serotonergic transporter; 5-HT<sub>2A</sub>R: serotonin 2A receptor; STR: striatum; CX: cortex; SNr: substantia nigra pars reticulata; PUT: putamen; PFC: prefrontal cortex; GP: globus pallidus; DM Caud: dorsomedial caudate nucleus; LID: L-DOPA-treated dyskinetic animals; LND: L-DOPA-treated nondyskinetic animals; R: correlation.

The potent increase in 5-HT levels observed in these studies has been suggested to account for a nonexocytotic efflux of 5-HT *via* 5-HT transporters due to the strong displacement of 5-HT from exocytosis vesicles by the newly synthesized DA [24]. However, recent data using systemic administration of L-DOPA at moderate doses (3–12 mg/kg) have reported distinct effects on 5-HT release depending on the dose and the brain region dialysated. While an acute injection of 12 mg/kg L-DOPA decreases 5-HT extracellular levels in the PFC and SNr, a biphasic effect was observed in the HIPP and no effect in the striatum [44, 45]. A transient increase in 5-HT levels has been observed only after the very high dose of 100 mg/kg in all brain regions (Navailles et al., unpublished observation; see Table 1). Different mechanisms could be triggered regarding the dose of L-DOPA used (exocytotic versus nonexocytotic) while the region-dependent effects of L-DOPA on 5-HT release may reflect the anatomo-functional heterogeneity of 5-HT terminals [24].

After a chronic L-DOPA treatment (12 mg/kg/day for 10 days), the reactivity of 5-HT terminals to a subsequent challenge of L-DOPA (3–12 mg/gk) was further modified in a region-dependent manner [45]. The inhibitory effect of L-DOPA at 3 and 12 mg/kg on 5-HT release was potentiated in the SNr and HIPP of L-DOPA-treated rats but not in the PFC. In the striatum of L-DOPA-treated rats, 5-HT release remained unaltered by L-DOPA whatever the dose

used. Interestingly, this region-dependent reactivity of 5-HT terminals appears to correlate with the ability of L-DOPA to increase DA release after a chronic treatment (see Section 2). Of particular interest, the lack of sensitivity of striatal 5-HT terminals to L-DOPA on 5-HT release is associated with a preserved increase in L-DOPA-induced DA release while the highest sensitivity of 5-HT terminals observed in the SNr leads to the most profound loss of efficacy of L-DOPA to increase DA release [45]. This imbalance between the striatum and the SNr could not be unmasked after a chronic treatment with L-DOPA at 6 mg/kg, which did not change the effect of L-DOPA on 5-HT release [48]. Nevertheless, it appears that, for a moderate though therapeutic dose of L-DOPA (12 mg/kg), its effects on DA transmission occur in detriment of 5-HT transmission [45]. The distinct molecular (variable expression and sensitivity to 5-HT<sub>1A/1B</sub> receptors, SERT, VGLUT3, cation channels) [90–97], anatomical (originating from the medial or dorsal raphe nuclei) [46, 98], ontogenesis (pet1-dependent versus pet1-resistant 5-HT neurons) [99, 100] characteristics of 5-HT neurons projecting to these different brain regions may participate in this region-dependent changes in 5-HT and DA releases [24]. In addition, chronic L-DOPA treatment by itself also alters the morphology of these 5-HT neurons and the synaptic plasticity in various brain regions (Table 1) [55, 56, 101–104], an effect that may participate in the

imbalanced 5-HT and DA transmissions within structures in the Parkinsonian brain and favor the onset of LIDs (see below).

Beyond the changes of 5-HT release (Table 1), chronic L-DOPA also alters 5-HT transmission by modifying the expression and function of numerous 5-HT receptors. Studies that aimed at improving L-DOPA's effects have focused on 5-HT<sub>1A/1B</sub> receptors. Although a decrease in 5-HT<sub>1A</sub> receptor expression in the dorsal raphe (and hippocampus) [105, 106] may be directly linked to the loss of 5-HT neurons in Parkinsonian patients [107–111], an increase has been described in the neocortex of Parkinson's disease patients [112], the putamen [105], caudate nucleus, and middle layers of premotor-motor cortices of MPTP-treated monkeys [77]. Although it remains difficult to attribute these effects to the progression of the disease or to L-DOPA therapy in Parkinsonian patients [107, 113], a massive increase in 5-HT<sub>1A</sub> receptor binding could be observed in the caudate nucleus of L-DOPA-treated MPTP-lesioned monkeys [77]. No alteration in 5-HT<sub>1B</sub> binding has been observed in the striatum and substantia nigra of Parkinsonian patients [114] and 6-OHDA rats [115] while an increase in 5-HT<sub>1B</sub> receptor expression in these brain regions has been reported after chronic L-DOPA treatment in 6-OHDA-lesioned rats [75]. Other 5-HT receptors such as 5-HT<sub>2A</sub> and 5-HT<sub>2C</sub> receptors have been proposed to improve L-DOPA therapy in Parkinson's disease [69, 79, 116–119]. These receptors are known to be sensitive to chronic alteration of DA transmission [119–122]. However, the few data available have reported conflicting results. 5-HT<sub>2A</sub> receptor expression has been shown to increase in the striatum of 6-OHDA rats [76] and the neocortex of Parkinsonian patients [112] while it did not change in the putamen and PFC of MPTP-treated monkeys [78]. Although L-DOPA reversed the increase in the striatum of 6-OHDA rats [76], it increased 5-HT<sub>2A</sub> receptor binding in the dorsomedial caudate nucleus of MPTP-treated monkeys [78]. 5-HT<sub>2C</sub> receptor expression was decreased in the striatum but not in the subthalamic nucleus of 6-OHDA rats without any change after L-DOPA treatment [76]. However, the increased expression of 5-HT<sub>2C</sub> receptors in the substantia nigra pars reticulata in Parkinsonian patients [80] appears to participate in the overactivity of this brain region by dampening the antiparkinsonian action of DA drugs in 6-OHDA rats primed with L-DOPA [120, 121]. Altogether, these data indicate that chronic L-DOPA treatment alters 5-HT function and the resulted changes in 5-HT markers have been mostly associated with the genesis and expression of LIDs.

**3.2. Biochemical, Morphological, and Molecular Changes in 5-HT Indexes Associated with LIDs.** Changes in 5-HT indexes have recently gained growing importance as they may reflect fluctuations in L-DOPA-induced DA release from 5-HT neurons that have been associated with the emergence of LIDs (Table 2) [61, 62]. In this attempt, most studies have focused on modifications of tissue and extracellular levels of 5-HT together with changes in 5-HT terminals density and morphology in 6-OHDA rats developing or not dyskinesias after a chronic treatment with 6 mg/kg of L-DOPA. Con-

flicting results, however, have emerged regarding 5-HT tissue and extracellular levels. Independently of the emergence of LIDs, chronic L-DOPA treatment either reduced [45, 48, 61], did not affect [72, 78], or tended to increase [84] 5-HT tissue and extracellular levels in the striatum. In most studies, however, tissue and extracellular 5-HT levels in the striatum and the cortex, but not the SNr, of rats developing LIDs were significantly higher than in nondyskinetic rats [48, 72, 84] suggesting a positive correlation to LIDs. Accordingly, Eskow et al. [85], by using selective 5,7-DHT lesions that are known to abolish both LIDs [61] and L-DOPA-induced DA release [31], could establish a positive correlation between striatal 5-HT levels and LIDs. These results are in contrast with the study by Gil et al. [72] in which 5-HT tissue levels were negatively correlated to axial, limb, and orolingual abnormal involuntary movements (AIMs). Interestingly, Carta et al. [84] could not establish a link between striatal 5-HT levels but did observe a positive correlation between 5-HT levels in the PFC and AIMs providing further evidence for a role of 5-HT function beyond the striatum in the emergence of LIDs.

In support of an increased 5-HT function in the genesis of LIDs, chronic L-DOPA treatment has been shown to increase AADC protein expression without increasing tyrosine hydroxylase expression in the lesioned-side striatum of dyskinetic rats [74]. This effect was associated with a higher 5-HT immunoreactivity compared to nondyskinetic animals, highlighting an increased 5-HT fiber density mediated by L-DOPA [74]. In line with this, Rylander et al. [55] have shown that chronic L-DOPA induced a dose-dependent increase in SERT-binding densities on the lesioned striatum (and motor-premotor cortices) that was associated with an increased number of striatal 5-HT varicosities but not with an increase in the number of 5-HT cell bodies or expression of SERT mRNA in raphe cells. Both striatal SERT binding and number of 5-HT varicosities correlated positively with the AIMs scores, showing that L-DOPA induced a sprouting of striatal 5-HT terminals in dyskinetic animals [55]. Furthermore, SERT-immunoreactive varicosities displayed larger synaptic incidence with striatal neurons and resulted in larger amount of stimulated (KCl evoked) [<sup>3</sup>H]-DA release in striatal slices from L-DOPA-treated dyskinetic rats [55]. However, Lundblad et al. [73] failed to correlate the higher 5-HT nerve density in the lesioned striatum of dyskinetic rats with the magnitude of KCl-evoked DA release measured *in vivo* by chronoamperometry after chronic L-DOPA treatment. Although SERT binding was decreased in the putamen and globus pallidus (GP) of MPTP-treated monkeys [55], a marked hyperinnervation of TPH-positive fibers (increase in number and diameter of TPH-positive axon varicosities) was observed in the dorsal caudate and putamen, but not the GP of MPTP-intoxicated monkeys [56]. Nevertheless, using both 5-HT markers, these studies have consistently shown an elevated SERT binding and a further increase in the number and enlargement of TPH positive axonal varicosities in caudate nucleus and putamen of MPTP-treated monkeys that develop LIDs [55, 56]. In Parkinsonian patients, SERT-binding levels were also significantly increased in both the putamen and GP of dyskinetic

patients [55]. Regarding the lifetime L-DOPA medication, results indicate that patients with highest levels of SERT binding were those developing LIDs earliest during their PD treatment [55]. However, by using another marker of serotonin transporter ( $^{11}\text{C}$ -DASP) in PET, Politis et al. [123] could not establish a correlation between  $^{11}\text{C}$ -DASP binding and exposure to dopaminergic therapy. Altogether, these data suggest that L-DOPA pharmacotherapy induced a maladaptive plasticity of 5-HT axon terminals that may predispose to LIDs. Indeed, the 5-HT hyperinnervation together with marked hypertrophy of 5-HT axon varicosities may worsen the fluctuations of L-DOPA-induced DA release [48, 55, 56, 59].

The combination of 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> agonists provides useful pharmacological manipulation to reduce the large increases in DA efflux and the occurrence of LIDs [48, 61, 124]. Their efficacy is reached when combining subthreshold doses of 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> agonists that are thought to activate presynaptic 5-HT<sub>1</sub> receptors and dampen the release of L-DOPA-derived DA from 5-HT neurons [64]. The stimulation of postsynaptic 5-HT<sub>1</sub> receptors on non-5-HT neurons may also contribute to their antidyskinetic effect by modulating GABA and glutamate release [64]. However, adverse effects involving the stimulation of postsynaptic 5-HT<sub>1A</sub> receptors could worsen their therapeutic efficacy [125, 126]. Some studies have identified specific changes induced by chronic L-DOPA treatment on 5-HT<sub>1B</sub> postsynaptic receptors that may be directly involved in the development of LIDs. Chronic L-DOPA treatment increased the expression of 5-HT<sub>1B</sub> receptors and its adaptor protein p11 at striatonigral neurons [75]. The ability of 5-HT<sub>1B</sub> agonist to reduce LIDs was p11 dependent [75]. Moreover, in L-DOPA-treated 6-OHDA rats that recovered from AIMs after a chronic treatment with citalopram (a selective serotonergic reuptake inhibitor, SSRI), the expression of 5-HT<sub>1B</sub> receptors in the striatum was almost fully abolished [127]. The authors could reveal a positive correlation between the decreased anxiety induced by citalopram and its ability to reduce AIMs that involves a marked reduction in 5-HT<sub>1B</sub> receptor expression (Table 2). However, in keeping with data obtained in the study by Zhang et al. [75], the reduction of LIDs by citalopram may not solely account for its effect on 5-HT<sub>1B</sub> receptor expression but may also involve the ability of citalopram to abolish L-DOPA-induced DA release [31]. Nevertheless, these data allow identifying a new association between 5-HT<sub>1B</sub> receptors and LIDs. A recent work could also highlight a relationship between 5-HT<sub>2A</sub> receptors and LIDs in MPTP-treated monkeys [78]. [ $^3\text{H}$ ]Ketanserin-specific binding to 5-HT<sub>2A</sub> receptors was increased in the dorsomedial caudate nucleus and anterior cingulate gyrus of dyskinetic L-DOPA-treated MPTP-intoxicated monkeys, an effect reversed by drugs inhibiting the expression of LIDs. The authors could reveal a positive correlation between the maximal dyskinesia scores at the end of L-DOPA treatment and 5-HT<sub>2A</sub> receptor-specific binding in the anterior and posterior caudate nucleus as well as the nucleus accumbens [78].

Despite the high degree of variability observed in the changes of 5-HT markers across studies performed in different animal models and Parkinsonian patients, the available data to date allow establishing a clear role of the 5-HT system in the induction and maintenance of LIDs. The numerous 5-HT indexes used could provide interesting insights into the mechanisms of action of L-DOPA in mediating LIDs. However, the failure to fully correlate one change in 5-HT markers with a complex behavior such as LIDs may encourage future studies to reconsider the heterogeneity and the widespread influence of the 5-HT system as a whole fundamental index in the genesis of LIDs. Indeed, the numerous changes in 5-HT function induced by L-DOPA in multiple brain regions may concur in synergy to an imbalanced DA transmission that may participate in the emergence of LIDs.

#### 4. Functional Outcomes of 5-HT Neuron-Mediated DA Transmission in LIDs

Because the 5-HT terminals are responsible for the gross increase in DA, leading to a homogeneous and ectopic release of DA in the brain, one may wonder the extent to which the striatum is involved in the therapeutic benefit of L-DOPA. It is far from our purpose to rule out many years of research centred on striatal DA transmission, but it is important to conceive that other brain regions play an important role in motor responses induced by L-DOPA. The main argument to look beyond the striatum is the success of the deep brain stimulation of the subthalamic nucleus in Parkinson patients, a surgical approach of the disease that does not rely on striatal DA release.

*4.1. Role of Imbalanced Cortical-Subcortical DA Transmission in Motor Output.* It is a common sense to reaffirm that DA transmission is altered in PD and that the relationships between DA transmission, symptoms severity, and medication coevolve with the deleterious progression of the disease. Nonetheless, adding the evidence that 5-HT neurons participate in the raise of extracellular DA offers a larger picture of the putative scenarios. In early stages of the disease, the presence of spared DA terminals and DAT in the striatum limits the excessive increase in DA extracellular levels induced by L-DOPA from 5-HT neurons. However, the increase in DA from 5-HT terminals in the striatum likely introduces a noise in the "coherent" DA transmission. Indeed, this aberrant release is not regulated while the "coherent" release from spared DA neurons is impaired due to the inhibitory effect of electrical activity of L-DOPA on DA neurons activity [19]. The more the disease progresses, the higher should be the contribution of 5-HT neurons in L-DOPA-induced DA release. Thus, even in the early stages of the disease, it is noticeable that L-DOPA is efficient to treat the core symptoms of the disease (tremor, bradykinesia, rigidity, posture) but has limited effects on precise coordinated movements or some impaired cognitive functions [128]. In the advanced stages of the disease, spared DA neurons are no longer able to buffer excessive swings of

DA released from 5-HT neurons, a condition favoring motor fluctuations and LIDs [61].

Whatever the stage of the disease, the small release of DA from 5-HT neurons has potentially a larger impact beyond the striatum where DAT are poorly expressed. The impact may also be magnified due to altered pattern of activities found in extrastriatal territories such as the cortex or the HIPP. Indeed, numerous studies in humans using functional imaging have reported changes in activities in several cortical territories and the HIPP [129]. These brain areas expressing substantial amount of DA receptors [50], the excessive increase in DA release induced by L-DOPA in these territories could have a higher impact on the functions exerted by these brain regions. Of note, it has been described for many years that an increase in cortical DA may counteract aberrant DA signaling in subcortical areas. For instance, the catalepsy induced by the DA antagonist haloperidol, a rat model of Parkinsonism, is reversed by the direct infusion of DA into the PFC. Moreover, the increase in DA release induced by L-DOPA is very high in the SN, one of the brain regions receiving the strongest 5-HT innervation [46], and it has been shown for several years that the SN directly participates in the motor effects of L-DOPA in the 6-OHDA rat model of Parkinson's disease [68, 130].

The minimal release of striatal DA after therapeutic doses of L-DOPA could be compensated by an increase in D<sub>2</sub> receptor efficiency. An increase in striatal D<sub>2</sub> receptors has been reported in early stages of the disease, but some data have reported that DA "replacement" therapy reduced the excessive expression of striatal D<sub>2</sub> receptors to levels comparable to matched controls [131, 132]. Based on the neurochemical data in the 6-OHDA rat model of Parkinson's disease, the benefit of L-DOPA could be an uneven release of DA or a hypodopaminergy in the striatum combined with an extrastriatal hyperdopaminergy.

**4.2. Role of Imbalanced Cortical-Subcortical DA Transmission in LIDs.** The increase in DA release induced by L-DOPA has been directly incriminated in LIDs [133]. Our data showing that chronic treatment with L-DOPA is associated with a dramatic loss of DA release in various rat brain areas compared to the striatum [45] points to an inverse schema. First, the inhibitory tone provided by cortical DA upon subcortical DA function would be lowered after chronic treatment, and subcortical DA release by 5-HT fibers would be preserved due to some sprouting of striatal 5-HT fibers [55, 56]. The situation is not known for several brain regions though it has been reported that LIDs in rodents is associated with an increase in c-Fos expression in the STN [134]. Besides, chronic L-DOPA treatment has been shown to increase c-Fos expression also in the cortex and globus pallidus [135, 136]. In addition, excessive DA tone in some brain regions other than the striatum may promote abnormal involuntary movement of the face, one clinical manifestation reported in LIDs in primates and rodents [7–9]. Second, the aberrant release of DA *via* 5-HT neurons would favor abnormal learning, at least in the striatum. Indeed, DA is critically involved in procedural learning, and LIDs is thought to result in part from aberrant molecular

events at the level of medium spiny neurons of the striatum that involve DA receptors [104, 137, 138]. The postulated pathological form of synaptic plasticity may occur in the different territories of the striatum. It has been reported in MPTP-treated monkeys that LID involved not only the sensorimotor part of the striatum, but also its associative and limbic territories [139]. Third, as noted above, 5-HT processes could be involved as well [122, 140], particularly in considering that the "coherent" 5-HT transmission is altered by L-DOPA [24]. As for DA transmission, alteration in 5-HT transmission occurring elsewhere than the sensorimotor part of the striatum may promote abnormal movements in rodents [122, 141].

In a therapeutic point of view, one possibility is to limit the excessive DA transmission by administering a neuroleptic at risk of generating Parkinsonism. Nevertheless, the atypical neuroleptic clozapine has been shown to limit dyskinesia without aggravating the motor score [117]. It is difficult to interpret the origin of its efficacy as clozapine or other atypical antipsychotic drugs may slightly block subcortical DA transmission and enhance cortical DA transmission [142, 143]. Similar effects could account for the proposed efficacy of the antipsychotic and partial DA agonist drug aripiprazole [144]. According to the hypothesis above, treatment that is enhancing DA transmission in the cortex, that would limit the impact of cortico-subcortical glutamate transmission [145], could be a focus of future strategy against LIDs. It is noticeable that blockers of the N-methyl-d-aspartate receptor such as amantadine can also limit LIDs in patients [146].

The direct control of striatal DA transmission *via* 5-HT drugs is difficult to manage because the biochemical and behavioral relationships between 5-HT receptors and DA transmission are not well understood [147]. The use of 5-HT drugs able to control 5-HT nerve activity, to control the output of DA from 5-HT neurons, is a great opportunity, and clinical trials are currently underwent to alleviate LIDs using this strategy. The limit of this approach is that 5-HT drugs used may also act directly on cells other than 5-HT neurons due to the widespread distribution of 5-HT receptors in the brain. Also, a general decrease in DA release from 5-HT neurons may counteract dyskinesia and aggravate Parkinsonism [148–153]. Again, 5-HT drugs could be used to reinforce the initial imbalance created by L-DOPA, namely, the quite homogeneous pattern of DA release induced by L-DOPA, through cortical mechanisms.

## 5. Conclusion

The consideration of the 5-HT system in the core mechanism of action of L-DOPA opens many opportunities to better apprehend LIDs and to propose diverse therapeutic strategies in the treatment of LIDs. The excess of striatal DA released by L-DOPA remains an important preoccupation, but the possibility to facilitate DA transmission in the cortex could be also an interesting strategy. Additional studies are warranted to further study the imbalance of DA transmission promoted by the intervention of 5-HT neurons in the mechanism of action of L-DOPA to propose additional brain targets.

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## References

- [1] E. R. Dorsey, R. Constantinescu, J. P. Thompson et al., “Projected number of people with Parkinson disease in the most populous nations, 2005 through 2030,” *Neurology*, vol. 68, no. 5, pp. 384–386, 2007.
- [2] O. Hornykiewicz, “Dopamine (3-hydroxytyramine) and brain function,” *Pharmacological Reviews*, vol. 18, no. 2, pp. 925–964, 1966.
- [3] H. Bernheimer, W. Birkmayer, O. Hornykiewicz, K. Jellinger, and F. Seitelberger, “Brain dopamine and the syndromes of Parkinson and Huntington. Clinical, morphological and neurochemical correlations,” *Journal of the Neurological Sciences*, vol. 20, no. 4, pp. 415–455, 1973.
- [4] G. C. Cotzias, “L-Dopa for Parkinsonism,” *New England Journal of Medicine*, vol. 278, no. 11, p. 630, 1968.
- [5] O. Hornykiewicz, “Dopamine in the basal ganglia. Its role and therapeutic implications (including the clinical use of L-DOPA),” *British Medical Bulletin*, vol. 29, no. 2, pp. 172–178, 1973.
- [6] J. E. Ahlskog and M. D. Muentner, “Frequency of levodopa-related dyskinesias and motor fluctuations as estimated from the cumulative literature,” *Movement Disorders*, vol. 16, no. 3, pp. 448–458, 2001.
- [7] J. Jankovic, “Motor fluctuations and dyskinesias in Parkinson's disease: clinical manifestations,” *Movement Disorders*, vol. 20, no. 11, pp. S11–S16, 2005.
- [8] M. Lundblad, M. Andersson, C. Winkler, D. Kirik, N. Wierup, and M. A. Cenci Nilsson, “Pharmacological validation of behavioural measures of akinesia and dyskinesia in a rat model of Parkinson's disease,” *European Journal of Neuroscience*, vol. 15, no. 1, pp. 120–132, 2002.
- [9] M. A. Cenci and K. E. Ohlin, “Rodent models of treatment-induced motor complications in Parkinson's disease,” *Parkinsonism and Related Disorders*, vol. 15, supplement 4, pp. S13–S17, 2009.
- [10] U. Ungerstedt, “6-hydroxy-dopamine induced degeneration of central monoamine neurons,” *European Journal of Pharmacology*, vol. 5, no. 1, pp. 107–110, 1968.
- [11] M. Olsson, G. Nikkhah, C. Bentlage, and A. Bjorklund, “Forelimb akinesia in the rat Parkinson model: differential effects of dopamine agonists and nigral transplants as assessed by a new stepping test,” *Journal of Neuroscience*, vol. 15, no. 5, pp. 3863–3875, 1995.
- [12] M. A. Cenci, “L-DOPA-induced dyskinesia: cellular mechanisms and approaches to treatment,” *Parkinsonism and Related Disorders*, vol. 13, no. 3, pp. S263–S267, 2007.
- [13] U. Ungerstedt, “Postsynaptic supersensitivity after 6-hydroxy-dopamine induced degeneration of the nigro-striatal dopamine system,” *Acta Physiologica Scandinavica*, vol. 367, supplement, pp. 69–93, 1971.
- [14] U. Ungerstedt, “Striatal dopamine release after amphetamine or nerve degeneration revealed by rotational behaviour,” *Acta Physiologica Scandinavica*, vol. 367, supplement, pp. 49–68, 1971.
- [15] T. Zetterstrom, M. Herrera-Marschitz, and U. Ungerstedt, “Simultaneous measurement of dopamine release and rotational behaviour in 6-hydroxydopamine denervated rats using intracerebral dialysis,” *Brain Research*, vol. 376, no. 1, pp. 1–7, 1986.
- [16] B. S. Bunney, G. K. Aghajanian, and R. H. Roth, “Comparison of effects of L dopa amphetamine and apomorphine on firing rate of rat dopaminergic neurones,” *Nature New Biology*, vol. 245, no. 143, pp. 123–125, 1973.
- [17] N. B. Mercuri, P. Calabresi, and G. Bernardi, “Responses of rat substantia nigra compacta neurones to L-DOPA,” *British Journal of Pharmacology*, vol. 100, no. 2, pp. 257–260, 1990.
- [18] D. G. Harden and A. A. Grace, “Activation of dopamine cell firing by repeated L-DOPA administration to dopamine-depleted rats: its potential role in mediating the therapeutic response to L-DOPA treatment,” *Journal of Neuroscience*, vol. 15, no. 9, pp. 6157–6166, 1995.
- [19] T. Maeda, K. Kannari, T. Suda, and M. Matsunaga, “Loss of regulation by presynaptic dopamine D2 receptors of exogenous L-DOPA-derived dopamine release in the dopaminergic denervated striatum,” *Brain Research*, vol. 817, no. 1–2, pp. 185–191, 1999.
- [20] D. W. Miller and E. D. Abercrombie, “Role of high-affinity dopamine uptake and impulse activity in the appearance of extracellular dopamine in striatum after administration of exogenous L-DOPA: studies in intact and 6-hydroxydopamine-treated rats,” *Journal of Neurochemistry*, vol. 72, no. 4, pp. 1516–1522, 1999.
- [21] S. Sarre, N. De Klippel, P. Herregodts, G. Ebinger, and Y. Michotte, “Biotransformation of locally applied L-dopa in the corpus striatum of the hemi-Parkinsonian rat studied with microdialysis,” *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 350, no. 1, pp. 15–21, 1994.
- [22] K. Kannari, H. Tanaka, T. Maeda, M. Tomiyama, T. Suda, and M. Matsunaga, “Reserpine pretreatment prevents increases in extracellular striatal dopamine following L-DOPA administration in rats with nigrostriatal denervation,” *Journal of Neurochemistry*, vol. 74, no. 1, pp. 263–269, 2000.
- [23] W. Y. Lee, J. W. Chang, N. L. Nemeth, and U. J. Kang, “Vesicular monoamine transporter-2 and aromatic l-amino acid decarboxylase enhance dopamine delivery after L-3,4-dihydroxyphenylalanine administration in parkinsonian rats,” *Journal of Neuroscience*, vol. 19, no. 8, pp. 3266–3274, 1999.
- [24] S. Navailles, M. Carta, M. Guthrie, and P. De Deurwaerdere, “L-DOPA and serotonergic neurons: functional implication and therapeutic perspectives in Parkinson's disease,” *Central Nervous System Agents in Medicinal Chemistry*. In press.
- [25] L. K. Ng, T. N. Chase, R. W. Colburn, and I. J. Kopin, “l-dopa-induced release of cerebral monoamines,” *Science*, vol. 170, no. 953, pp. 76–77, 1970.
- [26] F. Tison, N. Mons, M. Geffard, and P. Henry, “The metabolism of exogenous L-Dopa in the brain: an immunohistochemical study of its conversion to dopamine in non-catecholaminergic cells of the rat brain,” *Journal of Neural Transmission*, vol. 3, no. 1, pp. 27–39, 1991.
- [27] R. Arai, N. Karasawa, M. Geffard, and I. Nagatsu, “L-DOPA is converted to dopamine in serotonergic fibers of the striatum of the rat: a double-labeling immunofluorescence study,” *Neuroscience Letters*, vol. 195, no. 3, pp. 195–198, 1995.
- [28] H. Yamada, Y. Aimi, I. Nagatsu, K. Taki, M. Kudo, and R. Arai, “Immunohistochemical detection of l-DOPA-derived dopamine within serotonergic fibers in the striatum and the

- substantia nigra pars reticulata in Parkinsonian model rats," *Neuroscience Research*, vol. 59, no. 1, pp. 1–7, 2007.
- [29] L. K. Ng, R. W. Colburn, and I. J. Kopin, "Effects of L-dopa on accumulation and efflux of monoamines in particles of rat brain homogenates," *Journal of Pharmacology and Experimental Therapeutics*, vol. 183, no. 2, pp. 316–325, 1972.
- [30] L. K. Ng, T. N. Chase, R. W. Colburn, and I. J. Kopin, "L-dopa in Parkinsonism. A possible mechanism of action," *Neurology*, vol. 22, no. 7, pp. 688–696, 1972.
- [31] S. Navailles, B. Bioulac, C. Gross, and P. De Deurwaerdère, "Serotonergic neurons mediate ectopic release of dopamine induced by l-DOPA in a rat model of Parkinson's disease," *Neurobiology of Disease*, vol. 38, no. 1, pp. 136–143, 2010.
- [32] H. Tanaka, K. Kannari, T. Maeda, M. Tomiyama, T. Suda, and M. Matsunaga, "Role of serotonergic neuron in L-DOPA-derived extracellular dopamine in the striatum of 6-OHDA-lesioned rats," *NeuroReport*, vol. 10, no. 3, pp. 631–634, 1999.
- [33] K. Kannari, H. Yamato, H. Shen, M. Tomiyama, T. Suda, and M. Matsunaga, "Activation of 5-HT1A but not 5-HT1B receptors attenuates an increase in extracellular dopamine derived from exogenously administered L-DOPA in the striatum with nigrostriatal denervation," *Journal of Neurochemistry*, vol. 76, no. 5, pp. 1346–1353, 2001.
- [34] H. Yamato, K. Kannari, H. Shen, T. Suda, and M. Matsunaga, "Fluoxetine reduces L-DOPA-derived extracellular DA in the 6-OHDA-lesioned rat striatum," *NeuroReport*, vol. 12, no. 6, pp. 1123–1126, 2001.
- [35] J. S. Sprouse and G. K. Aghajanian, "Electrophysiological responses of serotonergic dorsal raphe neurons to 5-HT1A and 5-HT1B agonists," *Synapse*, vol. 1, no. 1, pp. 3–9, 1987.
- [36] F. J. Bosker, T. Y. C. E. De Winter, A. A. Klompmakers, and H. G. M. Westenberg, "Flesinoxan dose-dependently reduces extracellular 5-hydroxytryptamine (5-HT) in rat median raphe and dorsal hippocampus through activation of 5-HT1A receptors," *Journal of Neurochemistry*, vol. 66, no. 6, pp. 2546–2555, 1996.
- [37] D. A. Knobelman, H. F. Kung, and I. Lucki, "Regulation of extracellular concentrations of 5-hydroxytryptamine (5-HT) in mouse striatum by 5-HT(1A) and 5-HT(1B) receptors," *Journal of Pharmacology and Experimental Therapeutics*, vol. 292, no. 3, pp. 1111–1117, 2000.
- [38] M. Riad, S. Garcia, K. C. Watkins et al., "Somatodendritic localization of 5-HT1A and preterminal axonal localization of 5-HT1B serotonin receptors in adult rat brain," *Journal of Comparative Neurology*, vol. 417, no. 2, pp. 181–194, 2000.
- [39] T. Sharp, S. R. Bramwell, S. Hjorth, and D. G. Grahame-Smith, "Pharmacological characterization of 8-OH-DPAT-induced inhibition of rat hippocampal 5-HT release in vivo as measured by microdialysis," *British Journal of Pharmacology*, vol. 98, no. 3, pp. 989–997, 1989.
- [40] A. Adell, A. Carceller, and F. Artigas, "In vivo brain dialysis study of the somatodendritic release of serotonin in the raphe nuclei of the rat: effects of 8-hydroxy-2-(di-*n*-propylamino)tetralin," *Journal of Neurochemistry*, vol. 60, no. 5, pp. 1673–1681, 1993.
- [41] J. M. Casanovas and F. Artigas, "Differential effects of ipsapirone on 5-hydroxytryptamine release in the dorsal and median raphe neuronal pathways," *Journal of Neurochemistry*, vol. 67, no. 5, pp. 1945–1952, 1996.
- [42] L. Arborelius, G. G. Nomikos, P. Grillner et al., "5-HT(1A) receptor antagonists increase the activity of serotonergic cells in the dorsal raphe nucleus in rats treated acutely or chronically with citalopram," *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 352, no. 2, pp. 157–165, 1995.
- [43] Y. Temel, L. J. Boothman, A. Blokland et al., "Inhibition of 5-HT neuron activity and induction of depressive-like behavior by high-frequency stimulation of the subthalamic nucleus," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 43, pp. 17087–17092, 2007.
- [44] S. Navailles, A. Benazzouz, B. Bioulac, C. Gross, and P. De Deurwaerdère, "High-frequency stimulation of the subthalamic nucleus and L-3,4-dihydroxyphenylalanine inhibit in vivo serotonin release in the prefrontal cortex and hippocampus in a rat model of Parkinson's disease," *Journal of Neuroscience*, vol. 30, no. 6, pp. 2356–2364, 2010.
- [45] S. Navailles, B. Bioulac, C. Gross, and P. De Deurwaerdère, "Chronic L-DOPA therapy alters central serotonergic function and L-DOPA-induced dopamine release in a region-dependent manner in a rat model of Parkinson's disease," *Neurobiology of Disease*, vol. 41, no. 2, pp. 585–590, 2011.
- [46] E. C. Azmitia and M. Segal, "An autoradiographic analysis of the different ascending projections of the dorsal and median raphe nuclei in the rat," *Journal of Comparative Neurology*, vol. 179, no. 3, pp. 641–667, 1978.
- [47] H. W. Steinbusch, "Serotonin-immunoreactive neurons and their projections in the CNS," in *Handbook of Chemical Neuroanatomy—Classical Transmitters and Transmitters Receptors in the CNS Part II*, A. Björklund, T. Hökfelt, and M. J. Kuhar, Eds., pp. 68–125, Amsterdam, The Netherlands, 1984.
- [48] H. S. Lindgren, D. R. Andersson, S. Lagerkvist, H. Nissbrandt, and M. A. Cenci, "L-DOPA-induced dopamine efflux in the striatum and the substantia nigra in a rat model of Parkinson's disease: temporal and quantitative relationship to the expression of dyskinesia," *Journal of Neurochemistry*, vol. 112, no. 6, pp. 1465–1476, 2010.
- [49] J. A. Obeso, C. Marin, C. Rodriguez-Oroz et al., "The basal ganglia in Parkinson's disease: current concepts and unexplained observations," *Annals of Neurology*, vol. 64, no. 2, pp. S30–S46, 2008.
- [50] P. Seeman, "Brain dopamine receptors," *Pharmacological Reviews*, vol. 32, no. 3, pp. 229–313, 1980.
- [51] W. D. Brown, M. D. Taylor, A. D. Roberts et al., "FluoroDOPA PET shows the nondopaminergic as well as dopaminergic destinations of levodopa," *Neurology*, vol. 53, no. 6, pp. 1212–1218, 1999.
- [52] R. M. Kostrzewa, "Dopamine receptor supersensitivity," *Neuroscience and Biobehavioral Reviews*, vol. 19, no. 1, pp. 1–17, 1995.
- [53] J. Tong, P. S. Fitzmaurice, L. C. Ang, Y. Furukawa, M. Guttman, and S. J. Kish, "Brain dopamine-stimulated adenylyl cyclase activity in Parkinson's disease, multiple system atrophy, and progressive supranuclear palsy," *Annals of Neurology*, vol. 55, no. 1, pp. 125–129, 2004.
- [54] M. R. Ahmed, A. Berthet, E. Bychkov et al., "Lentiviral overexpression of GRK6 alleviates L-Dopa-induced dyskinesia in experimental parkinson's disease," *Science Translational Medicine*, vol. 2, no. 28, pp. 28–ra28, 2010.
- [55] D. Rylander, M. Parent, S. S. O-Sullivan et al., "Maladaptive plasticity of serotonin axon terminals in levodopa-induced dyskinesia," *Annals of Neurology*, vol. 68, no. 5, pp. 619–628, 2010.
- [56] B. Y. Zeng, M. M. Iravani, M. J. Jackson, S. Rose, A. Parent, and P. Jenner, "Morphological changes in serotonergic neurites in the striatum and globus pallidus in levodopa primed MPTP treated common marmosets with dyskinesia," *Neurobiology of Disease*, vol. 40, no. 3, pp. 599–607, 2010.

- [57] T. N. Chase, "Levodopa therapy: consequences of the non-physiologic replacement of dopamine," *Neurology*, vol. 50, supplement 5, pp. S17–S25, 1998.
- [58] C. W. Olanow and J. A. Obeso, "Pulsatile stimulation of dopamine receptors and levodopa-induced motor complications in Parkinson's disease: implications for the early use of COMT inhibitors," *Neurology*, vol. 55, supplement 4, no. 11, pp. S72–S77, 2000.
- [59] R. De La Fuente-Fernández, V. Sossi, Z. Huang et al., "Levodopa-induced changes in synaptic dopamine levels increase with progression of Parkinson's disease: implications for dyskinesias," *Brain*, vol. 127, no. 12, pp. 2747–2754, 2004.
- [60] M. A. Cenci and M. Lundblad, "Post- versus presynaptic plasticity in L-DOPA-induced dyskinesia," *Journal of Neurochemistry*, vol. 99, no. 2, pp. 381–392, 2006.
- [61] M. Carta, T. Carlsson, D. Kirik, and A. Björklund, "Dopamine released from 5-HT terminals is the cause of L-DOPA-induced dyskinesia in parkinsonian rats," *Brain*, vol. 130, no. 7, pp. 1819–1833, 2007.
- [62] M. Carta, T. Carlsson, A. Muñoz, D. Kirik, and A. Björklund, "Serotonin-dopamine interaction in the induction and maintenance of L-DOPA-induced dyskinesias," *Progress in Brain Research*, vol. 172, pp. 465–478, 2008.
- [63] A. Ulusoy, G. Sahin, and D. Kirik, "Presynaptic dopaminergic compartment determines the susceptibility to L-DOPA-induced dyskinesia in rats," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 29, pp. 13159–13164, 2010.
- [64] A. Muñoz, T. Carlsson, E. Tronci, D. Kirik, A. Björklund, and M. Carta, "Serotonin neuron-dependent and -independent reduction of dyskinesia by 5-HT1A and 5-HT1B receptor agonists in the rat Parkinson model," *Experimental Neurology*, vol. 219, no. 1, pp. 298–307, 2009.
- [65] C. Marin, E. Aguilar, M. C. Rodríguez-Oroz, G. D. Bartszyk, and J. A. Obeso, "Local administration of sarizotan into the subthalamic nucleus attenuates levodopa-induced dyskinesias in 6-OHDA-lesioned rats," *Psychopharmacology*, vol. 204, no. 2, pp. 241–250, 2009.
- [66] S. Sarre, P. Herregodts, D. Deleu et al., "Biotransformation of L-DOPA in striatum and substantia nigra of rats with a unilateral, nigrostriatal lesion: a microdialysis study," *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 346, no. 3, pp. 277–285, 1992.
- [67] S. Sarre, I. Smolders, K. Thorré, G. Ebinger, and Y. Michotte, "Biotransformation of locally applied precursors of dopamine, serotonin and noradrenaline in striatum and hippocampus: a microdialysis study," *Journal of Neural Transmission*, vol. 104, no. 11-12, pp. 1215–1228, 1997.
- [68] D. Orosz and J. P. Bennett, "Simultaneous microdialysis in striatum and substantia nigra suggests that the nigra is a major site of action of L-dihydroxyphenylalanine in the "hemiparkinsonian" rat," *Experimental Neurology*, vol. 115, no. 3, pp. 388–393, 1992.
- [69] V. Di Matteo, M. Pierucci, E. Esposito, G. Crescimanno, A. Benigno, and G. Di Giovanni, "Serotonin modulation of the basal ganglia circuitry: therapeutic implication for Parkinson's disease and other motor disorders," *Progress in Brain Research*, vol. 172, pp. 423–463, 2008.
- [70] D. B. Putterman, A. C. Munhall, L. B. Kozell, J. K. Belknap, and S. W. Johnson, "Evaluation of levodopa dose and magnitude of dopamine depletion as risk factors for levodopa-induced dyskinesia in a rat model of Parkinson's disease," *Journal of Pharmacology and Experimental Therapeutics*, vol. 323, no. 1, pp. 277–284, 2007.
- [71] K. Thorré, S. Sarre, I. Smolders, G. Ebinger, and Y. Michotte, "Dopaminergic regulation of serotonin release in the substantia nigra of the freely moving rat using microdialysis," *Brain Research*, vol. 796, no. 1-2, pp. 107–116, 1998.
- [72] S. J. Gil, C. H. Park, J. E. Lee, Y. K. Minn, and H. C. Koh, "Positive association between striatal serotonin level and abnormal involuntary movements in chronic L-DOPA-treated hemiparkinsonian rats," *Brain Research Bulletin*, vol. 96, no. 6, pp. 1718–1727, 2011.
- [73] M. Lundblad, S. Af Bjerkén, M. A. Cenci, F. Pomerleau, G. A. Gerhardt, and I. Strömberg, "Chronic intermittent L-DOPA treatment induces changes in dopamine release," *Journal of Neurochemistry*, vol. 108, no. 4, pp. 998–1008, 2009.
- [74] S. Gil, C. Park, J. Lee, and H. Koh, "The roles of striatal serotonin and l-amino-acid decarboxylase on l-DOPA-induced dyskinesia in a hemiparkinsonian rat model," *Cellular and Molecular Neurobiology*, vol. 30, no. 6, pp. 817–825, 2010.
- [75] X. Zhang, P. E. Andren, P. Greengard, and P. Svenningsson, "Evidence for a role of the 5-HT1B receptor and its adaptor protein, p11, in L-DOPA treatment of an animal model of Parkinsonism," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 6, pp. 2163–2168, 2008.
- [76] X. Zhang, P. E. Andren, and P. Svenningsson, "Changes on 5-HT2 receptor mRNAs in striatum and subthalamic nucleus in Parkinson's disease model," *Physiology and Behavior*, vol. 92, no. 1-2, pp. 29–33, 2007.
- [77] P. Huot, T. H. Johnston, J. B. Koprach, L. Winkelmolen, S. H. Fox, and J. M. Brotchie, "Regulation of cortical and striatal 5-HT(1A) receptors in the MPTP-lesioned macaque," *Neurobiology of Aging*. In press.
- [78] G. Riahi, M. Morissette, M. Parent, and T. Di Paolo, "Brain 5-HT2A receptors in MPTP monkeys and levodopa-induced dyskinesias," *European Journal of Neuroscience*, vol. 33, no. 10, pp. 1823–1831, 2011.
- [79] S. L. Nicholson and J. M. Brotchie, "5-Hydroxytryptamine (5-HT, serotonin) and Parkinson's disease—opportunities for novel therapeutics to reduce the problems of levodopa therapy," *European Journal of Neurology*, vol. 9, supplement 3, pp. 1–6, 2002.
- [80] S. H. Fox and J. M. Brotchie, "5-HT(2C) receptor binding is increased in the substantia nigra pars reticulata in Parkinson's disease," *Movement Disorders*, vol. 15, no. 6, pp. 1064–1069, 2000.
- [81] W. Meissner, P. Ravenscroft, R. Reese et al., "Increased slow oscillatory activity in substantia nigra pars reticulata triggers abnormal involuntary movements in the 6-OHDA-lesioned rat in the presence of excessive extracellular striatal dopamine," *Neurobiology of Disease*, vol. 22, no. 3, pp. 586–598, 2006.
- [82] S. H. Fox, R. Chuang, and J. M. Brotchie, "Serotonin and Parkinson's disease: on movement, mood, and madness," *Movement Disorders*, vol. 24, no. 9, pp. 1255–1266, 2009.
- [83] B. Scholtissen, F. R. J. Verhey, J. J. Adam, W. Weber, and A. F. G. Leentjens, "Challenging the serotonergic system in Parkinson disease patients: effects on cognition, mood, and motor performance," *Clinical Neuropharmacology*, vol. 29, no. 5, pp. 276–285, 2006.
- [84] M. Carta, H. S. Lindgren, M. Lundblad, R. Stancampiano, F. Fadda, and M. A. Cenci, "Role of striatal L-DOPA in the production of dyskinesia in 6-hydroxydopamine lesioned rats," *Journal of Neurochemistry*, vol. 96, no. 6, pp. 1718–1727, 2006.

- [85] K. L. Eskow, K. B. Dupre, C. J. Barnum, S. O. Dickinson, J. Y. Park, and C. Bishop, "The role of the dorsal raphe nucleus in the development, expression, and treatment of L-dopa-induced dyskinesia in hemiparkinsonian rats," *Synapse*, vol. 63, no. 7, pp. 610–620, 2009.
- [86] G. Bartholini, M. Da Prada, and A. Pletscher, "Decrease of cerebral 5-hydroxytryptamine by 3,4-dihydroxyphenylalanine after inhibition of extracerebral decarboxylase," *Journal of Pharmacy and Pharmacology*, vol. 20, no. 3, pp. 228–229, 1968.
- [87] G. M. Everett and J. W. Borcharding, "L-dopa: effect on concentrations of dopamine, norepinephrine, and serotonin in brains of mice," *Science*, vol. 168, no. 3933, pp. 849–850, 1970.
- [88] H. Tohgi, T. Abe, S. Takahashi, J. Takahashi, and H. Hamato, "Alterations in the concentration of serotonergic and dopaminergic substances in the cerebrospinal fluid of patients with Parkinson's disease, and their changes after L-dopa administration," *Neuroscience Letters*, vol. 159, no. 1-2, pp. 135–138, 1993.
- [89] F. Hery, G. Simonnet, S. Bourgoin et al., "Effect of nerve activity on the in vivo release of [3H]serotonin continuously formed from L-[3H]tryptophan in the caudate nucleus of the cat," *Brain Research*, vol. 169, no. 2, pp. 317–334, 1979.
- [90] P. Blier, A. Serrano, and B. Scatton, "Differential responsiveness of the rat dorsal and median raphe 5-HT systems to 5-HT<sub>1</sub> receptor agonists and p-chloroamphetamine," *Synapse*, vol. 5, no. 2, pp. 120–133, 1990.
- [91] D. S. Kreiss and I. Lucki, "Differential regulation of serotonin (5-HT) release in the striatum and hippocampus by 5-HT(1A) autoreceptors of the dorsal and median raphe nuclei," *Journal of Pharmacology and Experimental Therapeutics*, vol. 269, no. 3, pp. 1268–1279, 1994.
- [92] I. Hervás, N. Bel, A. G. Fernández, J. M. Palacios, and F. Artigas, "In vivo control of 5-hydroxytryptamine release by terminal autoreceptors in rat brain areas differentially innervated by the dorsal and median raphe nuclei," *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 358, no. 3, pp. 315–322, 1998.
- [93] R. Invernizzi, M. Carli, A. Di Clemente, and R. Samanin, "Administration of 8-hydroxy-2-(di-n-propylamino)tetralin in raphe nuclei dorsalis and medianus reduces serotonin synthesis in the rat brain: differences in potency and regional sensitivity," *Journal of Neurochemistry*, vol. 56, no. 1, pp. 243–247, 1991.
- [94] R. Invernizzi, C. Velasco, M. Bramante, A. Longo, and R. Samanin, "Effect of 5-HT(1A) receptor antagonists on citalopram-induced increase in extracellular serotonin in the frontal cortex, striatum and dorsal hippocampus," *Neuropharmacology*, vol. 36, no. 4-5, pp. 467–473, 1997.
- [95] J. M. Casanovas, M. Lésourd, and F. Artigas, "The effect of the selective 5-HT(1A) agonists alnespirone (S-20499) and 8-OH-DPAT on extracellular 5-hydroxytryptamine in different regions of rat brain," *British Journal of Pharmacology*, vol. 122, no. 4, pp. 733–741, 1997.
- [96] L. Romero and F. Artigas, "Preferential potentiation of the effects of serotonin uptake inhibitors by 5-HT(1A) receptor antagonists in the dorsal raphe pathway: role of somatodendritic autoreceptors," *Journal of Neurochemistry*, vol. 68, no. 6, pp. 2593–2603, 1997.
- [97] B. Amilhon, E. Lepicard, T. Renoir et al., "VGLUT3 (vesicular glutamate transporter type 3) contribution to the regulation of serotonergic transmission and anxiety," *Journal of Neuroscience*, vol. 30, no. 6, pp. 2198–2210, 2010.
- [98] R. McQuade and T. Sharp, "Functional mapping of dorsal and median raphe 5-hydroxytryptamine pathways in fore-brain of the rat using microdialysis," *Journal of Neurochemistry*, vol. 69, no. 2, pp. 791–796, 1997.
- [99] P. Gaspar, O. Cases, and L. Maroteaux, "The developmental role of serotonin: news from mouse molecular genetics," *Nature Reviews Neuroscience*, vol. 4, no. 12, pp. 1002–1012, 2003.
- [100] V. Kiyasova, S. P. Fernandez, J. Laine et al., "A genetically defined morphologically and functionally unique subset of 5-HT neurons in the mouse raphe nuclei," *Journal of Neuroscience*, vol. 31, no. 8, pp. 2756–2768, 2011.
- [101] B. Picconi, A. Pisani, I. Barone et al., "Pathological synaptic plasticity in the striatum: implications for parkinson's disease," *NeuroToxicology*, vol. 26, no. 5, pp. 779–783, 2005.
- [102] B. Picconi, V. Ghiglieri, and P. Calabresi, "L-3,4-dihydroxyphenylalanine-induced sprouting of serotonin axon terminals: a useful biomarker for dyskinesias?" *Annals of Neurology*, vol. 68, no. 5, pp. 578–580, 2010.
- [103] I. A. Prescott, J. O. Dostrovsky, E. Moro, M. Hodaie, A. M. Lozano, and W. D. Hutchison, "Levodopa enhances synaptic plasticity in the substantia nigra pars reticulata of Parkinson's disease patients," *Brain*, vol. 132, no. 2, pp. 309–318, 2009.
- [104] A. Berthet, G. Porras, E. Doudnikoff et al., "Pharmacological analysis demonstrates dramatic alteration of D1 dopamine receptor neuronal distribution in the rat analog of L-DOPA-induced dyskinesia," *Journal of Neuroscience*, vol. 29, no. 15, pp. 4829–4835, 2009.
- [105] D. Frechilla, A. Cobreros, L. Saldise et al., "Serotonin 5-HT<sub>1A</sub> receptor expression is selectively enhanced in the striosomal compartment of chronic Parkinsonian monkeys," *Synapse*, vol. 39, no. 4, pp. 288–296, 2001.
- [106] M. Doder, E. A. Rabiner, N. Turjanski, A. J. Lees, and D. J. Brooks, "Tremor in Parkinson's disease and serotonergic dysfunction: an 11C-WAY 100635 PET study," *Neurology*, vol. 60, no. 4, pp. 601–605, 2003.
- [107] S. J. Kish, "Biochemistry of Parkinson's disease: is a brain serotonergic deficiency a characteristic of idiopathic Parkinson's disease?" *Advances in Neurology*, vol. 91, pp. 39–49, 2003.
- [108] B. Scholtissen, F. R. J. Verhey, H. W. M. Steinbusch, and A. F. G. Leentjens, "Serotonergic mechanisms in Parkinson's disease: opposing results from preclinical and clinical data," *Journal of Neural Transmission*, vol. 113, no. 1, pp. 59–73, 2006.
- [109] K. A. Jellinger, "Pathology of Parkinson's disease: changes other than the nigrostriatal pathway," *Molecular and Chemical Neuropathology*, vol. 14, no. 3, pp. 153–197, 1991.
- [110] G. M. Halliday, P. C. Blumbergs, R. G. H. Cotton, W. W. Blessing, and L. B. Geffen, "Loss of brainstem serotonin- and substance P-containing neurons in Parkinson's disease," *Brain Research*, vol. 510, no. 1, pp. 104–107, 1990.
- [111] G. G. Kovacs, S. Klöppel, I. Fischer et al., "Nucleus-specific alteration of raphe neurons in human neurodegenerative disorders," *NeuroReport*, vol. 14, no. 1, pp. 73–76, 2003.
- [112] C. P. L. H. Chen, J. T. Alder, L. Bray, A. E. Kingsbury, P. T. Francis, and O. J. F. Foster, "Post-synaptic 5-HT(1A) and 5-HT(2A) receptors are increased in Parkinson's disease neocortex," *Annals of the New York Academy of Sciences*, vol. 861, pp. 288–289, 1998.
- [113] S. J. Kish, J. Tong, O. Hornykiewicz et al., "Preferential loss of serotonin markers in caudate versus putamen in Parkinson's disease," *Brain*, vol. 131, no. 1, pp. 120–131, 2008.

- [114] M. E. Castro, J. Pascual, T. Romón, J. Berciano, J. Figols, and A. Pazos, "5-HT(1B) receptor binding in degenerative movement disorders," *Brain Research*, vol. 790, no. 1-2, pp. 323–328, 1998.
- [115] R. Quirion and J. Richard, "Differential effects of selective lesions of cholinergic and dopaminergic neurons on serotonin-type 1 receptors in rat brain," *Synapse*, vol. 1, no. 1, pp. 124–130, 1987.
- [116] K. Ikeguchi and A. Kuroda, "Mianserin treatment of patients with psychosis induced by antiparkinsonian drugs," *European Archives of Psychiatry and Clinical Neuroscience*, vol. 244, no. 6, pp. 320–324, 1995.
- [117] F. Durif, M. Vidailhet, F. Assal, C. Roche, A. M. Bonnet, and Y. Agid, "Low-dose clozapine improves dyskinesias in Parkinson's disease," *Neurology*, vol. 48, no. 3, pp. 658–662, 1997.
- [118] G. Di Giovanni, V. Di Matteo, M. Pierucci, A. Benigno, and E. Esposito, "Serotonin involvement in the basal ganglia pathophysiology: could the 5-HT<sub>2c</sub> receptor be a new target for therapeutic strategies?" *Current Medicinal Chemistry*, vol. 13, no. 25, pp. 3069–3081, 2006.
- [119] P. De Deurwaerdère, L. Mignon, and M. F. Chesselet, "Physiological and pathophysiological aspects of 5-HT<sub>2c</sub> receptors in basal ganglia," in *The Pathophysiology of Central 5-HT<sub>2c</sub> Receptors*, G. Di Giovanni and K. Neve, Eds., The receptors series, Humana Press, Springer, New York, NY, USA, 2010.
- [120] S. H. Fox, B. Moser, and J. M. Brotchie, "Behavioral effects of 5-HT(2C) receptor antagonism in the substantia nigra zona reticulata of the 6-hydroxydopamine-lesioned rat model of Parkinson's disease," *Experimental Neurology*, vol. 151, no. 1, pp. 35–49, 1998.
- [121] S. H. Fox and J. M. Brotchie, "5-HT(2C) receptor antagonists enhance the behavioural response to dopamine D1 receptor agonists in the 6-hydroxydopamine-lesioned rat," *European Journal of Pharmacology*, vol. 398, no. 1, pp. 59–64, 2000.
- [122] P. De Deurwaerdère and M. F. Chesselet, "Nigrostriatal lesions alter oral dyskinesias and c-Fos expression induced by the serotonin agonist 1-(m-chlorophenyl)piperazine in adult rats," *Journal of Neuroscience*, vol. 20, no. 13, pp. 5170–5178, 2000.
- [123] M. Politis, K. Wu, C. Loane et al., "Staging of serotonergic dysfunction in Parkinson's Disease: an in vivo 11C-DASB PET study," *Neurobiology of Disease*, vol. 40, no. 1, pp. 216–221, 2010.
- [124] A. Muñoz, Q. Li, F. Gardoni et al., "Combined 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptor agonists for the treatment of L-DOPA-induced dyskinesia," *Brain*, vol. 131, no. 12, pp. 3380–3394, 2008.
- [125] G. M. Goodwin, R. J. De Souza, A. J. Wood, and A. R. Green, "The enhancement by lithium of the 5-HT(1A) mediated serotonin syndrome produced by 8-OH-DPAT in the rat: evidence for a post-synaptic mechanism," *Psychopharmacology*, vol. 90, no. 4, pp. 488–493, 1986.
- [126] J. Yamada, Y. Sugimoto, and K. Horisaka, "The behavioural effects of 8-hydroxy-2-(di-n-propylamino)tetralin (8-OH-DPAT) in mice," *European Journal of Pharmacology*, vol. 154, no. 3, pp. 299–304, 1988.
- [127] W. L. Kuan, J. W. Zhao, and R. A. Barker, "The role of anxiety in the development of levodopa-induced dyskinesias in an animal model of Parkinson's disease, and the effect of chronic treatment with the selective serotonin reuptake inhibitor citalopram," *Psychopharmacology*, vol. 197, no. 2, pp. 279–293, 2008.
- [128] L. F. Schettino, S. V. Adamovich, W. Hening, E. Tunik, J. Sage, and H. Poizner, "Hand preshaping in Parkinson's disease: effects of visual feedback and medication state," *Experimental Brain Research*, vol. 168, no. 1-2, pp. 186–202, 2006.
- [129] A. Dagher and A. Nagano-Saito, "Functional and anatomical magnetic resonance imaging in Parkinson's disease," *Molecular Imaging and Biology*, vol. 9, no. 4, pp. 234–242, 2007.
- [130] G. S. Robertson and H. A. Robertson, "Evidence that L-dopa-induced rotational behavior is dependent on both striatal and nigral mechanisms," *Journal of Neuroscience*, vol. 9, no. 9, pp. 3326–3331, 1989.
- [131] A. Antonini, J. Schwarz, W. H. Oertel, O. Pogarell, and K. L. Leenders, "Long-term changes of striatal dopamine D2 receptors in patients with Parkinson's disease: a study with positron emission tomography and [11C]raclopride," *Movement Disorders*, vol. 12, no. 1, pp. 33–38, 1997.
- [132] R. Kuriakose and A. J. Stoessl, "Imaging the nigrostriatal system to monitor disease progression and treatment-induced complications," *Progress in Brain Research*, vol. 184, pp. 177–192, 2010.
- [133] E. V. Encarnacion and R. A. Hauser, "Levodopa-induced dyskinesias in Parkinson's disease: etiology, impact on quality of life, and treatments," *European Neurology*, vol. 60, no. 2, pp. 57–66, 2008.
- [134] J. J. Soghomonian, "L-DOPA-induced dyskinesia in adult rats with a unilateral 6-OHDA lesion of dopamine neurons is paralleled by increased c-fos gene expression in the subthalamic nucleus," *European Journal of Neuroscience*, vol. 23, no. 9, pp. 2395–2403, 2006.
- [135] P. Svenningsson, J. Arts, L. Gunne, and P. E. Andren, "Acute and repeated treatment with L-DOPA increase c-jun expression in the 6-hydroxydopamine-lesioned forebrain of rats and common marmosets," *Brain Research*, vol. 955, no. 1-2, pp. 8–15, 2002.
- [136] Y. Xu, S. Sun, and X. Cao, "Effect of levodopa chronic administration on behavioral changes and fos expression in basal ganglia in rat model of PD," *Journal of Huazhong University of Science and Technology. Medical sciences*, vol. 23, no. 3, pp. 258–262, 2003.
- [137] A. Berthet and E. Bezard, "Dopamine receptors and L-dopa-induced dyskinesia," *Parkinsonism and Related Disorders*, vol. 15, supplement 4, pp. S8–S12, 2009.
- [138] P. Calabresi, P. Giacomini, D. Centonze, and G. Bernardi, "Levodopa-induced dyskinesia: a pathological form of striatal synaptic plasticity?" *Annals of Neurology*, vol. 47, no. 4, supplement 1, pp. S60–S69, 2000.
- [139] C. Guigoni, Q. Li, I. Aubert et al., "Involvement of sensorimotor, limbic, and associative basal ganglia domains in L-3,4-dihydroxyphenylalanine-induced dyskinesia," *Journal of Neuroscience*, vol. 25, no. 8, pp. 2102–2107, 2005.
- [140] L. Gong, R. M. Kostrzewa, R. W. Fuller, and K. W. Perry, "Supersensitization of the oral response to SKF 38393 in neonatal 6-OHDA-lesioned rats is mediated through a serotonin system," *Journal of Pharmacology and Experimental Therapeutics*, vol. 261, no. 3, pp. 1000–1007, 1992.
- [141] A. Beyeler, N. Kadir, S. Navailles et al., "Stimulation of serotonin<sub>2C</sub> receptors elicits abnormal oral movements by acting on pathways other than the sensorimotor one in the rat basal ganglia," *Neuroscience*, vol. 169, no. 1, pp. 158–170, 2010.
- [142] S. Kapur and G. Remington, "Serotonin-dopamine interaction and its relevance to schizophrenia," *American Journal of Psychiatry*, vol. 153, no. 4, pp. 466–476, 1996.

- [143] J. F. Liégeois, J. Ichikawa, and H. Y. Meltzer, "5-HT<sub>2A</sub> receptor antagonism potentiates haloperidol-induced dopamine release in rat medial prefrontal cortex and inhibits that in the nucleus accumbens in a dose-dependent manner," *Brain Research*, vol. 947, no. 2, pp. 157–165, 2002.
- [144] G. Meco, P. Stirpe, F. Editto et al., "Aripiprazole in l-dopa-induced dyskinesias: a one-year open-label pilot study," *Journal of Neural Transmission*, vol. 116, no. 7, pp. 881–884, 2009.
- [145] M. Carlsson and A. Carlsson, "Interactions between glutamatergic and monoaminergic systems within the basal ganglia—implications for schizophrenia and Parkinson's disease," *Trends in Neurosciences*, vol. 13, no. 7, pp. 272–276, 1990.
- [146] E. Wolf, K. Seppi, R. Katzenschlager et al., "Long-term antidyskinetic efficacy of amantadine in Parkinson's disease," *Movement Disorders*, vol. 25, no. 10, pp. 1357–1363, 2010.
- [147] S. Navailles and P. De Deurwaerdère, "Presynaptic control of serotonin on striatal dopamine function," *Psychopharmacology*, vol. 213, no. 2-3, pp. 213–242, 2010.
- [148] B. Gomez-Mancilla and P. J. Bedard, "Effect of nondopaminergic drugs on L-DOPA-induced dyskinesias in MPTP-treated monkeys," *Clinical Neuropharmacology*, vol. 16, no. 5, pp. 418–427, 1993.
- [149] C. G. Goetz, P. Damier, C. Hicking et al., "Sarizotan as a treatment for dyskinesias in Parkinson's disease: a double-blind placebo-controlled trial," *Movement Disorders*, vol. 22, no. 2, pp. 179–186, 2007.
- [150] K. L. Eskow, V. Gupta, S. Alam, J. Y. Park, and C. Bishop, "The partial 5-HT<sub>1A</sub> agonist buspirone reduces the expression and development of l-DOPA-induced dyskinesia in rats and improves l-DOPA efficacy," *Pharmacology Biochemistry and Behavior*, vol. 87, no. 3, pp. 306–314, 2007.
- [151] K. Kannari, K. Kurahashi, M. Tomiyama et al., "Tandospirone citrate, a selective 5-HT<sub>1A</sub> agonist, alleviates L-DOPA-induced dyskinesia in patients with Parkinson's disease," *Brain and Nerve*, vol. 54, no. 2, pp. 133–137, 2002.
- [152] M. M. Iravani, K. Tayarani-Binazir, W. B. Chu, M. J. Jackson, and P. Jenner, "In 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-treated primates, the selective 5-hydroxytryptamine 1a agonist (R)-(+)-8-OHDPAT inhibits levodopa-induced dyskinesia but only with increased motor disability," *Journal of Pharmacology and Experimental Therapeutics*, vol. 319, no. 3, pp. 1225–1234, 2006.
- [153] M. Tomiyama, T. Kimura, T. Maeda, K. Kannari, M. Matsunaga, and M. Baba, "A serotonin 5-HT<sub>1A</sub> receptor agonist prevents behavioral sensitization to L-DOPA in a rodent model of Parkinson's disease," *Neuroscience Research*, vol. 52, no. 2, pp. 185–194, 2005.