

## Research Article

# Toxoplasmosis Behind Bars: One Health Approach on Serosurvey Dynamics and Associated Risk Factors for Women Inmates, Correctional Officers, and In-Prison Feral Cats

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Brazil holds the third highest general and fifth female incarcerated population worldwide. Despite the incarceration ecosystem that may favor the spreading of zoonotic diseases, particularly when unattended animals are present, no comprehensive study has focused on toxoplasmosis dynamics in such environment. Accordingly, the present study has aimed to serologically assess anti-*Toxoplasma gondii* (IgG) antibodies by indirect immunofluorescent antibody test in inmates, correctional officers, and feral cats at the Women's State Penitentiary of Parana, southern Brazil. In overall, 230/506 (45.5%; CI 95%: 41.2–49.8) incarcerated women, 31/91 (34.1%; 95% CI: 25.2–44.3) correctional officers, and 23/39 (59.0%; CI 95%: 43.2–72.9) cats were seropositive to anti-*T. gondii* antibodies. Logistic regression revealed that seropositivity likelihood increased with consumption of raw meat ( $p = 0.040$ ) and decreased with elementary educational level ( $p = 0.001$ ). No statistical difference was found comparing seropositivity between inmates and correctional officers ( $p = 0.057$ ). As women inmates have been considered among the most vulnerable groups in disease morbidity and mortality, seropositivity observed herein may be directly related to vulnerability and high *T. gondii* oocyst exposure dispersed in cat feces during incarceration.

## 1. Introduction

Brazil has been ranked as the third highest general incarcerated population worldwide, with 811,000 persons deprived of liberty, surpassed only by China with 1.69 and the USA with around 2.0 million people [1]. In addition, Brazil holds the fifth largest female incarcerated population with 37,380 individuals, only behind the USA (205,400), China (103,766), Russia (53,304), and Thailand (44,751) [2]. Despite being considered

a small fraction of the total population worldwide, female incarceration has increased by 50% in the past 15 years [1, 3].

Incarcerated persons, along with refugees and homeless, have been considered the three most vulnerable populations in disease morbidity and mortality [4]. Incarceration environment may increase the likelihood of infectious diseases, as secluded populations may exacerbate vulnerability due to movement restriction, mutual convivence within confined spaces, and limited health assistance [5]. Such daily life

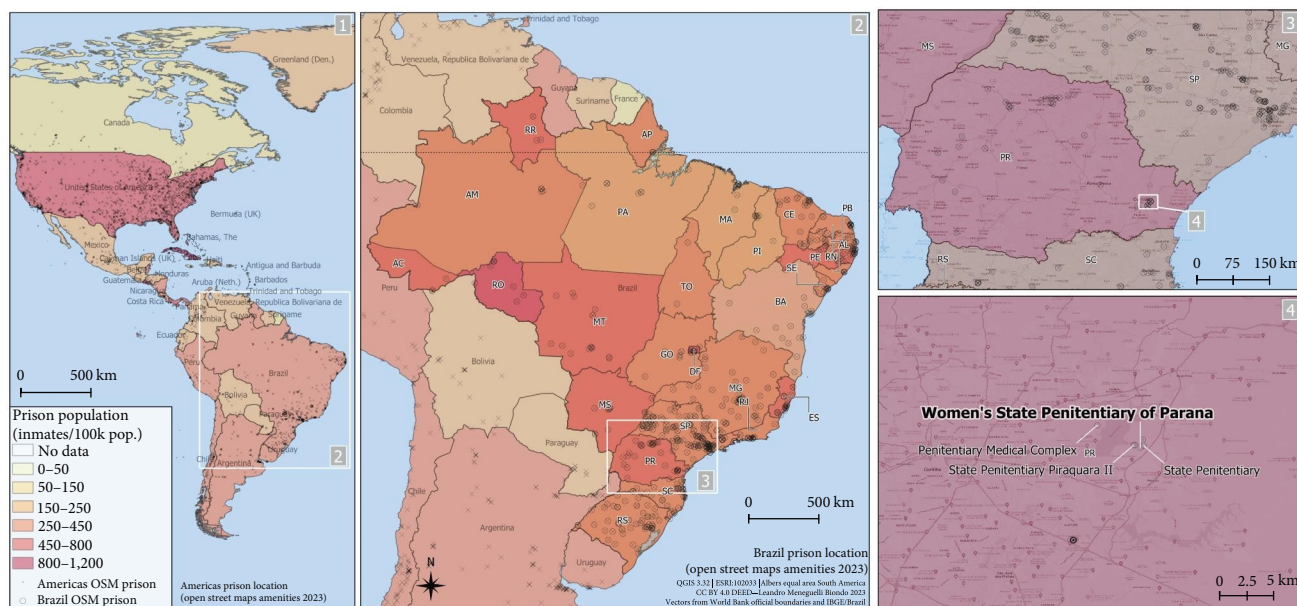


FIGURE 1: Sampling location of inmates, correctional officers, and feral cats in the Women's State Penitentiary of Parana, southern Brazil. Prison population distribution in Brazilian states was also highlighted, according to the Infopen system from the Public Ministry [16].

within overcrowded prisons and their precariousness has made the environment conducive to disease spreading [6]. For instance, the incidence of tuberculosis (TB) in prisons may be 10 times greater than the general population [7]. Susceptibility to infectious diseases may be even higher in immunocompromised populations, with four to fivefold more likelihood for toxoplasmosis in HIV-positive inmates [8, 9]. Furthermore, women inmates may have poor access to health care [3, 10].

Considered a ubiquitous protozoan parasite, *Toxoplasma gondii* has caused toxoplasmosis, a zoonotic infection with a worldwide distribution [11]. The disease has a complex epidemiology, being transmitted by ingestion of oocysts shed into feces of definitive feline hosts and contaminate water, soil, and crops, and by consumption of intracellular cysts in undercooked meat from intermediate hosts [12]. Despite usually being an asymptomatic disease in immunocompetent individuals, toxoplasmosis has been considered relevant to public health primarily within the context of congenital toxoplasmosis or postnatally acquired disease in immunocompromised patients [13].

The complexity of health problems worldwide, particularly in developing countries, has demanded a more holistic and systemic approach involving interrelations of medical and environmental sciences, in a One Health convergence [14]. One in 1,000 Latin American persons may be affected by a parasitic zoonosis, according to the World Health Organization, with toxoplasmosis indicated as among the most frequent and priority for public actions [15].

Despite the closed ecosystem of incarceration that may favor the spreading of zoonotic diseases, particularly when unattended animals are present, no comprehensive study has focused on toxoplasmosis dynamics in the women inmate population. Accordingly, the present study has aimed to

serologically assess anti-*T. gondii* (IgG) antibodies in all inmates, correctional officers, and feral cats at the Women's State Penitentiary of Parana, southern Brazil.

## 2. Materials and Methods

**2.1. Ethical Aspects.** The present study was performed in coordination with the administration of the Penitentiary Department of the Piraquara Complex, officially included as part of their official activities, along with COVID-19 sampling and testing.

**2.2. Study Design.** The present study was a cross-sectional serosurvey of anti-*T. gondii* antibodies (IgG) and associated risk factors in women deprived of liberty, correctional officers, and feral cats of a state penitentiary in southern Brazil. In addition, a longitudinal approach was performed to compare seroprevalence during the COVID-19 epidemics.

**2.3. Study Population and Area.** Both inmate and correctional officer populations were sampled in total twice, first in October 2020 and later in November 2021, while feral cats were captured and sampled during the study period. Socio-epidemiological information was obtained by signed questionnaires. The study was fully conducted at the Women's State Penitentiary of Parana State (Figure 1), part of the Piraquara Correctional Complex, third in population in the Brazilian incarceration system with around 7,000 men and women inmates held by 800 correctional officers and arranged in seven different units at the time. The complex, located in the Piraquara municipality ( $25^{\circ}24'59''\text{S}$ ,  $49^{\circ}04'46''\text{W}$ ), is part of the Curitiba metropolitan area, capital of Paraná State and the eighth largest Brazilian city with around 3.2 million habitants.

TABLE 1: Information for assessing the potential exposure and associated risk factors for toxoplasmosis in women inmates and correctional agents in a penitentiary of Paraná State, southern Brazil.

Topics	Gathered information	
	Inmates	Correctional agents
Socioeconomic characteristics	Age, ethnicity, educational level	Gender, age, ethnicity, educational level
Gestational characteristics	Pregnant, mother, history of miscarriage	History of miscarriage
Practices and hygienic habits in penitentiary	Hours in cell, contact with cats, frequency in solarium, washing hands before meals, contact with soil, biting nails	Contact with cats, meals provided by the penitentiary, consuming of water in the penitentiary
History	Cat owner before detention, raw meat ingestion	Raw meat ingestion

**2.4. Blood Sampling and Epidemiological Data.** Participants herein, including inmates and officers, were sampled after signing a consent form and filling out an epidemiological questionnaire. Approximately 8 mL of whole blood was collected by cephalic venipuncture in humans and by jugular venipuncture in cats, collected by certified nurses and veterinarians, respectively. Samples were placed in a tube with serum separator gel, centrifuged at 800 g for 5 min, and subsequently, serum separated and kept at  $-20^{\circ}\text{C}$  until processing.

Epidemiological data were obtained from inmates and correctional officers after informed notice, confidentiality of their identities, and right to refuse participation at any time. Participants were asked to voluntarily sign the Free and Informed Consent Term in compliance with the National Brazilian Health Council (resolution no. 441/2012). The epidemiological questionnaire was based on potential associated risk factors for toxoplasmosis (Table 1).

As counterpart and ethical research, the study herein included a free-of-charge assistance to feral cats at the penitentiary, conducted by the shelter medicine service at the Veterinary Teaching Hospital, Federal University of Paraná. Once per week, feral cats were trapped inside the buildings and courtyards, examined, treated (if necessary), dewormed, given anti-flea, vaccinated, microchipped, and taken for neutering/spaying. Kittens and docile cats were sent for adoption, while healthy feral cats were released back into the penitentiary and monitored.

**2.5. Serological Testing.** Serological testing of anti-*T. gondii* antibodies were performed by indirect immunofluorescent antibody test (IFAT) in both human and cat samples, using correspondent species-specific conjugates, as previously established [17]. Serial dilutions from 1 : 16 to 1 : 4,096 were applied in pH 7.2 phosphate-buffered saline solution, using a cutoff titer of  $\geq 16$  IU. Immunofluorescence slides were presensitized with 0.1% formaldehyde to inactivate *T. gondii* tachyzoites (RH strain), obtained from an intraperitoneal lavage in Swiss mice following 3-day inoculation.

**2.6. Statistical Analyses.** All statistical analyses were performed using R software. First, collected data were assessed for detecting missing values, outliers, and inconsistencies. Then, data were cleaned by excluding values based on the missingness proportion ( $>10\%$ ) and the nature of missing values. Continuous variables were transformed into binary

indicators or categories. Descriptive analyses were performed to summarize the data using contingency tables.

Univariate analyses were performed to assess the relationship between each independent variable and seropositivity, applying either the chi-square or Fisher's exact test. All calculations were performed independently by group, with the inclusion of information given by inmates (506) and correctional officers (91). Dependent variables were selected for inclusion in the logistic regression model based on the significance of univariate analysis ( $p < 0.2$ ). Multicollinearity among independent variables was checked to avoid high intrinsic correlation. Odds ratio and confidence intervals were calculated to quantify the strength of associations in the logistic regression model.

The logistic regression model was refined by adding or removing variables based on their statistical significance, effect size, and theoretical relevance. This refinement process continued until a final model with parsimonious and interpretable results was obtained. The *T. gondii* antibody dynamics were compared based on inmates (133) tested in both samplings.

The area under the ROC curve (AUC) was computed as a measure of the model's discriminative ability, ranging from 0.5 (no) to 1 (perfect) discrimination. Confidence intervals for the AUC were estimated to assess the statistical significance of the discriminatory power.

A significant level of 5% was adopted for all statistical tests.

### 3. Results

**3.1. Incarcerated Women.** A total of 506 incarcerated women were included in the study, with ages ranging from 18 to 62 years (median: 32), mostly self-declared as white (269/506; 53.2%), having elementary education (337/506; 66.6%), and at least one child (412/506; 81.4%). Two women referred to self-pregnancy at the time of questionnaire application.

Overall, 230/506 (45.5%; CI 95%: 41.2–49.8) incarcerated women were seropositive to anti-*T. gondii* antibodies. Out of the inmates tested again after a 1-year interval, 92/133 (69.2%) presented the same results, while 12/133 (9.0%) became seronegative and 29/133 (21.8%) seropositive (Table 2). A statistically significant difference in proportions was found in seropositive results when comparing the two serological tests (McNemar's *chi-squared* = 6.24, *df* = 1, *p*-value = 0.012).

TABLE 2: Serological results by indirect immunofluorescent antibody test (IFAT) in a 1-year interval for anti-*Toxoplasma gondii* antibodies among incarcerated women ( $N = 133$ ) in a penitentiary of Paraná State, southern Brazil.

IFAT results	Positive (%)	Negative (%)	Total
First testing	58 (43.6)	75 (56.4)	133
Second testing (1-year interval)	75 (56.4)	58 (43.6)	133

Risk factors associated to seropositivity for anti-*T. gondii* antibodies were assessed and presented (Table 3). Logistic regression revealed that seropositivity likelihood increased with consumption of raw meat (OR: 1.53;  $p = 0.040$ ) and decreased with elementary educational level (OR: 0.47;  $p = 0.001$ ).

Univariate analysis showed a significant proportion of seropositive women in the non-white group ( $p = 0.036$ ), but the variable was not retained in the logistic regression ( $p = 0.052$ ). Hours in cell and frequency in solarium were also fitted to be included in the logistic regression, but neither variable was retained in the final model.

All other assessed variables (age, cat owner before detention, contact with cats during detention, washing hands before meals, pregnant status, mother, historic of miscarriage, contact with soil during detention, biting nails) were not associated to seropositivity ( $p > 0.05$ ).

**3.2. Correctional Officers.** The group of correctional officers herein was constituted of 91 individuals aging from 21 to 63 (median: 44), mostly female (84/91; 92.3%), self-declared white (60/91; 65.9%), and with higher education (65/91; 71.4%).

Serological examination revealed 31/91 (34.1%; 95% CI: 25.2–44.3) seropositive officers for toxoplasmosis. No tested variable, including gender, ethnicity, educational level, contact with cats in the penitentiary, meals provided by the penitentiary, consumption of water in the penitentiary, historic of raw meat ingestion, and history of miscarriage, was associated to seropositivity (Table 4).

In addition, no statistical difference was found comparing seropositivity proportions between inmates and correctional officers (OR: 1.6; 95% CI: 1.010–2.575;  $p = 0.0572$ ).

**3.3. Feral Cats.** Out of the feral cats trapped in this study, 19/39 (48.7%) were males and 20/39 (51.3%) females, mostly adults. Overall, 23/39 (59.0%; CI 95%: 43.2–72.9) cats were seropositive for anti-*T. gondii* antibodies. No statistical difference (chi-square: 0.037;  $p = 0.848$ ) in seropositivity was verified considering cat age and gender.

## 4. Discussion

The present study represents the first serosurvey conducted to assess seroprevalence for toxoplasmosis in the female inmate population of Brazil. Previous studies worldwide have mostly focused on incarcerated pregnant women, revealing South America as the highest pooled metanalysis seroprevalence (56.2%; 50.5%–62.8%) for latent toxoplasmosis [18], similar to that observed in the general female population herein (45.5%). Even so, seroprevalence herein was higher than the male inmate population, with 43/170 (25.3%) in Malaysia [9] and 207/497 (41.6%) in Indonesia [8].

Such results should be carefully compared due to different cultural, socioeconomic, gender, and pathogen exposure found in various countries and inmate populations. Additionally, the antibody testing method and cutoff point adopted herein may also be associated with these differences.

Toxoplasmosis has been considered widely prevalent in Brazil [19], with the highest seroprevalence recorded to date revealed in a blood donor serosurvey (75.0%; CI 95%: 68.0–82.0) [20]. In addition, this neglected parasitic disease has been associated with poverty [21, 22], with a significantly higher prevalence of latent toxoplasmosis in pregnant women in low-income countries with low human development indexes [18]. Socioeconomic vulnerability has been associated to disease in Brazil [23] and is considered a contributing factor to the persistent occurrence of congenital toxoplasmosis [24]. The logistic analysis herein revealed that seropositivity was significantly reduced in inmates with high school, in agreement with low education as a risk factor for toxoplasmosis in puerperal women [25–27].

Ethnicity has been associated to toxoplasmosis in pregnant women, as observed in Brazil [28] and in the USA [22]. Although the univariate analysis herein has pointed out a statistically higher proportion of seropositive women in the non-white group, such a variable was not retained by logistic regression ( $p = 0.052$ ). Thus, future studies should be conducted with higher in mate samplings, if possible, in a multi-centric design, to fully establish such ethnicity as an associated risk factor in female inmates in Brazil and abroad.

Although only two inmates in the present study self-declared pregnant at the time, most of them referred to having at least one child. Despite no statistical significance was found between seropositivity and motherhood or miscarriage, the last has been reported as a consequence of toxoplasmosis in pregnant women [29, 30]. As miscarriage may be a result of a variety of disorders, a higher sampling with a survey of concomitant causes should be performed to pinpoint the role of toxoplasmosis, alone or combined.

As expected, ingestion of raw meat increased the odds of seropositivity in the inmates herein (OR: 1.53), a major risk factor for toxoplasmosis worldwide [19, 31, 32]. As the majority of female prisons in Brazil have been supplied with ultra-processed food, mostly fried chicken (79.2%) as a meat source [33], food during incarceration should not be an important source of *T. gondii* infection. Similarly, such “fast-food effect” due to diet habits of mostly processed food has also been shown in homeless [34] and animal hoarding [35] populations in Brazil, which also presented lower anti-*T. gondii* seroprevalence than the general population. Meat processing and cooking were also shown to reduce *T. gondii* levels in meat products in the USA [36]. As human anti-*T. gondii* antibodies

TABLE 3: Associated risk factors for anti-*Toxoplasma gondii* antibodies in incarcerated women (N= 506) in a penitentiary of Paraná State, southern Brazil.

	Serological results (%)		Univariate analysis		Multivariate analysis	
	Positive	Negative	OR (95% CI)	<i>p</i>	OR (95% CI)	<i>p</i>
	230 (45.5)	276 (54.5)	—	—	—	—
Age (years old)	—	—	—	0.301	—	—
18–26	55 (24.0)	68 (24.6)	1 (Reference)	—	—	—
27–31	58 (25.3)	65 (23.6)	1.10 (0.67–1.83)	—	—	—
32–38	45 (19.7)	72 (26.1)	0.77 (0.46–1.30)	—	—	—
>38	71 (31.0)	71 (25.7)	1.23 (0.76–2.01)	—	—	—
Ethnicity	—	—	—	0.036	—	—
Non-white	119 (52.0)	116 (42.2)	1 (Reference)	—	—	—
White	110 (48.0)	159 (57.8)	0.68 (0.47–0.96)	—	0.68 (0.45–1.00)	0.052
Educational level	—	—	—	0.001	—	—
Elementary	172 (74.8)	165 (60.0)	1 (Reference)	—	—	—
High school	44 (19.1)	93 (33.8)	0.46 (0.30–0.69)	—	0.47 (0.29–0.75)	0.001
Graduate	14 (6.09)	17 (6.18)	0.79 (0.37–1.67)	—	0.76 (0.32–1.71)	0.500
Hours in cell	—	—	—	0.007	—	—
All day	168 (75.7)	170 (63.9)	1 (Reference)	—	—	—
18 hr	54 (24.3)	96 (36.1)	0.57 (0.38–0.85)	—	0.95 (0.57–1.58)	0.841
Cat owner before detention	—	—	—	0.354	—	—
No	142 (66.0)	188 (70.4)	1 (Reference)	—	—	—
Yes	73 (34.0)	79 (29.6)	1.22 (0.83–1.80)	—	—	—
Contact with cats during detention	—	—	—	0.982	—	—
No	156 (72.9)	186 (72.4)	1 (Reference)	—	—	—
Yes	58 (27.1)	71 (27.6)	0.97 (0.65–1.46)	—	—	—
Frequency in solarium (per week)	—	—	—	0.067	—	—
No access	37 (17.6)	47 (18.7)	1 (Reference)	—	—	—
Once	64 (30.5)	100 (39.7)	0.81 (0.48–1.39)	—	0.89 (0.49–1.59)	0.687
Twice or more	109 (51.9)	105 (41.7)	1.32 (0.79–2.20)	—	1.40 (0.80–2.45)	0.235
Washing hands before meals	—	—	—	0.363	—	—
No	7 (3.08)	4 (1.48)	1 (Reference)	—	—	—
Yes	220 (96.9)	267 (98.5)	0.48 (0.12–1.65)	—	—	—
History of raw meat ingestion	—	—	—	0.021	—	—
No	127 (58.8)	183 (69.3)	1 (Reference)	—	—	—
Yes	89 (41.2)	81 (30.7)	1.58 (1.09–2.31)	—	1.53 (1.02–2.29)	0.040
Pregnant	—	—	—	1	—	—
No	216 (99.5)	268 (99.6)	1 (Reference)	—	—	—
Yes	1 (0.46)	1 (0.37)	1.24 (0.03–48.6)	—	—	—
Mother	—	—	—	0.364	—	—
No	34 (15.0)	50 (18.5)	1 (Reference)	—	—	—
Yes	192 (85.0)	220 (81.5)	1.28 (0.80–2.08)	—	—	—
History of miscarriage	—	—	—	0.545	—	—
No	144 (69.2)	168 (66.1)	1 (Reference)	—	—	—
Yes	64 (30.8)	86 (33.9)	0.87 (0.58–1.29)	—	—	—
Contact with soil during detention	—	—	—	0.791	—	—
No	207 (93.7)	252 (92.6)	1 (Reference)	—	—	—
Yes	14 (6.33)	20 (7.35)	0.86 (0.41–1.73)	—	—	—
Biting nails	—	—	—	0.485	—	—
No	151 (68.9)	173 (65.5)	1 (Reference)	—	—	—
Yes	68 (31.1)	91 (34.5)	0.86 (0.58–1.26)	—	—	—

TABLE 4: Associated risk factors for anti-*Toxoplasma gondii* antibodies in correctional officers ( $N=91$ ) in a penitentiary of Paraná, southern Brazil.

Variables	Serological results		Univariate analysis	
	Positive (%)	Negative (%)	OR (95% CI)	$p$ Overall
	31 (34.1)	60 (65.9)	—	—
Gender	—	—	—	0.224
Female	27 (87.1)	57 (95.0)	1 (Reference)	—
Male	4 (12.9)	3 (5.00)	2.75 (0.54–15.8)	—
Age (years old)	—	—	—	0.204
<38	9 (30.0)	12 (20.3)	1 (Reference)	—
38–43	6 (20.0)	15 (25.4)	0.54 (0.14–1.98)	—
44–50	4 (13.3)	18 (30.5)	0.31 (0.07–1.21)	—
>50	11 (36.7)	14 (23.7)	1.05 (0.32–3.48)	—
Ethnicity	—	—	—	0.695
Non-white	9 (29.0)	21 (35.6)	1 (Reference)	—
White	22 (71.0)	38 (64.4)	1.34 (0.53–3.59)	—
Educational level	—	—	—	0.389
Elementary	3 (9.68)	2 (3.33)	1 (Reference)	—
High school	8 (25.8)	13 (21.7)	0.43 (0.04–3.44)	—
Graduate	20 (64.5)	45 (75.0)	0.31 (0.03–2.18)	—
Contact with cats in the penitentiary	—	—	—	0.434
No	13 (48.1)	20 (36.4)	1 (Reference)	—
Yes	14 (51.9)	35 (63.6)	0.62 (0.24–1.60)	—
Meals provided by the penitentiary	—	—	—	0.242
No	12 (40.0)	15 (25.4)	1 (Reference)	—
Yes	18 (60.0)	44 (74.6)	0.52 (0.20–1.34)	—
Consuming of water in the penitentiary	—	—	—	0.988
No	7 (22.6)	12 (20.0)	1 (Reference)	—
Yes	24 (77.4)	48 (80.0)	0.85 (0.30–2.59)	—
Historic of raw meat ingestion	—	—	—	0.593
No	20 (66.7)	35 (58.3)	1 (Reference)	—
Yes	10 (33.3)	25 (41.7)	0.71 (0.27–1.76)	—
Historic of miscarriage	—	—	—	0.365
No	24 (88.9)	43 (79.6)	1 (Reference)	—
Yes	3 (11.1)	11 (20.4)	0.51 (0.10–1.86)	—

may provide a lifelong protective immunity [36], and infection via raw meat may have been acquired previously to incarceration.

Unwashed fruits and vegetables and untreated water may also be important routes of *T. gondii* transmission to women [12]. Although water and foodborne (unwashed fruits and vegetables) transmission have been reportedly among the most frequent routes of toxoplasmosis outbreaks in Brazil [36, 37], food distributed herein came from industrial kitchens commercially contracted outside the penitentiary (under tight inspection and food handling practices), as the in-prison kitchen was closed at the beginning of COVID-19 pandemics. Thus, unless contamination occurred during in-prison storage and handling, food and water consumed by inmates herein may be unlikely sources of infection during the survey.

The central point in the present study was the overlapping presence of 23/39 (59.0%) seropositive feral cats in the penitentiary, serving as an alert for the environmental source

of *T. gondii* infection during incarceration. The seropositivity observed in feral cats herein was significantly higher than the 16.3% (46/282) observed in owned cats in the same city [38] but lower than the 84.4% (49/58) found in other Paraná state areas. In addition, 21.92% (98/447) of domestic cats were seropositive in northern Brazil, with a statistically significant association between age and serology among cats over 1 year old [39]. Contact with cats or with soil containing infective *T. gondii* oocysts has already been established as a risk factor for toxoplasmosis [14]. However, no association was observed herein between seropositivity and cat or soil contact, probably due to the high number of women who declared no contact with cats or with soil during the prison period. Nonetheless, penitentiary outdoor sunlight areas, daily used by female inmates, were heavily contaminated with *Toxocara cati* eggs [40], suggesting daily inmate close contact with cat feces.

As the feral cat population herein was not fully neutered and spayed at the beginning of samplings, undesirable cat

litters may have been responsible for historical *T. gondii* oocyst shedding, transmission, and persistence. In addition, cats freely transit through inside and outside prison may have allowed contact with other sources of *T. gondii*, such as prey meat and contaminated water. Finally, the feral cat population herein has been maintained by food scraps of daily meals produced in prison prior to COVID-19, suggesting a potential sharing of infection sources.

Another strength of this study was to investigate the *T. gondii* seropositivity dynamics in a 1-year period. A fluctuation was observed in both seronegative and seropositive results, with a significant difference in the proportions of seropositive results comparing the two serological time tests. As tests were performed in duplicate, such findings herein may be attributed to IFAT sensitivity in general, which has ranged from 80.4% to 100%, and specificity from 91.4% to 95.8%, depending on the tested species [41]. Such findings could be attributed to IFAT sensitivity, but tests were performed in duplicate. Further, concurrent infection could lead to a disruption in immunocompetence and recrudescence of toxoplasmosis [42], with a positive correlation between serum anti-*T. gondii* IgG levels and in TB patients, for instance, as observed in Egypt [43]. In addition, an increase in TB has been associated to HIV in prison inmates in southern Brazil [44]. As information regarding HIV results were lost over 50%, no analysis was possible to draw between toxoplasmosis and HIV coinfection. Thus, further investigation should be conducted to evaluate the reciprocal influence of antibody dynamics for toxoplasmosis and concurrent infections.

The seroprevalence of correctional officers and potential associated risk factors for toxoplasmosis were also evaluated. The observed herein 34.1% seroprevalence was very close to the overall 35.0% observed in a global meta-analysis [45]. No tested variable was associated to seropositivity, and seropositivity proportions between inmates and correctional officers were not associated.

As limitations of this study, although feral cat prevalence found herein has indicated high cat exposure and a high degree of environmental contamination, no *T. gondii* oocyst recovery from soil, environment contamination, and genotyping analysis were performed, and should be further investigated. This finding needs to be interpreted with caution due to the self-declared information given by inmates, which could lead to a bias and misinterpretation of the results. In addition, the combined use of another serological method (e.g., ELISA) with IFAT could help to better understand the seroconversion dynamics observed herein. According to a meta-analysis, the use of enzyme-linked fluorescent immunoassay, consisting of an immunoenzymatic method with the final fluorescence detection, has provided a 92% sensitivity and 80% specificity for the IgG marker, highly improving the sensitivity accuracy [46].

Finally, as women deprived of liberty have been considered among the most vulnerable groups in disease morbidity and mortality [4], seropositivity observed herein may be directly related to vulnerability and high *T. gondii* oocyst exposure dispersed in cat feces during incarceration.

## 5. Conclusion

The study herein represented the first serosurvey conducted to assess seroprevalence for toxoplasmosis in the female inmate population of Brazil. The seropositivity observed herein may be directly related to vulnerability and high *T. gondii* oocyst exposure dispersed in cat feces during incarceration. In addition, as the feral cat population herein was not fully neutered and spayed at the beginning of samplings, undesirable cat litters may have been responsible for historical *T. gondii* oocyst shedding, transmission, and persistence. Finally, cats freely transit through inside and outside the prison and may have allowed contact with different sources of *T. gondii*, such as water and prey.

## Data Availability

All relevant data are within the manuscript and its supporting information files.

## Ethical Approval

The present study was approved by the Brazilian National Human Health Council (register 31676020.3.0000.0102, protocol number 4.177.728) through the Human Ethics Committee of the Federal University of Paraná. In addition, feral cat capture, transportation, treatment, neutering, and return were approved by the Ethics Committee of Animal Use at the Federal University of Paraná (protocol number 022/2022).

## Conflicts of Interest

The authors have declared that no competing interests exist.

## Authors' Contributions

Conceptualization was done by AWB; methodology was done by HL, LBK, and AWB; validation was done by VAS and HL; formal analysis was done by VAS, RTSF, DAF, GNM, LMB, RG, and HL; data curation was done by GLBP, JK, and AWB; writing—original draft preparation was done by GLBP, VAS, RG, LBK, and AWB; writing—review and editing was done by GLBP, VAS, JR, RTSF, DAF, GNM, JK, LMB, RG, HL, LBK, and AWB; supervision was done by AWB. GLBP and VAS contributed equally to this work.

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## References

- [1] World Prison Brief, "An online database comprising information on prisons and the use of imprisonment around the world," 2023, (n.d.), <https://www.prisonstudies.org/>.

- [2] M. d. Justiça, “Bem vindo—Ministério da Justiça e Segurança Pública,” 2023, (n.d.), <https://dados.mj.gov.br/>.
- [3] M. C. Van Hout and R. Mhlanga-Gunda, “Contemporary women prisoners health experiences, unique prison health care needs and health care outcomes in sub Saharan Africa: a scoping review of extant literature,” *BMC International Health and Human Rights*, vol. 18, pp. 1–12, 2018.
- [4] R. W. Aldridge, A. Story, S. W. Hwang et al., “Morbidity and mortality in homeless individuals, prisoners, sex workers, and individuals with substance use disorders in high-income countries: a systematic review and meta-analysis,” *The Lancet*, vol. 391, no. 10117, pp. 241–250, 2018.
- [5] M. J. Akiyama, A. C. Spaulding, and J. D. Rich, “Flattening the curve for incarcerated populations—Covid-19 in jails and prisons,” *New England Journal of Medicine*, vol. 382, no. 22, pp. 2075–2077, 2020.
- [6] F. Bernardes Filho, J. M. Santana, R. C. P. de Almeida et al., “Leprosy in a prison population: a new active search strategy and a prospective clinical analysis,” *PLOS Neglected Tropical Diseases*, vol. 14, no. 12, Article ID e0008917, 2020.
- [7] S. Charalambous, K. Velen, Z. Rueda et al., “Scaling up evidence-based approaches to tuberculosis screening in prisons,” *The Lancet Public Health*, vol. 8, no. 4, pp. e305–e310, 2023.
- [8] A. A. Prasetyo, Y. Sari, S. Haryati, and I. Raharjo, “Toxoplasma and viral antibodies among HIV patients and inmates in central java, Indonesia,” in *Toxoplasma and Blood Borne Viral Antibodies*, vol. 46, 2015.
- [9] L. Angal, Y. A. Lim, N. J. Yap et al., “Toxoplasmosis in HIV and non HIV prisoners in Malaysia,” *Tropical Biomedicine*, vol. 33, pp. 159–169, 2016.
- [10] P. Abbott, P. Magin, J. Davison, and W. Hu, “Medical homelessness and candidacy: women transiting between prison and community health care,” *International Journal for Equity in Health*, vol. 16, no. 1, Article ID 130, 2017.
- [11] L. Kamus, S. Belec, L. Lambrecht et al., “Maternal and congenital toxoplasmosis in Mayotte: prevalence, incidence and management,” *PLOS Neglected Tropical Diseases*, vol. 17, no. 3, Article ID e0011198, 2023.
- [12] N. C. Smith, C. Goulart, J. A. Hayward, A. Kupz, C. M. Miller, and G. G. van Dooren, “Control of human toxoplasmosis,” *International Journal for Parasitology*, vol. 51, no. 2-3, pp. 95–121, 2021.
- [13] G. Milne, J. P. Webster, and M. Walker, “*Toxoplasma gondii*: an underestimated threat ?” *Trends in Parasitology*, vol. 36, no. 12, pp. 959–969, 2020.
- [14] A. A. Aguirre, T. Longcore, M. Barbieri et al., “The One Health approach to toxoplasmosis: epidemiology, control, and prevention strategies,” *EcoHealth*, vol. 16, no. 2, pp. 378–390, 2019.
- [15] CDC, “Toxoplasmosis,” 2023, (accessed November 2, 2023), (n.d.), <https://www.cdc.gov/parasites/toxoplasmosis/index.html>.
- [16] Ministerio da Justicia e Seguridad Publica, “Bases de Datos—Secretaria Nacional de Políticas Penais,” 2023, <https://www.gov.br/senappen/pt-br/servicos/sisdepen/bases-de-dados/base-s-de-dados>.
- [17] M. E. Camargo, “Introdução às técnicas de imunofluorescência,” in *Introdução Às Técnicas Imunofluorescência*, Article ID 112, 1973.
- [18] A. Rostami, S. M. Riahi, H. R. Gamble et al., “Global prevalence of latent toxoplasmosis in pregnant women: a systematic review and meta-analysis,” *Clinical Microbiology and Infection*, vol. 26, no. 6, pp. 673–683, 2020.
- [19] S. Almeria and J. P. Dubey, “Foodborne transmission of *Toxoplasma gondii* infection in the last decade. An overview,” *Research in Veterinary Science*, vol. 135, pp. 371–385, 2021.
- [20] M. Foroutan-Rad, H. Majidiani, S. Dalvand et al., “Toxoplasmosis in blood donors: a systematic review and meta-analysis,” *Transfusion Medicine Reviews*, vol. 30, no. 3, pp. 116–122, 2016.
- [21] P. J. Hotez, “Neglected diseases and poverty in “The Other America”: the greatest health disparity in the United States ?” *PLoS Neglected Tropical Diseases*, vol. 1, no. 3, Article ID e149, 2007.
- [22] J. L. Jones, D. Kruszon-Moran, S. Elder et al., “*Toxoplasma gondii* infection in the United States, 2011–2014,” *The American Journal of Tropical Medicine and Hygiene*, vol. 98, no. 2, pp. 551–557, 2018.
- [23] M. Mareze, A. N. Benitez, A. P. D. Brandão et al., “Socioeconomic vulnerability associated to *Toxoplasma gondii* exposure in southern Brazil,” *PLOS ONE*, vol. 14, no. 2, Article ID e0212375, 2019.
- [24] B. B. De La Fuente Villar, E. de S. Neves, V. C. Louro et al., “Toxoplasmosis in pregnancy: a clinical, diagnostic, and epidemiological study in a referral hospital in Rio de Janeiro, Brazil,” *The Brazilian Journal of Infectious Diseases*, vol. 24, no. 6, pp. 517–523, 2020.
- [25] M. Laboudi, “Review of toxoplasmosis in Morocco: seroprevalence and risk factors for toxoplasma infection among pregnant women and HIV-infected patients,” *Pan African Medical Journal*, vol. 27, Article ID 269, 2017.
- [26] T. R. Olariu, S. Ursoniu, I. Hotea, V. Dumitrascu, D. Anastasiu, and M. A. Lupu, “Seroprevalence and risk factors of *Toxoplasma gondii* infection in pregnant women from western Romania,” *Vector-Borne and Zoonotic Diseases*, vol. 20, no. 10, pp. 763–767, 2020.
- [27] J. F. Medeiros, A. C. R. E. Silva, N. D. F. da Rocha et al., “Seroprevalence of toxoplasmosis in puerperal women treated at a tertiary referral hospital,” *Revista Brasileira de Ginecologia e Obstetrícia/RBGO Gynecology and Obstetrics*, vol. 45, no. 2, pp. 59–64, 2023.
- [28] A. L. Sartori, R. Minamisava, M. M. Avelino, and C. A. Martins, “Triagem pré-natal para toxoplasmose e fatores associados à soropositividade de gestantes em Goiânia, Goiás,” *Revista Brasileira de Ginecologia e Obstetrícia*, vol. 33, no. 2, pp. 93–98, 2011.
- [29] L. Simon, C. Trastour, A. Soler et al., “A case of congenital toxoplasmosis-associated miscarriage with maternal infection four months prior to conception,” *Parasitology International*, vol. 79, Article ID 102165, 2020.
- [30] K. Hurt, P. Kodym, D. Stejskal et al., “Toxoplasmosis impact on prematurity and low birth weight,” *PLOS ONE*, vol. 17, no. 1, Article ID e0262593, 2022.
- [31] C. Bieńkowski, M. Aniszewska, M. Kowalczyk et al., “Analysis of preventable risk factors for *Toxoplasma gondii* infection in pregnant women: case-control study,” *Journal of Clinical Medicine*, vol. 11, no. 4, Article ID 1105, 2022.
- [32] B. Hajimohammadi, S. Ahmadian, Z. Firoozki et al., “A meta-analysis of the prevalence of toxoplasmosis in livestock and poultry worldwide,” *EcoHealth*, vol. 19, no. 1, pp. 55–74, 2022.
- [33] C. Audi, S. M. Santiago, M. Andrade et al., “Ultra-processed foods consumption among inmates in a women’s prison in São Paulo,” *Revista Española de Sanidad Penitenciaria*, vol. 20, pp. 87–94, 2018.



- [34] L. G. Felipetto, P. I. Teider-Junior, F. F. V. da Silva et al., “Serosurvey of anti-*Toxoplasma gondii* antibodies in homeless persons of São Paulo City, southeastern Brazil,” *Frontiers in Public Health*, vol. 8, Article ID 732, 2020.
- [35] G. R. da Cunha, M. Pellizzaro, C. M. Martins et al., “Spatial serosurvey of anti-*Toxoplasma gondii* antibodies in individuals with animal hoarding disorder and their dogs in southern Brazil,” *PLOS ONE*, vol. 15, no. 5, Article ID e0233305, 2020.
- [36] M. Guo, R. L. Buchanan, J. P. Dubey et al., “Qualitative assessment for *Toxoplasma gondii* exposure risk associated with meat products in the United States,” *Journal of Food Protection*, vol. 78, no. 12, pp. 2207–2219, 2015.
- [37] L. S. Balbino, J. C. Bernardes, W. A. Ladeia et al., “Epidemiological study of toxoplasmosis outbreaks in Brazil,” *Transboundary and Emerging Diseases*, vol. 69, no. 4, pp. 2021–2028, 2022.
- [38] M. de A. Cruz, L. S. Ullmann, P. Y. Montaña, J. L. Hoffmann, H. Langoni, and A. W. Biondo, “Seroprevalence of *Toxoplasma gondii* infection in cats from Curitiba, Paraná, Brazil,” *Revista Brasileira de Parasitologia Veterinária*, vol. 20, no. 3, pp. 256–258, 2011.
- [39] K. de S. Rocha, M. de S. Lima, T. R. M. Monteiro et al., “Serological prevalence of *Toxoplasma gondii* infection in cats (Belém, Pará, Brazil),” *Revista Brasileira de Parasitologia Veterinária*, vol. 29, no. 2, 2020.
- [40] Even3 Publicações, “One health behind bars: toxocariasis in women inmates and correctional officers, feral cats, and environmental soil contamination,” 2023, <https://www.even3.com.br/anais/parasito-2023-280030/602502-one-health-behind-bars--toxocariasis-in-women-inmates-and-correctional-officers-feral-cats-and-environmental-soil/>.
- [41] Q. Liu, Z.-D. Wang, S.-Y. Huang, and X.-Q. Zhu, “Diagnosis of toxoplasmosis and typing of *Toxoplasma gondii*,” *Parasites & Vectors*, vol. 8, no. 1, Article ID 292, 2015.
- [42] S. Dian, A. R. Ganiem, and S. Ekawardhani, “Cerebral toxoplasmosis in HIV-infected patients: a review,” *Pathogens and Global Health*, vol. 117, no. 1, pp. 14–23, 2023.
- [43] M. Mashaly, N. Nabih, I. M. Fawzy, and A. A. El Henawy, “Tuberculosis/toxoplasmosis co-infection in Egyptian patients: a reciprocal impact,” *Asian Pacific Journal of Tropical Medicine*, vol. 10, no. 3, pp. 315–319, 2017.
- [44] C. Busatto, J. Mespaque, P. Schwarzbald et al., “Tuberculosis in prison inmates in southern Brazil: investigating the epidemiological and operational indicators,” *Revista da Sociedade Brasileira de Medicina Tropical*, vol. 55, 2022.
- [45] V. Rahmanian, K. Rahmanian, A. S. Jahromi, and S. Bokaie, “Seroprevalence of *Toxoplasma gondii* infection: an umbrella review of updated systematic reviews and meta-analyses,” *Journal of Family Medicine and Primary Care*, vol. 9, no. 8, pp. 3848–3855, 2020.
- [46] I. Souza, V. Siqueira, I. Ribeiro et al., “Molecular and serological diagnosis of toxoplasmosis: a systematic review and meta-analysis,” *Revista do Instituto de Medicina Tropical de São Paulo*, vol. 65, 2023.