

Research Article

CO₂ Laser and Topical Fluoride Therapy in the Control of Caries Lesions on Demineralized Primary Enamel

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This study evaluated the effect of CO₂ laser irradiation and topical fluoride therapy in the control of caries progression on primary teeth enamel. 30 fragments (3 × 3 × 2 mm) from primary canines were submitted to an initial cariogenic challenge that consisted of immersion on demineralizing solution for 3 hours and remineralizing solution for 21 hours for 5 days. Fragments were randomly assigned into three groups ($n = 10$): L: CO₂ laser ($\lambda = 10.6 \mu\text{m}$), APF: 1.23% acidulated phosphate fluoride, and C: no treatment (control). CO₂ laser was applied with 0.5 W power and 0.44 J/cm² energy density. Fluoride application was performed with 0.1 g for 1 minute. Cariogenic challenge was conducted for 5 days following protocol previously described. Subsurface Knoop microhardness was measured at 30 μm from the edge. Obtained data were subjected to analysis the variance (ANOVA) and Duncan test with significance of 5%. It was found that the L group showed greater control of deciduous enamel demineralization and were similar to those of APF group, while being statistically different from C group ($P \leq 0.05$) that showed the lowest microhardness values. It was concluded that CO₂ laser can be an additional resource in caries control progression on primary teeth enamel.

1. Introduction

Application of fluoride compounds has been used to control dental caries in primary teeth under different forms [1, 2] and different concentrations [3]. The mechanism of fluoride interferes in the process of mineral loss, promoting inhibition of demineralization, and enhancement dental substrate remineralization [4]. The ability of acidulated phosphate fluoride (APF) to become the primary teeth and more acid-resistant when exposed to cariogenic challenge was evidenced by Castellan et al. 2007 [1]. However, for an effective fluoride action controlling demineralization, it must be constantly in the oral cavity [5].

Higher incidence of dental caries in primary teeth associated with rapid progression of these lesions due to lower mineral content [6] leads to early loss of these teeth [7], factors that encourage more studies to improve existing

preventive treatments and to evaluate innovative techniques such as CO₂ laser irradiation [8, 9].

CO₂ laser irradiation is more appropriate to dental enamel because it produces radiation in the infrared region (9.3, 9.6, 10.3, and 10.6 μm) that coincides closely with some of apatite absorption bands, mainly phosphate and carbonate group absorption [10]. Therefore, higher effectiveness in caries prevention could be achieved with lower occurrence of harmful effects to dental tissues [10]. Using this laser, energy is absorbed in few micrometers of the external enamel surface and converted into heat, causing loss of carbonate from mineral and fusion of hydroxyapatite crystals, reducing the interprismatic spaces [11]. Furthermore, it increases its acid resistance, decreasing the mineral reactivity and promoting caries-preventive effect [9].

The CO₂ laser may control caries progression in permanent [12] and bovine enamel [13] when compared to fluoride

compounds [14]. The efficacy of this laser in caries control on demineralized primary enamel was also previously evaluated by Tagliaferro et al. 2006 [8] and da Silva Tagliaferro et al. 2009 [9]. However, in these studies, laser was applied on sound enamel. There are no studies in the literature evaluating the effect of CO₂ laser in previously demineralized primary enamel, simulating a patient with high cariogenic challenge and high caries risk.

As creation of an acid-resistant surface seems to be a promise in the control of caries lesions, the aim of this study was to evaluate *in vitro* the effect of CO₂ laser irradiation and topical fluoride therapy in control of caries progression on enamel of primary teeth by subsurface microhardness analysis.

2. Material and Methods

2.1. Experimental Design. The factor under investigation was surface treatment at 3 levels: L: CO₂ laser irradiation; APF: 1.23% acidulated phosphate fluoride; C: no treatment (control). The sample consisted of 30 fragments of human primary enamel distributed among three surface treatments ($n = 10$), according to a randomized and complete block design. The quantitative response variable was the subsurface Knoop microhardness (KHN) of the substrate subjected to the chemical demineralization *in vitro*.

2.2. Ethical Aspects. This research was approved by the Ethics in Research Committee of the School of Dentistry of Ribeirão Preto, University of São Paulo (Process number 2010.1.1373.58.9). Freshly extracted sound primary canines were obtained from Human Tooth Bank of the same institution.

2.3. Selection and Preparation of Samples. Primary teeth were hand scaled and cleaned with water/pumice slurry, in rotating bristle brushes at low speed (N270, Dabi Atlante, Ribeirão Preto, SP, Brazil) to remove calculus and surface-adhered debris and stored in 0.1% thymol solution. The absence of cracks, hypomineralization, and hypoplasia was confirmed under an $\times 20$ magnifier (Leica S6 D Stereozoom, Mycosystems Leica AG, Switzerland) and teeth with structural defects were discarded. Afterwards, the selected teeth were sectioned in the cement-enamel junction in precision cutter water-cooled (Isomet 1000, Buehler, Lake Bluff, IL, USA), to separate the root and coronal portions. The buccal surface of each tooth was sectioned to obtain a fragment of enamel measuring $3 \times 3 \times 2$ mm.

The fragments were fixed in acrylic resin blocks using melted wax (Wax Sculpture Fixed Prosthodontics, Aspheric Chemical Industry Ltda., São Caetano do Sul, SP, Brazil) with the subsurfaces facing the external environment. The subsurfaces were then flattened with #1200-grit silicon carbide paper in a water-cooled polishing machine (Politriz, DP-9U2, Struers A/S, Copenhagen, Denmark) (Hermes Abrasives Ltd., VA, USA) and polished with $0.3 \mu\text{m}$ alumina paste (Arotec S/A Ind. Com, SP, Brazil) by felt polisher (ATM, Altenkirchen, Germany) [15]. In order to obtain a sample

of patterned fragments, three readings were performed on the side of the fragments (subsurface) $30 \mu\text{m}$ from the edge and $100 \mu\text{m}$ of each other through a microhardness tester HVM-2000 (Shimadzu Corporation, Kyoto, Japan) with a diamond indenter for Knoop hardness (KHN) under 25 g load for 5 seconds [11]. The three readings were averaged and used as the microhardness value of each fragment. Specimens with microhardness values 20% above or below the mean value of all fragments were discarded [16]. Thirty fragments of primary enamel were selected based on initials Knoop hardness values of its fragments lateral side.

2.4. Initial Cariogenic Challenge. For obtaining initial microscopic lesions of standardized white spot lesion, simulating patients with high caries activity, an artificial caries challenge was performed in all fragments. The specimens were repositioned with the buccal surface facing the external environment in resin blocks and fixed with wax. All surfaces except the buccal were covered with melted wax and stored individually in plastic containers. The initial cariogenic challenge was performed during 5 days according to the protocol proposed by Argenta et al. 2003 [17]. Artificial caries lesions were produced by immersion of the fragments in demineralizing solution (pH 4.6) for 3 hours and remineralizing solution (pH 7.0) for 21 hours at 37°C. After the artificial carious lesions formation, the specimens were kept in humidity for 2 days at 4°C.

2.5. Surface Treatment. According to a complete block design and randomized, the specimens were divided according to treatment in three groups ($n = 10$): L: CO₂ laser, APF: 1.23% acidulated phosphate fluoride, and C: no treatment (control).

The CO₂ laser with $\lambda = 10.6 \mu\text{m}$ (PC 015-D CO₂ Laser System, Shanghai JueHua Laser Tech. Development Co., Ltd., Shanghai, China) was applied in ultrapulsed mode, 0.5 W average power, 0.44 J/cm^2 energy density measured with Power Meter (FieldMax II-TOP, Coherent Inc., Santa Clara, USA), $100 \mu\text{s}$ pulse duration, 0.001 sec interval between pulses, 0.4 mm beam diameter on the substrate surface, where the operator kept the laser tip perpendicularly to the substrate with distance tip/substrate of 4 mm [18] for 20 sec. Parameters used in the present study were able to produce only chemical and structural modification on primary enamel, without causing surface damage or tissue removal. After irradiation, the samples were kept in artificial saliva at 37°C for 24 hours. These were the components of artificial saliva, the reagent (213 mg of CaCl₂·H₂O, 738 mg of KH₂PO₄, 1.114 mg of KCl, 381 mg of NaCl, 12 g of Tris, 2.2 g of gastric mucin, and qsp 1 liter) weighed on an analytical balance (AB204-S/FACT, Mettler Toledo, Columbus, OH, USA) and subjected to agitation, adjusting the pH to 7.0.

0.1 g of 1.23% acidulated phosphate fluoride gel (DFL Industry, Rio de Janeiro, RJ, Brazil, pH 3.6) was weighed on analytical balance (AUW220D, SPLABOR, Presidente Prudente, SP, Brazil) and applied to the dry surface deciduous enamel using microbrush (KG Sorensen, Cotia, SP, Brazil). After 1 minute [19], the specimens were washed with deionized water for 10 seconds, dried with absorbent paper, and after stored in artificial saliva at 37°C for 24 hours.

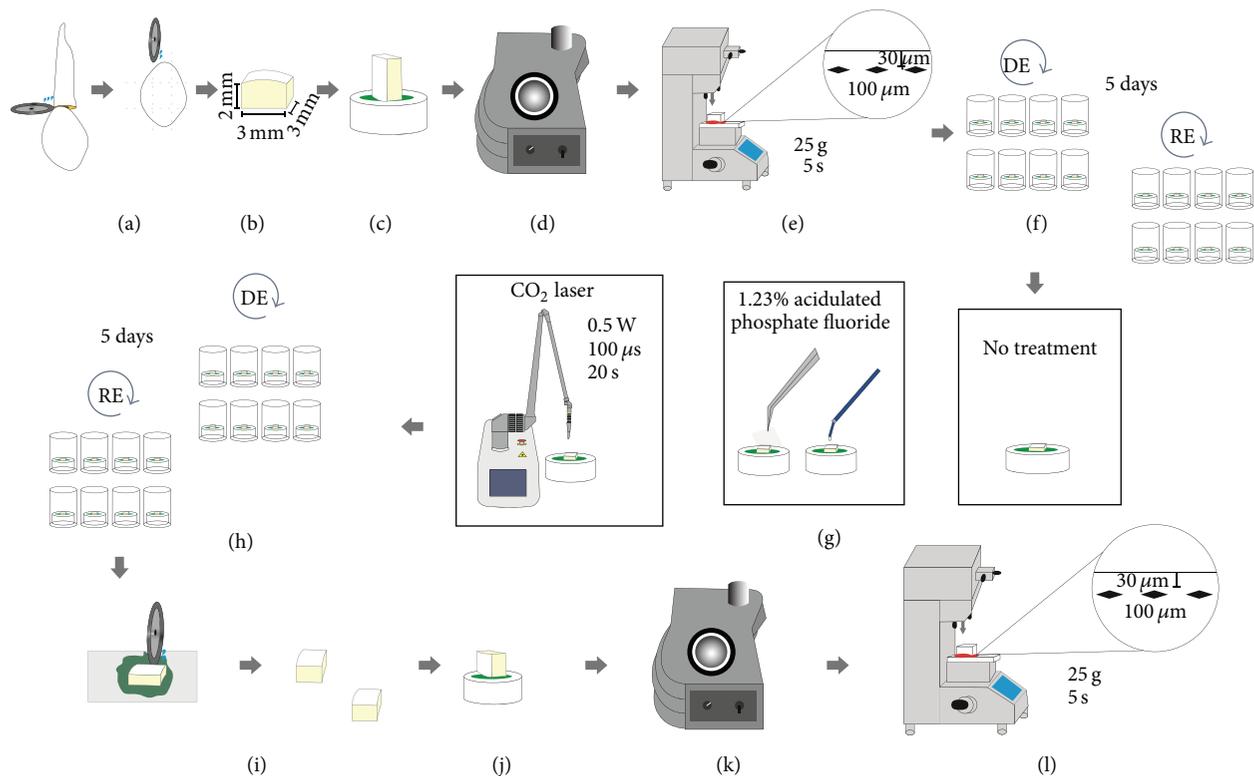


FIGURE 1: Schematic design of the methodology presented. (a) Section of the teeth. (b) Obtaining fragments. (c) Fixation of specimens in resin blocks. (d) Planning and polishing the enamel surface. (e) Selection of specimens. (f) Initial cariogenic challenge. (g) Surface treatments. (h) Cariogenic challenge after surface treatment. (i) Section of the fragments. (j) Fixing the fragments into blocks of acrylic resin. (k) Polishing the enamel surface. (l) Microhardness evaluation.

The control group did not receive any treatment, being kept in artificial saliva at 37°C for 24 hours.

2.6. Cariogenic Challenge Postsuperficial Treatment. The samples were replaced in plastic containers and all surfaces, except for the treated surface, and were covered with melted wax. The same pH cycling that was applied before the laser or the fluoride treatment was repeated 5 times, at a rhythm of one per day, in order to simulate the conditions of cariogenic severe challenge.

2.7. Microhardness Test. After cariogenic challenge period, specimens were sectioned longitudinally and fixed with melted wax and their internal side (sectional) was left exposed and polished in a polishing machine (DP-9U2; Struers S/A, Copenhagen, Denmark). After polishing, specimens were observed under an optical microscope to verify the superficial smoothness and were subjected to ultrasonic cleaning (Dabi Atlante, Ribeirão Preto, SP, Brazil) for two minutes to remove the debris. Then, impressions were made in one of the hemisections, keeping the long axis of the diamond indenter parallel to the external surface of the enamel using a static load of 25 g for 5 sec [1]. Three measurements were performed at the center of the fragment, with 100 μm in distance from one another, 30 μm from the edge, totalizing 3 indentations

per specimen. The readings were averaged and used as the microhardness value of each slab, using a microhardness tester HMV-2000 (Shimadzu Corporation, Kyoto, Japan).

The protocol used in this study is shown in Figure 1.

2.8. Statistical Analysis. The mean values of microhardness of each specimen were analyzed and showed a normal distribution and homogeneity of variance. Thus, analysis of variance (ANOVA) was employed. The Duncan test was used to investigate differences between the mean of surface treatment factor using SPSS 12.0 for Windows (SPSS Inc., Chicago, IL, USA) with a significant level of 5%.

3. Results

The results showed that microhardness of subsurface treatments performed on primary teeth enamel was statistically different ($P \leq 0.05$), as shown in Table 1.

Duncan test showed that surface treatment with CO₂ laser showed the highest microhardness values (KNH) on primary teeth enamel, but it was not statistically different from 1.23% acidulated phosphate fluoride application. However, a statistically significant difference from the control group that presented the lowest microhardness values was found.

TABLE 1: Microhardness values (mean and standard deviation) according to the superficial treatments in different experimental groups ($P = 0.03$).

Treatment	Mean	Standard deviation
CO ₂ laser	324.99 ^a	33.78
1.23% acidulated phosphate fluoride	309.30 ^a	68.42
No treatment (control)	209.86 ^b	67.03

Similar letters indicate statistical similarity.

4. Discussion

Acidulated phosphate fluoride [1, 2] and CO₂ laser radiation [8, 9] have been used to prevent caries in primary teeth in order to interfere the balance of deremineralization. The effects of laser irradiation on the tissue are closely related to wavelength, absorption of laser light by the irradiated tissue, laser power, emission mode, energy density, and frequency [20, 21].

CO₂ laser is responsible for increasing acid resistance on irradiated enamel [1, 8, 9]. On the other hand, fluoride is able to incorporate on dental substrate, preventing the development of carious lesions, inhibiting enamel demineralization, and enhancing remineralization through minerals gain [3].

In this study, surface treatment with CO₂ laser in primary enamel was statistically similar to 1.23% acidulated phosphate fluoride. The probable reason for the increased acid resistance of the primary enamel after CO₂ laser treatment is a consequence of thermal effect [22, 23]. Heating of tooth surface results in structural and chemical alterations in the irradiated dental substrates with melting point of hydroxyapatite [24], regarding calcium [25, 26] and phosphorus loss [27], calcium and phosphorus concentration on the surfaces [28], and alterations in organic matrix [29].

Thermal variations produced by using the CO₂ laser on enamel promote reduction of water and carbonate content [22] which is converted into phosphate followed by protein decomposition at temperatures of 100–650°C, thermal recrystallization (650 and 1.100°C), and destructive phenomena such as melting of hydroxyapatite (>1.100°C) [30]. CO₂ laser may decrease dental permeability and hinder diffusion of acids, due to the surface sealing [31], reducing the demineralization of dental structure [10]. The enamel irradiated using high energy densities revealed nonhydroxyapatite phases, apparently similar to tri- and tetracalcium phosphates [32].

The thermal effects are responsible for changes in the irradiated tooth surfaces while they may differ from the temperature observed at pulp chamber, due to the support structures present around the teeth and the blood flow of the pulp tissue; this heat could be dissipated [33, 34]. The pulp temperature increase, related to the use of high power lasers, is based on the amount of energy applied and therefore, the exposure time is fundamental. High energy densities in short periods of time cause less pulp damage [35], since the thermal relaxation is inversely proportional to the square of the irradiated volume [33].

The low thermal conductivity of the enamel and the rapid decrease in temperature in the lower layer of spent glaze can also contribute to the lack of pulp damage, due to high absorption of this substrate by the appropriate wavelength of 10.6 μm CO₂ laser [36]. The low energy density, used in this study, promoted thermal relaxation time of the deciduous enamel ranging between 1 and 60 μs, and the pulse duration of the laser CO₂ was 100 μs. Esteves-Oliveira et al. 2009 [37] using energy density 0.3 J/cm², similar to this study, were able to decrease enamel caries progression without causing surface and subsurface thermal damage.

CO₂ laser action on primary and permanent enamel can be distinct, due to the differences between these substrates. The mineralization, calcium, and phosphorus percentage is lower in primary teeth than in permanent teeth [6]. The thickness of primary enamel is almost half of the permanent enamel that may have an influence on the demineralization [6] and may provide greater temperature rise when compared to permanent teeth, since thicker structures of enamel and dentin promote smaller temperature change [35, 38–41].

Carbonate content reduction on permanent enamel, promoted by CO₂ laser irradiation [11], results in lower hydroxyapatite solubility. The increasing in crystals size [42], melting [23, 42], and fusion [11] of irradiated enamel have also reduced the enamel dissolution on permanent teeth against acid challenge, although melting of enamel tissues is not a necessity for laser radiation to inhibit caries formation in enamel [24]. In primary teeth, CO₂ laser is also able to reduce carbonate content of enamel [9], which may have led to increased resistance to demineralization in this study.

It has been reported that, after a professional fluoride application, calcium fluoride (CaF₂) is formed on enamel surface and fluoride is released to fluid phase. This effect promotes a consequent reduction of enamel demineralization. Also, a dose-response effect is observed between the concentration of CaF₂, reservoirs on enamel and fluoride released, to “plaque fluid” and the subsequent inhibition of enamel demineralization [5]. The findings of this study have shown that topical application of APF in primary teeth is effective in the demineralization process and caries control [1, 2].

The amount of fluoride formed in the enamel depends on the concentration and the pH of the product applied and how long it remains in contact with the enamel [19]. Tenuta et al. 2008 [5] stated that the constant presence of fluoride in the oral cavity is more important than its concentration for the final enamel absorption. Thus, topical application of more acidic and concentrated fluoride compounds could provide effective protection against demineralization of tooth enamel or caries lesion formation [43], with higher incorporation of fluoride on enamel [19], however, no difference in fluoride uptake by enamel [19] was observed when fluoride was applied by one minute compared to four minutes.

In the present study, as CO₂ laser was applied on previously demineralized primary enamel simulating a patient with high cariogenic challenge and high caries risk, it is difficult to make a direct comparison with these results to previous literary studies. Until now, there is no research that performed previously cariogenic challenge on primary

teeth enamel, targeting the demineralization controlling and not preventing demineralization, having sound as substrate. Besides, the higher the demineralization is, the more difficult caries control becomes.

5. Conclusion

CO₂ laser with $\lambda = 10.6 \mu\text{m}$ was effective in the control of demineralization on previously demineralized primary enamel, presenting some advantages on being a quick, comfortable, and simple method of applying, especially in children, considering the difficulty of using a fluoride. In this way, CO₂ laser can be a resource in the control of caries lesions progression on primary teeth enamel.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References

- [1] C. S. Castellan, A. C. Luiz, L. M. Bezinelli et al., "In vitro evaluation of enamel demineralization after Er:YAG and Nd:YAG laser irradiation on primary teeth," *Photomedicine and Laser Surgery*, vol. 25, no. 2, pp. 85–90, 2007.
- [2] H. Jiang, Z. Bian, B. J. Tai, M. Q. Du, and B. Peng, "The effect of a bi-annual professional application of APF foam on dental caries increment in primary teeth: 24-Month clinical trial," *Journal of Dental Research*, vol. 84, no. 3, pp. 265–268, 2005.
- [3] J. M. Ten Cate, "Contemporary perspective on the use of fluoride products in caries prevention," *British Dental Journal*, vol. 214, no. 4, pp. 161–167, 2013.
- [4] A. Tatevossian, "Fluoride in dental plaque and its effects," *Journal of Dental Research*, vol. 69, pp. 645–652, 1990.
- [5] L. M. A. Tenuta, R. V. Cerezetti, A. A. del bel Cury, C. P. M. Tabchoury, and J. A. Cury, "Fluoride release from CaF₂ and enamel demineralization," *Journal of Dental Research*, vol. 87, no. 11, pp. 1032–1036, 2008.
- [6] M. A. H. de Menezes Oliveira, C. P. Torres, J. M. Gomes-Silva et al., "Microstructure and mineral composition of dental enamel of permanent and deciduous teeth," *Microscopy Research and Technique*, vol. 73, no. 5, pp. 572–577, 2010.
- [7] T. Alsheneifi and C. V. Hughes, "Reasons for dental extractions in children," *Pediatric Dentistry*, vol. 23, no. 2, pp. 109–112, 2001.
- [8] E. P. S. Tagliaferro, L. K. A. Rodrigues, M. Nobre dos Santos, L. E. S. Soares, and A. A. Martin, "Combined effects of carbon dioxide laser and fluoride on demineralized primary enamel: an in vitro study," *Caries Research*, vol. 41, no. 1, pp. 74–76, 2006.
- [9] E. P. da Silva Tagliaferro, L. K. A. Rodrigues, L. E. S. Soares, A. A. Martin, and M. Nobre-Dos-Santos, "Physical and compositional changes on demineralized primary enamel induced by CO₂ laser," *Photomedicine and Laser Surgery*, vol. 27, no. 4, pp. 585–590, 2009.
- [10] J. D. B. Featherstone, N. A. Barrett-Vespone, D. Fried, Z. Kantorowitz, and W. Seka, "CO₂ laser inhibition of artificial caries-like lesion progression in dental enamel," *Journal of Dental Research*, vol. 77, no. 6, pp. 1397–1403, 1998.
- [11] A. L. L. Klein, L. K. A. Rodrigues, C. P. Eduardo, M. N. Dos Santos, and J. A. Cury, "Caries inhibition around composite restorations by pulsed carbon dioxide laser application," *European Journal of Oral Sciences*, vol. 113, no. 3, pp. 239–244, 2005.
- [12] P. Rechmann, D. A. Charland, B. M. T. Rechmann, C. Q. Le, and J. D. B. Featherstone, "In vivo occlusal caries prevention by pulsed CO₂-laser and fluoride varnish treatment - a clinical pilot study," *Lasers in Surgery and Medicine*, vol. 45, no. 5, pp. 302–310, 2013.
- [13] J. Cohen, J. D. B. Featherstone, C. Q. Le, D. Steinberg, and O. Feuerstein, "Effects of CO₂ laser irradiation on tooth enamel coated with biofilm," *Lasers in Surgery and Medicine*, vol. 46, no. 3, pp. 216–223, 2014.
- [14] A. Souza-Gabriel, V. Colucci, C. P. Turssi, M. C. Serra, and S. A. M. Corona, "Microhardness and SEM after CO₂ laser irradiation or fluoride treatment in human and bovine enamel," *Microscopy Research and Technique*, vol. 73, no. 11, pp. 1030–1035, 2010.
- [15] A. T. Hara, C. S. Queiroz, A. F. P. Paes Leme, M. C. Serra, and J. A. Cury, "Caries progression and inhibition in human and bovine root dentine in situ," *Caries Research*, vol. 37, no. 5, pp. 339–344, 2003.
- [16] A. T. Hara, C. P. Turssi, M. Ando et al., "Influence of fluoride-releasing restorative material on root dentine secondary caries in situ," *Caries Research*, vol. 40, no. 5, pp. 435–439, 2006.
- [17] R. M. O. Argenta, C. P. M. Tabchoury, and J. A. Cury, "A modified pH-cycling model to evaluate fluoride effect on enamel demineralization," *Brazilian Oral Research*, vol. 17, no. 3, pp. 241–246, 2003.
- [18] S. A. Tepper, M. Zehnder, G. F. Pajarola, and P. R. Schmidlin, "Increased fluoride uptake and acid resistance by CO₂ laser-irradiation through topically applied fluoride on human enamel in vitro," *Journal of Dentistry*, vol. 32, no. 8, pp. 635–641, 2004.
- [19] R. S. Villena, L. M. A. Tenuta, and J. A. Cury, "Effect of APF gel application time on enamel demineralization and fluoride uptake in situ," *Brazilian Dental Journal*, vol. 20, no. 1, pp. 37–41, 2009.
- [20] L. J. Miserendino, E. J. Neiburger, and R. M. Pick, "Current status of lasers in dentistry," *III Dental Journal*, vol. 56, no. 4, pp. 254–257, 1987.
- [21] Y. Kimura, K. Yonaga, M. Murakoshi, K. Yokoyama, H. Watanabe, and K. Matsumoto, "Effects on periradicular periodontal tissues of root canal irradiation with Er:YAG laser in rats," *Photomedicine and Laser Surgery*, vol. 22, no. 4, pp. 335–341, 2004.
- [22] N. D. Phan, D. Fried, and J. D. B. Featherstone, "Laser-induced transformation of carbonated apatite to fluorapatite in bovine enamel," in *Lasers in Dentistry V*, vol. 3593 of *Proceedings of SPIE*, pp. 233–239, 1999.
- [23] J. D. Featherstone and D. G. Nelson, "Laser effects on dental hard tissues," *Advances in Dental Research*, vol. 1, no. 1, pp. 21–26, 1987.
- [24] C.-Y. S. Hsu, T. H. Jordan, D. N. Dederich, and J. S. Wefel, "Effects of low-energy CO₂ laser irradiation and the organic matrix on inhibition of enamel demineralization," *Journal of Dental Research*, vol. 79, no. 9, pp. 1725–1730, 2000.

- [25] C. Apel, J. Meister, R. S. Ioana, R. Franzen, P. Hering, and N. Gutknecht, "The ablation threshold of Er:YAG and Er:YSGG laser radiation in dental enamel," *Lasers in Medical Science*, vol. 17, no. 4, pp. 246–252, 2002.
- [26] R. C. M. Cecchini, D. M. Zezell, E. de Oliveira, P. M. de Freitas, and C. D. P. Eduardo, "Effect of Er:YAG laser on enamel acid resistance: morphological and atomic spectrometry analysis," *Lasers in Surgery and Medicine*, vol. 37, no. 5, pp. 366–372, 2005.
- [27] L. E. Rodríguez-Vilchis, R. Contreras-Bulnes, O. F. Olea-Mejía, I. Sánchez-Flores, and C. Centeno-Pedraza, "Morphological and structural changes on human dental enamel after Er:YAG laser irradiation: AFM, SEM, and EDS evaluation," *Photomedicine and Laser Surgery*, vol. 29, no. 7, pp. 493–500, 2011.
- [28] M. Hossain, Y. Nakamura, Y. Murakami, Y. Yamada, and K. Matsumoto, "A comparative study on compositional changes and Knoop hardness measurement of the cavity floor prepared by Er:YAG laser irradiation and mechanical bur cavity," *Journal of Clinical Laser Medicine & Surgery*, vol. 21, no. 1, pp. 29–33, 2003.
- [29] Y. Liu and C.-Y. S. Hsu, "Laser-induced compositional changes on enamel: a FT-Raman study," *Journal of Dentistry*, vol. 35, no. 3, pp. 226–230, 2007.
- [30] C. P. Lin, B. S. Lee, S. H. Kok, W. H. Lan, Y. C. Tseng, and F. H. Lin, "Treatment of tooth fracture by medium energy CO₂ laser and DP-bioactive glass paste: thermal behavior and phase transformation of human tooth enamel and dentin after irradiation by CO₂ laser," *The Journal of Materials Science: Materials in Medicine*, vol. 11, no. 6, pp. 373–381, 2000.
- [31] M. Hossain, Y. Nakamura, Y. Kimura, Y. Yamada, M. Ito, and K. Matsumoto, "Caries-preventive effect of Er:YAG laser irradiation with or without water mist," *Journal of Clinical Laser Medicine and Surgery*, vol. 18, no. 2, pp. 61–65, 2000.
- [32] D. Fried, N. Ashouri, T. Breunig, and R. Shori, "Mechanism of water augmentation during IR laser ablation of dental enamel," *Lasers in Surgery and Medicine*, vol. 31, no. 3, pp. 186–193, 2002.
- [33] D. M. Zezell, S. C. M. Cecchini, C. P. Eduardo et al., "Experimental studies of the applications of the holmium laser in dentistry," *Journal of Clinical Laser Medicine and Surgery*, vol. 13, no. 4, pp. 283–289, 1995.
- [34] K. Takamori, "A histopathological and immunohistochemical study of dental pulp and pulpal nerve fibers in rats after the cavity preparation using Er:YAG laser," *Journal of Endodontics*, vol. 26, no. 2, pp. 95–99, 2000.
- [35] I. W. M. Jeffrey, B. Lawrenson, E. M. Saunders, and C. Longbottom, "Dentinal temperature transients caused by exposure to CO₂ laser irradiation and possible pulpal damage," *Journal of Dentistry*, vol. 18, no. 1, pp. 31–36, 1990.
- [36] M. Sabaeian and M. Shahzadeh, "Simulation of temperature and thermally induced stress of human tooth under CO₂ pulsed laser beams using finite element method," *Lasers in Medical Science*, pp. 1–7, 2013.
- [37] M. Esteves-Oliveira, D. M. Zezell, J. Meister et al., "CO₂ laser (10.6 μm) parameters for caries prevention in dental enamel," *Caries Research*, vol. 43, no. 4, pp. 261–268, 2009.
- [38] J. M. Ferreira, J. Palamara, P. P. Phakey, W. A. Rachinger, and H. J. Orams, "Effects of continuous-wave CO₂ laser on the ultrastructure of human dental enamel," *Archives of Oral Biology*, vol. 34, no. 7, pp. 551–562, 1989.
- [39] A. F. Paghdiwala, T. K. Vaidyanathan, and M. F. Paghdiwala, "Evaluation of erbium:YAG laser radiation of hard dental tissues: analysis of temperature changes, depth of cuts and structural effects," *Scanning Microscopy*, vol. 7, no. 3, pp. 989–997, 1993.
- [40] J. A. Von Fraunhofer and D. J. Allen, "Thermal effects associated with the Nd:YAG dental laser," *Angle Orthodontist*, vol. 63, no. 4, pp. 299–303, 1993.
- [41] J. M. White, M. C. Fagan, and H. E. Goodis, "Intrapulpal temperatures during pulsed Nd:YAG laser treatment of dentin, *in vitro*," *Journal of Periodontology*, vol. 65, no. 3, pp. 255–259, 1994.
- [42] D. G. Nelson, J. S. Wefel, W. L. Jongebloed, and J. D. Featherstone, "Morphology, histology and crystallography of human dental enamel treated with pulsed low-energy infrared laser radiation," *Caries Research*, vol. 21, no. 5, pp. 411–426, 1987.
- [43] A. C. B. Delbem and J. A. Cury, "Effect of application time of APF and NaF gels on microhardness and fluoride uptake of *in vitro* enamel caries," *American Journal of Dentistry*, vol. 15, no. 3, pp. 169–172, 2002.



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