

Review Article

Multitargeted Effects of Vitexin and Isovitexin on Diabetes Mellitus and Its Complications

Ibrahim Luru Abdulai,¹ Samuel Kojo Kwofie ,^{1,2} Winfred Seth Gbewonyo ,³
Daniel Boison ,⁴ Joshua Buer Puplampu ,⁴ and Michael Buenor Adinortey ⁴

¹West African Centre for Cell Biology of Infectious Pathogens, College of Basic and Applied Sciences, University of Ghana, P.O. Box LG 54, Legon, Accra, Ghana

²Department of Biomedical Engineering, School of Engineering Sciences, College of Basic and Applied Sciences, University of Ghana, P.O. Box LG77, Legon, Accra, Ghana

³Department of Biochemistry, Cell and Molecular Biology, School of Biological Sciences, University of Ghana, Legon, Accra, Ghana

⁴Department of Biochemistry, School of Biological Sciences, University of Cape Coast, Cape Coast, Ghana

Correspondence should be addressed to Michael Buenor Adinortey; madinortey@ucc.edu.gh

Received 14 October 2020; Accepted 19 March 2021; Published 12 April 2021

Academic Editor: Ahmad Mansour

Copyright © 2021 Ibrahim Luru Abdulai et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Till date, there is no known antidote to cure diabetes mellitus despite the discovery and development of diverse pharmacotherapeutic agents many years ago. Technological advancement in natural product chemistry has led to the isolation of analogs of vitexin and isovitexin found in diverse bioresources. These compounds have been extensively studied to explore their pharmacological relevance in diabetes mellitus. **Aim of the Study.** The present review was to compile results from in vitro and in vivo studies performed with vitexin and isovitexin derivatives relating to diabetes mellitus and its complications. A systematic online literature query was executed to collect all relevant articles published up to March 2020. **Results.** In this piece, we have collected data and presented it in a one-stop document to support the multitargeted mechanistic actions of vitexin and isovitexin in controlling diabetes mellitus and its complications. **Conclusion.** Data collected hint that vitexin and isovitexin work by targeting diverse pathophysiological and metabolic pathways and molecular drug points involved in the clinical manifestations of diabetes mellitus. This is expected to provide a deeper understanding of its actions and also serve as a catapult for clinical trials and application research.

1. Introduction

Diabetes mellitus is a disorder characterized by persistent hyperglycemia, resulting from partial or complete pancreas damage leading to the imbalance in carbohydrate, lipid, and protein homeostases. A hyperglycemic state is the effect of impaired insulin secretion, insulin resistance, or both [1]. Medicinal plants and their isolated compounds have received huge attention due to their reported use in the management of diabetes mellitus (DM). Though several discoveries have been made concerning pharmacotherapies used for managing diabetes mellitus, its management is still a

major challenge for healthcare providers. Several attempts to grapple with the disease have been confronted by humungous financial expenses. In the U.K, for instance, about 10% of the primary care prescribing budget is spent on treating diabetes, with a daily average of £2.2 m spent by the National Health Service (NHS) on prescriptions only [2]. The global economic cost as at 2014 was an estimated US\$612 billion. In urban Ghana, the economic burden of diabetes is high with catastrophic effects on households [3]. Studies conducted by [4] revealed that the overall total cost estimate for diabetes treatment by individuals in Ghana was USD 10,485.11 with a total mean cost of USD 46.81 on monthly basis.

Till date, there is no known antidote to cure diabetes mellitus despite the discovery and development of diverse pharmacotherapeutic agents many years ago. The high cost of these drugs and the unbearable side effect they come along with heighten the demand for natural antidiabetic products which may possess minimal to no side effects. Currently, consideration of two mono-*c*-glycosylflavones, vitexin and isovitexin, as drug candidates for diabetes mellitus and related complication [5], is a growing research interest amongst many scientists all over the world. Several reports are dotted on different publishing platforms, thus necessitating the need for a one-stop document that provides knowledge on past and recent advancement regarding studies on vitexin and its analogues (Figure 1) as drug candidates for diabetes mellitus. This review provides an appraisal of experimental data on vitexin and isovitexin in published works that could serve as the basis for research consideration in other preclinical studies and consequently clinical trials. In this piece, we methodically provide a comprehensive summary of the antidiabetic mechanisms involved in controlling persistent hyperglycemia to serve as evidence and also provide a deeper understanding of its actions. This is expected to act as a reference for further clinical applications and research.

2. Methodology

2.1. Search Strategy. A systematic online literature query was executed to collect all relevant articles published up to March 2020. The systematic search was conducted on selected databases such as ScienceDirect, PubMed, and Google Scholar with the singleword “vitexin” to obtain published studies to-date. Investigators were not contacted, and we did not attempt to identify unpublished data. The review was guided by Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) recommendations [6].

2.2. Articles Selection. The study articles were selected on the basis of an inclusion criteria stated as studies published in only English Language and those articles that have the defined keyword “vitexin” either in the title, abstract, whole text, or even the keywords. Articles on experimental screening with vitexin and isovitexin derivatives were of paramount importance. Review articles, abstracts, editorial/letters, conference, and proceedings were omitted in this systematic review. Also, articles unrelated to diabetes, except antioxidant, anti-inflammation, safety, and toxicological applications of vitexin were excluded. To further filter the articles, search results were screened manually and articles that comprise repeated data or studies duplicated by the search databases and some articles on phytochemical isolation of vitexin and isovitexin derivatives were excluded. The remaining articles represented the main source of information for the write-up. Figure 2 is a flowchart showing the study selection process.

3. Results and Discussion

3.1. Natural Sources and Chemical Structural Properties of Vitexin and Isoviteixin. Vitexin and isovitexin are mono-*C*-glycosylflavones found in diverse natural plant sources. Rapid technological advancement in natural product chemistry has led to the isolation of vitexin and its analogs. The isomer pair and their derivatives are ubiquitously available in natural sources such as insects [7], honey [8], fungus *Dichotomopilus funicola* Y3 [9], and even plant tissues, including *Tetrastigma hemsleyanum* leaves [10], buckwheat sprouts [11], leaves of *Echinodorus scaber* [12], and others.

Vitexin (apigenin-8-*C*- β -glucopyranoside) and isovitexin (apigenin-6-*C*- β -glucopyranoside) analogs are mono-*C*-glycosylflavones with one sugar moiety attached to the flavone skeleton. In part of the *C*-glycoside structure of vitexin and isovitexin, the sugar moiety is connected right to the C6–C3–C6 aglycone skeleton through a C–C bond, which is reported to resist acid hydrolysis [13, 14]. It is confirmed that the sugar ring in a flavone-glucosyl moiety is attached through C–C bonds to the 8 position of the flavone nucleus and is approximately perpendicular to the flavone plane [15]. Vitexin and isovitexin appear as yellow crystalline chemical substances and are soluble in ethanol and dimethyl sulfoxide (DMSO) and mostly give yellow color solution when dissolved in an alkali medium [16]. Several studies have reported the biosynthesis of these class of compounds in the laboratory [17, 18].

3.2. Impact of Vitexin and Isoviteixin on Drug Targets in Diabetes Mellitus and Its Complications. Vitexin and its analogs have been studied as an antihyperglycemic agent. Using rodents, oral administration of vitexin or isovitexin was reported to significantly decrease postprandial blood glucose content in a dose-dependent manner [19, 20]. Furthermore, isovitexin, in both normal and diabetic rats, demonstrated pronounced antihyperglycemic property and also attenuated a rise in blood glucose [21]. Hyperglycemia, caused mainly by insulin resistance and insensitive receptors, disturbs the metabolism of lipids and carbohydrates as well as proteins which can further progress to several micro and macrovascular complications. Examples of these complications include hypertension, dyslipidemia, neuropathy, nephropathy, and sexual impairments. Several targets are reported in literature to be useful points for antidiabetic drug candidates. These include pancreatic β -cell, lipid, and carbohydrate digesting enzymes, cell signaling molecules present in adipose tissue, angiotensin I-converting enzyme (ACE), free radicals, and glutamate transporters.

3.2.1. Pancreatic β -Cell. It is now well noted that pancreatic β -cell functional deficit is a common feature of both type 1 and type 2 diabetes (t1d and t2d, respectively) [22]. In order to examine whether vitexin can protect pancreatic β -cell injury and apoptosis, a study on pancreatic β -cell malfunction was carried out. It was reported

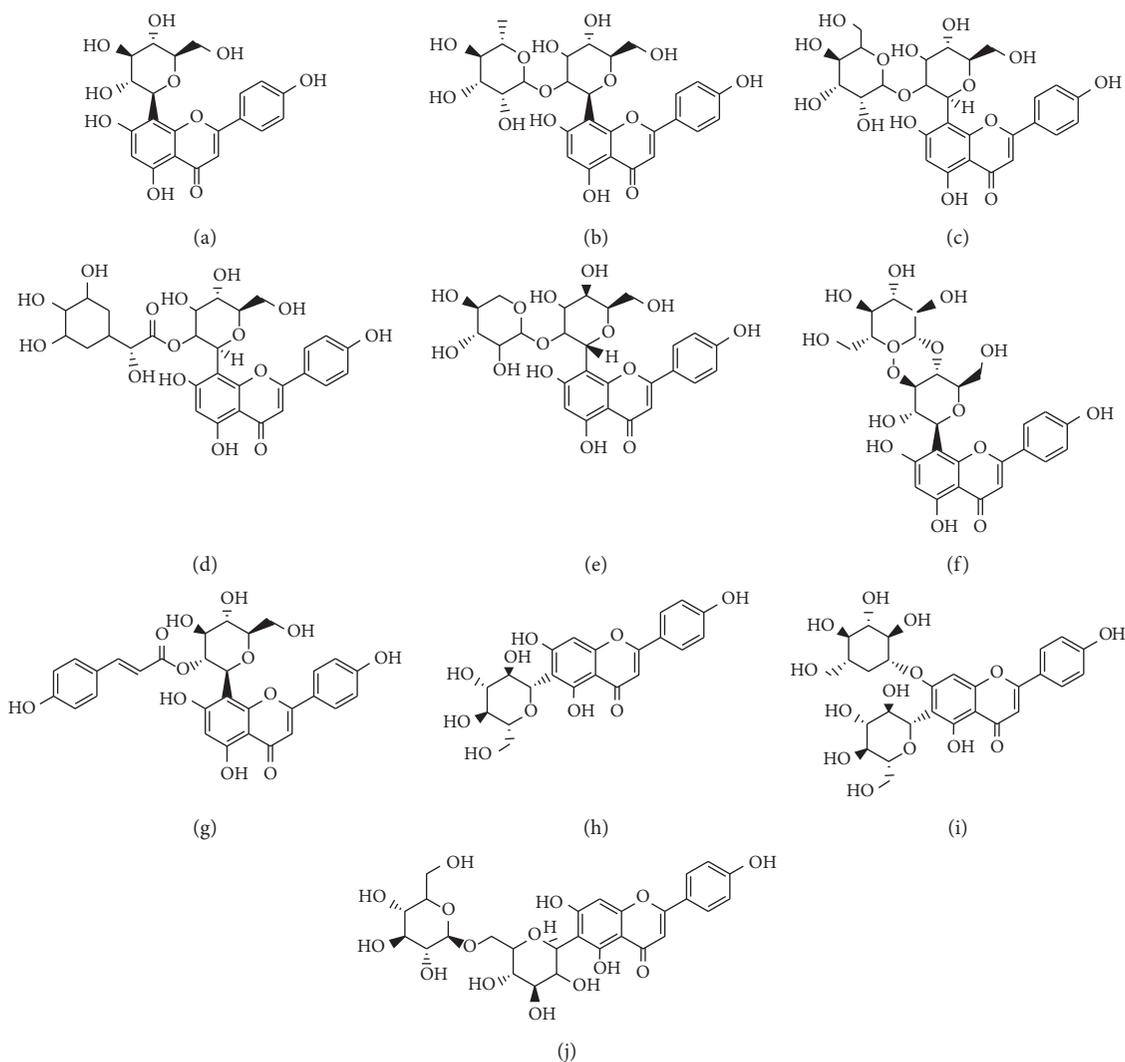


FIGURE 1: Structures of vitexin (apigenin-8-C- β -glucopyranoside) and its analogs. (a) Vitexin (apigenin-8-C- β -glucopyranoside). (b) Vitexin 2''-O-rhamnoside. (c) Vitexin 2''-O-glucoside. (d) 2''-O-galloylvitexin. (e) Vitexin 2-O-xyloside. (f) Vitexin 4''-O-glucoside. (g) Vitexin 2''-O-p-trans-coumarate. (h) Isovitexin (apigenin-6-C- β -glucopyranoside). (i) Isovitexin-7-O-glucoside. (j) Isovitexin 6''-O-glucoside.

that vitexin treatment alleviates lipopolysaccharide- (LPS-) induced apoptosis and damage in islet tissue of rats and INS-1 cells via diminishing the levels of proinflammatory cytokines, TNF- α , and the high mobility group box 1 (HMGB1). Additional experiments proved that vitexin treatment blocked the initiation of P38 MAPK signaling pathways in LPS-mediated INS-1 cells [23]. In NAFLD mice, vitexin also improved insulin signaling through upregulation of insulin receptor substrate-1 (IRS-1) and also its downstream target AKT [24]. In both in vivo and in vitro experiments, it has been demonstrated that vitexin improves pancreatic antioxidant enzyme (glutathione reductase (GR) and superoxide dismutase (SOD) activities, decreases apoptosis and damage induced to islet tissue, stimulates islet cell regeneration, and significantly suppresses fasting blood glucose [23, 25]. SIRT6 has been mentioned as a key regulator of glucose homeostasis and serves as a therapeutic target for obesity and insulin

resistant diabetes mellitus [26]. Singh et al. [27] identified vitexin as an active binder of the SIRT6 protein with binding affinity of 15.27. Later, vitexin acting as a SIRT6 modulator was confirmed using the affinity-based method [28]. Additionally, Rahim et al. [29] reported the antiobesity activities of isovitexins such as isovitexin, isovitexin-7-O-glucoside, and vitexin-2''-O-rhamnoside, which inhibited pancreatic lipase compared to vitexin which showed no activity. Macdonald [30] suggested that quinone reductase may participate in secretion of insulin. Following that, Boutin et al. [31] demonstrated the inhibition of quinone reductase 2 catalytic activity by flavonoids, such as vitexin-2''-O-rhamnoside, vitexin, and isovitexin. Available scientific experimental data have demonstrated that vitexin acts to protect against pancreatic β -cell injury and decreases apoptosis. It exercises this role by impacting positively on endogenous antioxidant status which mop out free radicals responsible for increased cell death.

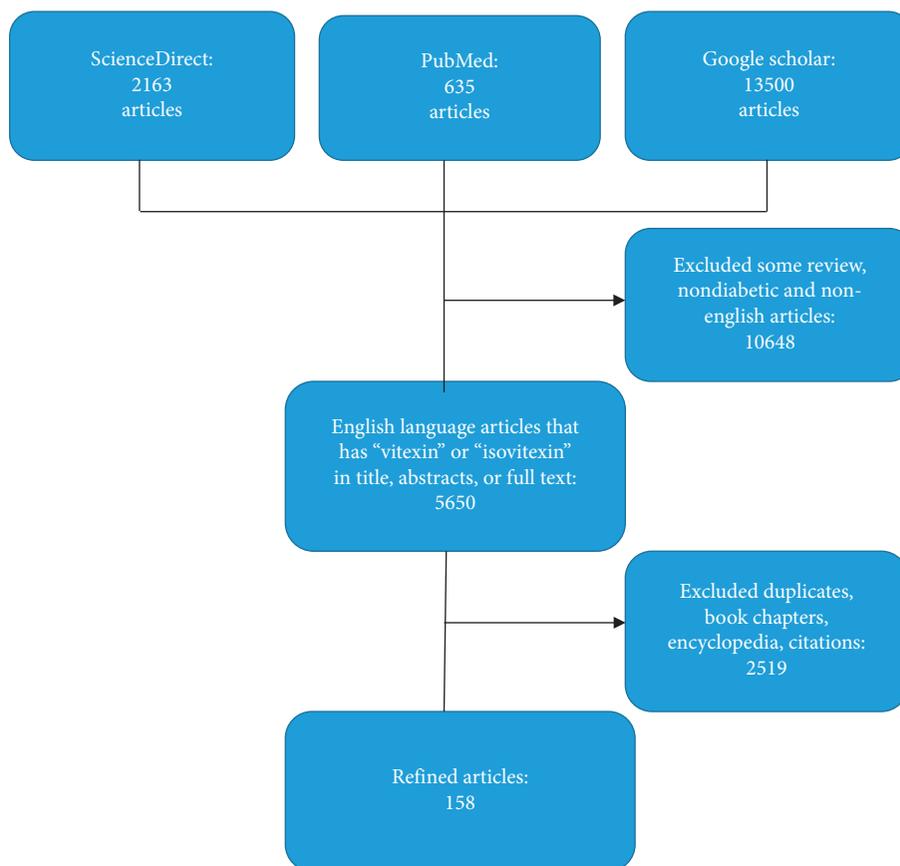


FIGURE 2: Flow chart of the search method and manuscripts selection.

3.2.2. *Alpha Amylase, Glucosidase, Aldose Reductase, Protein Tyrosine Phosphatase 1B, and Lipase.* Enzymes are the main players in glucose metabolism and DM. Treatment of diabetes and its complications is achievable via obstructing key digestive enzymes that participate in lipid and starch digestion as well as metabolism [32]. There are reports on the use of vitexin against drug target enzymes in DM. In an experiment, isovitexin and vitexin revealed potency against α -amylase by reducing its activity, and vitexin binding to α -amylase showed stronger affinity (-7.54 kcal/mol) and stability than that of isovitexin (-5.61 kcal/mol) [33]. Several other studies suggest that vitexin, isovitexin-4'-methyl ether, isovitexin, and 2''-o-xylopyranosyl vitexin may serve as natural hypoglycemic and antidiabetic drugs due to their pronounced activities in α -glucosidase and blood glucose inhibition studies [19, 20, 34–39]. Additionally, acarbose and vitexin presented a synergistic inhibition on α -glucosidase, while vitexin alone uncompetitively repressed α -glucosidase activity and binding of vitexin brought about conformational changes in the enzyme structure [40]. Aldose reductase is one of the principal enzymes in the polyol metabolic pathway that catalyzes the reductive conversion of glucose to sorbitol, which is followed by fructose production catalyzed by the enzyme NADPH-dependent sorbitol dehydrogenase. Increased formation of fructose leads to increased dicarbonyl

moiety which is a key player in AGE formation known to contribute to diabetic complications. It is also reported that excess intracellular sorbitol consequently causes osmotic damage to cells and leads to diabetic cataract complication [41]. Antidiabetic evaluation of C-glycosides revealed that vitexin and isovitexin displayed inhibitory activity against rat lens aldose reductase and human recombinant aldose reductase [42–44]. Protein tyrosine phosphatase 1B (PTP-1B) stimulates negative regulation of insulin and leptin signal transduction; thus, inhibition of this enzyme could serve as a target for diabetes mellitus control. A study by Choi and colleagues in 2013 demonstrated that vitexin and isovitexin possess antidiabetic potentials by its inhibiting effects on PTP-1B enzyme [42].

Generally, lipase enzymes catalyze the hydrolysis of ester bonds of triglycerides, and elevated serum lipase activity is implicated in t2ds [45]. Using the ligand fishing technique, isovitexin was identified as a lipase enzyme inhibitor [46]. Also, in lipase inhibitory assay, vitexin 2''-O-glucoside and vitexin 2''-O-rhamnoside exhibited antilipase activities at $131.48 \mu\text{M}$ for vitexin 2''-O-glucoside and $114.49 \mu\text{M}$ for vitexin 2''-O-rhamnoside [47]. Data enumerated allude to vitexin as a strong drug candidate for consideration in further drug discovery studies and consequently clinical trials for identifications of enzyme inhibitors.

3.2.3. Adipose Tissue. Adipose tissue functions as the primary energy storage site and play additional role as an endocrine organ. Adipose tissue secretes adipokines (adiponectin, resistin, and leptin) which regulate energy homeostasis [48]. However, excessive adiposity is associated with adverse conditions such as obesity and DM [49]. In quest for the antidiabetic agent, vitexin was evaluated for its role in obstructing the pentose phosphate pathway as a novel therapeutic strategy for reducing the prevalence of obesity. Vitexin exhibited marked potency and reduced bodyweight, adipose accumulation and lipid storage in the liver, sugar consumption, and triglyceride production and regulated glycolysis by activation of the ERK 1/2 MAPK signaling pathway [50, 51].

One of the causes of obesity is unregulated genes, such as the CCAAT-enhancer-binding proteins (C/EBP- α), and peroxisome proliferator activated receptor γ (PPAR- γ) which stimulates lipid/glucose metabolism and crucial regulator of adipocyte differentiation. A study showed that vitexin had the highest activity and decreased C/EBP- α and PPAR- γ protein expression levels in 3T3-L1 cells [52]. Another study demonstrated that vitexin suppressed the phosphorylation of p38 and ERK involved in the discharge of inflammatory adipocytokines, although the expression of PPAR- γ was upregulated [53]. The report further mentioned that vitexin treatment increased phosphorylation of AMPK, TNF- α , and aP2 mRNA expression; whereas, the mRNA expression of C/EBP and SREBP1 was unaffected. Recently, Peng et al. [54] mentioned that vitexin significantly reduced lipid levels to normal, as well as reduced adipocyte size mediated by high-fat diet and consequently regulated lipid metabolism via AMPK- α , C/EBP- α , and fatty acid synthase in white adipose tissue and also significantly repressed fat buildup in adipocytes. In addition, vitexin was able to regulate intracellular lipogenesis and adipogenesis through anti-inflammatory mechanisms and the MEK/ERK pathway in the KK-Ay mouse model, and further in vitro studies with 3T3-L1 cells showed that vitexin inhibited inflammation-mediated lipogenesis with highly reduced levels of IL-6 and monocyte chemoattractant protein-1 (MCP-1) after vitexin treatment [55]. The evidences affirm the role of vitexin as an antiobesity drug agent.

3.2.4. Angiotensin I-Converting Enzyme (ACE). Hypertension extensively promotes the risk of both microvascular and macrovascular illnesses, including coronary artery disease and possibly neuropathy. Generally, inhibition of angiotensin I-converting enzyme (ACE) is currently considered to be a useful therapeutic approach in the treatment of high blood pressure. In an in vitro ACE-inhibitory activity experiment, isovitexin showed moderate activity (46%) and was more active than its 8-C-analog, vitexin (21%) [56]. In another study, the hypertensive action of vitexin was accompanied by an enhanced increase in rate and depth of respiration and a slight increase in mean arterial pressure of anaesthetized normotensive rats [57]. It was suggested that vitexin sustained hypotension due to its ganglion-inhibitory and anti-inflammatory, antihistaminic, antibradykinin, and

antiserotonin effects. Vitexin is shown to have haemodynamic effects and caused decrease of mean aortic pressure, arterial and pulmonary capillary pressure, and heart rate [58]. According to Lu et al. [59], vitexin possesses the potential to protect against cardiac hypertrophy via Ca²⁺-induced calcineurin-NFATc3 and CaMKII pathways.

More recently, isovitexin vasodilatory effects were evaluated in an experiment by Tirloni and colleagues who reported that its potency was dependent on endothelium NO release, small conductance Ca²⁺-activated (SK KCa) potassium channels, and Kir6.1 ATP-sensitive K⁺ channels for vascular smooth muscle upregulation [60]. Also, vitexin strongly decreased blood pressure in volunteers and rabbits and increased urine flow, urinary sodium, and potassium excretions; hence, the hypertensive effect of vitexin was suggested to be most likely through its diuretic effects [61]. Ragone et al. [62] reported that the spasmolytic effect of *Aloysia citrodora* on rat duodenum is partly due to its vitexin, since vitexin noncompetitively inhibited the dose-response curves of acetylcholine and Ca²⁺ influx with higher affinity. Vitexin exhibits its spasmolytic properties via K⁺ channel activation and causes a concentration-dependent relaxation of the spontaneous contractions [63]. Experiments performed with purified isovitexin established that it successfully attenuated PDGF-mediated ERK1/2 activation and proliferation of rat aortic smooth muscle cells in culture, thus a novel candidate for atherosclerosis [64]. Furthermore, vitexin with antiarrhythmic potential significantly reduced isoproterenol-stimulated beating incidences at 3 and 10 mg/ml [65]. Mixing isovitexin with orientin produced a slightly significant vascular relaxation at high concentration of 60 mg/mL [66]. Data amassed signify the pharmacological role of vitexin and isovitexin in hypertension management which warrants further clinical research in that regard.

3.2.5. Free Radicals and Glutamate Transporters (NMDAR, mGluR1, and mGlu5). Diabetic neuropathy (DN) is one of the debilitating complications of DM. Several molecular and metabolic pathways responsible for diabetic neuropathy progression have been proposed for therapeutic investigations. The interruption in these pathways by vitexin and its derivatives redefine the medicinal application of these compounds in managing diabetic neuropathy. Vitexin and its derivatives mitigate DN via the following mechanisms: (1) antioxidant, (2) anti-inflammatory, and (3) gene or protein inhibitory activities. In PC12 cells, vitexin exhibited its neuroprotection via attenuation of cell apoptosis, neuroinflammation, oxidative stress, cytosolic calcium, and enhanced β -secretase 1 content [67, 68]. In addition, vitexin selectively modulated the expression of genes involved in the antioxidant pathway, glutamate transporters (NMDAR, mGluR1, and mGlu5), maintained cholesterol homeostasis and reduced ER stress to offer neuroprotection [69, 70]. Helms found that vitexin interacted with E2-2K, implicated in neurodegenerative diseases, at its E1 interacting site [71]. Also, according to Yang et al. [72], vitexin attenuates neuronal cell death through modification of Bcl-2 and Bax expressional balance, cleavage of poly (ADP-ribose)

polymerase and caspase 3, and interferes with the calcium influx pathway in cultured cortical neurons. Further investigations showed that vitexin acts through PI3K/Akt, HIF-1 α , VEGF, and p38 MAPK signaling paths, improves phosphorylation of PI3K and Akt, and concurrently blocks Bax/Bcl-2 ratio and caspase-3 activity in mice and SH-SY5Y cells [72]. Also, the use of vitexin prevented bradykinesia [73, 74]. Additionally, vitexin and isovitexin caused activation of Nrf2, enhanced the activity of HO-1, inhibited neutrophils activation and elevation of inflammatory cytokine (MAPK and NF- κ B) and ROS, and suppressed activated nucleotide-binding domain and NLRP3 inflammasome [75, 76]. In neuro-2a cells, vitexin blocks oxidative stress, facilitated damage by boosting the expression of the Nrf-2/HO-1 pathway, inducing glutamate transporter expression, downregulation of calpain and NMDAR, inhibition of glutamate-stimulated Bax expression, and also effectively docked to NMDAR and GSK-3 β [77].

In Alzheimer's disease (AD), vitexin and its analogs have been demonstrably shown to possess novel medicinal benefits. Cholinesterase (ChE) breakdown acetylcholine (ACh) in the central nervous system, and the reduced levels of ACh in the brain are implicated in AD. Substantial data have shown that vitexin-4'-O-glucoside, vitexin, isovitexin, and isovitexin-4'-O-glucoside (isosaponarin) suppressed the catalytic role of acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) [42, 78–80]. Further docking analysis explained that vitexin interacts with AChE through hydrogen bonds [78]. In addition, vitexin favorably attenuated the function of β -site amyloid precursor (APP) cleaving enzyme 1 (neuropathological marker of AD) and generation of ROS [42, 78, 79]. Malar et al. [77] illustrated that vitexin downregulates glutamate transporters and protect neuro-2a cells from glutamate toxicity involved in AD. Furthermore, vitexin showed protection against memory impairment by enhancing memory retrieval and also reversed the shorter step-through latencies in AD [81]. More so, the anticonvulsant effect of vitexin was noticed in its possible ability to modulate minimal clonic seizures in pentylenetetrazol-induced rats and generalized tonic clonic seizures by delaying seizure appearance time [81, 82]. It is clear that there is some scientific proof to support the antineuropathy properties of vitexin and isovitexin analogs.

3.2.6. Hypothalamus-Pituitary-Gonadal Axis and Dopamine D-2 Receptors. Recently, sexual dysfunction and fertility impairment cases have attracted attention as one of the major complications of diabetes. The hypothalamus-pituitary-gonadal axis have been suggested as the major pathway linking sexual dysfunction and fertility impairment to diabetes in male animals and humans [83]. The work by Li et al. [84] revealed that vitexin rectifies sexual dysfunction and fertility impairments in male diabetic mice possibly via modulating the hypothalamus-pituitary-gonadal axis. According to this report, vitexin significantly enhanced the sexual behavior, fertility levels, testicular pathological structure damage, and reproductive organ weight and

restored sperm quality compared with the diabetic control group. Further downstream analysis indicated that vitexin increased secretion of serum testosterone, follicular-stimulating hormone, and luteinizing hormone contents, whereas the gonadotropin-releasing hormone level was decreased. Also, in a study involving the treatment of premenstrual symptoms, vitexin and isovitexin were screened against dopamine D-2 receptors that regulate prolactin synthesis, and it was observed that vitexin and isovitexin inhibited dopamine D-2 receptor at higher concentration [85]. These experimental results provide some indications as to the impact of these flavonoids on endocrine disorders which result in persistent hyperglycemia and as such, further investigations are on its use as a possible drug agent in sexual dysfunction and fertility impairment management. The multifaceted medicinal effects of these flavonoids deserve more investigative studies to further understand its pharmacological role in sexual dysfunction and fertility impairment.

3.2.7. AGEs, Kidney Injury Molecule-1, and Neutrophil Gelatinase-Associated Lipocalin. Diabetic nephropathy (DN) is a leading cause of end-stage renal failure and contributes to most chronic complications of DM. Blocking the formation of advanced glycation endproducts (AGE) remains one of the therapeutic metabolic strategies in preventing DN and vascular diseases. Fortunately, it has been reported that isovitexin showed excellent inhibitory activities towards the formation of AGEs [43]. Additionally, other published studies have shown that vitexin and isovitexin exhibit their antidiabetic effects by inhibiting the formation of AGEs in vitro [42, 86].

Furthermore, the protective effect of vitexin on LPS-induced acute kidney injury and renal failure was recently observed and reported by Wang and colleagues [87]. The mechanistic studies illustrated that vitexin strongly reduced contents of serum creatinine and blood urea nitrogen, kidney injury molecule-1, and neutrophil gelatinase-associated lipocalin. Also, it upregulated AMPK/FOXO3a signaling via stimulating p-AMPK to AMPK ratio and protein content of FOXO3a in the nucleus while deactivating FOXO3a expression in cytoplasm. In DN, changes in MMPs expression add to extracellular matrix accumulation and glomerular hypertrophy which eventually cause proteinuria and renal insufficiency [88]. Testing for MMPs inhibitors, isovitexin and vitexin displayed the most potent inhibitory activity against MMP-1 expression at 100lg/ml [89]. Moreover, vitexin and isovitexin were evaluated on the subtypes of MMPs (MMP-2, MMP-8, and MMP-9) using experimental and computational approaches. The two compounds showed pronounced inhibition activity towards the MMPs tested, and docking analysis revealed that vitexin and isovitexin bind to the active sites of the three tested MMPs [33]. In the I/R injury model, vitexin also protected BBB integrity, enhanced tight junction proteins synthesis, as well as inhibited MMP involved in regulating junction adhesion proteins [90]. Data portray the significance of vitexin and isovitexin as drug candidates for management of kidney damage.

3.2.8. *IL-1 β , NF- κ B, and JUN N-Terminal Kinase (JNK) Pathways as Inflammation Targets.* Diabetes mellitus triggers inflammation and oxidative stress which causes injury to body organs, resulting in numerous diabetic complications [91]. Existing evidence propose a possible role for inflammation in the pathogenesis of diabetes mellitus. The suggested mechanisms linking inflammation to diabetes comprise activation of IL-1 β , NF- κ B, and JUN N-terminal kinase (JNK) pathways and recruitment of immune cells resulting in insulin resistance and dysfunction insulin secretion [92]. Therefore, targeting and blocking any of these inflammatory mechanisms represents a novel therapeutic approach for DM. Immune response modulatory property of vitexin, vitexin-2-O-rhamnoside, and vitexin-2-O-xyloside has been illustrated [93]. More so, topical anti-inflammatory activity of vitexin and isovitexin was previously stated [94]. The anti-inflammatory potential of vitexin measured in albumen denaturation inhibition assay was 54.2% compared to aspirin (55.6%), while vitexin inhibited proteinase by 57.8%, and the anti-inflammatory effect was obvious in vivo [95]. In animal studies, vitexin mitigated LPS-induced recruitment of neutrophils, maintained cytokine content, and activated nucleotide-binding domain and leucine-rich repeat PYD-containing protein 3 (NLRP3) inflammasome [75]. Better still, using a bioassay, the flavonoid 6'''-(3-hydroxy-3-methylglutaroyl)-2''-O- β -D-galactopyranosyl vitexin inhibited complement activation on the classic pathway in vitro [96].

The NF- κ B molecule is a transcription factor that controls the transcription of DNA for the perpetuation of the inflammatory immune response and acts as a switch for turning inflammation on and off in the body [92]. The NF- κ B proteins are localized in the cytoplasm. Inhibitors interact with these proteins and hijack them from directing DNA to express inflammatory proteins [92]. As an anti-inflammatory agent, administration of vitexin drastically attenuates the I/R-induced increased myocardia NF- κ Bp65 proteins content compared with control mice [86, 97].

It was previously reported that mast cells are activated by physical contact with activated T cells [98]. Following activation, mast cells discharge a group of inflammation mediators, including IL-1, IL-6, IL-8, IL-10, and TNF- α [98], and excessive induction is implicated in numerous metabolic disorders. Targeting inflammatory factors using IL-1 antagonists in patients with T2D, during clinical trials, showed promising results and support the role of inflammation in diabetes [92]. In both Jurkat T lymphocytes and RBL-2H3 mast cells, it was shown that vitexin inhibited the calcium release-activated calcium channel currents (ICRAC) and may inhibit mast cell degranulation and T cell recruitment [99]. Independently, vitexin and 2''-O- α -L-rhamnopyranosyl vitexin markedly decreased the migration of leukocyte in vivo and reduced particularly the peritoneal lavage neutrophils expression [5, 100]. In both in vivo and in vitro models, vitexin and 2''-O- α -L-rhamnopyranosyl vitexin treatment suppressed the secretion of activator-inflammatory cytokines TNF, IL-6, and IL-1 β proteins' content compared with controls [5, 55, 97, 100–103]. Yang et al. [104] suggested that the vitexin inhibitory effect on IL-1 β -

induced inflammatory responses and may be attributed partly to the inhibition of the HIF-1 α pathway. In the inflammation model, vitexin exhibited pronounced activity in blocking the expression of TNF- α and the subsequent neutrophil activation [74, 105, 106]. Hashiguchi et al., in 2017 [107], reported that treatment of macrophage cells with vitexin alone did not change the expression level of IL-6; however, a combination of mung bean coat extracts with additional vitexin suppressed IL-6 expression in cells. Interleukin-10 (IL-10) is a potent anti-inflammatory cytokine with properties that suppress the overprovocation of host immune response, averting injury to host tissues and normalization of tissue homeostasis [108]. IL-10 dysregulation, especially during potent immune response, is related to increased immunopathology as well as high risk for acquisition of autoimmune syndromes [108]. Nevertheless, vitexin possessed the property to directly stimulate the synthesis of IL-10 to attenuate the mounted inflammation process either in inflammatory models or in vitro [5, 101].

The major pathways such as extracellular-regulated protein kinases 1/2 (ERK1/2), P38, and c-Jun N-terminal kinase (JNK) signals in the MAPK signaling pathway are associated with the pathogenesis of hyperinsulinemia in DM. Dysregulation of ERK1/2, P38, and JNK became a logic treatment step in chronic inflammation [109]. Vitexin has the property to reduce the expression of p-p38, p-ERK1/2, and p-JNK in macrophages [5]. Vitexin decreased progression of neutrophils to inflammation by targeted downmodulating proinflammatory regulators achieved through suppression of p38, ERK1/2, and JNK classical pathways [110].

The medicinal target, 5-lipoxygenase, catalyzes the biosynthesis of inflammatory mediators (leukotrienes), in chronic inflammatory diseases [111]. In an invitro study, 5-lipoxygenase enzyme was shown to be moderately inhibited by vitexin with IC₅₀ = 5.1 μ M, referenced to indomethacin (IC₅₀ = 0.98 M) [112]. Vitexin exerted a powerful reduction of adenosine deaminase and myeloperox expression as they relate to the release of cytokines (TNF- α and IL-17) in mice [113]. Myeloperoxidase (MPO) is noticed to be a mediator of inflammation and tissue injury, resulting in autoimmune syndromes. Multiple studies have observed the reduction of myeloperoxidase function using saponarin, vitexin, and isovitexin in vitro [106, 114].

Cyclooxygenases (COX-1 and COX-2) and prostaglandin E2 (PGE2) are inflammatory mediators. The COX enzymes catalyse the breakdown of arachidonic acid into prostaglandins, and attenuation of COX with NSAIDs leads to declined PGE2 release and decreased inflammation. The anti-inflammatory effect of vitexin via COX-1, COX-2, and PGE2 modulation has been studied. In macrophages, vitexin and isovitexin suppressed COX-1 and COX-2 mRNAs expression [103, 115–117]. A recent study illustrated that vitexin displayed moderate inhibition more towards COX-2 compared to COX-1 enzyme, and the flavonoids interacted with hydrogen bonding more at the cyclooxygenase catalytic sites just like celecoxib, the known inhibitor [118]. An in silico study disclosed that vitexin had the highest fitness score against the COX-2 (63.49) and for enzyme COX-1

(60.43) and showed tactical hydrogen bonding against COX-1 and COX-2 [119]. Furthermore, vitexin and isovitexin have the property to suppress PGE₂ in macrophages [5, 117]. Available evidence affirms the role of vitexin and isovitexin in inflammation in the pathogenesis of diabetes mellitus. Thus, they could be considered in clinical trials research as a prophylaxis drug agent in diabetes mellitus.

3.2.9. Reactive Species. The wide-ranging involvement of reactive species or free radicals and accompanied declined antioxidant capacity in most diabetic complications is well established. Reactive species are derivatives of normal cell function, and overproduction accelerates hyperglycemia-mediated oxidative tissue damage implicated in human diseases including metabolic and cardiovascular disorders [120]. Data from earlier studies showed that antioxidants diminish the rate of developing diabetic-related complication and recover insulin sensitivity [120]. The mechanistic approach to antioxidant activity by vitexin and its derivatives appears in three ways. One is by limiting the expression and/or the activities of enzymes that directly catalyse the production of oxidants, second, upregulate the expression and/or activity of enzymes involved in the generation of cellular antioxidants, and finally, acts as ROS scavengers themselves.

During excessive ROS discharge, the expression levels of lactate dehydrogenase (LDH) and creatine kinase (CK) are upregulated. Treatment with vitexin alleviates oxidative stress via decreasing the release of ROS and downregulates the synthesis of LDH and CK enzymes [73, 121–124]. In the course of lipid peroxidation, malondialdehyde (MDA) production increases due to increased oxidative stress; nonetheless, vitexin and vitexin-4''-O-glucoside have the properties to suppress lipid peroxidation and significantly reverse the enhanced production of MDA [73, 121–123, 125]. In neuro-2a cells, elevated production of protein carbonyl content (PCC) and MDA and endoplasmic reticulum stress gene (Gadd153) induce ROS-mediated toxicity. However, pretreatment with vitexin restored PCC and MDA levels, significantly downregulated Gadd153 (0.93 fold) leading to the reduction of free radicals, suppressed ROS-mediated lipid peroxidation and protein oxidation, and protects membrane potential [70]. Several studies have confirmed the lipid peroxidation properties of vitexin, vitexin-4''-O-glucoside, and isovitexin in vitro [126–129]. NO synthase (NOS) catalyse the breakdown of L-arginine to form nitric oxide (NO) which reacts with reactive superoxide free radical to form toxic peroxy nitrite which has tissue damaging properties. Culturing lipopolysaccharide- (LPS-) triggered macrophages in the presence of vitexin attenuates stimulated NO release and synthesis of nitric oxide synthase (iNOS) in the macrophages [130]. Intracellular NO and ONOO⁻ production are modulated by eNOS and iNOS activities, and vitexin demonstrates its antioxidant effect via powerful reduction of LDH and iNOS function as well as escalating eNOS phosphorylation and activity in cells [90]. During in silico docking experiment to NOS and xanthine oxidase (superoxide-generating enzyme), isovitexin presented great potential in the docking and binding to the

enzymes targeted; whereas, vitexin docked and bound with only NOS [131]. Furthermore, myeloperoxidase-derived oxidants serve as a vital contributor to tissue damage, and reports showed the reduction of myeloperoxidase and horseradish peroxidase catalyzed oxidation by saponarin, vitexin, and isovitexin [106, 114]. Vitexin and isovitexin have potent antioxidant activities with effective inhibition of advanced glycation end products [86]. Also, xanthine oxidase, 15-lipoxygenase, and nitric oxide synthase enzymes induce oxidative modification by generating ROS during their catalytic activities, leading to uncontrolled tissue damage. Isovitexin has been reported to inhibit these enzymes activities [132–134].

In the body, protection against oxidative stress is achieved by antioxidants generally generated enzymatically including catalase (CAT), SOD, glutathione peroxidase (GPX), glutathione reductase (GR), and glutathione oxidase (GPO) or via nonenzymatic sources glutathione (GSH), vitamin C, vitamin E, selenium, Zn, carotene, and beta-carotene. However, under oxidative stressed conditions, these enzymes are significantly repressed, exposing tissues and organs to injury. One of the antioxidant mechanisms of vitexin is to increase the activities of these enzymes, thereby discharging antioxidant that efficiently quench reactive species and limit ROS content in the system. In vivo, it was observed that pretreatment with vitexin and vitexin-4''-O-glucoside significantly promoted CAT and SOD activities [73, 122, 125, 135–137]. Also, upregulation of FOXO3a stimulates the production of antioxidant enzymes, and vitexin is known to activate the expression of p-FOXO3a in the rat model [124], thus demonstrating a medicinal effect against free radicals-induced body injury. The protein NADPH acts as an indicator of oxidative status and has a positive effect on the cellular antioxidant potentials; meanwhile, vitexin has the property to increase NADPH protein expression levels and exerts lipid peroxidation impact in I/R injury models [121]. Vitexin nullifies oxidative stress via decreasing the discharge of ROS whilst promoting the activity of glutathione peroxidase [73]. Nuclear factor erythroid-2-related factor 2 (Nrf2) is believed to control expression of antioxidant enzymes, e.g., heme oxygenase-(HO-)1 which protects organs against oxidative damage [138]. In vivo and in vitro, vitexin and isovitexin enhanced the expression of Nrf2 and HO-1 activity and attenuated oxidative stress injury [75, 76]. Work performed by Malar et al. illustrated that vitexin mitigated induce ROS-mediated toxicity by significantly activating endoplasmic reticulum stress gene, Grp78 (0.84 fold) [70].

In the third antioxidant mechanism, vitexin and its derivatives exhibit their antioxidant capacities by rapidly quenching the attacking ROS or NOS and thereby alleviating their destructive power. Several studies on antioxidant potentials of these glycosides in different antiradical assays have been reported. Growing evidence support that vitexin scavenge NO in vivo or in vitro, thereby eliminating its destructive pathways [5, 74, 102, 106, 139]. In the DPPH method, absorbance of DPPH radical decreases as the radical is scavenged by antioxidants through donation of hydrogen to form the stable DPPH-H. The more rapid the decrease in

absorbance, the more potent the antioxidant activity of the compound in terms of hydrogen donating ability [140]. Antioxidant activity of vitexin and its analogs were investigated using the DPPH assay and isovitexin-6''-O- α -L-glucoside, vitexin, vitexin-2''-O-rhamnoside, isovitexin, 2''-O-rhamnosyl vitexin, vitexin-2-O-rhamnoside, vitexin 2''-O-xyloside, isovitexin-7-O-glucoside, isovitexin 6''-O-E-p-coumarate, and vitexin 2''-O-glucoside exhibited significant DPPH• scavenging activities [128, 132, 141–147]. Vitexin and isovitexin DPPH radical scavenging activity is suggested to be attributed to the phenolic hydroxyl group in the 4' position on the B-ring [33, 38, 143, 146, 148–157]. Moreover, isovitexin-6''-O- α -L-glucoside, vitexin, vitexin-2''-O-rhamnoside, and isovitexin demonstrated moderate to high affinity to generated free ABTS•+ in the ABTS assay [44, 141–145, 156–162]. In superoxide radical-scavenging assay, isovitexin and vitexin showed a protective effect against the oxidative-induced reactions by scavenging generated free superoxide anion radicals [37, 133, 147, 150, 156, 163, 164]. Vitexin and vitexin-2''-O-p-trans-coumarate have anti-ROS protective influence against H₂O₂-mediated oxidative injury, sensitive in neutralizing free oxygen radical, and also offers protection to the antioxidant enzymes [133, 161, 165, 166]. The 7-OH and 4'-OH groups were suggested to be important in the activity [167]. Furthermore, in β -carotene bleaching assay, isovitexin and vitexin showed moderate to strong antioxidant effects in inhibition of β -carotene bleaching [146, 151, 168]. Isovitexin and vitexin have also demonstrated ferric reducing capacities [128, 133, 151, 169] and Fe II chelating abilities [160, 161]. Vitexin 2''-O- β -D-xylosyl vitexin and 2''-O-galloyl vitexin exhibited the strongest antioxidant activity in ORAC assay [170, 171].

Nevertheless, other studies claimed that vitexin and isovitexin did show weak to no significant free DPPH radical scavenging abilities [33, 84, 127, 141, 142, 153, 158, 159, 169, 172–181]. Also, poor antioxidant activity of vitexin in scavenging ABTS•+ have been observed [153, 161]. In addition, vitexin and isovitexin were weak scavengers of ClO⁻ in copper-phenanthroline assay [161]. Vitexin-2-O-rhamnoside showed low power in scavenging freely generated radicals in tyrosine nitration inhibition assay [142]. Whereas, studies by von Gadow et al. [182] and Leong et al. [152] observed isovitexin and vitexin to be weak antioxidants in the β -carotene bleaching test and further explained that the observed is due to isovitexin and vitexin not having the 3',4'-dihydroxy configuration or m-5,7-dihydroxylation of the A-ring, which provides good electron delocalization and stabilization of phenoxy radicals and the compounds with only one hydroxyl group on the B-ring.

The chemical structures of vitexin and its analogs greatly influence the antiradical properties of these compounds. It is proposed that the antioxidant capacity of these compounds depend on the extent of hydroxylation, position of glycosylation, and the kind of glycosylation [94, 156, 183, 184]. Additionally, glycosylation of the A-ring and/or loss of the 3-OH distinctly affects the antioxidant activity of vitexin, while mono OH substitution at

4' of vitexin has stronger antioxidant activity [164, 171, 185, 186]. Moreover, glycosylation at C-6 of the A-ring makes greater uniform distribution of spin density as well as enhances stability of free radicals resulting in superior antioxidant capability [187]. In contrast, Lespade and Bercion [188] reported that glycosylation causes charge changes on the oxygen atoms of the hydroxyl groups and generally leads to a decrease in the antioxidant ability of a compound; thus, glucose substitution at position 7 leads to a decrease in negative charge on the oxygen atom at position 3, which is not exciting for potent antioxidants even if it leads to a slightly better antioxidant potency. The most potent antioxidants have a large negative charge on the oxygen atom which gives out its hydrogen [188]. Also, Ma et al. [189] proposed that introducing an acyl group into isovitexin significantly improved its lipophilicity but reduced its antiradical activity, particularly, acylation at the isovitexin primary hydroxyl group of glucose moiety. This lengthened exposition on the antioxidant properties of vitexin and isovitexin clearly affirms their role as antioxidants. Thus, they could be considered for further research as alternatives to some synthetic antioxidants reported to have many adverse effects.

3.2.10. Cell Signaling Molecules Involved in Apoptosis, Autophagy, Mitochondrial, and Platelet Aggregation. In diabetic wounds, cell proliferation, differentiation, and angiogenesis are profoundly disturbed which impairs wound healing [190]. The wound healing property of vitexin was recently published [191]. The wound healing effect of chitosan-based gel formulation containing vitexin was assessed, and the gel potently accelerated wound healing both in vivo and in vitro. Histological examinations confirmed that vitexin formulation triggered skin regeneration and that the wound healing assay showed vitexin-containing gel to strongly activate cell proliferation.

Diabetes occurs more often in individuals with chronic obstructive pulmonary disease (COPD) than in the general population; for that matter, diabetes has been proposed as the risk factor of COPD [192]. Managing COPD is recognized as a logic step in reducing some of the morbidities of diabetes. Flavone C-glycosides, vitexin, and isovitexin exhibited potent inhibitory activities against IL-8 production and matrix metalloproteinase-1 (MMP-1) expression, an inflammatory marker for COPD [89]. In both assays at 100 μ g/ml, the two compounds demonstrated their potential therapeutic role in the treatment of COPD by preventing the pathological disruption of the lung extracellular matrix observed in this disease.

DM individuals pose approximately four times higher risk for stroke, and cardiometabolic risk factors such as hypertension, obesity, and dyslipidemia [193]. A study reported that vitexin could suppress the autophagy dysfunction by attenuating middle cerebral artery occlusion-induced cerebral ischemic stroke via the mechanistic target of rapamycin, the mTOR/Ulk1 pathway, thus alleviating oxidative injury and reducing the production of proinflammatory mediators [101].

Diabetes triggers inflammation and oxidative stress which cause injury to body organs, resulting in numerous diabetic complications [91]. The pharmaceutical effect of vitexin on DM-induced cardiovascular diseases is accomplished through protein kinases, the key cellular modulators of glucose homeostasis and AMPK [137], PI3K/Akt/mTOR [123], and MAPKs [194] signaling pathways. AMP-activated protein kinase (AMPK) is a glucose-sensing enzyme that is triggered when cellular glucose levels are low, and it signals to stimulate glucose metabolism in muscles, fatty acid oxidation in adipose tissues, and decreases hepatic energy accumulation [195]. AMPK activation prompts insulin-sensitizing effects, making it an ideal therapeutic target for t2ds. Multiple evidences suggest that AMPK is dysregulated in DM; however, vitexin directly induced AMPK activity at low μM concentrations, which suppressed intracellular and plasma cholesterol contents in HepG2 cells and mice [135]. In the AMPK signaling pathway, vitexin prevented cell injury by prompting autophagy, enhanced cell viability, and alleviated apoptosis in the cells [137]. At the cellular level, vitexin enhanced autophagy via stimulating the synthesis of p-AMPK while reducing p-mTOR expression, and the protective effects of vitexin were blocked by applying autophagy inhibitor to cells. In addition, vitexin suppressed the expression of proapoptotic markers such as Bax, cleaved caspase-3, and cleaved caspase9; whereas, Bcl-2 expression was enhanced. Furthermore, vitexin also elicited the anti-inflammatory effect through inhibiting the overexpression of VCAM-1, ICAM-1, TNF- α , E-selectin, IL-1, and IL-6, and finally, vitexin alleviates oxidative stress by promoting cellular antioxidant activities [137].

The apoptotic protective property of vitexin also works by using the MAPK signaling pathway in vitro and in vivo. Lyu et al. [73] indicated that vitexin treatment activates expression of the hypoxia inducible factor-1 α subunit (HIF-1 α) and vascular endothelial growth factor (VEGF) proteins and blocked expression of phosphorylated-p38 MAP kinase (p38) protein in rat. Better still, vitexin suppressed Bax, cleaved caspase-3, NF- κB , and TNF- α production and nullifies oxidative stress via stimulating the activities of antioxidant enzymes [73, 77, 122]. The cardioprotective effect of vitexin is reported to be achieved by inhibiting the initial activation of ERK and p38 MAPK pathways through suppressing the activation of NFATc1, c-Fos, and phospho-c-Jun and at the same time enhancing the expression of phospho-ERK [122, 196]. Bhardwaj et al. [194] suggested that during heat stress, vitexin binds to HSF-1, enhances cell viability, upregulates Hsp90 expression, and thereby activates ER-stress induced autophagy, consequently modulating MAPKs expression and leading to vacuole accumulation and autophagic flux in cells. In HCT-116 cells, exposure to vitexin recruits HSF-1 target proteins downstream and inhibits heat shock proteins [197]. Further experiments identified HSF-1 as a possible molecular target of vitexin. It binds to the DNA-binding domain in HSF-1 protein, which suppresses oligomerization and activation of HSF-1 [197].

In preventing cardiovascular injury, vitexin mitigated apoptosis and autophagy in myocardium cells by the use of PI3K/Akt/mTOR signaling pathway [123]. In H9c2 cells, vitexin inhibited autophagy by suppressing apoptosis and reducing Beclin1 and LC3I/II expression, while p62 expression was upregulated. Vitexin suppressed apoptosis by upregulating Bcl-2, whereas the synthesis of cleaved caspase-3, Epac1, Rap1, and Bax was suppressed [121, 123]. In addition, vitexin also activated the cellular antioxidant pathway [121, 123, 124]. Again, vitexin inhibited the expression of inflammatory cytokines, attenuated increased levels of IL-6, TNF- α , and IL-1 β , and downregulated NF- κB p65 [97, 124].

Vitexin protected mitochondrial dysfunction by promoting mitochondrial activity, its membrane activity, and the ATP level, while it ultimately mitigated mitochondrial injury and cardiomyocyte apoptosis through enhancing the expression of MFN2, blocked the induction of Drp1, and significantly mitigated ROS content [198]. Moreover, in vitro, treating genotoxic-induced larvae with vitexin prompted its antigenotoxicity effects and decreased the occurrence of mutant spots, compared to control [199].

Vitexin protection of RGC-5 cells from oxidative stress injury and death has been illustrated. Treatment of vitexin to RGC-5 cells induced with either l-buthionine-(S,R)-sulfoximine and glutamate or H_2O_2 prevented cell death by mitigating ROS oxidative effects, but the activity was low [200]. Hence, the therapeutic use of vitexin in retinal degenerative disease is repurposing. Lei and Yang reported the cure of the atherosclerosis rat model by feeding with vitexin for 30 days [201]. Further experimental analysis showed that feeding with vitexin or pravastatin or in combination attenuated increase in serum IL-1b, IL-6, TNF- α , total cholesterol, atherogenic index, low-density lipoprotein, ICAM-1, MCP-1, VCAM-1, aortic nitro tyrosine, and liver HMG-CoA-reductase activity. Also, vitexin restored endothelial function and decreased in serum high-density lipoprotein, % HTR (HDL/TC ratio), PON1, homocysteine thiolactonase, aortic SOD, GPx, and CAT.

Platelet aggregation is the drug target in the cure of most vascular diseases such as acute coronary syndrome, stroke, ischemic attacks, peripheral artery occlusive, and coronary disease in DM [202]. The management of these diseases usually involve the use of platelet aggregation inhibitors that attenuate activation and/or aggregation of thrombocyte [202]. Several studies have reported the platelet aggregation inhibitory properties of vitexin and its analogs. The anti-platelet effects of vitexin was tested, and vitexin showed pronounced biological activity by suppression of platelet aggregation mediated by collagen or ADP [203]. In rat plasma and in transgenic zebrafish, vitexin and 2''-O-rhamnosyl vitexin markedly inhibited platelet aggregation; furthermore, 2''-O-rhamnosyl vitexin inhibited thrombus production induced by FeCl_3 , and the compounds anti-platelet activities were considered to be acting through the adenosine diphosphate pathway [204]. Additionally, isovitexin 6''-O-glucoside and isovitexin produced moderate

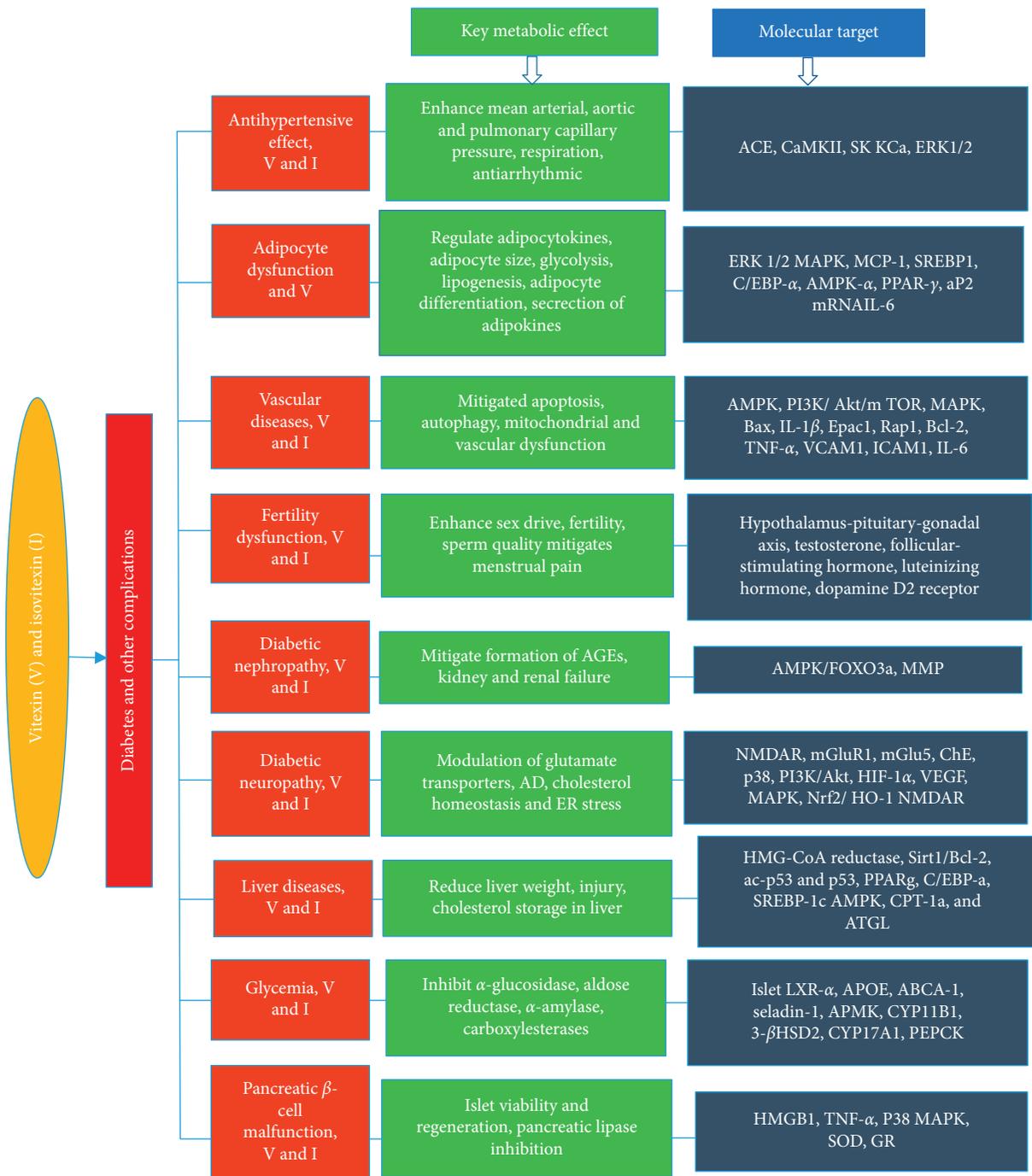


FIGURE 3: Schematic diagram of metabolic effects and molecular targets of vitexin and isovitexin in diabetes mellitus and its complication. V, vitexin; I, isovitexin.

antiplatelet activity and suppressed arachidonic acid, collagen, thrombin, and platelet-activating factor in rabbit platelets [205].

3.3. Safety and Toxicity Profile of Vitexin and Isovitexin. The elucidation of the safety and toxicity profile of small molecules is an integral stage of drug discovery and development. Substantial and growing data now exist

concerning the toxicity and safety profiles of vitexin and its derivatives. The toxicity of vitexin and its derivatives have been observed by (1) studies in vitro using cells or cell lines and (2) in vivo experimentation in animal models. The cytotoxicity of vitexin and isovitexin has been assessed in human leukemia CCRF-CEM cells [206], macrophages cells (RAW264.7) [134, 207], T lymphocytes and RBL-2H3-mast cells [99], N9 microglial cells [74], neutrophils [106], breast carcinoma cell lines, either T47D or MCF-7 [203], MCF-7,

A549, HepG2, and HT-29 cells [208], HepG2 cells [209], J774 macrophages cells [139], and human keratinocyte (HaCaT) cells [144], and they only exerted negligible to no cytotoxic effects. In addition, vitexin-2''-O-rhamnoside and vitexin-4''-O-glucoside caused no cytotoxic effect on human adipose-derived stem cells, A2780 and MCF-7 cells [210, 211]. Studies in neuro-2a cells revealed that A- β 25-35 significantly affected the cell viability by inducing ROS-mediated toxicity and apoptosis. However, pretreatment of neuro-2a cells with vitexin (50 μ M) significantly restored the cell viability up to $92.86 \pm 5.57\%$ [70]. On the contrary, 100 and 500 μ M isovitexin concentrations were found to be toxic to Caco-2 cells after 4 h of incubation [212]. The acute cytotoxicity of vitexin and isovitexin was evaluated in both normoglycemic mice and induced diabetic rat, and the results show that at 2 g/kg wt, neither the normoglycemic mice nor diabetic rat showed any overt signs of toxicity after 24 h and 14 days of observation, and the weight of the rodents was not significantly different with no mortality recorded throughout the 14-day monitoring [19, 20].

For the genetic toxicity test, the results showed that vitexin insignificantly induced few mutant spots and exhibited no genotoxic activity in vitro [199]. The antimutagenic potentials of flavones, isovitexin, and vitexin were compared in the *Salmonella typhimurium* mutagenicity assay using tester strains TA100 and TA98 and mutagens such as aflatoxin B1 (AFB1) and 2-acetamido-fluorene (2-AAF) alongside with metabolic stimulation. From data, vitexin showed a protective effect at 0.4 mM but, however, lost this protection at 0.08 and 0.8 mM against 2-AAF. On the other hand, isovitexin in a dose-dependent manner enriched 2-AAF mutagenicity at concentrations greater than 0.4 mM. Both flavones have protective activities on AFB1 at higher concentrations ≥ 0.4 mM [213]. Structural features that affect antimutagenic capacities of natural flavonoids include physicochemical parameters such as hydrophilicity or lipophilicity which is dependent on the degree of hydroxylation as well as O-methylation, glycosylation on rings A and B, C4-keto moiety, and the double bond at C2-C3 [213]. Furthermore, the in vivo genotoxic test, SMART, in *Drosophila melanogaster* wings demonstrated that vitexin has no genotoxic activity [199]. The metabolite vitexin was investigated to establish genotoxic profiles in viable human lymphocyte cells in vitro for 48 h, and the results show that the planar flavones vitexin enhances the sister chromatid exchange frequency by a factor of around 4-5 at a concentration of 100 μ g/ml and induces micronuclei and a small increase in polyploid cells [214].

4. General Discussion

Despite the fact that isovitexin and vitexin both possess a similar molecular weight of 432.381 g/mol, their analogs vary considerably in terms of their huge masses. This characteristic could contribute to the differences observed among these compounds. The current review sums up the efficacies of vitexin and its analogs for therapeutic application in diabetes and its complications considering multiple drug targets that may be involved. Numerous reports reviewed outlined the

molecular mechanisms of vitexin and isovitexin analogs in diabetic-related complications. There appears to be clear evidence that vitexin and its analogs possess many pharmacological benefits in attenuation of diabetic complications including adipose tissue dysfunction, sexual and fertility impairment, pancreatic β -cell malfunction, hyperglycemia, diabetic neuropathy, liver disorders, diabetic nephropathy, vascular disease, platelet aggregation, and hypertension. Data points to the fact that the antidiabetic mechanism of vitexin and its analogs acts mainly by alleviation of cell apoptosis, targeting point in hypothalamus-gonadal axis and other organs affected by persistent hyperglycemia. Additionally, oxidative stress due to their inflammation is also controlled (Figure 3). Again, substantial evidence has proven the safety and low toxicity profiles of these compounds in vivo and in vitro. Further studies are required to outline additional scientific evidence of vitexin and its analogs for clinical trials in the treatment of diabetes and associated sicknesses. For now, vitexin research on retinopathy is limited, and hence, more studies in that regard would add to knowledge. There are no clinical trial studies reported on only vitexin and isovitexin in diabetes mellitus. The only available work was the inclusion of vitexin in a dietary supplement named Ritmonutra® used to prevent benign supraventricular and ventricular arrhythmias in people who are free of a specific heart disease (<http://www.ClinicalTrials.gov>).

5. Concluding Remarks

It is clear that vitexin and its analogs possess several pharmacological relevances in attenuation of diabetes mellitus and its complications. In addition, the antidiabetic mechanism of vitexin and its analogs is through alleviation of cell apoptosis and oxidative stress due to their anti-inflammation and antioxidant properties. Also, ample evidence has showed the safety of these compounds in vivo and in vitro. Available published reports support the multifaceted medicinal role of vitexin and its analogs in managing diabetes mellitus and its complications.

Data Availability

The datasets used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

MBA and ILA conceived idea. MBA, ILA, SKK, SG, DB, and JBP searched and reviewed the literature, discussed the relevant articles, and drafted portions of the manuscript. All approved the final draft before submission.

References

- [1] N. Sarwar, P. Gao, S. R. Seshasai et al., "Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective

- studies," *Lancet (London, England)*, vol. 375, no. 9733, pp. 2215–2222, 2010.
- [2] J. E. Shaw, R. A. Sicree, and P. Z. Zimmet, "Global estimates of the prevalence of diabetes for 2010 and 2030," *Diabetes Research and Clinical Practice*, vol. 87, no. 1, pp. 4–14, 2010.
- [3] F. Pei, "Managing diabetes in urban Ghana: is it affordable?" Master's thesis, Duke Global Health Institute Duke University, Durham, NC, USA, 2015.
- [4] E. A. Quaye, E. O. Amporful, P. Akweongo, and M. K. Aikins, "Analysis of the financial cost of diabetes mellitus in four cocoa clinics of Ghana," *Value in Health Regional Issues*, vol. 7, pp. 49–53, 2015.
- [5] S. I. G. Rosa, F. Rios-Santos, S. O. Balogun, and D. T. de Oliveira Martins, "Vitexin reduces neutrophil migration to inflammatory focus by down-regulating pro-inflammatory mediators via inhibition of p38, ERK1/2 and JNK pathway," *Phytomedicine*, vol. 23, no. 1, pp. 9–17, 2016.
- [6] D. Moher, A. Liberati, J. Tetzlaff, and D. G. Altman, "Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement," *PLoS Medicine*, vol. 6, no. 7, Article ID e1000097, 2009.
- [7] A. Wilson, "Flavonoid pigments of butterflies in the genus *Melanargia*," *Phytochemistry*, vol. 24, no. 8, pp. 1685–1691, 1985.
- [8] C. Y. Chew, L. S. Chua, N. Soontorngun, and C. T. Lee, "Discovering potential bioactive compounds from Tualang honey," *Agriculture and Natural Resources*, vol. 52, no. 4, pp. 361–365, 2018.
- [9] C. B. GuH. Ma et al., "Characterization, culture medium optimization and antioxidant activity of an endophytic vitexin-producing fungus *Dichotomopilusfunicola* Y3 from pigeon pea [*Cajanuscajan* (L.) Millsp.]," *Journal of Applied Microbiology*, vol. 125, no. 4, pp. 1054–1065, 2018.
- [10] Y. SunQ. Hui et al., "Apoptosis in human hepatoma HepG2 cells induced by the phenolics of *Tetrastigmahemsleyanum* leaves and their antitumor effects in H22 tumor-bearing mice," *Journal of Functional Foods*, vol. 40, pp. 349–364, 2018.
- [11] H. Jeong, J. Sung, J. Yang, Y. Kim, H. S. Jeong, and J. Lee, "Effect of sucrose on the functional composition and antioxidant capacity of buckwheat (*Fagopyrum esculentum* M.) sprouts," *Journal of Functional Foods*, vol. 43, pp. 70–76, 2018.
- [12] S. I. G. RosaF. Rios-Santos et al., "Hydroethanolic extract from *Echinodorusscaber*Rataj leaves inhibits inflammation in ovalbumin-induced allergic asthma," *Journal of Ethnopharmacology*, vol. 203, pp. 191–199, 2017.
- [13] Z. Wang, M. Cui, F. Song, L. Lu, Z. Liu, and S. Liu, "Evaluation of flavonoids binding to DNA duplexes by electrospray ionization mass spectrometry," *Journal of the American Society for Mass Spectrometry*, vol. 19, no. 7, pp. 914–922, 2008.
- [14] W. LiuY. Kong et al., "Determination and quantification of active phenolic compounds in pigeon pea leaves and its medicinal product using liquid chromatography-tandem mass spectrometry," *Journal of Chromatography A*, vol. 1217, no. 28, pp. 4723–4731, 2010.
- [15] F. A. Journak and D. H. Templeton, "The crystal structure and absolute configuration of a bromohexaacetyl derivative of vitexin," *ActaCrystallographica Section B Structural Crystallography and Crystal Chemistry*, vol. 31, no. 5, pp. 1304–1309, 1975.
- [16] M. Singh, M. Kaur, and O. Silakari, "Flavones: an important scaffold for medicinal chemistry," *European Journal of Medicinal Chemistry*, vol. 84, pp. 206–239, 2014.
- [17] K. Marinova, K. Kleinschmidt, G. Weissenböck, and M. Klein, "Flavonoid biosynthesis in barley primary leaves requires the presence of the vacuole and controls the activity of vacuolar flavonoid transport," *Plant Physiology*, vol. 144, no. 1, pp. 432–444, 2007.
- [18] S. A. Osmani, S. Bak, and B. L. Møller, "Substrate specificity of plant UDP-dependent glycosyltransferases predicted from crystal structures and homology modeling," *Phytochemistry*, vol. 70, no. 3, pp. 325–347, 2009.
- [19] C. Y. Choo, N. Y. Sulong, F. Man, and T. W. Wong, "Vitexin and isovitexin from the leaves of *Ficus deltoidea* with in-vivo α -glucosidase inhibition," *Journal of Ethnopharmacology*, vol. 142, no. 3, pp. 776–781, 2012.
- [20] N. A. Y. BintiSulong, "Bioflavonoids from the leaves of *Ficus deltoidea* jack with α -glucosidase inhibition," July 2016, <http://ir.uitm.edu.my/id/eprint/14099>.
- [21] P. Folador, L. H. Cazarolli, A. C. Gazola, F. H. Reginatto, E. P. Schenkel, and F. R. M. B. Silva, "Potential insulin secretagogue effects of isovitexin and swertisin isolated from *Wilbrandiaebracteata* roots in non-diabetic rats," *Fitoterapia*, vol. 81, no. 8, pp. 1180–1187, 2010.
- [22] Y. Saisho, " β -cell dysfunction: its critical role in prevention and management of type 2 diabetes," *World Journal of Diabetes*, vol. 6, no. 1, pp. 109–124, 2015.
- [23] F. Wang, J. Yin, Y. Ma, H. Jiang, and Y. Li, "Vitexin alleviates lipopolysaccharide-induced islet cell injury by inhibiting HMGB1 release," *Molecular Medicine Reports*, vol. 15, no. 3, pp. 1079–1086, 2017.
- [24] S. Inamdar, A. Joshi, S. Malik, R. Boppana, and S. Ghaskadbi, "Vitexin alleviates non-alcoholic fatty liver disease by activating AMPK in high fat diet fed mice," *Biochemical and Biophysical Research Communications*, vol. 519, no. 1, pp. 106–112, 2019.
- [25] S. NurdianaY. M. Goh et al., "Changes in pancreatic histology, insulin secretion and oxidative status in diabetic rats following treatment with *Ficus deltoidea* and vitexin," *BMC Complementary and Alternative Medicine*, vol. 17, no. 1, pp. 1–17, 2017.
- [26] J. Kuang, L. Chen, Q. Tang, J. Zhang, Y. Li, and J. He, "The role of Sirt6 in obesity and diabetes," *Frontiers in Physiology*, vol. 9, pp. 1–9, 2018.
- [27] N. Singh, S. Ravichandran, D. D. Norton, S. D. Fugmann, and R. Moaddel, "Synthesis and characterization of a SIRT6 open tubular column: predicting deacetylation activity using frontal chromatography," *Analytical Biochemistry*, vol. 436, no. 2, pp. 78–83, 2013.
- [28] N. Singh, S. Ravichandran, K. Spelman, S. D. Fugmann, and R. Moaddel, "The identification of a novel SIRT6 modulator from *Trigonella foenum-graecum* using ligand fishing with protein coated magnetic beads," *Journal of Chromatography B*, vol. 968, pp. 105–111, 2014.
- [29] A. T. M. A. Rahim, Y. Takahashi, and K. Yamaki, "Mode of pancreatic lipase inhibition activity in vitro by some flavonoids and non-flavonoid polyphenols," *Food Research International*, vol. 75, pp. 289–294, 2015.
- [30] M. J. Macdonald, "Quinone reductase enzyme activity in pancreatic islets," *Endocrinology*, vol. 129, no. 3, pp. 1370–1374, 1991.
- [31] J. A. Boutin, F. Chatelain-Egger, F. Vella, P. Delagrangé, and G. Ferry, "Quinone reductase 2 substrate specificity and

- inhibition pharmacology," *Chemico-Biological Interactions*, vol. 151, no. 3, pp. 213–228, 2005.
- [32] K. Hamden, K. Mnafigui, Z. Amri, A. Aloulou, and A. Elfeki, "Inhibition of key digestive enzymes-related to diabetes and hyperlipidemia and protection of liver-kidney functions by trigonelline in diabetic rats," *Scientia Pharmaceutica*, vol. 81, no. 1, pp. 233–246, 2013.
- [33] A. R. Abu Bakar, T. Manaharan, A. F. Merican, and S. B. Mohamad, "Experimental and computational approaches to reveal the potential of *Ficus deltoidea* leaves extract as α -amylase inhibitor," *Natural Product Research*, vol. 32, no. 4, pp. 473–476, 2018.
- [34] S. M. Ezzat and M. M. Salama, "A new α -glucosidase inhibitor from *Achillea fragrantissima* (Forssk.) Sch. Bip. growing in Egypt," *Natural Product Research*, vol. 28, no. 11, pp. 812–818, 2014.
- [35] H. S. Mohammed M. M. Abdel-Aziz et al., "Antibacterial and potential antidiabetic activities of flavone C-glycosides isolated from *Beta vulgaris* subspecies *Cicla* L. var. *Pharmaceus* (amaranthaceae) cultivated in Egypt," *Current Pharmaceutical Biotechnology*, vol. 20, no. 7, pp. 595–604, 2019.
- [36] H. Li, F. Song, J. Xing, R. Tsao, Z. Liu, and S. Liu, "Screening and structural characterization of α -glucosidase inhibitors from hawthorn leaf flavonoids extract by ultrafiltration LC-DAD-MS n and SORI-CID FTICR MS," *Journal of the American Society for Mass Spectrometry*, vol. 20, no. 8, pp. 1496–1503, 2009.
- [37] M. Shibano, K. Kakutani, M. Taniguchi, M. Yasuda, and K. Baba, "Antioxidant constituents in the dayflower (*Commelina communis* L.) and their α -glucosidase-inhibitory activity," *Journal of Natural Medicines*, vol. 62, no. 3, pp. 349–353, 2008.
- [38] J.-B. Wei X. Li et al., "Characterization and determination of antioxidant components in the leaves of *Camellia chrysantha* (Hu) Tuyama based on composition-activity relationship approach," *Journal of Food and Drug Analysis*, vol. 23, no. 1, pp. 40–48, 2015.
- [39] Y. Yao, X. Cheng, L. Wang, S. Wang, and G. Ren, "A determination of potential α -glucosidase inhibitors from Azukibeans (*Vigna angularis*)," *International Journal of Molecular Sciences*, vol. 12, no. 10, pp. 6445–6451, 2011.
- [40] M. Ni, X. Hu, D. Gong, and G. Zhang, "Inhibitory mechanism of vitexin on α -glucosidase and its synergy with acarbose," *Food Hydrocolloids*, vol. 105, pp. 105824–105851, 2020.
- [41] S. K. Srivastava, K. V. Ramana, and A. Bhatnagar, "Role of aldose reductase and oxidative damage in diabetes and the consequent potential for therapeutic options," *Endocrine Reviews*, vol. 26, no. 3, pp. 380–392, 2005.
- [42] J. S. Choi, M. Nurul Islam, M. Yousof Ali, E. J. Kim, Y. M. Kim, and H. A. Jung, "Effects of C-glycosylation on anti-diabetic, anti-Alzheimer's disease and anti-inflammatory potential of apigenin," *Food and Chemical Toxicology*, vol. 64, pp. 27–33, 2014.
- [43] J. M. Kim D. S. Jang et al., "Aldose-reductase- and protein-glycation-inhibitory principles from the whole plant of *Duchesneachrysantha*," *Chemistry & Biodiversity*, vol. 5, no. 2, pp. 352–356, 2008.
- [44] H. Li, S. Hwang, B. Kang, J. Hong, and S. Lim, "Inhibitory effects of *Colocasia esculenta* (L.) Schott constituents on aldose reductase," *Molecules*, vol. 19, no. 9, pp. 13212–13224, 2014.
- [45] N. E. J. Tanvi, Q. S. Akhter, S. Nahar, M. N. Sumi, and M. Hosen, "Serum amylase and lipase levels in type 2 diabetes mellitus," *Journal of Bangladesh Society of Physiologist*, vol. 12, no. 2, pp. 52–53, 2018.
- [46] H. Guo, Y. Chen, N. Song, X. Yang, S. Yao, and J. Qian, "Screening of lipase inhibitors from bamboo leaves based on the magnetic ligand fishing combined with HPLC/MS," *Microchemical Journal*, vol. 153, Article ID 104497, 2020.
- [47] Y. Tao, H. Cai, W. Li, and B. Cai, "Ultrafiltration coupled with high-performance liquid chromatography and quadrupole-time-of-flight mass spectrometry for screening lipase binders from different extracts of *Dendrobium officinale*," *Analytical and Bioanalytical Chemistry*, vol. 407, no. 20, pp. 6081–6093, 2015.
- [48] S. M. Majka K. E. Fox et al., "De novo generation of white adipocytes from the myeloid lineage via mesenchymal intermediates is age, adipose depot, and gender specific," *Proceedings of the National Academy of Sciences*, vol. 107, no. 33, pp. 14781–14786, 2010.
- [49] A. Gastaldelli, M. Gaggini, and R. A. Defronzo, "Role of adipose tissue insulin resistance in the natural history of type 2 diabetes: results from the San Antonio metabolism study," *Diabetes*, vol. 66, no. 4, pp. 815–822, 2017.
- [50] W. Lee, S.-K. Ku, and J.-S. Bae, "Vascular barrier protective effects of orientin and isoorientin in LPS-induced inflammation in vitro and in vivo," *Vascular Pharmacology*, vol. 62, no. 1, pp. 3–14, 2014.
- [51] A. Seyedan, Z. Mohamed, M. A. Alshagga, S. Koosha, and M. A. Alshawsh, "*Cynometra cauliflora* Linn. attenuates metabolic abnormalities in high-fat diet-induced obese mice," *Journal of Ethnopharmacology*, vol. 236, pp. 173–182, 2019.
- [52] J. Kim I. Lee et al., "Vitexin, orientin and other flavonoids from *Spirodelapolyrhiza* inhibit adipogenesis in 3T3-L1 cells," *Phytotherapy Research*, vol. 24, no. 10, pp. 1543–1548, 2010.
- [53] H.-R. Wi, M.-J. Choi, S.-L. Choi, A.-J. Kim, and M.-S. Lee, "Effects of vitexin from mung bean on 3T3-L1 adipocyte differentiation and regulation according to adipocytokine secretion," *Journal of the Korean Society of Food Science and Nutrition*, vol. 41, no. 8, pp. 1079–1085, 2012.
- [54] Y. Peng Q. Sun et al., "Vitexin ameliorates high fat diet-induced obesity in male C57BL/6J mice via the AMPK α -mediated pathway," *Food & Function*, vol. 10, no. 4, Article ID 1940, 2019.
- [55] I. Kang S. Choi et al., "Effects of mung bean (*Vignaradiata* L.) ethanol extracts decrease proinflammatory cytokine-induced lipogenesis in the KK-Ay diabetes mouse model," *Journal of Medicinal Food*, vol. 18, no. 8, pp. 841–849, 2015.
- [56] M. A. Lacaille-Dubois, U. Franck, and H. Wagner, "Search for potential angiotensin converting enzyme (ACE)-inhibitors from plants," *Phytomedicine*, vol. 8, no. 1, pp. 47–52, 2001.
- [57] M. Prabhakar, H. Bano, I. Kumar, M. Shamsi, and S. Khan, "Pharmacological investigations on vitexin," *Planta Medica*, vol. 43, no. 12, pp. 396–403, 1981.
- [58] F. Occhiuto, C. Circosta, A. De Pasquale, and F. Briguglio, "Comparative haemodynamic effects of the flavonoids rhoifolin and vitexin in the dog," *Phytotherapy Research*, vol. 4, no. 3, pp. 118–120, 1990.
- [59] C.-c. Lu Y.-q. Xu et al., "Vitexin protects against cardiac hypertrophy via inhibiting calcineurin and CaMKII signaling pathways," *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 386, no. 8, pp. 747–755, 2013.
- [60] C. A. S. Tirloni R. A. C. Palozzi et al., "Influence of *Luehea divaricata* Mart. extracts on peripheral vascular resistance

- and the role of nitric oxide and both Ca²⁺-sensitive and Kir6.1 ATP-sensitive K⁺ channels in the vasodilatory effects of isovitexin on isolated perfused mesenteric beds," *Phyto-medicine*, vol. 56, pp. 74–82, 2019.
- [61] A. Al-Jeboory and K. f. Dizaye, "Cardiovascular effects of vitexin isolated from *Prosopis farcta*," *Iraqi Journal of Pharmaceutical Sciences*, vol. 6, no. 1, pp. 14–19, 2006.
- [62] M. I. Ragone, M. Sella, P. Conforti, M. G. Volonté, and A. E. Consolini, "The spasmolytic effect of *Aloysia citriodora*, Palau (South American cedrón) is partially due to its vitexin but not isovitexin on rat duodenums," *Journal of Ethno-pharmacology*, vol. 113, no. 2, pp. 258–266, 2007.
- [63] A. H. Gilani, A.-u. Khan, M. N. Ghayur, S. F. Ali, and J. W. Herzig, "Antispasmodic effects of Rooibos tea (*Aspalathus linearis*) is mediated predominantly through K⁺-channel activation," pp. 365–373, 2006.
- [64] R. Kesavan, U. R. Potunuru, B. Nastasijević, T. Avaneesh, and M. Dixit, "Inhibition of vascular smooth muscle cell proliferation by *Gentiana lutea* root extracts," *PLoS One*, vol. 8, no. 4, Article ID e61393, 2013.
- [65] S. Pahlavan, M. S. Tousi et al., "Effects of hawthorn (*Crataegus pentagyna*) leaf extract on electrophysiologic properties of cardiomyocytes derived from human cardiac arrhythmia-specific induced pluripotent stem cells," *The FASEB Journal*, vol. 32, no. 3, pp. 1440–1451, 2018.
- [66] R. Ramos Almeida, J. Montani, Raimundo et al., "Activity of *Cecropialyratiloba* extract on contractility of cardiac and smooth muscles in Wistar rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 33, no. 1-2, pp. 109–113, 2006.
- [67] L. Chen, B. Zhang, S. Shan, and X. Zhao, "Neuroprotective effects of vitexin against isoflurane-induced neurotoxicity by targeting the TRPV1 and NR2B signaling pathways," *Molecular Medicine Reports*, vol. 14, no. 6, pp. 5607–5613, 2016.
- [68] K. C. Dos Santos, T. V. Borges, G. Olescowicz, F. K. Ludka, C. A. D. M. Santos, and S. Molz, "Passiflora actinia hydroalcoholic extract and its major constituent, isovitexin, are neuroprotective against glutamate-induced cell damage in mice hippocampal slices," *Journal of Pharmacy and Pharmacology*, vol. 68, no. 2, pp. 282–291, 2016.
- [69] G. S. B. Aseervatham, U. Suryakala, Doulethunisha, S. Sundaram, P. C. Bose, and T. Sivasudha, "Expression pattern of NMDA receptors reveals antiepileptic potential of apigenin 8-C-glucoside and chlorogenic acid in pilocarpine induced epileptic mice," *Biomedicine & Pharmacotherapy*, vol. 82, pp. 54–64, 2016.
- [70] D. S. Malar, V. Suryanarayanan, M. I. Prasanth, S. K. Singh, K. Balamurugan, and K. P. Devi, "Vitexin inhibits Aβ₂₅₋₃₅ induced toxicity in Neuro-2a cells by augmenting Nrf-2/HO-1 dependent antioxidant pathway and regulating lipid homeostasis by the activation of LXR-α," *Toxicology in Vitro*, vol. 50, pp. 160–171, 2018.
- [71] K. Helms, "Characterization of vitexin as an inhibitor of the human ubiquitin-conjugation enzyme E2-2K," *Journal of Experimental Psychology: General*, vol. 136, no. 1, pp. 23–42, 2012.
- [72] L. Yang, Z.-m. Yang, N. Zhang, Z. Tian, S.-b. Liu, and M.-g. Zhao, "Neuroprotective effects of vitexin by inhibition of NMDA receptors in primary cultures of mouse cerebral cortical neurons," *Molecular and Cellular Biochemistry*, vol. 386, no. 1-2, pp. 251–258, 2014.
- [73] Z. Lyu, J. Cao, J. Wang, and H. Lian, "Protective effect of vitexin reduces sevoflurane-induced neuronal apoptosis through HIF-1α, VEGF and p38 MAPK signaling pathway in vitro and in newborn rats," *Experimental and Therapeutic Medicine*, vol. 15, no. 3, pp. 3117–3123, 2018.
- [74] Z. Wang, J. Cai et al., "Anti-inflammatory activities of compounds isolated from the rhizome of anemarrhena asphodeloides," *Molecules*, vol. 23, no. 10, Article ID 2631, 2018.
- [75] Y. Lu, T. Yu, J. Liu, and L. Gu, "Vitexin attenuates lipopolysaccharide-induced acute lung injury by controlling the Nrf2 pathway," *PLoS One*, vol. 13, no. 4, Article ID e0196405, 2018.
- [76] H. Lv, Z. Yu et al., "Isovitexin exerts anti-inflammatory and anti-oxidant activities on lipopolysaccharide-induced acute lung injury by inhibiting MAPK and NF-κB and activating HO-1/Nrf2 pathways," *International Journal of Biological Sciences*, vol. 12, no. 1, pp. 72–86, 2016.
- [77] D. S. Malar, M. I. Prasanth, R. B. Shafreen, K. Balamurugan, and K. P. Devi, "Grewia tiliaefolia and its active compound vitexin regulate the expression of glutamate transporters and protect Neuro-2a cells from glutamate toxicity," *Life Sciences*, vol. 203, pp. 233–241, 2018.
- [78] D. S. Malar, R. B. Shafreen, S. K. Pandian, and K. P. Devi, "Cholinesterase inhibitory, anti-amyloidogenic and neuro-protective effect of the medicinal plant Grewia tiliaefolia—an in vitro and in silico study," *Pharmaceutical Biology*, vol. 55, no. 1, pp. 381–393, 2017.
- [79] H. A. Jung, S. Karki, J. H. Kim, and J. S. Choi, "BACE1 and cholinesterase inhibitory activities of *Nelumbonucifera* embryos," *Archives of Pharmacol Research*, vol. 38, no. 6, pp. 1178–1187, 2015.
- [80] D. N. Olennikov, "Chemical investigation of anagallidium dichotomum and anticholinesterase activity of its constituents," *Chemistry of Natural Compounds*, vol. 49, no. 6, pp. 1137–1139, 2014.
- [81] E. Abbasi, M. Nassiri-asl, M. Shafeei, and M. Sheikhi, "Neuroprotective effects of vitexin, a flavonoid, on pentylenetetrazole-induced seizure in rats," *Chemical Biology & Drug Design*, vol. 80, no. 2, pp. 274–278, 2012.
- [82] E. Abbasi, M. Nassiri-Asl, M. Sheikhi, and M. Shafeei, "Effects of vitexin on scopolamine-induced memory impairment in rats," *The Chinese Journal of Physiology*, vol. 56, no. 3, pp. 184–189, 2013.
- [83] A. Kaprara and I. T. Huhtaniemi, "The hypothalamus-pituitary-gonad axis: tales of mice and men," *Metabolism*, vol. 86, pp. 3–17, 2018.
- [84] Z.-M. Li, N. Liu et al., "Vitexin alleviates streptozotocin-induced sexual dysfunction and fertility impairments in male mice via modulating the hypothalamus-pituitary-gonadal axis," *Chemico-Biological Interactions*, vol. 297, pp. 119–129, 2019.
- [85] B. Meier, D. Berger, E. Hoberg, O. Sticher, and W. Schaffner, "Pharmacological activities of vitexagnus-castus extracts in vitro," *Phytomedicine*, vol. 7, no. 5, pp. 373–381, 2000.
- [86] X. Peng, Z. Zheng et al., "Inhibitory effect of mung bean extract and its constituents vitexin and isovitexin on the formation of advanced glycation endproducts," *Food Chemistry*, vol. 106, no. 2, pp. 475–481, 2008.
- [87] C. Wang, T. Li, L. Zhang, and Y. Zhu, "Vitexin alleviates lipopolysaccharide-induced acute kidney injury via triggering AMPK/FOXO3a signaling pathway in newborn rats," *Latin American Journal of Pharmacy*, vol. 38, no. 3, pp. 558–564, 2019.
- [88] C. Tsioufis, I. Bafakis, A. Kasiakogias, and C. Stefanadis, "The role of matrix metalloproteinases in diabetes mellitus,"

- Current Topics in Medicinal Chemistry*, vol. 12, no. 10, pp. 1159–1165, 2012.
- [89] G. FloresK. Dastmalchi et al., “Antioxidants of therapeutic relevance in COPD from the neotropical blueberry *Anthopteris wardii*,” *Food Chemistry*, vol. 131, no. 1, pp. 119–125, 2012.
- [90] Y.-h. Cui, X.-q. Zhang, N.-d. Wang, M.-d. Zheng, and J. Yan, “Vitexin protects against ischemia/reperfusion-induced brain endothelial permeability,” *European Journal of Pharmacology*, vol. 853, pp. 210–219, 2019.
- [91] W. Wei, Q. Liu, Y. Tan, L. Liu, X. Li, and L. Cai, “Oxidative stress, diabetes, and diabetic complications,” *Hemoglobin*, vol. 33, no. 5, pp. 370–377, 2009.
- [92] M. Y. Donath and S. E. Shoelson, “Type 2 diabetes as an inflammatory disease,” *Nature Reviews Immunology*, vol. 11, no. 2, pp. 98–107, 2011.
- [93] R. Iglesias, L. Citores, A. Di Maro, and J. M. Ferreras, “Biological activities of the antiviral protein BE27 from sugar beet (*Beta vulgaris* L.),” *Planta*, vol. 241, no. 2, pp. 421–433, 2014.
- [94] I.-T. Hong, B. J. Kim, D. C. Yu et al., “Antioxidative and anti-inflammatory activities of *Phaseolus aureus*,” *Journal of Society of Cosmetic Scientists of Korea*, vol. 22, no. 2, pp. 41–51, 1996.
- [95] M. G. Raghu and P. Agrawal, “Evaluation of in-vitro and in-vivo anti-inflammatory activities of apigenin and vitexin,” *Journal of Pharmaceutical Sciences and Research*, vol. 8, no. 12, pp. 1349–1352, 2016.
- [96] J.-Y. Liu, H.-S. Yu et al., “Selective hydrolysis of flavonoid glycosides by *Curvularia lunata*,” *Chinese Journal of Natural Medicines*, vol. 11, no. 6, pp. 684–689, 2013.
- [97] L. Dong, Y. Fan, X. Shao, and Z. Chen, “Vitexin protects against myocardial ischemia/reperfusion injury in Langendorff-perfused rat hearts by attenuating inflammatory response and apoptosis,” *Food and Chemical Toxicology*, vol. 49, no. 12, pp. 3211–3216, 2011.
- [98] I. Shefler, P. Salamon, T. Reshef, A. Mor, and Y. A. Mekori, “T cell-induced mast cell activation: a role for microparticles released from activated T cells,” *The Journal of Immunology*, vol. 185, no. 7, pp. 4206–4212, 2010.
- [99] H. J. Kim, Y. R. Nam, E.-J. Kim, J. H. Nam, and W. K. Kim, “*Spirodela polyrrhiza* and its chemical constituent vitexin exert anti-allergic effect via ORAI1 channel inhibition,” *The American Journal of Chinese Medicine*, vol. 46, no. 6, pp. 1243–1261, 2018.
- [100] D. F. de Santana Aquino, A. C. Piccinelli, F. L. P. Soares, A. C. Arena, M. J. Salvador, and C. A. L. Kassuya, “Anti-hyperalgesic and anti-inflammatory activity of *Alternanthera maritima* extract and 2''-O- α -l-rhamnopyranosylvitexin in mice,” *Inflammation*, vol. 38, no. 6, pp. 2057–2066, 2015.
- [101] J. Jiang, J. Dai, and H. Cui, “Vitexin reverses the autophagy dysfunction to attenuate MCAO-induced cerebral ischemic stroke via mTOR/Ulk1 pathway,” *Biomedicine & Pharmacotherapy*, vol. 99, pp. 583–590, 2018.
- [102] N. Liu, K. S. Wang et al., “Vitexin compound 1, a novel extraction from a Chinese herb, suppresses melanoma cell growth through DNA damage by increasing ROS levels,” *Journal of Experimental & Clinical Cancer Research*, vol. 37, no. 1, pp. 1–16, 2018.
- [103] X. Zhang, P. Shang et al., “Chemical composition and antioxidative and anti-inflammatory properties of ten commercial mung bean samples,” *LWT—Food Science and Technology*, vol. 54, no. 1, pp. 171–178, 2013.
- [104] H. Yang, J. Huang, Y. Mao, L. Wang, R. Li, and C. Ha, “Vitexin alleviates interleukin-1 β -induced inflammatory responses in chondrocytes from osteoarthritis patients: involvement of HIF-1 α pathway,” *Scandinavian Journal of Immunology*, vol. 90, no. 2, pp. 1–10, 2019.
- [105] G. O. De Melo, M. F. Muzitano et al., “C-glycosylflavones from the aerial parts of *Eleusine indica* inhibit LPS-induced mouse lung inflammation,” *Planta Medica*, vol. 71, no. 4, pp. 362–363, 2005.
- [106] B. A. Nikfarjam, F. Hajiali, M. Adineh, and M. Nassiri-asl, “Anti-inflammatory effects of quercetin and vitexin on activated human peripheral blood neutrophils: the effects of quercetin and vitexin on human neutrophils,” vol. 20, no. 2, pp. 127–131, 2017.
- [107] A. Hashiguchi, K. Hitachi, W. Zhu, J. Tian, K. Tsuchida, and S. Komatsu, “Mung bean (*Vignaradiata* (L.)) coat extract modulates macrophage functions to enhance antigen presentation: a proteomic study,” *Journal of Proteomics*, vol. 161, pp. 26–37, 2017.
- [108] S. S. Iyer and G. Cheng, “Role of interleukin 10 transcriptional regulation in inflammation and autoimmune disease,” *Critical Reviews in Immunology*, vol. 32, no. 1, pp. 23–63, 2013.
- [109] J.-A. Teng, S.-G. Wu et al., “The activation of ERK1/2 and JNK MAPK signaling by insulin/IGF-1 is responsible for the development of colon cancer with type 2 diabetes mellitus,” *PLoS One*, vol. 11, no. 2, Article ID e0149822, 2016.
- [110] J. Tester, “Anti-inflammatory action of the flavonoid, vitexin,” *Australian Journal of Herbal Medicine*, vol. 28, no. 3, p. 90, 2016.
- [111] L. H. Boudreau, G. Lassalle-Claux et al., “New hydroxycinnamic acid esters as novel 5-lipoxygenase inhibitors that affect leukotriene biosynthesis,” *Mediators of Inflammation*, vol. 2017, Article ID 6904634, 12 pages, 2017.
- [112] S. R. M. Ibrahim, G. A. Mohamed, K. Z. Alshali, R. A. A. Haidari, A. A. El-Kholy, and M. F. Zayed, “Lipoxygenase inhibitors flavonoids from *Cyperus rotundus* aerial parts,” *Revista Brasileira de Farmacognosia*, vol. 28, no. 3, pp. 320–324, 2018.
- [113] G. O. Dos Reis, G. Vicente et al., “*Croton antisiphiliticus* Mart. attenuates the inflammatory response to carrageenan-induced pleurisy in mice,” *Inflammopharmacology*, vol. 22, no. 2, pp. 115–126, 2014.
- [114] I. Ielciu, A. Mouithys-Mickalad et al., “Flavonoid composition, cellular antioxidant activity and (myelo)peroxidase inhibition of a *Bryonia alba* L. (Cucurbitaceae) leaves extract,” *Journal of Pharmacy and Pharmacology*, vol. 71, no. 2, pp. 230–239, 2018.
- [115] G.-L. Chen, M.-X. Fan, J.-L. Wu, N. Li, and M.-Q. Guo, “Antioxidant and anti-inflammatory properties of flavonoids from lotus plumule,” *Food Chemistry*, vol. 277, pp. 706–712, 2019.
- [116] A. Dongmo, T. Miyamoto, K. Yoshikawa, S. Arihara, and M.-A. Laccaille-Dubois, “Flavonoids from *Acacia pennata* and their cyclooxygenase (COX-1 and COX-2) inhibitory activities,” *Planta Medica*, vol. 73, no. 11, pp. 1202–1207, 2007.
- [117] S.-T. Huang, C.-T. Chen et al., “Inhibitory effects of a rice hull constituent on tumor necrosis factor α , prostaglandin E₂, and cyclooxygenase-2 production in lipopolysaccharide-activated mouse macrophages,” *Annals of the New York Academy of Sciences*, vol. 1042, no. 1, pp. 387–395, 2005.
- [118] S. Abubakar, M. A. Al-Mansoub, V. Murugaiyah, and K.-L. Chan, “The phytochemical and anti-inflammatory

- studies of *Dillenia suffruticosa* leaves,” no. 3, pp. 660–675, 2019.
- [119] A. Islam M. Kabir et al., “Virtual screening for potential COX-inhibiting constituents from *Mimosa pudica*,” *Journal of Applied Pharmaceutical Science*, vol. 5, no. 7, pp. 71–75, 2015.
- [120] D. Rajendiran, S. Packirisamy, and K. Gunasekaran, “A review on role of antioxidants in diabetes,” *Asian Journal of Pharmaceutical and Clinical Research*, vol. 11, no. 2, pp. 48–53, 2018.
- [121] X. Che X. Wang et al., “Vitexin exerts cardioprotective effect on chronic myocardial ischemia/reperfusion injury in rats via inhibiting myocardial apoptosis and lipid peroxidation,” *American Journal of Translational Research*, vol. 8, no. 8, pp. 3319–3328, 2016.
- [122] L.-Y. Dong, S. Li, Y.-L. Zhen, Y.-N. Wang, X. Shao, and Z.-G. Luo, “Cardioprotection of vitexin on myocardial ischemia/reperfusion injury in rat via regulating inflammatory cytokines and MAPK pathway,” *The American Journal of Chinese Medicine*, vol. 41, no. 6, pp. 1251–1266, 2013.
- [123] Z. Tang, L. Yang, and X. Zhang, “Retracted article: vitexin mitigates myocardial ischemia reperfusion-induced damage by inhibiting excessive autophagy to suppress apoptosis via the PI3K/Akt/mTOR signaling cascade,” *RSC Advances*, vol. 7, no. 89, pp. 56406–56416, 2017.
- [124] Z. Sun B. Yan et al., “Vitexin attenuates acute doxorubicin cardiotoxicity in rats via the suppression of oxidative stress, inflammation and apoptosis and the activation of FOXO3a,” *Experimental and Therapeutic Medicine*, vol. 12, no. 3, pp. 1879–1884, 2016.
- [125] H.-b. Li, X.-x. Ying, and J. Lu, “The mechanism of vitexin-4''-O-glucoside protecting ECV-304 cells against tertbutyl hydroperoxide induced injury,” *Natural Product Research*, vol. 24, no. 18, pp. 1695–1703, 2010.
- [126] D. Dietrych-Szostak, “Flavonoids in hulls of different varieties of buckwheat and their antioxidant activity,” in *Proceedings of the 9th International Symposium on Buckwheat*, pp. 621–625, Prague, Czech Republic, August 2004.
- [127] Z. Hohmann, P. Forgo, T. Martinek et al., “Diterpenoids and flavonoids from the fruits of *Vitex agnus-castus* and antioxidant activity of the fruit extracts and their constituents,” *Phytotherapy Research*, vol. 21, no. 4, pp. 391–394, 2007.
- [128] E. M. Hassan, R. A. Hassan, S. A. El-Toumy, S. M. Mohamed, and E. A. Omer, “Phenolic metabolites and antioxidant activity of *Codiaeum variegatum* CV. Spirale,” *Journal of Pharmacy Research*, vol. 8, no. 5, pp. 619–623, 2014.
- [129] X.-x. Ying H.-b. Li et al., “HPLC determination of malondialdehyde in ECV304 cell culture medium for measuring the antioxidant effect of vitexin-4''-O-glucoside,” *Archives of Pharmacal Research*, vol. 31, no. 7, pp. 878–885, 2008.
- [130] J. H. Yang, M. H. Choi, S. H. Yang et al., “Potent anti-inflammatory and antiadipogenic properties of bamboo (*Sasa coreana* Nakai) leaves extract and its major constituent flavonoids,” *Journal of Agricultural and Food Chemistry*, vol. 65, no. 31, pp. 6665–6673, 2017.
- [131] R. Narayanaswamy, A. Isha, L. Wai, and I. Ismail, “Molecular docking analysis of selected *Clinacanthus nutans* constituents as xanthine oxidase, nitric oxide synthase, human neutrophil elastase, matrix metalloproteinase 2, matrix metalloproteinase 9 and squalene synthase inhibitors,” *Pharmacognosy Magazine*, vol. 12, no. 45, pp. S21–S26, 2016.
- [132] A. Pham, C. Nguyen, K. Malterud, D. Diallo, and H. Wangensteen, “Bioactive flavone-C-glycosides of the African medicinal plant *Biophytum umbraculum*,” *Molecules*, vol. 18, no. 9, pp. 10312–10319, 2013.
- [133] C.-M. Lin, C.-T. Chen, H.-H. Lee, and J.-K. Lin, “Prevention of cellular ROS damage by isovitexin and related flavonoids,” *Planta Medica*, vol. 68, no. 4, pp. 365–367, 2002.
- [134] C.-M. Lin S.-T. Huang et al., “Isovitexin suppresses lipopolysaccharide-mediated inducible nitric oxide synthase through inhibition of NF-kappa B in mouse macrophages,” *Planta Medica*, vol. 71, no. 8, pp. 748–753, 2005.
- [135] J. H. Lee S. Y. Lee et al., “Barley sprout extract containing policosanols and polyphenols regulate AMPK, SREBP2 and ACAT2 activity and cholesterol and glucose metabolism in vitro and in vivo,” *Food Research International*, vol. 72, pp. 174–183, 2015.
- [136] S. Nurdiana Y. M. Goh et al., “Improvement of spatial learning and memory, cortical gyriification patterns and brain oxidative stress markers in diabetic rats treated with *Ficus deltoidea* leaf extract and vitexin,” *Journal of Traditional and Complementary Medicine*, vol. 8, no. 1, pp. 190–202, 2018.
- [137] S. Zhang, C. Guo, Z. Chen, P. Zhang, J. Li, and Y. Li, “Vitexin alleviates ox-LDL-mediated endothelial injury by inducing autophagy via AMPK signaling activation,” *Molecular Immunology*, vol. 85, pp. 214–221, 2017.
- [138] I. Bellezza, I. Giambanco, A. Minelli, and R. Donato, “Nrf2-Keap1 signaling in oxidative and reductive stress,” *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, vol. 1865, no. 5, pp. 721–733, 2018.
- [139] M. Hämäläinen, R. Nieminen, P. Vuorela, M. Heinonen, and E. Moilanen, “Anti-inflammatory effects of flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF-κB activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF-κB activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages,” *Mediators of Inflammation*, vol. 2007, Article ID 045673, 10 pages, 2007.
- [140] S. B. Kedare and R. P. Singh, “Genesis and development of DPPH method of antioxidant assay,” *Journal of Food Science and Technology*, vol. 48, no. 4, pp. 412–422, 2011.
- [141] Y. Bai J. Chang et al., “Antioxidant and myocardial preservation activities of natural phytochemicals from mung bean (*Vignaradiata* L.) seeds,” *Journal of Agricultural and Food Chemistry*, vol. 64, no. 22, pp. 4648–4655, 2016.
- [142] J. Bernatoniene, R. Masteikova, D. Majiene et al., “Free radical-scavenging activities of crataegus monogyna extracts,” *Medicina (B. Aires)*, vol. 44, no. 9, pp. 706–712, 2008.
- [143] D. Cao H. Li et al., “Antioxidant properties of the mung bean flavonoids on alleviating heat stress,” *PLoS One*, vol. 6, no. 6, Article ID e21071, 2011.
- [144] V. Chowjarean, T. Prueksasit, K. Joyjamras, and P. Chanvorachote, “Isovitexin increases stem cell properties and protects against PM2.5 in keratinocytes,” *In Vivo*, vol. 33, no. 6, pp. 1833–1841, 2019.
- [145] R. Erenler B. Meral et al., “Bioassay-guided isolation, identification of compounds from *Origanum rotundifolium* and investigation of their antiproliferative and antioxidant activities,” *Pharmaceutical Biology*, vol. 55, no. 1, pp. 1646–1653, 2017.
- [146] W. Nan, K. Fu, Y.-J. Fu et al., “Antioxidant activities of extracts and main components of pigeonpea [*Cajanus cajan* (L.) Millsp.] leaves,” *Molecules*, vol. 14, no. 3, pp. 1032–1043, 2009.

- [147] C. Sridhar, K. V. Rao, and G. V. Subbaraju, "Flavonoids, triterpenoids and a lignan from *Vitex altissima*," *Phytochemistry*, vol. 66, no. 14, pp. 1707–1712, 2005.
- [148] W. A. El-Kashak, A. I. Elshamy et al., "Rumpictuside A: unusual 9,10-anthraquinone glucoside from *Rumex pictus* Forsk," *Carbohydrate Research*, vol. 448, pp. 74–78, 2017.
- [149] E. Joubert, P. Winterton, T. J. Britz, and D. Ferreira, "Superoxide anion and α,α -diphenyl- β -picrylhydrazyl radical scavenging capacity of rooibos (*Aspalathus linearis*) aqueous extracts, crude phenolic fractions, tannin and flavonoids," *Food Research International*, vol. 37, no. 2, pp. 133–138, 2004.
- [150] J. H. Kim, B. C. Lee et al., "The isolation and antioxidative effects of vitexin from acer palmatum," *Archives of Pharmacological Research*, vol. 28, no. 2, pp. 195–202, 2005.
- [151] M.-H. Liu, C.-H. Ko, N. Ma, P.-W. Tan, W.-M. Fu, and J.-Y. He, "Chemical profiles, antioxidant and anti-obesity effects of extract of *Bambusa textilis* McClure leaves," *Journal of Functional Foods*, vol. 22, pp. 533–546, 2016.
- [152] A. C.-N. Leong, Y. Kinjo, M. Tako, H. Iwasaki, H. Oku, and H. Tamaki, "Flavonoid glycosides in the shoot system of Okinawa Taumu (*Colocasia esculenta* S.)," *Food Chemistry*, vol. 119, no. 2, pp. 630–635, 2010.
- [153] Y. Sun, H. Yuan et al., "Enrichment and antioxidant properties of flavone C-glycosides from trollflowers using macroporous resin," *Food Chemistry*, vol. 141, no. 1, pp. 533–541, 2013.
- [154] W. Wätjen, N. Weber et al., "Prenylation enhances cytotoxicity of apigenin and liquiritigenin in rat H4IIE hepatoma and C6 glioma cells," *Food and Chemical Toxicology*, vol. 45, no. 1, pp. 119–124, 2007.
- [155] Y. Sun, K. Yang et al., "Homogenate-assisted vacuum-powered bubble extraction of mosobamboo flavonoids for on-line scavenging free radical capacity analysis," *Molecules*, vol. 22, no. 7, Article ID 1156, 2017.
- [156] D. Zielińska and H. Zieliński, "Antioxidant activity of flavone C-glycosides determined by updated analytical strategies," *Food Chemistry*, vol. 124, no. 2, pp. 672–678, 2011.
- [157] Y. Zhang, D. Wang, L. Yang, D. Zhou, and J. Zhang, "Purification and characterization of flavonoids from the leaves of *Zanthoxylum bungeanum* and correlation between their structure and antioxidant activity," *PLoS One*, vol. 9, no. 8, Article ID e105725, 2014.
- [158] L.-S. Lee, E.-J. Choi et al., "Contribution of flavonoids to the antioxidant properties of common and tartary buckwheat," *Journal of Cereal Science*, vol. 68, pp. 181–186, 2016.
- [159] R. Masteikova, J. Bernatoniene, R. Bernatoniene, and S. Velziene, "Antiradical activities of the extract of *Passiflora incarnata*," *Acta Polonica Pharmaceutica*, vol. 65, no. 5, pp. 577–583, 2008.
- [160] P. W. Sinjman, E. Joubert, D. Ferreira et al., "Antioxidant activity of the dihydrochalcones aspalathin and nothofagin and their corresponding flavones in relation to other rooibos (*Aspalathus linearis*) flavonoids, epigallocatechin gallate, and Trolox," *Journal of Agricultural and Food Chemistry*, vol. 57, no. 15, pp. 6678–6684, 2009.
- [161] M. A. Soobrattee, V. S. Neeragheem, A. Luximon-Ramma, O. I. Arooma, and T. Bahorun, "Phenolics as potential antioxidant therapeutic agents: mechanism and actions," *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 579, no. 1–2, pp. 200–213, 2005.
- [162] Y. Sun, R. Tsao et al., "The phytochemical composition, metabolites, bioavailability and in vivo antioxidant activity of *Tetragymnahemsleyanum* leaves in rats," *Journal of Functional Foods*, vol. 30, pp. 179–193, 2017.
- [163] P. S. Bustos, R. Deza-Ponzio, P. L. Páez, J. L. Cabrera, M. B. Virgolini, and M. G. Ortega, "Flavonoids as protective agents against oxidative stress induced by gentamicin in systemic circulation. Potent protective activity and microbial synergism of luteolin," *Food and Chemical Toxicology*, vol. 118, pp. 294–302, 2018.
- [164] J. Kang, Z. Li, T. Wu, G. S. Jensen, A. G. Schauss, and X. Wu, "Anti-oxidant capacities of flavonoid compounds isolated from acai pulp (*Euterpe oleracea* Mart.)," *Food Chemistry*, vol. 122, no. 3, pp. 610–617, 2010.
- [165] F. An, X. Cao, H. Qu, and S. Wang, "Attenuation of oxidative stress of erythrocytes by the plant-derived flavonoids vitexin and apigenin," *Die Pharmazie*, vol. 70, no. 11, pp. 724–732, 2015.
- [166] A. Ugusman, Z. Zakaria, C. K. Hui, N. A. Nordin, and Z. A. Mahdy, "Flavonoids of *Piper sarmentosum* and its cytoprotective effects against oxidative stress," *EXCLI Journal*, vol. 11, pp. 705–714, 2012.
- [167] G.-R. Wang, W.-Z. Tang, Q.-Q. Yao, H. Zhong, and Y.-J. Liu, "New flavonoids with 2BS cell proliferation promoting effect from the seeds of *Trigonella foenum-graecum* L.," *Journal of Natural Medicines*, vol. 64, no. 3, pp. 358–361, 2010.
- [168] C. Bergeron, A. Marston, R. Gauthier, and K. Hostettmann, "Iridoids and secoiridoids from *Gentiana linearis*," *Phytochemistry*, vol. 44, no. 4, pp. 633–637, 1997.
- [169] A. Ndhalla, M. Aderogba, B. Ncube, and J. Van Staden, "Anti-oxidative and cholinesterase inhibitory effects of leaf extracts and their isolated compounds from two closely related *Croton* species," *Molecules*, vol. 18, no. 2, pp. 1916–1932, 2013.
- [170] S. K. Gabr, R. O. Bakr, E. S. Mostafa, A. M. El-Fishawy, and T. S. El-Alfy, "Antioxidant activity and molecular docking study of *Erythrina × neillii* polyphenolics," *South African Journal of Botany*, vol. 121, pp. 470–477, 2019.
- [171] L. Wen, Y. Zhao et al., "Identification of a flavonoid C-glycoside as potent antioxidant," *Free Radical Biology and Medicine*, vol. 110, pp. 92–101, 2017.
- [172] M. A. Aderogba, L. J. McGaw, M. Bezabih, and B. M. Abegaz, "Isolation and characterisation of novel antioxidant constituents of *Croton zambesicus* leaf extract," *Natural Product Research*, vol. 25, no. 13, pp. 1224–1233, 2011.
- [173] R. Caidan, L. Cairang, J. Pengcuo, and L. Tong, "Comparison of compounds of three *Rubus* species and their antioxidant activity," *Drug Discoveries & Therapeutics*, vol. 9, no. 6, pp. 391–396, 2015.
- [174] Z.-Z. Du, X.-W. Yang, H. Han, X.-H. Cai, and X.-D. Luo, "A new flavone C-glycoside from *Clematis rehderiana*," *Molecules*, vol. 15, no. 2, pp. 672–679, 2010.
- [175] T. K. Girish, K. A. Kumar, and U. J. S. Prasada Rao, "C-Glycosylated flavonoids from black gram husk: protection against DNA and erythrocytes from oxidative damage and their cytotoxic effect on HeLa cells," *Toxicology Reports*, vol. 3, pp. 652–663, 2016.
- [176] B. C. K. Hirota, "C-glycosyl flavones and a comparative study of the antioxidant, hemolytic and toxic potential of *Jatropha multifida* leaves and bark," *International Journal of Phytomedicine*, vol. 4, pp. 1–5, 2012.
- [177] K. R. Joshi, H. P. Devkota, T. Nakamura, T. Watanabe, and S. Yahara, "Chemical constituents and their DPPH radical scavenging activity of nepalese crude drug *Begonia picta*," *Records of Natural Products*, vol. 9, no. 3, pp. 446–450, 2015.

- [178] S. Subhadhirasakul, N. Keawpradub, C. Promwong, and S. Yuenyongsawad, "Natural product communications free radical scavenging and cytoprotective activity of *Salacia euphlebia*Merr." *Natural Product Communications*, vol. 3, no. 2, pp. 211–214, 2008.
- [179] S. PerveenA. M. El-Shafae et al., "Antioxidant and urease inhibitory C-glycosylflavonoids from *Celtis africana*," *Journal of Asian Natural Products Research*, vol. 13, no. 9, pp. 799–804, 2011.
- [180] L. S. M. VelozoM. J. P. Ferreira et al., "C-glycosyl flavones from *Peperomiablanda*," *Fitoterapia*, vol. 80, no. 2, pp. 119–122, 2009.
- [181] H. YaoY. Chen et al., "Screening and quantitative analysis of antioxidants in the fruits of *Livistona chinensis*R. Br using HPLC-DAD-ESI/MS coupled with pre-column DPPH assay," *Food Chemistry*, vol. 135, no. 4, pp. 2802–2807, 2012.
- [182] A. von Gadow, E. Joubert, and C. F. Hansmann, "Comparison of the antioxidant activity of aspalathin with that of other plant phenols of rooibos tea (*Aspalathuslinearis*), α -tocopherol, BHT, and BHA," *Journal of Agricultural and Food Chemistry*, vol. 45, pp. 632–638, 1997.
- [183] W. Bors, W. Heller, C. Michel, and M. Saran, "Flavonoids as antioxidants: determination of radical-scavenging efficiencies," *Methods in Enzymology*, vol. 186, pp. 343–355, 1990.
- [184] A. GordonA. P. G. Cruz et al., "Chemical characterization and evaluation of antioxidant properties of Açai fruits (*Euterpe oleraceae* Mart.) during ripening," *Food Chemistry*, vol. 133, no. 2, pp. 256–263, 2012.
- [185] R. Praveena and K. Sadasivam, "Theoretical simulations on the antioxidant mechanism of naturally occurring flavonoid: a DFT approach," *AIP Conference Proceedings*, vol. 1728, no. 1, 2016.
- [186] M. SimpsonD. Hjelmqvist et al., "Anti-peroxyl radical quality and antibacterial properties of rooibos infusions and their pure glycosylated polyphenolic constituents," *Molecules*, vol. 18, no. 9, pp. 11264–11280, 2013.
- [187] L. Zhu, J. Chen, J. Tan, X. Liu, and B. Wang, "Flavonoids from *Agrimoniapilosa*Ledeb: free radical scavenging and DNA oxidative damage protection activities and analysis of bioactivity-structure relationship based on molecular and electronic structures," *Molecules*, vol. 22, no. 3, pp. 195–211, 2017.
- [188] L. Lespade and S. Bercion, "Theoretical investigation of the effect of sugar substitution on the antioxidant properties of flavonoids," *Free Radical Research*, vol. 46, no. 3, pp. 346–358, 2012.
- [189] X. Ma, R. Yan, S. Yu, Y. Lu, Z. Li, and H. Lu, "Enzymatic acylation of isoorientin and isovitexin from bamboo-leaf extracts with fatty acids and antiradical activity of the acylated derivatives," *Journal of Agricultural and Food Chemistry*, vol. 60, no. 43, pp. 10844–10849, 2012.
- [190] X. ZhengS. Narayanan et al., "Triggering of a Dll4-notch1 loop impairs wound healing in diabetes," *Proceedings of the National Academy of Sciences*, vol. 116, no. 14, pp. 6985–6994, 2019.
- [191] N. Bektas, B. Şnela, E. Yenilmez, O. Özatik, and R. Arslan, "Evaluation of wound healing effect of chitosan-based gel formulation containing vitexin," *Saudi Pharmaceutical Journal*, vol. 28, no. 1, pp. 87–94, 2020.
- [192] S. F. Ehrlich, C. P. Quesenberry, S. K. van den Eeden, J. Shan, and A. Ferrara, "Patients diagnosed with diabetes are at increased risk for asthma, chronic obstructive pulmonary disease, pulmonary fibrosis, and pneumonia but not lung cancer," *Diabetes Care*, vol. 33, no. 1, pp. 55–60, 2010.
- [193] N. N. Tun, G. Arunagirinathan, S. K. Munshi, and J. M. Pappachan, "Diabetes mellitus and stroke: a clinical update," *World Journal of Diabetes*, vol. 8, no. 6, pp. 235–248, 2017.
- [194] M. Bhardwaj, S. Paul, R. Jakhar, and S. C. Kang, "Potential role of vitexin in alleviating heat stress-induced cytotoxicity: regulatory effect of Hsp90 on ER stress-mediated autophagy," *Life Sciences*, vol. 142, pp. 36–48, 2015.
- [195] K. A. Coughlan, R. J. Valentine, N. B. Ruderman, and A. K. Saha, "AMPK activation: a therapeutic target for type 2 diabetes?" *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 7, pp. 241–253, 2014.
- [196] J. JiangY. Jia et al., "Vitexin suppresses RANKL-induced osteoclastogenesis and prevents lipopolysaccharide (LPS)-induced osteolysis," *Journal of Cellular Physiology*, vol. 234, no. 10, pp. 17549–17560, 2019.
- [197] M. BhardwajS. Paul et al., "Vitexin confers HSF-1 mediated autophagic cell death by activating JNK and ApoL1 in colorectal carcinoma cells," *Oncotarget*, vol. 8, no. 68, pp. 112426–112441, 2017.
- [198] W. XueX. Wang et al., "Vitexin attenuates myocardial ischemia/reperfusion injury in rats by regulating mitochondrial dysfunction induced by mitochondrial dynamics imbalance," *Biomedicine & Pharmacotherapy*, vol. 124, Article ID 109849, 2020.
- [199] L. M. Fernandes, Z. da Rosa Guterres, I. V. Almeida, and V. E. P. Vicentini, "Genotoxicity and antigenotoxicity assessments of the flavonoid vitexin by the *Drosophila melanogaster*somatic mutation and recombination test," *Journal of Medicinal Food*, vol. 20, no. 6, pp. 601–609, 2017.
- [200] H. J. LeeK.-A. Kim et al., "The compound isolated from the leaves of *Phyllostachys nigra* protects oxidative stress-induced retinal ganglion cells death," *Food and Chemical Toxicology*, vol. 48, no. 6, pp. 1721–1727, 2010.
- [201] X. Lei and Y. Yang, "Vitexin and an HMG-Co A reductase inhibitor prevent the risks of atherosclerosis in high-fat atherogenic diet fed rats," *Journal of King Saud University—Science*, vol. 32, no. 3, pp. 2088–2095, 2020.
- [202] M. Meinshausen, A. Rieckert, A. Renom-Guiteras et al., "Effectiveness and patient safety of platelet aggregation inhibitors in the prevention of cardiovascular disease and ischemic stroke in older adults—a systematic review," *BMC Geriatrics*, vol. 17, no. 1, Article ID 225, 2017.
- [203] F. U. Afifi and R. Abu-Dahab, "Phytochemical screening and biological activities of *Eminiumspiculatum*(Blume) Kuntze (family Araceae)," *Natural Product Research*, vol. 26, no. 9, pp. 878–882, 2012.
- [204] L.-Z. LiP.-Y. Gao et al., "Monoterpenes and flavones from the leaves of *Crataegus pinnatifida* with anticoagulant activities," *Journal of Functional Foods*, vol. 12, pp. 237–245, 2015.
- [205] C.-N. Lin, S.-H. Kuo, M.-I. Chung, F.-N. Ko, and C.-M. Teng, "A new flavone C-glycoside and antiplatelet and vasorelaxing flavones from *Gentiana arisanensis*," *Journal of Natural Products*, vol. 60, no. 8, pp. 851–853, 1997.
- [206] M. OzarowskiA. Piasecka et al., "Comparison of bioactive compounds content in leaf extracts of *Passiflora incarnata*, *P. caerulea* and *P. alata* and in vitro cytotoxic potential on leukemia cell lines," *Revista Brasileira de Farmacognosia*, vol. 28, no. 2, pp. 179–191, 2018.
- [207] L. Liu, X. Hu, L. Guo, R. Wang, and Q. Zhao, "Anti-inflammatory effect of the compounds from the flowers of *Trollius chinensis*," *Pakistan Journal of Pharmaceutical Sciences*, vol. 131, no. 5, pp. 1951–1957, 2018.

- [208] L. GuoS. Qiao et al., "Investigation of the effective components of the flowers of *Trollius chinensis* from the perspectives of intestinal bacterial transformation and intestinal absorption," *Pharmaceutical Biology*, vol. 55, no. 1, pp. 1747–1758, 2017.
- [209] S. Khole, N. A. Panat, P. Suryawanshi, S. Chatterjee, T. Devasagayam, and S. Ghaskadbi, "Comprehensive assessment of antioxidant activities of apigenin isomers: vitexin and isovitexin," *Free Radicals and Antioxidants*, vol. 6, no. 2, pp. 155–166, 2016.
- [210] W. WeiX. Ying et al., "Effects of vitexin-2''-O-rhamnoside and vitexin-4''-O-glucoside on growth and oxidative stress-induced cell apoptosis of human adipose-derived stem cells," *Journal of Pharmacy and Pharmacology*, vol. 66, no. 7, pp. 988–997, 2014.
- [211] A. PickH. Müller et al., "Structure-activity relationships of flavonoids as inhibitors of breast cancer resistance protein (BCRP)," *Bioorganic & Medicinal Chemistry*, vol. 19, no. 6, pp. 2090–2102, 2011.
- [212] G. Hithamani, D. Kizhakayil, and K. Srinivasan, "Uptake of phenolic compounds from plant foods in human intestinal Caco-2 cells," *Journal of Biosciences*, vol. 42, no. 4, pp. 603–611, 2017.
- [213] P. W. Snijman, S. Swanevelder, E. Joubert, I. R. Green, and W. C. A. Gelderblom, "The antimutagenic activity of the major flavonoids of rooibos (*Aspalathus linearis*): some dose-response effects on mutagen activation-flavonoid interactions," *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, vol. 631, no. 2, pp. 111–123, 2007.
- [214] R. Popp and O. Schimmer, "Induction of sister-chromatid exchanges (SCE), polyploidy, and micronuclei by plant flavonoids in human lymphocyte cultures. A comparative study of 19 flavonoids," *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 246, no. 1, pp. 205–213, 1991.